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ARTICLE

Why Genetic Modification of Lignin Leads to Low-Recalcitrance Biomass

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Genetic modification of plants *via* down-regulation of cinnamyl alcohol dehydrogenase leads to incorporation of aldehyde groups in the lignin polymer. The resulting lignocellulosic biomass has increased bioethanol yield. However, a molecular-scale explanation of this finding is currently lacking. Here, we perform molecular dynamics simulation of the copolymer with hemicellulose of wild type and the genetically modified lignin, in aqueous solution. We find that the non-covalent association with hemicellulose of lignin containing aldehyde groups is reduced compared to the wild-type. This phase separation may increase the cell wall porosity in the mutant plants, thus explaining their easier deconstruction to biofuels. The thermodynamic origin of the reduced lignin-hemicellulose association is found to be a more favorable self-interaction energy and less favorable interaction with hemicellulose for the mutant lignin. Furthermore, reduced hydration water density fluctuations are found for the mutant lignin, implying a more hydrophobic lignin surface. The results provide a detailed description of how aldehyde incorporation makes lignin more hydrophobic and reduces its association with hemicellulose, thus suggesting that increased lignin hydrophobicity may be an optimal characteristic required for improved biofuel production.

Introduction

Plant biomass is composed of cellulose fibers embedded in a matrix of lignin and hemicellulose polymers.[1] Lignins are hydrophobic heteropolymers, which can be branched, that provide plants with mechanical rigidity and help regulate water conduction.[2-5] They are composed of three monomers or monolignols: syringyl (S), guaiacyl (G), and p-hydroxyphenyl (H), radically coupled to form a lignin polymer chain.[5] The polymerization of lignin is random, resulting in differences between lignins among taxa and species as well as cell types in the same plant.[5]

Lignin protects plants against enzymatic degradation by forming a covalently bonded network with hemicellulose that blocks enzymatic access to cellulose.[2, 5, 6] A biological route of cellulosic biofuel production involves enzymatic break down of cellulose into fermentable sugars.[7, 8] Thermochemical pretreatment of biomass increases the effectiveness of subsequent cellulose hydrolysis by increasing access of cellulolytic enzymes to cellulose. This is achieved by either partial removal of lignin and hemicellulose or by phase separation of those two polymers.[7, 9] However, currently, pretreatment is energy intensive, time consuming, and costly. Therefore, improving the pretreatment process is an essential step in making plant biomass a more economically viable biofuel source.

Recent advances in genetic engineering have shown that precursors along the biosynthetic pathway of monolignols can be incorporated into lignin, sometimes without detrimental effects on plant growth.[2, 5, 10, 11] For example, down-regulation of cinnamyl alcohol dehydrogenase (CAD), which catalyzes the final step in biosynthesis of monolignols, leads to incorporation of cinnamic aldehydes into the lignin polymer.[4, 5, 12-15] Hydroxycinnamyl and cinnamic aldehydes differ from wild type monolignols by the presence of aldehyde groups on the allyl chain, which is thus more hydrophobic than with the equivalent wild type hydroxyl groups.

Reduction in CAD activity leads to improved lignocellulosic biofuel feedstocks due to reduced lignin content and/or modified lignin structure.[16] For example, mutants with reduced CAD activity are more susceptible to enzymatic hydrolysis (saccharification) than wild-type plants in *Brachypodium*[16] and switchgrass.[17] Another study showed down-regulation of CAD to improve ethanol yield in maize.[18] However, a molecular-level understanding of how aldehyde precursor inclusion may lead to less recalcitrant biomass is lacking.

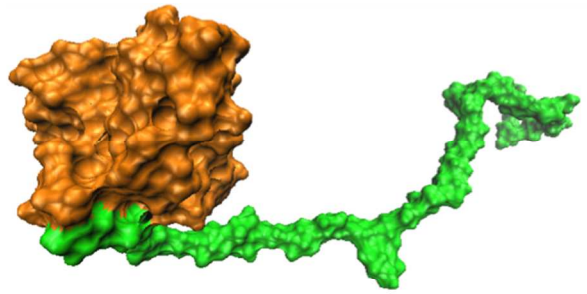

Fig. 1 Snapshot of the simulated lignin (orange)-hemicellulose (green) copolymer.

