

# Environmental Science Processes & Impacts

Accepted Manuscript



This is an *Accepted Manuscript*, which has been through the Royal Society of Chemistry peer review process and has been accepted for publication.

*Accepted Manuscripts* are published online shortly after acceptance, before technical editing, formatting and proof reading. Using this free service, authors can make their results available to the community, in citable form, before we publish the edited article. We will replace this *Accepted Manuscript* with the edited and formatted *Advance Article* as soon as it is available.

You can find more information about *Accepted Manuscripts* in the [Information for Authors](#).

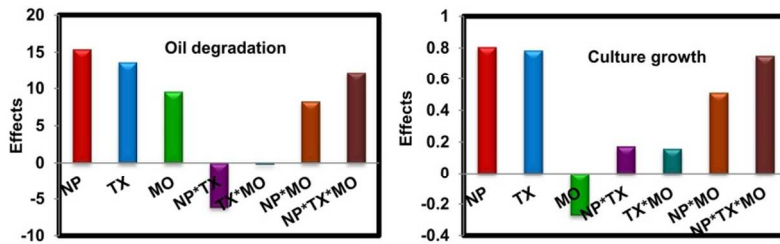
Please note that technical editing may introduce minor changes to the text and/or graphics, which may alter content. The journal's standard [Terms & Conditions](#) and the [Ethical guidelines](#) still apply. In no event shall the Royal Society of Chemistry be held responsible for any errors or omissions in this *Accepted Manuscript* or any consequences arising from the use of any information it contains.



[rsc.li/process-impacts](http://rsc.li/process-impacts)

**Graphical Abstract:**

While each of the strategies nutrient addition (NP), surfactant addition (TX) and bioaugmentation with microorganisms (MO) can enhance oily sludge biodegradation, employing these strategies simultaneously leads to enhanced biodegradation through synergistic effects.



**Environmental Impact:**

Proper management of oily sludge is essential for preventing soil and groundwater contamination. In this study, a  $2^3$  full factorial design is used to evaluate the impact of various bioremediation strategies for oily sludge decontamination, i.e., bioaugmentation with indigenous microorganisms (MO), biostimulation with nutrients (NP) and biostimulation with surfactants (TX). This design reveals effective strategies while minimizing the experimental runs. Batch biodegradation studies were conducted over a period of 30 days. The effects were computed based on the  $2^3$  design and statistically significant effects were identified based on ANOVA. The main effects of nutrients, surfactant and bioaugmentation were all positive and significant for oil degradation. Significant synergistic effects among the various strategies were also observed.

1 **Evaluation of Bioaugmentation and Biostimulation Effects on the Treatment of Refinery**  
2 **Oily Sludge using 2<sup>n</sup> Full Factorial Design**

3  
4  
5  
6  
7 **by**

8  
9  
10  
11 **Jublee Jasmine and Suparna Mukherji\***

12 Centre for Environmental Science and Engineering (CESE)  
13 Indian Institute of Technology, Bombay, Powai, Mumbai-400076.  
14

15 Revised Version

16 Submitted to

17  
18 **Environmental Science: Processes and Impacts**

19  
20 April, 2014

21  
22  
23  
24  
25 \*Corresponding Author. Professor, CESE, IIT Bombay, Powai, Mumbai 400076;  
26 Email: mitras@iitb.ac.in; Phone: 91-(022)-2576-7854; Fax: 91-(022)-2576-4650  
27

28        **Abstract**

29        Bioremediation approaches for treatment of oily sludge from a refinery were evaluated  
30        using the  $2^3$  factorial design. The three strategies tested were bioaugmentation with  
31        indigenous microbial consortia (MO) isolated from oily sludge, biostimulation with  
32        nutrients (NP) and biostimulation with the surfactant Triton X-100 (TX). Eight  
33        experimental runs were conducted in triplicate with factor settings +/- (high/low) as per  
34        the  $2^3$  design. The main effects and various interaction effects of the factors on oil  
35        degradation and microbial growth in suspension were evaluated during a 30 day study.  
36        Multifactor ANOVA could reveal the significant effects while the normal order score  
37        approach failed in this scenario. The main effect of biostimulation with nutrients in the  
38        form of nitrate and phosphate as well as biostimulation with Triton X-100 was positive  
39        and significant when both oil degradation and microbial growth in suspension were  
40        chosen as the response variable. However, the main effect of bioaugmentation was only  
41        significant for oil degradation but was insignificant for microbial growth at 90%  
42        confidence level. The MO-NP binary interaction and the MO-NP-TX ternary interaction  
43        were positive and significant indicating the synergistic effect of these strategies on oil  
44        degradation and microbial growth. All other binary interactions were found to be  
45        insignificant.

46

47        **Keywords:** ANOVA; Bioaugmentation; Biostimulation; Indigenous microbial  
48        consortium; Refinery oily sludge

49

50

## 51           **1. Introduction**

52

53           Oily sludge is generated from petroleum refineries in huge quantities as a byproduct of  
54           various processes and operations. Massive amount of oily sludge is also disposed off  
55           from the crude oil storage tank bottoms during cleaning and maintenance. Handling and  
56           disposal of such huge volume of waste becomes a major challenge. Oily sludge is a  
57           complex mixture of petroleum hydrocarbons and other solids including heavy metals that  
58           are carcinogenic and potent immunotoxicants. Apart from physicochemical treatment  
59           technologies, bioremediation is a clean, environmentally friendly treatment technology  
60           that can be applied for degradation of oily sludge generated in oil refineries <sup>1-3</sup>. Although  
61           successfully applied, landfarming approaches has certain limitations in terms of large  
62           space and time requirements and air pollution due to emission of volatile organic  
63           compounds. In contrast, slurry phase degradation can provide rapid and extensive  
64           degradation of oil by enhancing mass transfer rates and promoting interaction among  
65           microorganisms, pollutants and nutrients <sup>4-6</sup>.

66           Two approaches can be used to enhance the bioremediation process i.e., bioaugmentation  
67           with native or tailored microbial consortium and biostimulation with nutrients and  
68           surfactants in controlled batch slurry systems <sup>4,7-11</sup>. However, effect of such approaches is  
69           not beneficial in all scenarios. The possible reasons could be site specific features such as  
70           soil or sludge type, distribution of contaminated hydrocarbons and presence/distribution  
71           of indigenous microorganisms capable of degrading the contaminants <sup>8,12,13</sup>.

