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The Photoformation of a Phthalide: A Ketene Intermediate Traced by $FSRS^{\dagger}$

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The photo-isomerization of *o*-acetylbenzaldehyde (oABA) in acetonitrile was studied by femto- and nanosecond transient absorption spectroscopy. Spectroscopic signatures are assigned with the aid of TD-DFT, TD-CAM-DFT and DFT-MRCI computations. The isomerization yields a lactone, 3-methylphthalide (3MP), with a quantum yield of 0.3 (30%). As evidenced by femtosecond stimulated Raman spectroscopy (FSRS), the isomerization proceeds via a ketene intermediate. It is formed within \sim 2-3 ps after photo-excitation. Intersystem crossing (ISC) populating the triplet state of oABA seems to compete with the ketene formation. Experiments on the non-reactive *meta*- and *para*-derivatives, which undergo efficient ISC with time constants of 5 ps, support this statement. The triplet state of oABA also contributes to the ketene formation, presumably involving a biradical intermediate. The ketene exhibits a lifetime of 1.4 μ s and generates an additional intermediate in the cascade towards the lactone.

1 Introduction

Intramolecular hydrogen transfers in the excited state are crucial elementary processes in photochemistry and photobiology¹. The transfer may occur in an adiabatic way yielding electronically excited transfer products². After radiative or non-radiative decay the transfer usually is reversed and the starting material is rapidly (< 10 ns) recovered^{3,4}. In a nonadiabatic process, transfer and non-radiative decay are coupled; the transfer product is formed in the electronic ground state. Often, these products are more stable with respect to the reverse transfer than those formed adiabatically. As a consequence secondary processes yielding stable products are more likely.

Blancafort et al.^{5,6} invoked symmetry arguments and correlation diagrams to explain why for a certain molecule the transfer is reversible or not. In the irreversible case the hydrogen transfer is accompanied by the relocation of two electrons from an *n*- to a π -orbital. This renders the reverse transfer orbital-forbidden and therefore slow. For reversible transfers the character of the orbitals involved does not alter which enables a fast reverse transfer. The model was first developed for the photochemistry of *ortho*-nitrobenzaldehyde (oNBA), a compound on which we conducted a series of femtosecond spectroscopy measurements^{7–11}. In oNBA, photoexcitation triggers the transfer of a hydrogen atom from the aldehyde substituent to the nitro group. The transfer occurs within a few 100 fs and results in the formation of a ketene intermediate¹² in its electronic ground state. In line with that, high level quantum chemistry computations find a barrier-less path from the Franck-Condon region of photo-excited oNBA to the ketene intermediate^{5,13}. The computed barrier for the reverse transfer amounts to 0.8 eV⁵. This explains the experimental finding^{7,14} that the ketene intermediate transforms quantitatively into a stable photoproduct, *ortho*-nitrosobenzoic acid.

To widen the scope of their model Blancafort et al.⁶ addressed a related molecule, *ortho*-phthalaldehyde (oPA, Scheme 1). This molecule is known to be photo-reactive, forming among other products a phthalide¹⁵. As the initial process of this reaction, the computations predict a non-adiabatic hydrogen transfer resulting in a ketene⁶. A ketene intermediate was indeed observed via IR spectroscopy in a matrix isolation experiment¹⁶. However, this experiment gave neither information on the time scale of the formation nor on its yield. Time resolved spectroscopy on oPA and related compounds^{15,17,18} lacked the temporal resolution to trace the ketene formation.

Here, we will present a comprehensive femto- and nanosecond spectroscopy study on *ortho*-acetylbenzaldehyde (oABA, Scheme 1). We have opted for oABA instead of oPA since for the latter at least two rotamers (denoted as EZ and EE) coexist¹⁵. These rotamers are expected to differ in their photo-reactivity which renders the observed kinetics heterogeneous¹⁵. For sterical reasons, in oABA the rotamer de-

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picted in Scheme 1 is expected to predominate. As oPA, oABA is known to photo-transform into a phthalide (3-methylphthalide, 3MP, Scheme 1)¹⁸.

The computations by Blancafort et al. on oPA did not take intersystem crossing (ISC) to triplet states into account. Most aromatic carbonyl compounds, however, undergo efficient ISC^{2,19}. So to set signatures of ISC apart from the ones of hydrogen transfer, results on the *meta-* and *para-*isomers (mABA and pABA) are included. Both exhibit a much lower photochemical reactivity (see below).



Scheme 1 Structures of oPA, oABA, 3MP and of diverse intermediates, which have been discussed in the photochemistry of oABA and analogous compounds.

In the remainder of this paper, the phototransformation of oABA to a phthalide will first be characterized by steady state illumination. Femto- and nanosecond transient absorption will reveal the characteristic times for the processes occurring in mABA, pABA and oABA. By means of femtosecond stimulated Raman spectroscopy (FSRS)²⁰ evidence for the ketene formation will be given. Spectral assignments will receive support from quantum chemistry. The results will finally be discussed in the light of earlier findings on oNBA and the computations on oPA.

2 Experimental and theoretical methods

2.1 Steady state illumination

The steady state illumination was performed in 1 cm fused silica cells (Hellma). For excitation at 266 nm, the frequency doubled output of a pulsed Nd:YAG laser (Continuum NY60, pulse duration 10 ns (FWHM), repetition rate 50 Hz), which was converted to 266 nm via an additional β -barium borate (BBO) crystal, was used. Residual fundamental and second harmonic light was removed by two dichroic mirrors. For all other excitation wavelengths, the xenon lamp and excitation monochromator of a Fluorolog 3 spectrometer (Horiba) were employed.

2.2 Femtosecond spectroscopy

The experimental set-ups for the FSRS and transient absorption (TA) experiments in the femto- to nanosecond range were described before^{8,21}. Here, only deviations from these descriptions and the relevant experimental parameters are summarized. For all measurements the frequency tripled output (266 nm) of a 1 kHz femtosecond laser/amplifier system (Coherent Libra) served as actinic pump. Pump-induced spectroscopic changes were probed by a single-filament white light continuum generated in CaF₂. Its diameter amounted to 25 μ m at the sample position. The time resolution of both techniques was about 200 fs. In all measurements, the relative orientation of the polarization vectors of actinic pump and probe light was at magic angle.

2.2.1 Transient absorption The energy of the actinic pump pulses amounted to 1 μ J. The respective beam diameter (FWHM) at the sample location was 200 μ m. For recording of the TA data, at each delay setting signal was accumulated for 2 s per scan (1000 actinic pump pulses) and 4 scans of the delay line were averaged. Signals of the neat solvent were recorded in separate measurements and subtracted with proper scaling²². The instrumental time zero shift was determined as a function of the detection wavelength via the optical Kerr effect and was corrected for.