Here, we conduct atomistic molecular dynamics (MD) simulations to characterize how aldehyde precursor inclusion in lignin affects its association with hemicellulose. We find that the increase in hydrophobicity of lignin from incorporation of the aldehyde groups reduces its association with hemicellulose, with more favorable intra-lignin interactions and less favorable interactions with hemicellulose. This work illustrates how MD simulations can be a useful tool for predicting the effects of changes in plant genotype to cell wall phenotype in efforts to enhance the production of biofuels or other bioproducts.

Methods

Lignin and hemicellulose models

A previously-built crosslinked lignin–hemicellulose copolymer model,[9] constructed based on experimental data of the composition of Aspen lignin and hemicellulose, was modified for use in this study. A structural model of a covalently linked lignin-hemicellulose copolymer was generated by using available experimental information on the average chemical composition of Aspen lignin and hemicellulose.[19] Aspen lignin is composed primarily of guaiacyl (G) and syringyl (S) monomeric units, in the ratio S/G~ 1.86, connected by various linkages. Here, a linear lignin polymer was constructed that was composed of 11 G and 20 S units connected via 22 β-O-4 (β-aryl ether), four 4-O-5 (bithenyl ether), two β-β (pinoresinol) and two β-1 linkages, similar to the experimentally determined average inter-linkage composition of Aspen.[19] Hemicelluloses are branched polymers composed of sugar residues. Here, the hemicellulose polymer consisted of a (1→4) linked backbone consisting of 28 β-xylose and four β-mannose monomers. Four 4-O-methyl-glucuronic acid (4-O-MeGlcA) side-chain monomers were bonded to the xylose backbone via a (1→2) link. The lignin and hemicellulose polymers were connected end-to-end via an ether bond between the γ carbon of the lignin and the O1 oxygen of the hemicelluose, resembling a ferulate lignin-hemicellulose crosslink. The lignin-hemicellulose polymer was then hydrated in a cubic box and four Na^+ ions added to neutralize the system (4-O-MeGlcA, with pKa value of 3, was deprotonated). The system is shown in Figure 1.

Here, hydrophobicity is defined by the magnitude and distribution of the partial charges of functional groups on monolignols.. The partial charge on the oxygen atom of the hydroxyl OH group (-0.66) is more negative than that of the methoxy MeO (-0.28) and aldehyde groups (-0.40) respectively. The variance of the distribution of the atomistic partial charges of three functional groups (which contain four, three, and two atoms, respectively), C_q is 0.02, 0.13 and 0.22 for the methoxy, aldehyde and hydroxyl groups, respectively. C_q provides a quantitative measure of the distribution of the magnitude of partial charges across atoms of an overall neutral functional group. A large C_q indicates a high degree of polarization and stronger electrostatic dipole interactions.

Three models of lignin are considered here, constructed by altering partial charges of specific functional groups on S and G (Fig. 2). The following modifications were made on these models.

5-OH Model. A more hydrophilic 5-OH group replaces the 5-MeO group at position 5 present on wild-type S lignin. The oxygen atom of the 5-OH group has its partial charge decreased from -0.28 to -0.54, which is the charge of a phenolic oxygen in the CHARMM force field.[20, 21] Accordingly, the carbon atom at

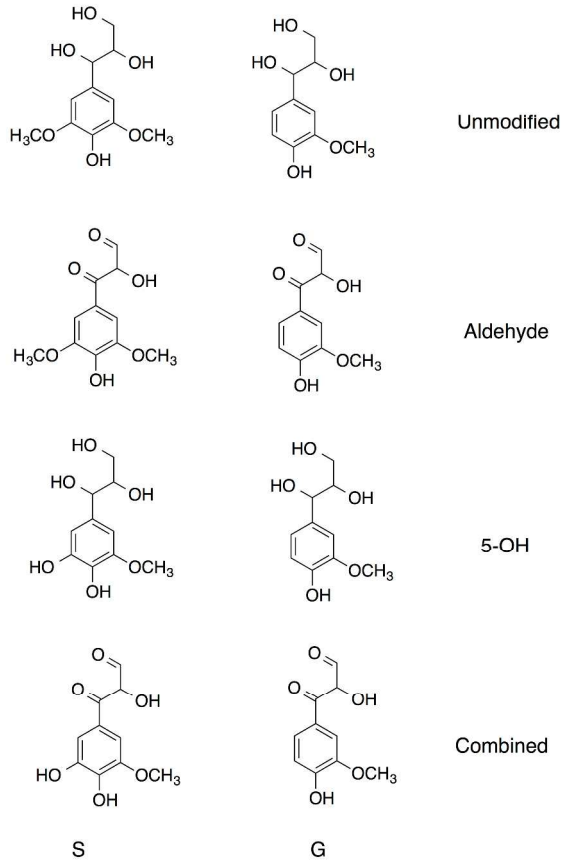

Fig. 2 Monomers of the lignin polymers simulated here, S are syringyl-like and G guiacyl-like. The full simulated system is shown in Figure 1.