72           A mixed consortium of microorganisms may exhibit various modes of hydrocarbon  
73           uptake, such as, uptake of soluble hydrocarbons, uptake of emulsified forms or direct  
74           interfacial uptake facilitated by development of hydrophobic cell surfaces.  
75           Bioaugmentation with mixed microbial consortia is preferred due to their wide metabolic  
76           networks, through which they can easily assimilate the complex hydrocarbons in oily

77 sludge or oil contaminated soils <sup>9,14-18</sup>. Moreover, enrichment of indigenous  
78 microorganisms and bioaugmentation with these enriched microorganisms well adapted  
79 to the contaminated environment has been recommended by various researchers <sup>10,15</sup>.  
80 Biostimulation with surfactants may increase the bioavailability of hydrocarbons by  
81 emulsification and also by altering the microbial cell surface properties so as to increase  
82 the interaction between the microbes and hydrocarbon contaminants <sup>2,11,12,17-20</sup>. Often oil  
83 contaminated soil or oily sludge is found to have much higher carbon content in  
84 comparison to nitrogen (N) and phosphorous (P). Adequate supply of N and P is essential  
85 for microbial growth and contaminant degradation. Moreover, during the course of  
86 natural attenuation nutrient depletion may lead to reduction in the indigenous microbial  
87 population. Thus, biostimulation with nutrients in the form of N and P is often found to  
88 induce contaminant degradation by the native microbial population while the activity of  
89 bioaugmented cultures is also enhanced <sup>2,11,12,21</sup>.

90 Laboratory feasibility tests for bioremediation are essential to determine the potential of  
91 indigenous microbes to degrade the pollutants and to evaluate strategies for optimizing  
92 the rate and extent of degradation before pilot/full scale design of in-situ or ex-situ  
93 treatment schemes. Various studies focusing on comparative treatment studies of  
94 different bioremediation strategies make use of statistical tools to evaluate the significant  
95 differences between treatments <sup>3,11,12</sup>. In this study a 2<sup>n</sup> full factorial design was used to  
96 investigate the effect of various factors so as to identify an appropriate treatment scheme  
97 for bioremediation of oily sludge from a refinery. This design can be used for screening a  
98 number of independent factors while minimizing the number of experimental runs.  
99 Statistical analysis of the results can provide useful information on bioremediation  
100 strategies. Three factors likely to influence biodegradation of oily sludge in laboratory  
101 batch systems were identified as bioaugmentation with indigenous microorganisms,

102 biostimulation with nutrients and biostimulation with surfactants. Two different response  
103 variables were chosen, i.e., oil degradation in the system and microbial growth in  
104 suspension after 30 days. A key objective was to determine the main effect of each of  
105 these factors and the interaction effect between various factors so as to reveal possible  
106 synergism/antagonism among the three strategies. Although these strategies are  
107 commonly employed for oily sludge bioremediation, synergistic/antagonistic interactions  
108 among these strategies have not been explored by other researchers. The effects were  
109 quantified based on the  $2^3$  design and the significance of these effects was determined  
110 based on Analysis of variance (ANOVA).

111

## 112 **2. Materials and Methods**

### 113 **2.1 Source of chemicals**

114 The chemical surfactant TritonX-100 was procured from SD Fine Chemicals Pvt. Ltd.  
115 (Mumbai, India). The various chemicals used for preparation of mineral media, nutrient  
116 broth and bacteriological agar were procured from SD Fine Chemicals Pvt. Ltd., SRL  
117 industries Ltd., Merck and Hi Media Pvt. Ltd. High purity dichloromethane (DCM) used  
118 for extraction was obtained from Merck (India).

### 119 **2.2 Source of oily sludge**

120 The oily sludge was obtained from the weathering pits of a petroleum refinery in Mumbai  
121 (India) during August, 2010. The sludge was dewatered and centrifuged under high  
122 pressure to recover almost 90% of the crude oil prior to its disposal in the weathering  
123 pits. The sludge was collected from the refinery site, dried, sieved, homogenized and  
124 stored at 4 °C. The batch biodegradation studies reported here were conducted almost two  
125 years after the sludge was collected (October-November, 2012).

### 126 **2.3 Isolation and Enrichment of microorganisms from refinery oily sludge**



127 Laboratory experiments were conducted to isolate and enrich microorganisms from the  
128 oily sludge. Enrichment was conducted in 500 mL flasks containing 100 mL mineral  
129 medium<sup>17</sup> with 0.5% (w/v) oil extracted from the sludge as the sole substrate. Isolation of  
130 pure cultures was carried out both by spread plating and streak plating in nutrient agar  
131 plates. The isolates were identified based on 16S rDNA sequencing (Macrogen, Inc.,  
132 Korea) followed by BLAST analysis (NCBI) and were identified based on closest match  
133 with available sequences in the database. Five pure cultures were combined and used for  
134 bioaugmentation in the oily sludge biodegradation studies.

#### 135 **2.4 2<sup>n</sup> Full Factorial Design**

136 A 2<sup>3</sup> full factorial design was utilized in which eight runs were conducted at appropriate  
137 setting of each factor. The three factors were chosen as bioaugmentation with indigenous  
138 cultures isolated from sludge (MO), biostimulation with nutrients (NP) and  
139 biostimulation with Triton X-100 (TX), a chemical surfactant. The effect of the three  
140 factors were studied on two response variables i.e., % degradation of total petroleum  
141 hydrocarbons (TPH) in sludge over 30 days and increase in viable count in the aqueous  
142 phase ( $\ln(N/N_0)$ , where  $N_0$  and  $N$  are viable count in the aqueous phase at 0 and at 30  
143 days). Eight runs were conducted as per the design matrix for 2<sup>3</sup> full factorial design to  
144 determine the main effects, and binary and ternary interaction effects<sup>22</sup>. The level of the  
145 three factors (+/-) in each run was controlled as per the design matrix. The normal plot  
146 approach and analysis of variance (ANOVA) approach were both used to evaluate the  
147 significant main effect and interaction effects. The ANOVA was performed using  
148 STATISTICA ver.8.

#### 149 **2.5 Biodegradation Study**

150 A thirty day long biodegradation study was set-up as per the full factorial design matrix.  
151 For each run as specified in this matrix, triplicate batches were set-up. Each batch flask

152 (500 mL) contained 10% (w/v) oily sludge in 100 mL mineral media (MM).  
153 Biostimulation consisted of addition of nutrients in the form of nitrate-nitrogen ( $\text{NO}_3\text{-N}$ )  
154 and phosphate-phosphorous ( $\text{PO}_4\text{-P}$ ). The MM already contained some N and P at the  
155 baseline level (-) of 222.4 mg/L N as  $\text{NH}_4\text{-N}$  and 198.66 mg/L P as  $\text{PO}_4\text{-P}$  such that the  
156 N:P ratio was 1.21:1 (mass basis). For nutrient addition (NP +) additional N and P i.e., 70  
157 mg/L  $\text{NO}_3\text{-N}$  and 31.2 mg/L  $\text{PO}_4\text{-P}$  was supplemented in the medium in the form of  
158  $\text{KNO}_3$  and  $\text{KH}_2\text{PO}_4$ , respectively. A nonionic surfactant, TritonX-100 was also used to  
159 study its effect on the extent of oily sludge degradation. Flasks biostimulated with Triton  
160 X-100 (TX +) contained the surfactant at twice the critical micelle concentration (CMC).  
161 For bioaugmentation (MO +), 5 mL of the 5-membered reconstituted consortium adjusted  
162 to unit absorbance at 600 nm was added as inoculum. These cultures enriched and  
163 isolated from the oily sludge were maintained in the laboratory using 0.5% (w/v) oil  
164 extracted from the sludge. The reconstituted consortium was prepared by addition of each  
165 of the strain in equal proportion after growing them up to end of log phase.