2.2.2 Femtosecond stimulated Raman spectroscopy The energy of the actinic pump pulses amounted to 4 μ J. The beam diameter (FWHM) at the sample location was 140 μ m. Stimulated Raman processes in the FSRS experiment were induced by laser pulses centered at 400 nm, which were generated via second harmonic spectral compression (SH-SC) in a 25 mm BBO crystal²³. As recently suggested²⁴, the output of the BBO was filtered in a double pass spectral filter (50 cm 2f set-up with a 3600 l/mm grating from Newport (10HG3600-300-1)). Via a variable slit the Raman pump pulses were confined to a spectral width of 12 cm^{-1} (FWHM). Pulse energy and beam diameter were 3 μ J and 50 μ m. The attained spectral resolution of the FSRS experiment was 16 cm⁻¹. Polarizations of the Raman pump and the white light continuum pulses were parallel. At each delay setting signal was accumulated for 14 s per scan (7000 actinic pump pulses) and 10 scans of the delay line were averaged. In contrast to the description in ref. 5, no chopper was used in the actinic pump branch.

FSRS data processing was performed as follows. After subtraction of the dark background *bgr*, the change in stimulated Raman signal upon photoexcitation $S(\tilde{v},t,s)$ as a function of wavenumber \tilde{v} was calculated for each delay setting *t* and scan *s* according to

$$S(\tilde{\mathbf{v}},t,s) = \frac{I_{wl+Rp+exc}(\tilde{\mathbf{v}},t,s) - bgr}{I_{wl+exc}(\tilde{\mathbf{v}},t,s) - bgr}.$$
(1)

^{2 |} Journal Name, 2010, [vol],1-13

Here, $I_{wl+Rp+exc}(\tilde{v},t,s)$ is the measured spectrum when all three pulses reach the sample and $I_{wl+exc}(\tilde{v},t,s)$ when the Raman pump pulse is blocked. To correct for baseline fluctuations, $S(\tilde{v},t,s)$ was first processed with a Savitzky-Golay filter²⁵ (4th order polynom, 2 points left/right) to yield $S_{Sav}(\tilde{v},t,s)$. Then the baseline was fitted with a spline function $SF(\tilde{v},t,s)$. This spline function was subtracted from the data to give the baseline corrected spectra $S_{corr}(\tilde{v},t,s)$

$$S_{corr}(\tilde{\mathbf{v}},t,s) = S_{Sav}(\tilde{\mathbf{v}},t,s) - SF(\tilde{\mathbf{v}},t,s).$$
(2)

Since the absorption of the Raman pump pulse from excited state molecules does not only lead to baseline fluctuations, but also to an apparent decrease in Raman intensity, a correction factor $C(\tilde{v}_{ref}, t, s)$ for this decrease is required. We compute this correction factor by relying on a solvent peak at \tilde{v}_{ref} , which is assumed to have an intensity independent of the photoexcitation of the solute²¹. $C(\tilde{v}_{ref}, t, s)$ is the ratio of this peak's background corrected intensity with (delay settings *t*) and without (negative delay setting $t_{neg} = -4$ ps) photoexcitation of the solute

$$C(\tilde{\mathbf{v}}_{ref}, t, s) = \frac{I_{wl+Rp}(\tilde{\mathbf{v}}_{ref}, t_{neg}, s) - bgr}{I_{wl+Rp+exc}(\tilde{\mathbf{v}}_{ref}, t, s) - bgr}.$$
(3)

The final transient Raman spectra $R(\tilde{v}, t)$ were then obtained by computing

$$R(\tilde{\mathbf{v}},t,s) = S_{corr}(\mathbf{v},t,s) \cdot C(\tilde{\mathbf{v}}_{ref},t,s) - S_{corr}(\tilde{\mathbf{v}},t_{neg},s)$$
(4)

and subsequent averaging over all scans *s*. Here, $S_{corr}(\tilde{v}, t_{neg}, s)$ equals the ground state FSRS spectrum of the sample for each scan.

2.3 Nanosecond transient absorption

The nanosecond transient absorption setup (Applied Photophysics, LSK 6.0) utilizes the output of a frequency quadrupled (266 nm) pulsed Nd:YAG laser (Innolas, Spitlight 600) for photoexcitation²⁶. The sample was excited with 38 mJ pump energy at 12 ns laser pulse duration and 1 Hz repetition rate. The pump beam of 0.8 cm diameter was directed into the sample (a standard 3.5 ml cuvette, pathlength 1 cm) at right angle to the probe beam. The sample was purged with nitrogen to remove dissolved oxygen. For quenching experiments the sample was purged with pure oxygen. For sensitive measurements of absorbance changes we used a high-intensity pulsed xenon lamp (150 W, 1.5 ms, Osram) as probe source. The pulsed output of the lamp remains flat in terms of intensity for $\sim 400 \ \mu s$. This temporal window was relied on in the experiments. Behind the sample the probe light was dispersed by a grating monochromator for wavelength selection. The output signal was detected by a photomultiplier (1P28, 250-600 nm), digitized by an oscilloscope (Agilent Infinium) and transferred to a workstation (Iyonix). Typically 10 laser pulses were averaged to record a kinetic trace at a selected wavelength.

2.4 Quantum chemical computations

Quantum chemical computations were performed using Gaussian09²⁷ and Turbomole²⁸. Optimized ground state geometries and vibrational modes in acetonitrile were obtained using Gaussian DFT B3LYP, the TZVP basis set and the PCM reaction field method²⁹. The calculated frequencies were scaled by a factor of 0.965. Excited state absorptions and oscillator strengths were calculated for the first ten excited states with TD-B3LYP and TD-CAM-B3LYP using Gaussian. DFT-MRCI^{30,31} values were computed in Turbomole using COSMO³² for solvation effects.

Gaussian09 computes Raman activities $S_{comp}(\tilde{v})$ for linearly polarized incident light and 90° unpolarized detection³³. Different conditions apply for the FSRS experiments here. While the incident light still has linear polarization, the detection is performed at 180°. Also, due to the stimulated Raman effect, the detection "selects" light scattered with parallel polarization relative to the incident light (Raman pump and Raman probe polarizations are parallel, see above). So, to obtain intensity values which are directly comparable to the relative intensities in $R(\tilde{v},t)$ (eq. 4), two corrections have to be applied to $S_{comp}(\tilde{v})$. First, the depolarization ratio ρ ("Depolar (P)" in the Gaussian output) has to be considered (eq. 5).³⁴ Second, the calculated activities have to be corrected for the intrinsic frequency dependency in stimulated Raman spectroscopy³⁵. Therefore, $S_{comp}(\tilde{v})$ has to be multiplied by the absolute wavenumbers $\tilde{v}_{S,abs}$ of the Stokes shifted bands. ³⁶ The incident light wavenumber $\tilde{v}_{0,abs}$ was set to 25000 cm⁻¹ (400 nm) since our experiment was performed under these conditions. Considering far-from-resonance conditions, application of both corrections yields Raman activities $S_{comp,corr}(\tilde{v})$, which are directly comparable to our experimental signal heights:

$$R(\tilde{\mathbf{v}},t) \sim S_{comp,corr}(\tilde{\mathbf{v}}) = \frac{S_{comp}(\tilde{\mathbf{v}}) \cdot \tilde{\mathbf{v}}_{S,abs}}{\rho + 1}$$
(5)

Finally, the corrected stick spectra were convoluted with Lorentzian functions (FWHM = 15 cm^{-1}).