position 4 on the phenyl ring has its partial charge increased by 0.07 to 0.11. This modification makes the phenol ring of S lignin less hydrophobic than in wild type plants, and reflects 5-hydroxy-guaiacyl lignin found in mutant plants.[22, 23]

ALDEHYDE MODEL. OH groups on the α, β, and γ carbon atoms of the allyl of both S and G lignin have their partial charges modified in order to model the presence of aldehyde functional groups. The partial charges of the oxygen atom of the allyl are increased from -0.66 for a hydroxyl oxygen to -0.40 for an aldehyde oxygen in the CHARMM General force field.[24] The carbon and hydrogen partial charges are also taken from the aldehyde groups in CHARMM General force field.[24] This modification makes the allyl chain of lignin more hydrophobic than in wild type plants and reflects coniferaldehyde and sinapaldehyde lignin found in mutant plants.[12, 13] A PDB file of the model is provided in the Supporting Information.

COMBINED MODEL. For the final model, the two modifications discussed above were combined to form a lignin with a 5-OH substitution and aldehyde groups on the allyl chain that reflects, 5-hydroxyconiferaldehyde, a lignin precursor that could be incorporated into lignin.[2]

For all models, β-O-4 (β-aryl ether) and β-β (pinoresinol) linkages were modified in order to preserve overall charge neutrality. Comparison is made to the unmodified. Wild-type model of Ref. 9.

Molecular dynamics simulation

MD simulations were performed using the NAMD 2.9 software[25] by employing the CHARMM carbohydrate[26, 27] and lignin[21] force fields and the TIP3P water model.[28] The Particle Mesh Ewald method[29, 30] was used with a grid spacing of 1Å and a force-switching function to smoothly transition Leonard Jones forces to zero over the range of 9-10Å. Multiple time steps of 2fs were used for bonded and short-range non-bond forces, and 6fs for long-range electrostatic forces. The neighbor list was updated every 10 steps with a pair-list distance of 11Å. Constant temperature was maintained by using the Langevin dynamics algorithm with a damping coefficient of 5 ps^{-1}. The pressure was maintained at 1atm using the Nose-Hoover Langevin piston algorithm[31, 32] that employed a piston oscillation period of 200fs and a piston damping decay time of 100 fs.

All models were solvated in a water box of size (126x126x125) Å. The solvated systems were then ionized in order to maintain the system charge neutral. The 5-OH, Aldehyde, and Combined lignin models were simulated for 388ns, 340ns, and 361ns respectively at temperature T=298K. All calculations were performed on the Titan Cray XK7 supercomputer at the Oak Ridge Leadership Computing Facility.

Analysis of all MD trajectories was performed with the VMD software[33] by using local scripts. An atomic contact was defined as two atoms separated by less than 3 Å. An atom is defined as hydrophobic when having a partial charge $|q|<0.3$ and hydrophilic otherwise. In comparison, the partial charges of apolar moieties in molecular mechanics force fields of proteins have $|q|<0.4$.[34] A sphere radius of 1.4 Å was employed in the computation of the solvent-accessible surface area (SASA). The surface water density, ρ is given by:

$$\rho = \frac{N}{S\,d} \quad (1)$$

where N is the number of water molecules at a distance less than 4.5Å from the lignin, S is the solvent accessible surface area of lignin, and d is the thickness of the hydration shell, which is taken as 4.5Å.[35] The surface energy density γ is given by:

$$\gamma = \frac{E}{S} \quad (2)$$

where E is the interaction energy between lignin and water. Lastly, the number of hydrogen bonds was calculated over the last 10ns of simulation time of the Aldehyde and Combined models at intervals of 1ps. Here, the cutoff distance between donor and acceptor is 3.5 Å and the angle formed by donor, hydrogen, and acceptor must be less than 30°. This geometric definition has been shown to roughly reproduce experimental water hydrogen bond activation energies[36] and to match experimentally determined water hydrogen bond geometries.[37]

Results

Structure and contacts.