166 The flasks were sacrificed initially (i.e., 0 day) and after 30 days of incubation at 35°C  
167 and 150 rpm in a rotary shaker. The sludge slurry contained in the flasks was filtered. The  
168 residual sludge collected in the filter paper was further analyzed for the residual oil  
169 content at initial i.e., 0 day and after 30 days. The aqueous suspension obtained was  
170 analyzed for pH, viable cell count and total organic carbon (TOC) content. Moreover,  
171 liquid-liquid extraction using dichloromethane as solvent (in the ratio of 1:1)<sup>19</sup> was also  
172 performed to estimate oil/total petroleum hydrocarbon (TPH) in the aqueous suspension.  
173 Residual TPH in sludge was estimated by soxhlet extraction using dichloromethane as  
174 solvent followed by gravimetric analysis and %degradation of oil was computed based on  
175 the zero day values. Standard error (SE) was based on propagation of error for studies  
176 conducted in triplicate setup. Viable counts in the aqueous phase of the sludge slurry set-

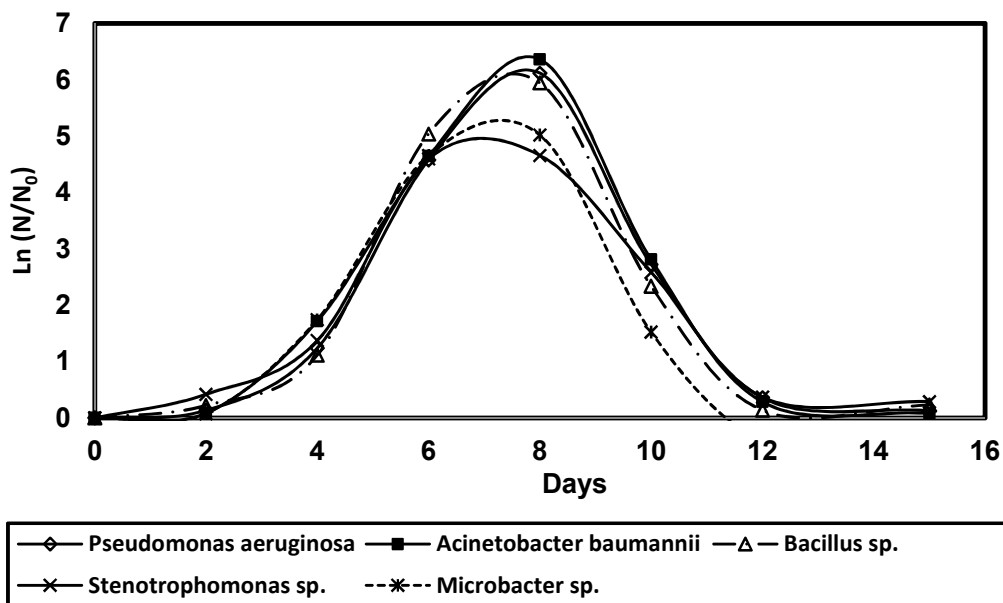
177 up were determined through standard plate count procedure at 0 and 30 days, respectively  
178 and  $\ln(N/N_0)$  was determined as a measure of culture growth in suspension.

179

### 180 **3. Results and Discussion**

181 Oily sludge is a natural material that is often reported to have microbial cultures  
182 associated with it. In batch biodegradation studies with this oily refinery sludge Jasmine  
183 and Mukherji <sup>23</sup> have earlier demonstrated that in batch systems where no microbial  
184 cultures were added (un-spiked controls) a large variability in oil biodegradation was  
185 observed ( $44 \pm 10\%$  over 30 days). In contrast, systems containing 0.1% sodium azide  
186 added as a biocide showed negligible oil degradation ( $6 \pm 4.5\%$  over 30 days). Loss of oil  
187 in the un-spiked controls was found to increase progressively with time and this was  
188 coupled with increase in culture count in the aqueous phase. In some cases,  
189 biodegradation in the un-spiked controls were almost comparable to systems where  
190 specific aliphatic and aromatic hydrocarbon degrading *Burkholderia* cultures were added  
191 extraneously. The predominant cultures found in the aqueous phase of the un-spiked  
192 controls were isolated and identified. The large variability observed in the unspiked  
193 controls indicated heterogeneity in distribution of these microorganisms in the sludge.  
194 The strains isolated from oily sludge were identified through 16 S rRNA analysis. The  
195 predominant cultures were identified as *Microbacter* sp., *Bacillus* sp., *Pseudomonas*  
196 *aeruginosa*, *Acinetobacter baumannii* and *Stenotrophomonas* sp. Each of these strains  
197 exhibited good growth when 0.5 %(w/v) of the oil extracted from oily sludge was  
198 provided as sole source of carbon and energy (Fig. 1) and end of log phase was reached  
199 within 7 days. Ongoing studies have revealed remarkable ability of these cultures to utilize  
200 hydrocarbons across various groups as sole substrate, including n-alkanes, cycloalkanes,  
201 and 2-, 3- and 4-ring PAHs (unpublished results). Although degradation of hydrocarbons,

202 crude oil and oily sludge by *Pseudomonas* sp. *Acinetobacter* sp. and *Bacillus* sp. is widely  
 203 reported <sup>7, 14, 24-27</sup>, no previous studies have reported the role of *Microbacter* sp. and  
 204 *Stenotrophomonas* sp. in oily sludge degradation.



205

206 **Fig. 1 Growth of pure cultures isolated from oily sludge on 0.5%(w/v) of extracted**  
 207 **oil provided as sole substrate in batch cultures**  
 208

209 In this study, a reconstituted consortium comprised of these strains was used for  
 210 bioaugmentation. The use of mixed indigenous microbial consortia may be advantageous  
 211 due to broader metabolic capacity, synergistic effects and co-metabolic effects <sup>7,21</sup>. These  
 212 indigenous microorganisms are expected to be better acclimatized to the oily sludge and  
 213 may exhibit better tolerance to co-contaminants <sup>11</sup> compared to other extraneous oil  
 214 degrading cultures. Nitrate was supplemented as nitrate-nitrogen in the batches with  
 215 nutrient supplementation since an excess of ammonium nitrogen is known to be toxic to  
 216 some microorganisms. An excess of phosphates is also sometimes reported to be toxic to  
 217 microorganisms, such that microbial degradation may be adversely affected in nutrient  
 218 supplemented systems. The nonionic surfactant Triton X-100 was used in this study since

219 nonionic surfactants are comparatively less toxic and Triton X-100 has been reported to  
220 enhance microbial degradation of oil<sup>17,19,20</sup>.