2.5 Materials

o-Acetylbenzaldehyde (95%), *m*-acetylbenzaldehyde (95%), and *p*-acetylbenzaldehyde (97%) were supplied by Aldrich and used as received. *trans*-Stilbene (98%) was supplied by TCI Europe. Spectroscopic grade acetonitrile (> 99%, water content <0.03%) from Acros Organics and acetonitrile-d₃ (99.8% deuteration) from Euriso-Top were used as solvents.

3 Results

3.1 Steady state illumination

The UV/Vis absorption spectrum of oABA dissolved in acetonitrile (Figure 1) exhibits clearly discernible bands at 288 nm (absorption coefficient $\varepsilon = 1720 \text{ M}^{-1}\text{cm}^{-1}$), 247 nm (7560 M⁻¹cm⁻¹), and 215 nm (23200 M⁻¹cm⁻¹) (for spectra of mABA and pABA see ESI Figure S1). Closer inspection of the spectrum reveals an additional band at ~ 350 nm with a small absorption coefficient of ~ 200 M⁻¹cm⁻¹. Illumination of the solution with 266 nm light mostly results in a reduction of the absorption except for spectral regions around 230 and 270 nm. Here small increases are seen. The spectral changes fit well with a photo-induced formation of 3MP. A difference spectrum derived from the illumination experiment matches the one computed from the spectra of oABA and phthalide. The parent compound phthalide was used as a "stand-in" for 3-methylphthalide which was not available.



Fig. 1 UV/Vis absorption spectrum of oABA dissolved in acetonitrile (initial concentration of 0.2 mM (black line), optical path length of 1 cm) and its changes upon 266 nm illumination. Upper panel: Spectra for several illumination times. Lower panel: Difference spectrum computed from the spectra in the upper panel (solid line) in comparison to a difference obtained by subtracting the oABA from the phthalide spectrum (dashed line).

An illumination experiment with FTIR detection (see ESI Figure S2) and GC/MS spectra (data not shown) also support the formation of 3MP as the only product. In the IR experiment a strong band at 1765 cm^{-1} is seen to rise.

This wavenumber is characteristic for the carbonyl stretching mode of the lactone motif³⁷ in 3MP. The reaction quantum yield ϕ_r was determined relying on UV/Vis and FTIR spectroscopy. In either experiments the absorption at the illumination wavelength was high enough and the illumination time short enough that constant absorbed light flux may be assumed. Under these conditions the concentrations change linearly with illumination time and the slope of the respective plots is proportional to ϕ_r . In the UV/Vis experiment the reduction of the absorption at 291 nm served as a measure for the consumption of oABA. At 291 nm the absorption of 3MP is negligible. In the IR experiment the consumption was monitored at 1697 cm^{-1} . Here, one carbonyl stretching mode of oABA is located. The UV/Vis experiment afforded a yield ϕ_r of 0.31 (31%) for an illumination wavelength of 288 nm. For IR detection and a slightly different illumination wavelength of 266 nm a value of 0.30 was determined. Since the formation of a lactone is impossible in mABA and pABA, the quantum yields for their consumption are by a factor of ~ 50 smaller than the one for oABA. No attempts were made to clarify the nature of mABA and pABA photoproducts. Recording fluorescence properties of oABA failed since during the acquisition the fluorescence increased in intensity. Obviously, in the fluorimeter the photo-reaction occurs and the 3MP formed exhibits a much higher fluorescence quantum yield than the starting material.

3.2 Transient absorption spectroscopy

3.2.1 meta- and para-Acetylbenzaldehyde mABA and pABA dissolved in acetonitrile were excited with femtosecond laser pulses centered at 266 nm. The difference absorption in response to that excitation was recorded as function of the detection wavelength and delay time (for pABA data see Figure 2, for mABA see ESI, Figure S3). In the pABA experiment, the contour plot reveals an ultrafast decay of bands in the range of 400-470 nm and 600-670 nm within \sim 200 fs (see also Figure 2, time trace B). Shortly after time zero a spectrally broad positive difference absorption with a pronounced band at 340 nm and a weaker one at 580 nm is observed. Within a few picoseconds the difference absorption around 580 nm decays close to zero and the band around 340 nm narrows and increases in height. From ~ 10 ps onwards the difference spectrum features an intense band with a maximum at 345 nm with a shoulder at 390 nm and a weak, broad absorption tail up to ~ 650 nm (see magnification in Figure 2, right panel). These features persist to the end of the time window (3 ns) and recur in a nanosecond transient absorption experiment (see ESI Figure S4).

The initial ultrafast decay occurs with a time constant below our instrumental response function (< 200 fs) and is attributed to internal conversion from the initially excited to



Fig. 2 Femtosecond transient absorption on pABA (3 mM) dissolved in acetonitrile upon excitation at 266 nm. A contour representation of the data is given in the central panel. Dashed vertical lines mark spectral positions for the time traces plotted in the left panel. Dashed horizontal lines represent delay times for the difference spectra plotted in the right panel. For the difference spectrum at 1 ns an additional ten-fold magnification is shown (grey dashed line). Note that the time axis is linear until 1 ps and logarithmic thereafter.



Fig. 3 Femtosecond transient absorption on oABA (17 mM) dissolved in acetonitrile upon excitation at 266 nm. A contour representation of the data is given in the central panel. Dashed vertical lines mark spectral positions for the time traces plotted in the left panel. Dashed horizontal lines represent delay times for the difference spectra plotted in the right panel. Note that the time axis is linear until 1 ps and logarithmic thereafter.

the lowest excited singlet state. Monoexponential fits of the time traces around 345 and 580 nm (traces A and C in Figure 2) yield identical time constants of 5 ps, describing the decay of the species featuring the 340 and 580 nm bands and population of the one featuring the 345 nm band. Around 450 nm the time traces are more complex. This is ascribed to vibrational cooling of the species formed within 5 ps. The lifetime of this species, as observed in a nanosecond experiment ($c_{pABA} = 0.14$ mM), proves to be oxygen sensitive (see ESI Figure S4). In a nitrogen purged solution the lifetime is ~ 2 µs. In an oxygen saturated solution it is lower by an order of magnitude (~ 0.2 µs). Analyses of the mABA data sets yield similar time constants (5 ps, ~ 2 µs (N₂) and ~ 0.2 µs (O₂), see ESI Figures S3 & S5).