The radius of gyration (R_g) of each model is shown as a function of simulation time in Fig. 3. The Aldehyde model initially expands slightly from its initial structure, then relaxes into a more compact state after about 60 ns (Fig. 3, blue) and on average is the most compact of the models and has the smallest fluctuations of R_g (Table 2). The 5-OH model displays considerably larger variations in R_g than the others and elongates near the end of the simulation to a greater extent (Fig.

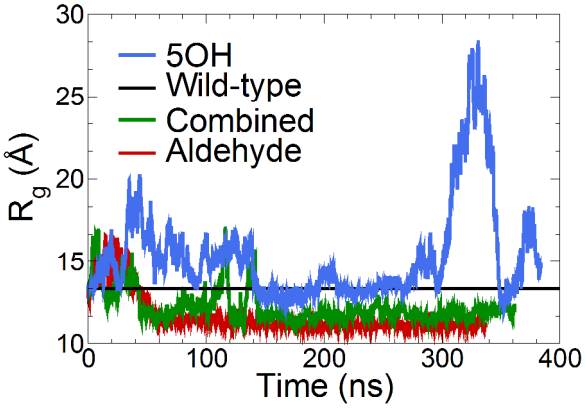

Fig. 3 Radius of gyration of the lignin polymers (R_g) as a function of time for each model of lignin. The black line represents the average R_g of the Wild-type (unmodified) model of lignin from Ref. 9.

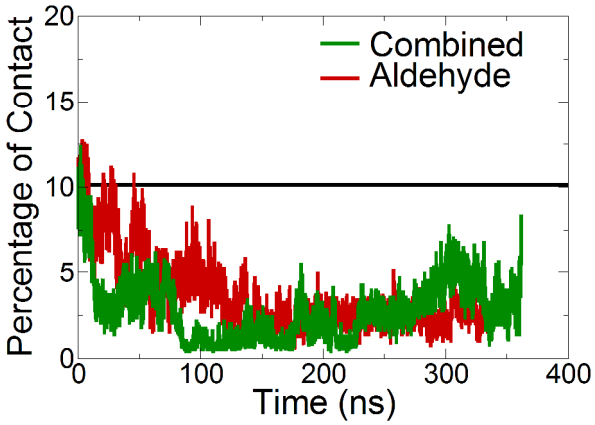

Fig. 4 Percentage of lignin atoms forming contacts with hemicellulose, defined here as the number of atoms of lignin within 3 Å of hemicellulose divided by the total number of lignin atoms. The black line is for the Wild-type (unmodifed) model from Ref. 9.

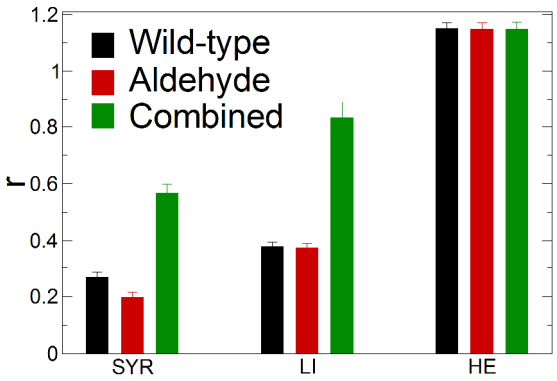

Fig. 5 Ratio of average solvent-accessible hydrophilic surface area to hydrophobic surface area, *r*, for each model of lignin (LI), hemicellulose (HE), and syringyl (SYR) monomer. Data averaged over the last 150ns.

3, red). Since the shape of the 5-OH model did not converge in simulation, it is not discussed further here. The Combined model shows a slightly greater R_g than the Aldehyde model, and has a more extended and variable structure (Fig. 3, purple). Comparison with the wild-type (unmodified) model (Table 1) indicates that inclusion of aldehyde groups reduces the size of the lignin.

Fig. 4 quantifies the proportion of lignin contacts with hemicellulose, defined here as the percentage of lignin atoms at a distance less than 3 Å from the hemicellulose. Compared to the Wild-type model, in the aldehyde-containing models contacts with hemicellulose are reduced over the course of the simulation.

The ratio of lignin-hemicellulose hydrogen bonds of the Combined model to the Aldehyde model over the last 1 0ns of simulation time is 2.4 ± 1.3. In comparison, the ratio of the total number of possible hydrogen-bond acceptors/donors on a monolignol monomer between the Aldehyde and Combined models is 1:1.25. Therefore, there is a significantly greater amount of hydrogen-bond saturation in the Combined than in the Aldehyde model.

Table 1 Average radius of gyration and percentage of lignin atoms in contact with hemicellulose for each lignin model averaged over the last 150 ns of each trajectory.