221 The study was designed as per 2<sup>3</sup> factorial design. The conditions prevailing in each  
222 run/treatment is illustrated in Table 1. The oily sludge contained 9-10.5% oil on dry  
223 weight basis. Significant variation was observed in the experiments due to heterogeneous  
224 distribution of oil and microorganisms in the sludge as illustrated by the SE values. TPH  
225 associated with the aqueous phase was consistently found to be very low ( $0.35 \pm 0.17\%$ ),  
226 hence, %degradation was estimated solely based on TPH determined by soxhlet  
227 extraction of the sludge. This study revealed that bioaugmentation with microorganisms  
228 and biostimulation with nutrients and surfactants increased the extent of degradation of  
229 oil in oily sludge to 57 ( $\pm 9.3$ ) % over 30 days. In contrast, the individual treatments (i.e.,  
230 only addition of microorganisms or only addition of nutrients or only addition of  
231 surfactants) yielded much lower degradation of oil. Oil degradation over 30 days was  
232 only  $6.7 \pm 3.1\%$  in the controls where neither bioaugmentation nor biostimulation was  
233 performed. Interestingly, for studies conducted with the sludge within a year of sludge  
234 collection, TPH degradation in the un-inoculated controls was much higher, i.e.,  
235  $44 \pm 10\%$ <sup>23</sup>. Thus, prolonged storage at 4°C decreased the activity of the indigenous  
236 cultures. With addition of nutrients only, oil degradation with respect to initial was  $32.4$   
237  $\pm 9.7\%$  whereas only addition of Triton X 100 increased oil degradation to  $39.1 \pm 4.6\%$   
238 over 30 days. Only bioaugmentation with microbial consortium caused  $20.5 \pm 3.0\%$  oil  
239 degradation over 30 days. Further interpretation regarding the impact of using multiple  
240 strategies, i.e., simultaneous addition of nutrients and surfactants and simultaneous  
241 bioaugmentation and biostimulation with nutrients were interpreted using factorial design  
242 and ANOVA concepts.

243

244

245

246

**Table 1. Oil degradation and culture growth in suspension over 30 days for various treatments for the oily sludge biodegradation study as per 2<sup>3</sup> design**

Run	Factors			Response variables			
	NP	TX	MO	% Degradation		Ln(N/N <sub>0</sub> )	
				Mean	SE	Mean	SE
1	-	-	-	6.7	3.1	0.40	0.28
2	+	-	-	32.4	9.7	1.28	0.32
3	-	+	-	39.1	4.6	1.61	0.37
4	+	+	-	27.8	8.8	1.33	0.39
5	-	-	+	20.5	3	0.22	0.32
6	+	-	+	38.2	8.9	0.61	0.31
7	-	+	+	28.0	4.4	0.23	0.06
8	+	+	+	57.8	9.3	2.48	0.49

247

248

249

250

251

252

253

254

255

256

257

258

259

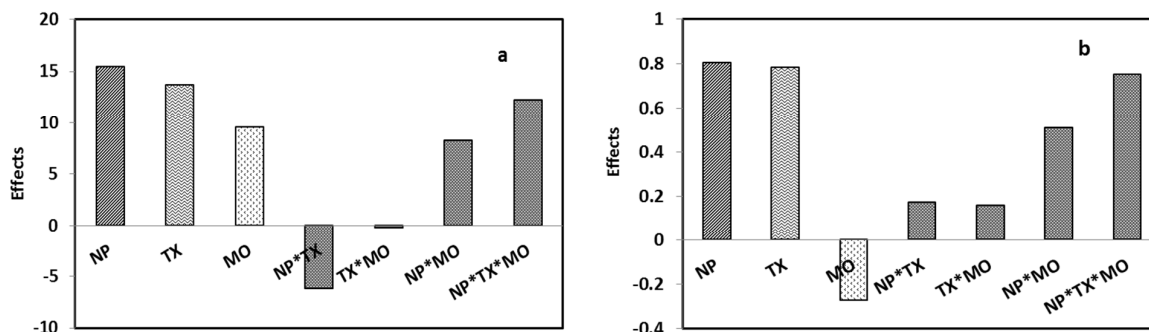
260

261

262

□ NP: nutrients, TX: Triton X 100, MO: microorganisms, SE: standard error, N: Viable count of microorganisms at 30 day, N<sub>0</sub>: Viable count of microorganisms at 0 day

The influence of the various factors i.e., bioaugmentation with indigenous cultures isolated from sludge (MO), biostimulation with nutrients (NP) and biostimulation with Triton X-100 (TX) could be determined for both set of response variables, i.e., oil degradation and culture growth in suspension over 30 days based on the 2<sup>3</sup> factorial design as illustrated in Fig. 2a-b. The average oil degradation over 30 days across all the experimental runs was 31.3% and average culture growth in suspension (Ln(N/N<sub>0</sub>)) was 1.02. All the main effects and interaction effects elevated oil degradation over 30 days except for the binary interactions between nutrient and surfactant addition and bioaugmentation and surfactant addition. Surprisingly, the main effect of bioaugmentation was to reduce culture growth in suspension. All the other main effects and binary and ternary interaction effects were positive when culture growth in suspension was used as the response variable.

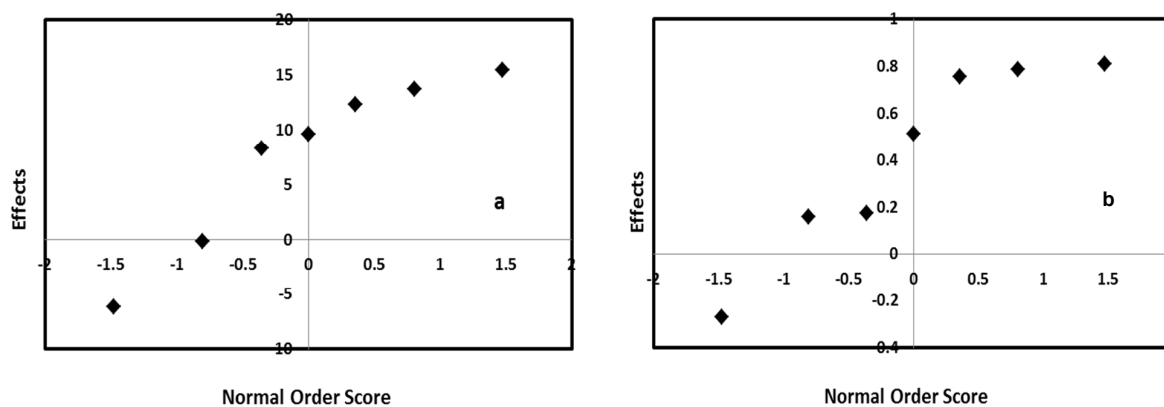


**Fig. 2 Magnitude of main effects and binary and ternary interaction effects for the oily sludge biodegradation study as per  $2^3$  factorial design for (a) oil degradation and (b) culture growth in suspension as the response variable**