The time resolved data on mABA and pABA lend to the interpretation that the compounds undergo efficient ISC to the triplet state. For a closely related system — acetophenone dissolved in acetonitrile - a triplet absorption spectrum similar to the ones observed here was reported³⁸. The lifetimes of $\sim 2 \ \mu s$ recorded in the nanosecond experiments are in the range typical for organic carbonyls in solution³⁹. The "building blocks" of mABA and pABA, benzaldehyde and acetophenone, exhibit longer triplet lifetimes in acetonitrile of 17 μ s and 130 μ s, respectively⁴⁰. We stress, however, that the lifetime values of $\sim 2 \ \mu s$ do not need to be the "intrinsic" lifetimes of the states. Concentration quenching, i.e. an interaction between a triplet excited and a ground state molecule, and triplet-triplet annihilation cannot be excluded. The former effect comes into play for benzaldehyde and acetophenone dissolved in acetonitrile⁴⁰. The lifetime reduction by oxygen translates into a bi-molecular quenching constant k_{q,O_2} of $5.0 \cdot 10^8$ M⁻¹s⁻¹ for pABA. Hereby, a published value for oxygen concentration in oxygen saturated acetonitrile (9.1 mM) was relied on³⁹. The quenching constant is clearly below the one for the diffusion limit $(1.9 \cdot 10^{10} \text{ M}^{-1} \text{s}^{-1})^{39}$. The rate constant for quenching of the acetophenone triplet by oxygen is also below the diffusion limit⁴¹. This observation was ascribed to spin statistics⁴¹. All this is in favor of assigning the transients observed at delay times > 10 ps to the lowest triplet states of mABA and pABA, which are both populated with a time constant of 5 ps.

3.2.2 *ortho*-Acetylbenzaldehyde The same set of transient absorption experiments as for mABA and pABA were conducted for oABA dissolved in acetonitrile. The spectrotemporal behavior differs substantially from the one of pABA. The difference absorption induced by 266 nm femtosecond excitation is spectrally very broad with peaks around time zero at 620, 390, and 345 nm (Figure 3).

Within the instrumental response time (200 fs) the difference absorption decreases in magnitude throughout the spectral region covered. On the time scale of picoseconds a decay in signal strength is observed, predominately around 380 nm. Within ~ 10 ps a band around 400 nm is seen to narrow in spectral width. Further kinetic compounds on the time scale of 100 ps and 1 ns are discernible. On the nanosecond time scale signal increases as well as decreases are seen. The difference spectrum recorded for the largest delay time (3 ns) in the femtosecond experiment exhibits peaks at 520, 405, and ~ 320 nm.



Fig. 4 Contour representation of nanosecond transient absorption data on oABA (0.12 mM) dissolved in nitrogen purged acetonitrile upon excitation at 266 nm. At early times the absorption maxima of the fs-UV/Vis offset reoccur. At later times a transient absorption around 370 nm is formed.

The same features are observed in the earliest difference spectra recorded with the nanosecond set-up (Figure 4). After $\sim 1 \ \mu s$ these signatures decay giving way to another species with a spectrum peaking at 370 nm and a ground state bleach around 280 nm becomes discernible. This species does not fully decay within the time range covered (up to 400 μs), so that at the given noise level the time constant for its decay of about 100 μs is afflicted with a rather high uncertainty. This lifetime proves to be mildly oxygen dependent (lifetime quenched by $\sim 30\%$ in presence of oxygen, see ESI Figure S6).

The kinetic analysis of the oABA data set is more challenging than for the ones of mABA and pABA. The femto- and nanosecond data sets were analyzed using a global fitting routine⁴². To model the femtosecond data set, trial functions with different numbers of exponential terms were tested. Visible inspection of the time traces (Figure 3) reveals that at least four terms are involved. It turned out that five exponential terms and an offset are required to describe the data set. The corresponding decay associated spectra (DAS) and time constants are shown in Figure 5. To cross-check the number of components, a singular value decomposition (SVD)⁴² was performed. To avoid the instrumental rise showing up in the SVD



Fig. 5 Decay associated spectra obtained from an analysis of the data on oABA plotted in Figure 3. The first time constant τ_1 lies below our instrument response function and is therefore afflicted with large uncertainty.

results, the procedure was restricted to the range 500 fs to 3 ns. In this range four dominant singular values were found. Con-

sidering that one time constant ($\tau_1 = 0.02$ ps, Figure 5) of the global analysis is outside of this range, the SVD procedure justifies using five terms. The nanosecond data could be reproduced using two major kinetic components and an offset. The resulting time constants are 1.4 μ s and 106 μ s. We will postpone the assignment of time constants to the Discussion since the FSRS data covered now will greatly facilitate this assignment.

3.3 Femtosecond stimulated Raman spectroscopy

Raman resonances of the solvent impair measurements on the solute at the spectral position of these resonances. Deuteration shifts the solvent resonances and thereby opens additional spectral windows. So, to cover a broad range of Raman shifts in the FSRS experiment, oABA dissolved in acetonitrile and in deuterated acetonitrile was investigated. As in the experiments described above, the sample was excited with 266 nm laser pulses. The Raman probe pulse was centered at 400 nm and thereby in resonance with the transient species observed in the transient absorption experiment (cf. Figure 3). The excitation results in increases of the Raman gain due to species populated by the excitation and decreases due to depletion of the starting material (Figure 6). Due to resonance Raman enhancement positive signatures are often stronger in magnitude than the negative ones. Positive signatures are located at Raman shifts of 1170, 1525, 2080, and 3600 cm^{-1} . The intense band at 1525 cm⁻¹ is present from ~ 1 ps onwards. The other bands are clearly discernible from 10-20 ps onwards. All bands do not change in strength or position thereafter. The band at 1525 cm^{-1} is by far the strongest. Thus, we assume that this apparent difference in rise time is caused solely by the fact that the weaker bands surmount the noise floor at later delay times. Early on the most intense band is centered at a lower wavenumber of 1500 cm^{-1} and features a larger bandwidth (40 cm⁻¹ FWHM) than later on (22 cm⁻¹). So within a few picoseconds the band shifts to larger wavenumbers and narrows, which is characteristic for vibrational cooling of a nascent species 43-45. To separate these spectral dynamics from a "genuine" rise, the band integral was computed and plotted as a function of delay time (Figure 7, upper panel). The resulting time trace can be modeled with an exponential rise and time constant of 2.2 ps. Plotting the peaks of the Raman band as obtained from Gaussian fits versus delay time reveals a slower response (Figure 7, lower panel). From a single exponential fit a relaxation time of 11.2 ps is obtained for this shift.