Model	R_g (Å)	Contacts (%)
Wild-type	13.3 ± 0.2	10.1 ± 1.3
Aldehyde	11.1 ± 0.1	2.4 ± 0.7
Combined	11.8 ± 0.3	2.8 ± 1.4

Surface polarity

Ratios of solvent accessible hydrophilic to hydrophobic surface areas given by *r*,

$$r = \frac{SASA_{philic}}{SASA_{phobic}} \quad (3)$$

are shown in Fig. 5. The ratios are given for the lignin polymer (LI), an isolated syringyl monomer (SYR), and the hemicellulose polymer (HE). The ratio *r* is higher for LI than for SYR for all models, reflecting the preferential exposure of hydrophilic groups to the solvent in the lignin polymer.[35] For SYR, the Wild-type model has a higher *r* than the Aldehyde model (Fig. 5, black and blue) because of the larger number of hydrophobic moieties introduced in the latter. However, the ratio for LI is the same for the Wild-type and Aldehyde polymers, indicating that the addition of aldehyde groups does not change the overall polarity of the lignin surface. The values of *r* for the Combined model of both SYR and LI are larger than the other models, reflecting the addition of hydrophilic moieties, and indicates an increase in the overall polarity of the lignin surface (Fig. 5, purple). The *r* for hemicellulose is the same for all models.

Surface hydrophobicity

The normalized variance of the water density, *Var(ρ)*, was used to quantitatively compare the hydrophobicity of the surface of each lignin model (Table 2). Here,

$$\mathrm{Var}(\rho) = \frac{\sigma_\rho^2}{\langle \rho \rangle} \quad (4)$$

where σ_ρ is the standard deviation and $\langle\rho\rangle$ is the mean of the water density over a period of 10ns. The higher *Var(ρ)*, i.e. the

Table 2 Normalized variance of water density fluctuations of each lignin model. Values are an average of five 10ns segments calculated over the last 50ns of the simulation.

Model	*Var(*ρ) (×10^{-4})
Wild-type	0.04 ± 0.02
Aldehyde	0.13 ± 0.05
Combined	0.13 ± 0.06

higher the density fluctuations in the hydration shell, the more hydrophobic is the solute surface.[38, 39] *Var(ρ)* for the Aldehyde and Combined models are statistically similar (Table 2). However, the Wild-type model has a lower value for *Var(*ρ*)*, which indicates that the hydrophobicity of the lignin surface is increased by the presence of the aldehyde groups.

Interaction energies

Non-bonded interaction energies are listed in Table 3. Both the Combined and the Aldehyde models have more favorable lignin-to-lignin (LI-LI) interactions and less favorable lignin-to-hemicellulose (LI-HE) interactions compared to the Wild-type model (Table 3). The Aldehyde model has the energetically least favorable interaction with hemicellulose, which reflects the smaller LI-HE contact reported in Fig. 5.

Both the Aldehyde and the Combined models interact less favorably with water (Table 4). Values for the lignin-water surface energy density γ, given by Equation 2, are shown in Table 4. The highest values for surface energy density are for the Aldehyde and Combined Models.

Discussion

The recalcitrance of lignocellulose biomass to hydrolysis into fermentable sugars is the main barrier to economically viable second-generation biofuel production. It has been shown that plants with reduced activity of the CAD enzyme, which catalyzes the last step of monolignol biosynthesis, integrally incorporate aldehydes into lignin[12] and also show improved yields of ethanol from biomass compared to wild-type.[17, 18] However, a molecular-scale explanation of this improvement in

Table 3 Average interaction energies of each lignin model with lignin (LI-LI) and hemicellulose (LI-HE). Data averaged over the last 10ns of the simulation.

Model	LI-LI (kcal/mol)	LI-HE (kcal/mol)
Wild-type	1083 ± 22	-136 ± 16
Aldehyde	687 ± 21	-27 ± 7
Combined	685 ± 21	-48 ± 13

Table 4 Average surface energy density of each lignin model during the last 10ns of simulation time. Obtained as the lignin-water interaction energy divided by the lignin SASA.