After the average values of main effect, the two way interaction effects and the three way interaction effect was determined, an attempt was first made to determine if the normal order score approach could highlight the significant effects. However, this approach failed to provide much insight on the significant main and interaction effects as illustrated in Fig. 3a-b. In the normal order score approach, the effects that are random and normally distributed fall on a straight line when effects are plotted against the normal order score. In contrast, significant effects that are not randomly distributed fall off the straight line joining the random effects<sup>22</sup>. A straight line passing through the origin could not be fitted for the data illustrated in Fig. 3a-b. However, since each run was conducted in triplicate the significant and insignificant effects could be determined based on ANOVA. The ANOVA approach yields more insight since it reveals information contained in the replicates which is missing in the normal order score approach.

Multifactor ANOVA was performed using STATISTICA and the results are summarized in Table 2. The significance of main and interaction effects may be determined based on F-statistics and p-value. Statistically significant main effects of NP and Triton X-100 addition were found for both the measured response variables at 90% confidence level.

285 However, the main effect of bioaugmentation was significant for TPH degradation but  
 286 insignificant in case of microbial growth in suspension.



287

288

289 **Fig. 3 Normal plot of effects for the 2<sup>3</sup> factorial design with (a) oil degradation and**  
 290 **(b) culture growth in suspension over 30 days as response variables**

291

292 Addition of nutrients had significant effects on the extent of degradation and also on  
 293 microbial growth in the aqueous phase. Thus, the nutrients present in the baseline media  
 294 poses a limitation in bioremediation of oily sludge over 30 day duration. Interestingly,  
 295 most of the two way interactions other than that between bioaugmentation and nutrient  
 296 addition were insignificant while the three way interaction term between all the three  
 297 factors were significant. Thus, simultaneous addition of nutrients and surfactants along  
 298 with bioaugmentation with indigenous cultures may offer significant benefit in  
 299 bioremediation of this refinery sludge. Thus, complex interactions affect oil  
 300 biodegradation and culture growth in the aqueous phase in this oily sludge  
 301 bioremediation scenario. Various other researchers have utilized the analysis of variance  
 302 approach for demonstrating the impact of various treatments on extent of biodegradation  
 303 and microbial activity<sup>3,11,12,21</sup>.

304 From the ANOVA results some variation is found between the significant and insignificant  
 305 effects for the two response variables chosen, i.e., oil degradation and culture growth in



306 suspension. The main effect of bioaugmentation with microorganisms was to increase oil  
307 degradation and this effect was found to be significant.

308  
309  
310  
311

**Table 2. Significant and insignificant effects at 90% confidence level based on ANOVA**

Response	Factors	Significant		Factors	Insignificant	
		F-Statistics	P-value		F-Statistics	P-value
% Degradation	NP	10.869	0.005	NP*TX	1.677	0.214
	TX	9.896	0.006	TX*MO	0.012	0.914
	MO	4.227	0.056			
	NP*MO	3.225	0.091			
	NP*TX*MO	7.235	0.016			
Ln (N/N <sub>0</sub> )	NP	10.192	0.006	MO	2.442	0.138
	TX	11.853	0.003	NP*TX	1.062	0.318
	NP*MO	4.156	0.058	TX*MO	0.369	0.552
	NP*TX*MO	11.133	0.004			

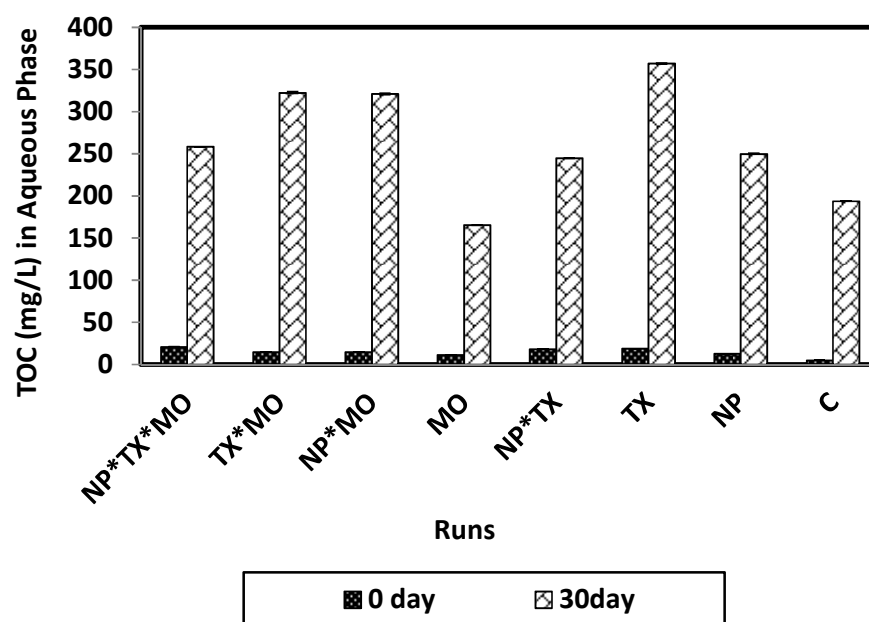
312 □ NP: nutrients, TX: Triton X 100, MO: microorganisms, N: Viable count of microorganisms at 30 day, N<sub>0</sub>: Viable count of  
313 microorganisms at 0 day

314  
315  
316  
317  
318  
319  
320  
321  
322  
323  
324  
325  
326

In contrast, the main effect of bioaugmentation on culture growth in suspension was to reduce culture growth. However this reduction was found to be insignificant based on the ANOVA results. Increase in oil degradation corresponding with negligible increase in culture growth in suspension may be because of increase in N<sub>0</sub> due to bioaugmentation causing nutrient deficiency. The enzymes produced by the microorganisms present at high concentration could have promoted higher oil degradation. It may also be due to different uptake mechanisms exhibited by the microbes responsible for oil degradation. In such sludges, oil mostly exists as sorbed oil rather than free phase oil. Microorganisms also associate with particulate matter in sludge and some sorbed hydrocarbons may have been utilized through direct interfacial uptake. Thus, viable count in the aqueous phase is not a true measure of microbial activity in the system. Degradation of oil is a better measure of the activity of oil degraders in this system although obtaining estimates on

327 degradation is more laborious and time consuming. Biostimulation with nutrients and  
328 surfactants was found to cause increase in growth of microorganisms in the aqueous  
329 phase. Surfactants are known to facilitate micellar solubilization of oil and emulsification  
330 of oil and is thus likely to cause desorption of oil. Moreover, surfactants are known to  
331 alter microbial cell surface hydrophobicity<sup>19,20</sup> such that distribution of microorganisms  
332 in the system may be altered. Simultaneous addition of nutrients possibly promoted oil  
333 degradation by microorganisms suspended in the aqueous phase thereby leading to higher  
334 culture growth in the aqueous phase.