The FSRS pattern and temporal behavior is very similar to the one observed for the ketene derived from oNBA⁸. Quantum chemistry will give further support that indeed a ketene intermediate is also formed here.



Fig. 6 FSRS on oABA (17 mM) dissolved in deuterated acetonitrile (left panel) and acetonitrile (central and right panel). The excitation was tuned to 266 nm and the Raman pump pulse to 400 nm. The contour representations depict changes in Raman gain as a function of the Raman shift and the delay time. Red (blue) coloring represents increased (decreased) Raman gain induced by the excitation. In the panels on top the ground state FSRS spectrum of oABA (dashed blue line, in a.u.) and a time resolved spectrum at large delay times (solid red line) are shown. The latter was averaged from 1.5 to 3.1 ns and processed with a Savitzky-Golay filter (4pt, 2nd order polynomial). Note that there is an *apparent* difference in rise times for the bands. Presumably, this is due to the weaker bands taking longer to surmount the noise.



Fig. 7 The rise behavior of the 1525 cm⁻¹ resonance in the FSRS data on oABA in deuterated acetonitrile (see left panel in Fig. 6). The upper panel gives the spectral integral over the band (black bullets), plotted as a function of the delay time. In the lower panel, the peak frequency of the band (black squares) as obtained from Gaussian fits of the respective transient spectra is plotted versus delay time. Single exponential fits are represented by red lines and the resulting time constants τ_{ket} for ketene formation and τ_{vc} for vibrational cooling are shown.

3.4 Quantum chemical computations

To affirm the formation of a ketene intermediate upon photo excitation of oABA, the structures of oABA and potential intermediates (Scheme 1) were geometry optimized considering solvent effects (for coordinates see ESI Tables S1-S5). In the *E*-ketene-enol the methyl group must point towards the ketene group. Optimization of this compound yielded a structure with a distorted six-membered ring due to the sterical demand of both substituents. The free energy *F* of formation of this structure is much larger than the one of the *Z*-ketene-enol (*F_E* - *F_Z* = 16.7 kJ/mol, T = 298 K, cf. ESI). So, an *E*-ketene-enol intermediate is unlikely to be formed here.

Based on the optimized structures, the respective Raman spectra in acetonitrile were computed (for computational details see Section 2.4). A comparison of the FSRS data with the calculated spectra confirms the formation of the Z-ketene-enol intermediate (Figure 8). As expected from vibrational spectroscopy on other ketenes⁴⁶ only this intermediate exhibits a band around 2050 cm^{-1} . The resonance is caused by the stretching mode of the ketene motif. The most intense band in the experimental spectrum at 1525 cm^{-1} , which is reproduced by the computation, is due to an in-phase stretching vibration of the butadiene scaffold in the ketene intermediate. Finally, the band at 3600 cm^{-1} seen in experiment and computation is caused by an OH-stretching mode and unequivocally shows that a hydrogen atom has been transferred. Some computed bands are not visible in the experiment. This can in most of the cases be explained by selective enhancement of some bands in resonance Raman spectroscopy.⁴⁷ The experiment was con-

Structure	Method	State	Wavelength λ
			(Oscillator strength f)
Z-Ketene-enol	DFT-MRCI	S ₁	576 nm $(5 \cdot 10^{-4})$
		S_2	380 nm (0.146)
	TD-CAM-DFT	S_1	466 nm (0.000)
		S_2	393 nm (0.160)
	TD-DFT	S_1	521 nm (0.000)
		S_2	429 nm (0.119)
Cyclic enol	DFT-MRCI	S_1	351 nm (0.129)
	TD-CAM-DFT	S_1	357 nm (0.135)
	TD-DFT	S_1	388 nm (0.099)
Biradical	TD-CAM-DFT	S_1	578 nm (0.055)
		S_2	460 nm (0.001)
		S_3	366 nm (0.014)
		S_4	329 nm (0.019)
	TD-DFT	S_1	636 nm (0.040)
		S_2	$515 \text{ nm} (7 \cdot 10^{-4})$
		S_3	369 nm (0.014)
		S_4	351 nm (0.030)

Table 1 Lowest energy vertical excitation wavelengths λ and oscillator strengths *f* for possible intermediates in acetonitrile obtained from different computational methods.

ducted under resonance conditions whereas the computations were not. In particular, the totally symmetric A-term resonances according to Albrecht's notation⁴⁸ become stronger compared to off-resonance scattering. This may explain the high intensity of the mode at 1525 cm^{-1} , which represents a totally symmetric vibration regarding the butadiene scaffold. In other cases, rising bands are compensated by the bleach of ground state bands as in the case of the intense 1625 cm^{-1} resonance. Taking differences between experimental and computed spectra into account, it is evident that within a few picoseconds after excitation the *Z*-ketene-enol intermediate is formed.

For the interpretation of the transient absorption spectra, transition energies and oscillator strengths of the intermediates were computed (Table 1). The FSRS experiment has shown that the 405 nm absorption band observed in the fs-UV/Vis data is caused by the ketene intermediate. In the computations we find that this band is the ketene $S_0 \rightarrow S_2$ transition. Note that the computed position of this band varies depending on the method employed. This may be explained by the differing quality of the description of dispersion interactions between solvent and solute in the different methods. Nevertheless the calculations deliver the following trends:

• The Z-ketene-enol exhibits a weak $S_0 \rightarrow S_1$ absorption at > 460 nm and an intense band around 400 nm, as seen in our experiments. The oscillator strength of the latter is ~ 0.15 .



Fig. 8 Comparison of experimental (lower panel, data as in top panel of Figure 6) and computed Raman spectra $S_{comp,corr}$ (top and central panel, see eq. 5). The calculated Z-ketene-enol spectrum (red) is in best agreement with the experimental spectrum compared to the biradical (green) and the cyclic enol (blue) spectra. Note that the OH-stretching mode of the biradical is shifted below 3200 cm⁻¹ (not shown).

- The cyclic enol exhibits an $S_0 \rightarrow S_1$ absorption with an intensity comparable to the $S_0 \rightarrow S_2$ transition of the ketene. Its position is shifted to shorter wavelengths by roughly 30 nm with respect to the latter.
- The biradical exhibits multiple low intensity absorption bands in the spectral window of our experiments.

This knowledge assists in assigning molecular processes to the kinetic components in our transient absorption experiments (see Discussion). Furthermore, the computed oscillator strengths allow for estimates of intermediate concentrations and their quantum yields.