Model	γ (kcal/mol/Å^2)
Wild-type	-0.27 ± 0.08
Aldehyde	-0.05 ± 0.06
Combined	-0.06 ± 0.07

biomass conversion to ethanol has not been provided until now. Decreasing the non-covalent association between lignin and hemicellulose is a potential explanation for the increased cellulose hydrolysis, by increasing the accessibility to cellulose of cellulolytic enzymes.[9]

Inclusion of the hydrophobic aldehyde groups in lignin reduces lignin-hemicellulose contact (Table 1). Generalizing this finding, we suggest that any genetic modification that renders lignin more hydrophobic may impact favorably biomass conversion to biofuel. A thermodynamic explanation of this finding is obtained by considering changes in lignin interaction with itself and with hemicellulose. The average interaction energies (Table 3) imply that the presence of aldehyde moieties leads to a variant lignin that interacts more favorably with itself than does wild-type lignin. Additionally, the lignin-hemicellulose interaction energy per contact is more favorable for wild-type lignin (-13.5 kcal/mol) than for the Aldehyde model (-11.3 kcal/mol). Finally, lignin:water interaction energy densities (Table 4) show the wild-type model to interact more favorably with water, which is chemically more similar to the hydrophilic hemicellulose than the hydrophobic lignin. Therefore, compared to wild-type lignin, inclusion of aldehydes in the variant lignin thermodynamically promotes self-association over non-covalent contact with hemicellulose.

A quantitative measure of the hydrophobicity of a surface is the magnitude of the density fluctuations of its hydration water, *Var(*ρ*)*: the greater the fluctuations, the more hydrophobic the surface.[38] *Var(ρ)* at the surface of the wild-type model is lower than for the Aldehyde and Combined models, (Table 2) thus suggesting that the genetic modification introduced by downregulation of CAD increases the hydrophobicity of the lignin surface. This is consistent with the differences in the water-lignin surface energy density. Although the surface energy density differs between the Wild-type and Aldehyde models, their respective SASA ratios are similar; this may be due to sharp cut-off used here to differentiate between a hydrophilic and hydrophobic atom. The ratio of the hydrophobic to hydrophilic SASA of the lignin polymer (LI) is higher than that of an isolated syringyl monomer (SYR). This implies that lignin adopts conformations that maximize exposure of its hydrophilic components.

Previous computational studies have focused on wild type lignin.[3] Quantum chemical studies have examined bond dissociation enthalpies relevant to lignin deconstruction,[40-44] radical coupling reactions in lignin biosynthesis[45-47] and dilignol interaction energies with ionic liquid ions.[48] MD simulations of wild-type lignin have probed its shape and configurations[35, 49, 50] as well as its interaction with cellulose[51-53] and hemicellulose.[9] The present study shows that mutating lignin *in silico* can yield macromolecular properties that are consistent with the phenotype of mutant plants. Therefore, the use of MD simulation as a predictive tool for the effect of genetic modifications on lignin structure could be further pursued in future work.

Conclusions

Effects of down regulation of the CAD enzyme in plants were modelled by introducing aldehyde functional groups in a lignin:hemicellulose copolymer. MD simulations of the model show the modified lignin to be more hydrophobic and to associate less with hemicellulose than does wild-type lignin. The looser coupling of lignin and hemicellulose could lead to increased accessibility of cellulose in plant biomass. Therefore, the results of this study explain why plants with reduced CAD activity are more easily deconstructed for biofuel production. This work suggests the potential for using MD simulation as a tool to connect genotype to phenotype in genetic modifications to plants.

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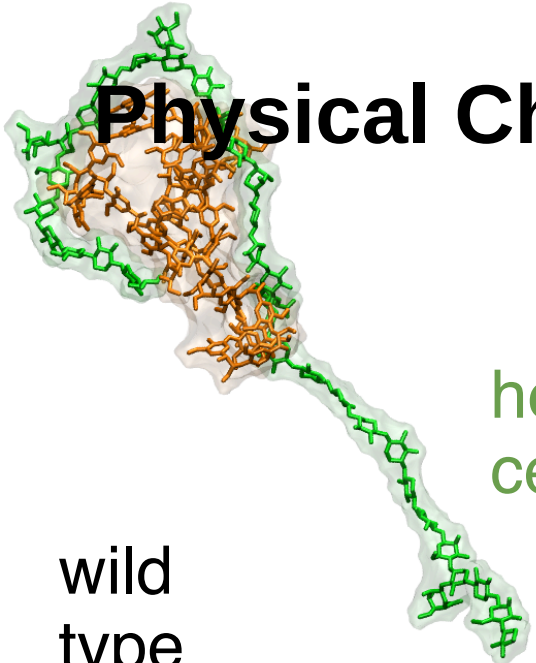

hemi
cellulose

wild
type

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CAD
mutant