335 Microbial activity in the aqueous phase led to change in color of the aqueous suspension  
336 after 30 days. This phenomenon may be indicative of oil/hydrocarbons leaching out over  
337 time or possibly due to accumulation of intermediates formed during microbial  
338 degradation of oil. While the oil/hydrocarbon content in the aqueous suspension across  
339 the various treatments was negligible both initially and after 30 days, the TOC in the  
340 aqueous phase was significantly higher after 30 days (Fig. 4). Increase in TOC was also  
341 observed for the un-inoculated controls. Increase in TOC suggests accumulation of  
342 soluble intermediates formed during biodegradation of oil associated with the sludge. In  
343 most cases increase in TOC per unit decrease in TPH was found to vary from 4% to 12%  
344 while it was found to be 30% for the un-inoculated controls. Thus, the bioaugmentation  
345 and biostimulation strategies used led to more complete biodegradation and  
346 comparatively lower accumulation of intermediates. Increase in TOC per unit decrease  
347 in TPH was least (4%) where all the three strategies (bioaugmentation, addition of  
348 nutrients and addition of surfactant) were employed simultaneously.



**Fig. 4 Variation in TOC in the aqueous phase for the various treatments in the oily sludge biodegradation study as per  $2^3$  design**

349

350

351

352

353

354

355

356

357

358

359

360

361

362

363

364

Various researchers have shown beneficial effects of bioaugmentation on degradation of oil associated with sludges and soil <sup>1,15,28</sup> as observed in this study. Bioaugmentation works best in scenarios where the indigenous population is very low <sup>3</sup>. In many scenarios, bioaugmented microorganisms have been found to be unable to adapt to the conditions prevailing in the contaminated environment. Bento *et al.*<sup>12</sup> demonstrated how presence of indigenous cultures in the soil limited the activity of bioaugmented microorganisms in a diesel contaminated soil system. They were unable to establish a correlation between TPH degradation, various treatments and activity of diesel degrading microorganisms since the indigenous microbial population degraded diesel more efficiently than the microbial consortium introduced during bioaugmentation. Tahhan *et al.*<sup>28</sup> also demonstrated that indigenous microorganisms could degrade hydrocarbons when oily sludge was added to soil without bioaugmentation. However, successive bioaugmentation with the enriched

365 indigenous consortium could improve the TPH removal rates. Alici *et al.*<sup>15</sup> reported that  
366 bioaugmentation with tailored microbial consortia could facilitate bioremediation of soil  
367 co-contaminated with diesel and heavy metals and up to 75% removal could be achieved  
368 in 42 days. Moriano *et al.*<sup>3</sup> reported that various amendments could enhance TPH  
369 removal to 45.5% over a period of 55 days; however, bioaugmentation with non-  
370 indigeneous cultures had no significant beneficial effect on TPH removal. Ayotamuno *et*  
371 *al.*<sup>29</sup> reported that bioaugmentation with extraneous microorganisms along with regular  
372 mixing and watering resulted in 63.7% - 84.5% reduction in TPH over a duration of six  
373 weeks. In general bioaugmentation is reported to be more successful in scenarios where  
374 the indigenous microorganisms native to the soil/sludge was enriched and added. Similar  
375 finding are revealed through our study. Low oil degradation (in run 1) due to depletion of  
376 the indigenous microorganisms originally present in the sludge during prolonged storage  
377 of the oily sludge could be partially overcome by bioaugmentation with indigenous  
378 microorganisms.

379 In the present study, addition of nutrients alone or in combination with bioaugmentation  
380 could enhance oil/TPH degradation significantly. Impact of nutrient addition on oil  
381 biodegradation is reported to vary widely and is possibly dependent on system specific  
382 conditions. Gallego *et al.*<sup>2</sup> reported significant enhancement such that upon addition of  
383 inorganic nitrogen and phosphorous up to 90% degradation of diesel was observed under  
384 laboratory conditions. In 12 week long laboratory studies on degradation of petroleum  
385 sludge and contaminated soil, Cvijovic *et al.*<sup>11</sup> demonstrated that addition of nutrients  
386 (NPK) offered greater beneficial effect compared to surfactant addition when culture  
387 growth in the aqueous phase was used as the response variable. Admon *et al.*<sup>34</sup> found that  
388 degradation of oily sludge contaminated soil occurred only after application of nutrients  
389 in the ratio of C:N:P = 50:10:1. Liu *et al.*<sup>35</sup> found that the addition of manure (as

390 nutrients) to oily sludge significantly increased the microbial activity and diversity;  
391 TPH in the treated sludge decreased by 58.2% over 365 days of bioremediation in  
392 comparison to only 15.6% in the control plot. Machin-Ramirez *et al.*<sup>36</sup> demonstrated  
393 that addition of commercial fertilizers enhanced the degradation of weathered oily sludge  
394 with removal of 24% TPH over a duration of 25 days. In contrast, Tahhan *et al.*<sup>28</sup>  
395 demonstrated inhibition of oily sludge biodegradation upon addition of nutrients possibly  
396 due to higher concentration of nitrogen and phosphorus already present in the sludge.

397 In addition to nutrients, TPH biodegradation is often limited due to hydrophobicity and  
398 low aqueous solubility of the constituent hydrocarbons. Surfactants may help in  
399 increasing the bioavailability of sorbed oil through micellar solubilization and  
400 emulsification. However, the use of surfactants in bioremediation experiments has been  
401 reported to both stimulate as well as inhibit hydrocarbon degradation influenced by  
402 various chemical properties of the surfactants and its interaction with  
403 microorganisms<sup>17,19,20,37</sup>. In the present study, the main effect of addition of surfactants  
404 was statistically significant both for microbial growth in suspension and oil degradation.

405 The binary interactions of surfactant addition and nutrient addition and that of surfactant  
406 addition and bioaugmentation were insignificant for both the response variables chosen.

407 The only significant binary interaction was that of nutrient addition and bioaugmentation  
408 with microorganisms, however it was only marginally significant ( $0.1 > p\text{-value} > 0.05$ ).