3.5 Quantum yield of the ketene formation

Based on the experimental and computational results an estimate of the quantum yield for ketene formation $\phi_{r,k}$ will be given. The measured difference absorption $\Delta A(v)$ as a function of frequency v after ketene formation is related to this yield via

$$\Delta A(\mathbf{v}) = \phi_{r,k} \Delta \varepsilon(\mathbf{v}) c^* d \approx \phi_{r,k} \varepsilon_k(\mathbf{v}) c^* d \tag{6}$$

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Hereby, $\Delta \varepsilon(v) = \varepsilon_k(v) - \varepsilon_{oABA}(v)$ is the difference absorption coefficient for the ketene as function of frequency. Since in the spectral range of interest the oABA starting material does not absorb (cf. Figures 1 and 3), it may be approximated by $\varepsilon_k(v)$. c^* stands for the effective concentration of initially excited molecules and *d* is the effective path length. To connect with the computed oscillator strength f_k , eq. (6) is spectrally integrated

$$\int \Delta A(\mathbf{v}) d\mathbf{v} = \phi_{r,k} \int \varepsilon_k(\mathbf{v}) d\mathbf{v} c^* d = \phi_{r,k} \frac{f_k}{const} c^* d, \quad (7)$$

whereby reference is made to the definition of the oscillator strength (see e.g. ref.⁴⁹). *const* subsumes constants and amounts to $1.44 \cdot 10^{-18} \text{ mol} \cdot \text{s}/\text{m}^{249}$. The evaluation of the left hand side of eq. (7) relied on a transient spectrum at a delay time of 3 ns (Figure 3). The complete spectrum was plotted as a function of frequency and reconstructed by three Gaussians. The integral of the Gaussian centered at $7.4 \cdot 10^{14} \text{ s}^{-1}$ (or 405 nm), which describes the $S_0 \rightarrow S_2$ band, was taken as spectral integral.

The determination of the yield further requires knowledge on the product c^*d . It was evaluated by a comparison. To this end, a femtosecond transient absorption experiment on a reference molecule was performed under conditions identical to those for oABA. trans-Stilbene dissolved in acetonitrile was used for this purpose. It has been thoroughly characterized by femtosecond spectroscopy and forms cis-stilbene with a quantum yield of 0.5 within less than 50 ps after excitation⁵⁰. Since the extinction coefficients of both *trans*- and cis-stilbene in acetonitrile are known⁵¹, one can compute the product c^*d from the persistent ground state bleach at large delay times (> 100 ps). This experiment affords a product c^*d of $1.8 \cdot 10^{-6}$ M \cdot cm for oABA. An estimate based on the energy per pump pulse, the diameters of pump and probe beams and the absorption at the pump wavelength yields a value in the range of the former one $(2.6 \cdot 10^{-6} \text{ M} \cdot \text{cm})$. With these inputs and $f_k = 0.146$ (DFT-MRCI, Table 1), the quantum yield of ketene formation $\phi_{r,k}$ is computed to be ~ 0.55 (± 0.2).

4 Discussion

A combination of FSRS with quantum chemical computations has allowed to unambiguously identify and characterize the Z-ketene-enol intermediate in the photoreaction of oABA (a summarizing kinetic scheme of the photoreaction of oABA to 3MP is given in Scheme 2). The ketene is formed with vibrational excess energy within ~ 2.5 ps after excitation. The excess energy is dissipated with a time constant of ~ 11 ps. Similar time constants ($\tau_2 = 2.8$ ps and $\tau_3 = 15$ ps) are retrieved from the UV/Vis data (see Figure 5).

In addition to these two processes, the three remaining time constants in this experiment have to be assigned. The first one, τ_1 , lies below our instrumental response function and is therefore afflicted with a rather large uncertainty. However, it can be assigned to the relaxation to the lowest excited singlet state since our excitation wavelength is in resonance with an upper singlet state.

As can be seen in the time traces (Figure 3, traces B and C) and the DAS of τ_5 (Figure 5), the ketene signature features an additional low amplitude rise with the characteristic time constant τ_5 . From the DAS a contribution of roughly 0.15 (15%) to the overall ketene yield can be estimated. This shows that on the early nanosecond timescale the ketene is populated by a path additional to the singlet channel. The amplitude of this rise is within the noise of the FSRS experiment and is therefore not clearly discernible there. We assume that the additional path involves the triplet state of oABA for the following reason. The fs-UV/Vis experiments on the oABA isomers mABA and pABA both delivered a time constant for triplet population of \sim 5 ps. The excited singlet state of oABA decays with a faster time constant of ~ 2.5 ps due to the additional reactive decay channel. Assuming that the rate constants for ISC are similar in all three isomers, the triplet state must still be partly populated within the 2.5 ps excited singlet state lifetime of oABA.

This triplet state then decays with a time constant of 150 ps (τ_4) and forms a triplet biradical via hydrogen transfer. The corresponding DAS features several minima and maxima due to simultaneous triplet decay and biradical formation. Due to the formation of the biradical, the DAS exhibits weak minima around 460 and 370 nm which conforms with our quantum chemical results. The most pronounced maximum is situated at ~ 340 nm and agrees with a decay of a triplet spectrum similar to the ones observed for mABA and pABA. The rather low positive amplitudes in this spectrum may be explained by a partial compensation of positive (triplet decay) and weaker negative (biradical formation) contributions. Furthermore, a sub-nanosecond triplet decay and biradical formation have already been discussed for the photochemistry of oPA15 and have been found in analogous reactions of ortho-substituted nitrobenzenes¹⁰. It is this biradical which we assume to be responsible for the additional low amplitude rise of the ketene signature.

The Z-ketene-enol persists until the end of the time window of our fs-experiments. As seen in the nanosecond experiment (Figure 4), an additional species with an absorption around 370 nm is formed upon decay of the Z-ketene with a time constant of 1.4 μ s. This species exhibits a lifetime of >100 μ s, which is only slightly reduced by oxygen. We presume that here we observe the same species as reported for similar compounds^{17,18} (the mild oxygen dependence has also been observed there and was attributed to oxidation of the species). There, the short lived components were ascribed to Z-keteneenol and the long lived to *E*-ketene-enol intermediates. We

^{10 |} Journal Name, 2010, [vol], 1–13



Scheme 2 Kinetic scheme of the photo reaction of oABA to 3MP in acetonitrile upon excitation at 266 nm. The indicated times represent lifetimes of the states. Species depicted in grey (biradical and cyclic enol) are suggested as intermediates for reasons given in the text. They were not structurally verified within this work.

agree with the first assignment, but digress from the latter. The quantum chemical computations place the *E*- energetically well above the *Z*-isomer. Thus, the *Z*-ketene-enol should not transform into the *E*-isomer. We cannot exclude that the triplet biradical partly recombines to the *E*-ketene-enol. However, this being a minor channel on the early nanosecond time scale, it cannot explain the formation of the 370 nm band. Instead, we suggest that the cyclic enol intermediate may be formed here. It has been discussed as an intermediate in the reaction of oPA¹⁵. For this structure, our computations predict an absorption band which is shifted by ~ 30 nm to lower wavelengths compared to the S₀ \rightarrow S₂ ketene absorption and exhibits a comparable oscillator strength. This is in agreement with our nanosecond data.