409 In contrast, the 3-way interaction between surfactant addition, nutrient addition and  
410 bioaugmentation was significant even at 98% confidence level for both the measured  
411 response variables. Thus, the effect of simultaneous addition of nutrients, surfactants and  
412 indigenous microorganisms provided a synergistic effect on both oil degradation and  
413 culture growth.

414 Although synergism/antagonism has not been specifically explored by other researchers,  
415 several studies have reported enhanced TPH degradation when surfactant addition is  
416 combined with other strategies. Cameotra and Singh<sup>38</sup> reported that the effects of  
417 addition of nutrients with bacterial consortia and crude biosurfactant with bacterial  
418 consortia resulted in less TPH degradation (91-95%) in comparison to when nutrients and  
419 surfactants were added together (98% TPH degradation). Rahman *et al.*<sup>21</sup> reported that  
420 addition of rhamnolipid biosurfactant increased the degradation of light n-alkanes in a  
421 scenario where soil mixed with oily sludge was treated. However, simultaneous  
422 supplementation with inorganic nutrients and rhamnolipid biosurfactant resulted in more  
423 complete degradation.

424

## 425 **5. Conclusion**

426 Addition of enriched indigenous microbes, addition of nutrients and addition of surfactant  
427 significantly enhanced oil degradation in sludge over and above that observed in the  
428 controls. Sludge biodegradation studies using 2<sup>3</sup> factorial design with oil degradation and  
429 microbial growth over 30 days as response variables revealed that the main effect of  
430 nutrient addition and surfactant addition enhanced oil degradation and culture growth in  
431 suspension and these effects were significant at 90% confidence level. However, the  
432 main effect of bioaugmentation only enhanced oil degradation but did not increase  
433 microbial growth in suspension. This may indicate the possible role of microorganisms  
434 attached on to sludge solids in degrading sorbed oil. The binary interactions are found to  
435 be insignificant except for that between nutrient addition and bioaugmentation which was  
436 found to have a synergistic effect. No other studies have revealed such a high ternary  
437 interaction effect between surfactant addition, nutrient addition and bioaugmentation  
438 indicating significant synergistic interaction among these strategies on oil degradation

439 and microbial growth. Thus, the effect of the various factors is not solely additive. The  
440 characteristics of the sludge, the characteristics and composition of the oil in sludge and  
441 the nature of the indigenous microorganisms present possibly affected the results.

442

443 **Acknowledgements:** The authors will like to acknowledge Mr A. D. Vyawahare,  
444 Manager, BPCL (Mumbai, India) for providing the sludge samples. Funding for this  
445 work was provided by IRCC, IIT Bombay.

446

447 **References:**

- 448 1 N. Vasudevan and P. Rajaram, Bioremediation of oily sludge contaminated soil,  
449 *Environ. Int.*, 2001, 26, 409-411.
- 450 2 J.L.R. Gallego, J. Loredó, J.F. Ilasmas, F. Vazquez and J. Sanchez, Bioremediation  
451 of diesel contaminated soils: Evaluation of potential in situ techniques by study of  
452 bacterial degradation, *Biodegradation*, 2001, 12, 325-335.
- 453 3 A.P. Moriano, A.P.de.A.G. Kataoka, D.de.F.de. Angelis and D.M. Bonotto,  
454 Laboratory study on the biodegradation of diesel oil contaminated soil from a petrol  
455 station, *Braz. J. Microbiol.*, 2007, 38, 346-353.
- 456 4 M.D. Ferrari, E. Neirotti, C. Albornoz, M.R. Mostazo and M. Cozzo, Biotreatment  
457 of hydrocarbons from petroleum tank bottom sludges in soil slurries, *Biotechnol.*  
458 *Lett.*, 1996, 18, 1241-1246.
- 459 5 M.S. Kuyukina, I.B. Ivshina, M.I. Ritchova, J.C. Philp, C.J. Cunningham and N.  
460 Christofi, Bioremediation of crude oil contaminated soil using slurry-phase  
461 biological treatment and landfarming techniques, *Soil Sediment Contam.*, 2003, 12,  
462 85-89.
- 463 6 I.V. Robles-Gonzalez, F. Fava and H.M. Poggi-Varaldo, A review on slurry  
464 bioreactors for bioremediation of soils and sediments, *Microb. Cell Fact.*, 2008, 7,  
465 1-16.
- 466 7 S. Mishra, J. Jyot, K.R. Chander and B. Lal, In situ bioremediation potential of a  
467 oily sludge –degrading bacterial consortium, *Curr. Microbiol.*, 2001, 43, 328-335.
- 468 8 S.E. Fantroussi and S.N. Agathos, Is bioaugmentation a feasible strategy for  
469 pollutant removal and site remediation?, *Curr. Opin. Microbiol.*, 2005, 8, 268-275.
- 470
- 471
- 472
- 473
- 474
- 475
- 476
- 477

- 478 9 J.L.R. Gallego, M.J. Garcia-Martinez, J.F. Llamas, C. Bellosch, A.I. Pelaez and J.  
479 Sanchez, Biodegradation of oil tank bottom sludge using microbial consortia,  
480 *Biodegradation*, 2007, 18, 269-281.  
481
- 482 10 R. Hosokawa, M. Nagai, M. Morikawa and H. Okuyama, Autochthonous  
483 bioaugmentation and its possible application to oil spills, *World J. Microbiol.*  
484 *Biotechnol.*, 2009, 25, 1519-1528.  
485
- 486 11 G.D. Cvijovic, J.S. Milic, T.M. Solevic, V.P. Beskoski, M.V. Ilic, L.S. Djokic,  
487 T.M. Narancic and M.M. Vrvic, Biodegradation of petroleum sludge and petroleum  
488 polluted soil by a bacterial consortium: a laboratory study, *Biodegradation*, 2012,  
489 23, 1-14.  
490
- 491 12 F.M. Bento, F.A.O. Camargo, B.C. Okeke and W.T. Frankenberger, Comparative  
492 bioremediation of soils contaminated with diesel oil by natural attenuation,  
493 biostimulation and bioaugmentation, *Bioresource Technol.*, 2005, 96, 1049-1055.  
494
- 495 13 P.W.G. Liu, L.M. Whang, T.C. Chang, I.C., Tseng, P.T. Pan and S.S. Cheng,  
496 Verification of necessity for bioaugmentation lessons from two batch case studies  
497 for bioremediation of diesel contaminated soils, *J. Chem. Technol. Biotechnol.*,  
498 2009, 84, 808-819.  
499
- 500 14 F.M. Ghazali, R.N.Z.A. Rahman, A.B. Salleh and M. Basri, Biodegradation of  
501 hydrocarbons in soil by microbial consortia, *Int. Biodeterior. Biodegrad.*, 2004, 54,  
502 61-67.  
503
- 504 15 C. Alici, R. Musella, F. Tasso, C. Ubaldi, S. Manzo, C. Creminini and A.R.  
505 Sprocati, Bioremediation of diesel oil in a co-contaminated soil by bioaugmentation  
506 with a microbial formula tailored with native strains selected for heavy metals  
507 resistance, *Sci.Total Environ.*, 2009, 407, 3024-3032.  
508
- 509 16 E.B. de. Moraes and S.M. Tauk-Tornisielo, Biodegradation of oil refinery residues  
510 using mixed culture of microorganisms isolated from a landfarming, *Braz. Arch.*  
511 *Biol. Technol.*, 2009, 52, 1571-1578.  
512
- 513 17 G. Mohanty and S. Mukherji, Effect of an emulsifying surfactant on diesel  
514 degradation by cultures exhibiting inducible cell surface hydrophobicity, *J. Chem*  
515 *Technol. Biotechnol.*, 2007, 82, 1004-1011.  
516
- 517 18 C.C. Lai, Y.C. Huang, Y.H. Wei and J.S. Chang, Biosurfactant –enhanced removal  
518 of total petroleum hydrocarbons from contaminated soils, *J. Hazard. Mater.*, 2009,  
519 167, 609-614.  
520
- 521 19 S. Mohanty and S. Mukherji, Alteration in cell surface properties of *Burkholderia*  
522 *spp.* during surfactant aided biodegradation of petroleum hydrocarbons, *Applied*  
523 *Microbiology and Biotechnology*, 2012, 94, 193-204.  
524
- 525 20 S. Mohanty and S. Mukherji, Surfactant aided biodegradation of NAPLs by  
526 *Burkholderia multivorans*: comparison between Triton X-100 and rhamnolipid  
527 JBR-515, *Colloids and Surfaces B: Biointerfaces.*, 2013, 102, 644-652.



- 528  
529 21 K.S.M. Rahman, T.J. Rahman, Y. Kourkoutas, I. Petsas, R. Marchant and I.M.  
530 Banat, Enhanced bioremediation of n-alkanes in petroleum sludge using bacterial  
531 consortium amended with rhamnolipid and micronutrients. *Bioresource Technol.*,  
532 2003, 90, 159-168.  
533  
534 22 P.M. Berthouex and L.C. Brown, *Statistics for Environmental Engineers*, 2nd ed.  
535 Lewis Publisher, Boca Raton, 2002, pp. 233–259.  
536  
537 23 J. Jasmine and S. Mukherji, Sustainable management of tank bottom sludge from  
538 refineries: role of bioaugmentation with n-alkane and naphthalene degrading  
539 Burkholderia cultures, *Int. J. Sustain. Innovat.*, 2013, 3 (1), 9-16.  
540  
541 24 N. Arvanitis, E. Katsifas, K.I. Chalkou, C. Meintanis and A.D. Karagouni, A  
542 refinery sludge deposition site: presence of nahH and alkJ genes and crude oil  
543 biodegradation ability of bacterial isolates, *Biotechnol. Lett.*, 2008, 30, 2105–2110.  
544  
545 25 O. Ward, A. Singh and J. Van Hamme, Accelerated biodegradation of petroleum  
546 hydrocarbon waste, *J. Ind. Microbiol. Biotechnol.*, 2003, 30, 260–270.  
547  
548 26 U.J.J. Ijah and S.P. Antai, Removal of Nigerian light crude oil in soil over a 12-  
549 month period, *Int. Biodeterior. Biodegrad.*, 2003, 51, 93–99.  
550  
551 27 S. Verma, R. Bhargava, and V. Pruthi, Oily sludge degradation by bacteria from  
552 Ankleshwar, India, *Int. Biodeterior. Biodegrad.*, 2006, 57, 207–213.  
553  
554 28 R.A. Tahhan, T.G. Ammari, S.J. Goussous and H.I. Al-shdaifat, Enhancing the  
555 biodegradation of total petroleum hydrocarbons in oily sludge by a modified  
556 bioaugmentation strategy, *Int. Biodeterior. Biodegrad.*, 2011, 65, 130-134.  
557  
558 29 M.J. Ayotamuno, R.N. Okparanma, E.K. Nweneka, S.O.T. Ogaji, S.D. Probert,  
559 Bio-remediation of a sludge containing hydrocarbons, *Appl. Energ.*, 2007, 84, 936-  
560 943.  
561  
562 30 N. Arvanitis, E. Katsifas, K.I. Chalkou, C. Meintanis and A.D. Karagouni, A  
563 refinery sludge deposition site: presence of nahH and alkJ genes and crude oil  
564 biodegradation ability of bacterial isolates, *Biotechnol. Lett.*, 2008, 30, 2105–2110.  
565  
566 31 O. Ward, A. Singh and J. Van Hamme, Accelerated biodegradation of petroleum  
567 hydrocarbon waste, *J. Ind. Microbiol. Biotechnol.*, 2003, 30, 260–270.  
568  
569 32 U.J.J. Ijah and S.P. Antai, Removal of Nigerian light crude oil in soil over a 12-  
570 month period, *Int. Biodeterior. Biodegrad.*, 2003, 51, 93–99.  
571  
572 33 S. Verma, R. Bhargava, and V. Pruthi, Oily sludge degradation by bacteria from  
573 Ankleshwar, India, *Int. Biodeterior. Biodegrad.*, 2006, 57, 207–213.  
574  
575 34 S. Admon, M. Green and Y. Avnimelech, Biodegradation kinetics of hydrocarbons in  
576 soil during land treatment of oily sludge, *Biorem. J.*, 2001, 5, 193-209.

- 577  
578  
579  
580  
581  
582  
583  
584  
585  
586  
587  
588  
589  
590  
591  
592  
593  
594  
595  
596  
597
- 35 W.X. Liu, Y.M. Luo, Y. Teng, Z.G. Li and L.Q. Ma, Bioremediation of oily sludge-contaminated soil by stimulating indigenous microbes, *Environ. Geochem. Hlth.*, 2010, 32, 23-29.
- 36 C. Machín-Ramirez, A.I. Okoh, D. Morales, K. Mayolo-Deloisa, R. Quintero, M.R. Trejo-Hernandez, Slurry-phase biodegradation of weathered oily sludge waste, *Chemosphere*, 2008, 70, 737-744.
- 37 S. Mohanty, J. Jasmine and S. Mukherji, "Practical considerations and challenges involved in surfactant enhanced bioremediation of Oil" *Biomedical Research International*, 2013, 2013, Article ID 328608, Doi: 10.1155/2013/328608.
- 38 S.S. Cameotra and P. Singh, Bioremediation of oil sludge using crude biosurfactants, *Int. Biodeter. Biodegrad.*, 2008, 62, 274-280.