Since all measurements were conducted in spectroscopic grade acetonitrile without further purification, it cannot be excluded that some steps of the kinetics may be influenced by water impurities. Water molecules may for example act as catalysts for certain reaction steps. However, since these are bimolecular reactions and the concentrations of the involved species are low, they should not affect the kinetics of our femtosecond data. In contrast, an influence on the microsecond kinetics cannot be excluded. The influence of water on the nano- to microsecond kinetics of a similar reaction has been investigated before¹⁷. It was observed that addition of water shortens the lifetime of the long-lived transient without leading to additional photoproducts. Thus, one has to keep in mind that the microsecond time constants we obtained apply for the given solvent conditions.

Regarding the computations of Blancafort et al.⁶, we can confirm the formation of a *Z*-ketene-enol intermediate via a singlet channel in the photoreaction of oABA. A triplet path

was not considered in these calculations. For oPA, an energy barrier of 0.3 eV was calculated for the formation of the ketene. Assuming a pre-exponential factor of $\sim 10^{14} \text{ s}^{-1}$ close to the maximum for hydrogen transfers, this translates into a formation on the nanosecond timescale according to the Arrhenius equation. For oABA, we observe a faster population in the picosecond range, which translates into a barrier in the range of ~ 0.14 eV. Additionally, the computations predict the occurence of a singlet biradical precursor for which we do not find any evidence. However, the calculations were performed for the gas phase while our experiments were conducted in solution. This may alter the barrier heights significantly and so our results are not in conflict with the computations on oPA. Additionally, Blancafort et al. predict that the ketene is formed by an irreversible hydrogen transfer⁶. We performed an estimate of the quantum yield of ketene formation (Section 3.5) based on the computed oscillator strength of the ketene absorption and the fs-UV/Vis signatures. The estimated ketene yield of ~ 0.55 is about two times larger than the overall reaction quantum yield to 3MP (\sim 30%). Unfortunately, this estimate does neither confirm nor exclude that the ketene is formed irreversibly. First, the estimated value is afflicted with a large uncertainty due to the uncertainties of the experimental and computed parameters that enter the calculation. Second, the ketene forms another intermediate on the way to 3MP so that a reaction of this intermediate back to oABA may lower the reaction quantum yield at a later time.

5 Conclusions

Extending our studies on excited state hydrogen transfers to nitro-free compounds, we have characterized the photoreac-

tion of an ortho-substituted acylarene, oABA. Up to now, literature provides an incomplete picture of the photochemistry of such compounds and is blurred by tentative assignments. Employing modern techniques of time-resolved spectroscopy, the reaction path has been followed from the earliest processes after photo excitation for the first time. Quantum chemistry aided in assignments of spectral signatures, so that an ultrafast excited state hydrogen transfer could unequivocally be verified. It was found that the kinetic behavior and reaction path of oABA closely resemble the ones found for ortho-substituted nitrobenzenes^{3,10,52}. Surprisingly, changing a substituent from a nitro- to a carbonyl-group does not appear to have a dramatic effect on the photochemical processes. The complex mechanism of these reactions appears to be conferrable to acylarenes containing hydrogen atoms in ortho-position.

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References

- 1 Hydrogen Bonding and Transfer in the Excited State, Volume I & II, ed. K.-L. Han and G.-J. Zhao, John Wiley & Sons, Inc., 2010.
- 2 P. Klan and J. Wirz, *Photochemistry of Organic Compounds. From Concepts to Practice*, Wiley, Chichester, 2009.
- 3 T. Schmierer, S. Laimgruber, K. Haiser, K. Kiewisch, J. Neugebauer and P. Gilch, *Phys. Chem. Chem. Phys.*, 2010, **12**, 15653–15664.
- 4 M. Dittmann, F. F. Graupner, B. Maerz, S. Oesterling, R. de Vivie-Riedle, W. Zinth, M. Engelhard and W. Lüttke, *Angew. Chem. Int. Ed.*, 2014, 53, 591–594.
- 5 A. Migani, V. Leyva, F. Feixas, T. Schmierer, P. Gilch, I. Corral, L. Gonzalez and L. Blancafort, *Chem. Commun.*, 2011, 47, 6383–6385.
- 6 Q. Li, A. Migani and L. Blancafort, Phys. Chem. Chem. Phys., 2012, 14, 6561–6568.
- 7 S. Laimgruber, W. J. Schreier, T. Schrader, F. Koller, W. Zinth and P. Gilch, *Angew. Chem. Int. Ed.*, 2005, **44**, 7901–7904.
- 8 S. Laimgruber, T. Schmierer, P. Gilch, K. Kiewisch and J. Neugebauer, *Phys. Chem. Chem. Phys.*, 2008, **10**, 3872–3881.
- 9 B. Heinz, T. Schmierer, S. Laimgruber and P. Gilch, J. Photochem. Photobiol. A, 2008, **199**, 274–281.
- 10 T. Schmierer, W. Schreier, F. Koller, T. Schrader and P. Gilch, *Phys. Chem. Chem. Phys.*, 2009, **11**, 11596–11607.
- 11 T. Schmierer, G. Ryseck, T. Villnow, N. Regner and P. Gilch, *Photochem. Photobiol. Sci.*, 2012, **11**, 1313–1321.
- 12 A. D. Allen and T. T. Tidwell, Chem. Rev., 2013, 113, 7287–7342.

- 13 V. Leyva, I. Corral, F. Feixas, A. Migani, L. Blancafort, J. Gonzalez-Vazquez and L. Gonzalez, *Phys. Chem. Chem. Phys.*, 2011, 13, 14685– 14693.
- 14 R. W. Yip and D. K. Sharma, Res. Chem. Interm., 1989, 11, 109–116.
- 15 J. C. Scaiano, M. V. Encinas and M. V. George, J. Chem. Soc.-Perkin Trans. 2, 1980, 724–730.
- 16 J. Gebicki and S. Kuberski, J. Chem. Soc. Chem. Commun., 1988, 1364– 1365.
- 17 J. C. Netto-Ferreira and J. C. Scaiano, Can. J. Chem.-Rev. Can. Chim., 1993, 71, 1209–1215.
- 18 L. Plistil, T. Solomek, J. Wirz, D. Heger and P. Klan, J. Org. Chem., 2006, 71, 8050–8058.
- 19 N. Turro, V. Ramamurthy and J. Scaiano, *Modern Molecular Photochemistry of Organic Molecules*, University Sciences Books, Sausalito, California, 2010.
- 20 P. Kukura, D. W. McCamant and R. A. Mathies, Annu. Rev. Phys. Chem., 2007, 58, 461–488.
- 21 S. Laimgruber, H. Schachenmayr, B. Schmidt, W. Zinth and P. Gilch, *Appl. Phys. B-Lasers Opt.*, 2006, 85, 557–564.
- 22 M. Lorenc, M. Ziolek, R. Naskrecki, J. Karolczak, J. Kubicki and A. Maciejewski, *Appl. Phys. B-Lasers Opt.*, 2002, 74, 19–27.
- 23 M. A. Marangoni, D. Brida, M. Quintavalle, G. Cirmi, F. M. Pigozzo, C. Manzoni, F. Baronio, A. D. Capobianco and G. Cerullo, *Opt. Express*, 2007, **15**, 8884–8891.
- 24 E. Pontecorvo, C. Ferrante, C. G. Elles and T. Scopigno, *Opt. Express*, 2013, **21**, 6866–6872.
- 25 A. Savitzky and M. Golay, Analytical Chem., 1964, 36, 1627.
- 26 D. Löwenich, K. Kleinermanns, V. Karunakaran and S. A. Kovalenko, *Photochem. Photobiol.*, 2008, 84, 193–201.
- 27 M. J. Frisch, G. W. Trucks, H. B. Schlegel, G. E. Scuseria, M. A. Robb, J. R. Cheeseman, G. Scalmani, V. Barone, B. Mennucci, G. A. Petersson, H. Nakatsuji, M. Caricato, X. Li, H. P. Hratchian, A. F. Izmaylov, J. Bloino, G. Zheng, J. L. Sonnenberg, M. Hada, M. Ehara, K. Toyota, R. Fukuda, J. Hasegawa, M. Ishida, T. Nakajima, Y. Honda, O. Kitao, H. Nakai, T. Vreven, J. J. A. Montgomery, J. E. Peralta, F. Ogliaro, M. Bearpark, J. J. Heyd, E. Brothers, K. N. Kudin, V. N. Staroverov, R. Kobayashi, J. Normand, K. Raghavachari, A. Rendell, J. C. Burant, S. S. Iyengar, J. Tomasi, M. Cossi, N. Rega, J. M. Millam, M. Klene, J. E. Kratmann, O. Yazyev, A. J. Austin, R. Cammi, C. Pomelli, J. W. Ochterski, R. L. Martin, K. Morokuma, V. G. Zakrzewski, G. A. Voth, P. Salvador, J. J. Dannenberg, S. Dapprich, A. D. Daniels, Ö. Farkas, J. B. Foresman, J. V. Ortiz, J. Cioslowski and D. J. Fox, *Gaussian09 Revision D.01*, Gaussian Inc. Wallingford CT 2009.
- 28 R. Ahlrichs, M. K. Armbruster, R. A. Bachorz, M. Bar, H. P. Baron, R. Bauernschmitt, F. A. Bischoff, S. Bocker, N. Crawford and P. Deglmann, *TURBOMOLE 6.3*, 2010.
- 29 J. Tomasi, B. Mennucci and R. Cammi, *Chem. Rev.*, 2005, **105**, 2999– 3094.
- 30 S. Grimme and M. Waletzke, J. Chem. Phys., 1999, 111, 5645-5655.
- 31 M. Kleinschmidt, C. M. Marian, M. Waletzke and S. Grimme, J. Chem. Phys., 2009, 130, 044708.
- 32 A. Schäfer, A. Klamt, D. Sattel, J. C. Lohrenz and F. Eckert, *Phys. Chem. Chem. Phys.*, 2000, 2, 2187–2193.
- 33 S. D. Williams, T. J. Johnson, T. P. Gibbons and C. L. Kitchens, *Theor. Chem. Acc.*, 2007, **117**, 283–290.
- 34 D. A. Long, *Raman Spectroscopy*, McGraw-Hill International Book Company, 1977.
- 35 D. McCamant, P. Kukura and R. Mathies, Appl. Spectrosc., 2003, 57, 1317–1323.
- 36 E. C. L. Ru and P. G. Etchegoin, Principles of Surface-Enhanced Raman Spectroscopy, Elsevier, 2009.
- 37 E. Pretsch, P. Bühlmann and M. Badertscher, Structure Determination of

Organic Compounds: Tables of Spectral Data, Springer, 2009.

- 38 H. Lutz, E. Breheret and L. Lindqvist, J. Phys. Chem., 1973, 77, 1758– 1762.
- 39 M. Montalti, A. Credi, L. Prodi and T. Gandolfi, *Handbook of Photochemistry, Third Edition*, Taylor & Francis, 2006.
- 40 L. Giering, M. Berger and C. Steel, J. Am. Chem. Soc., 1974, 96, 953–958.
- 41 S. Chattopadhyay, C. Kumar and P. Das, J. Photochem., 1985, 30, 81–91.
- 42 H. Satzger and W. Zinth, Chem. Phys., 2003, 295, 287-295.
- 43 E. T. J. Nibbering, H. Fidder and E. Pines, *Annu. Rev. Phys. Chem.*, 2005, 56, 337–367.
- 44 P. Hamm, S. Ohline and W. Zinth, J. Chem. Phys., 1997, 106, 519-529.
- 45 T. Schrader, A. Sieg, F. Koller, W. Schreier, Q. An, W. Zinth and P. Gilch, *Chem. Phys. Lett.*, 2004, **392**, 358–364.
- 46 N. B. Colthup, L. H. Daly and S. E. Wiberley, *Introduction to Infrared* and Raman Spectroscopy, Academic Press, 3rd edn, 1990.
- 47 E. Smith and G. Dent, *Modern Raman Spectroscopy*, John Wiley & Sons Ltd, 2005.
- 48 A. C. Albrecht, J. Chem. Phys., 1961, 34, 1476–1484.
- 49 W. Parson, *Modern Optical Spectroscopy, Student Edition*, Springer, Dordrecht, Heidelberg, London, New York, 2009.
- 50 S. Kovalenko, A. Dobryakov, I. Ioffe and N. Ernsting, *Chem. Phys. Lett.*, 2010, **493**, 255–258.
- 51 V. V. Semionova, E. M. Glebov, V. V. Korolev, S. A. Sapchenko, D. G. Samsonenko and V. P. Fedin, *Inorg. Chim. Acta*, 2014, 409, Part B, 342–348.
- 52 T. Schmierer, F. Bley, K. Schaper and P. Gilch, J. Photochem. Photobiol. A-Chem., 2011, 217, 363–368.