

Metallomics

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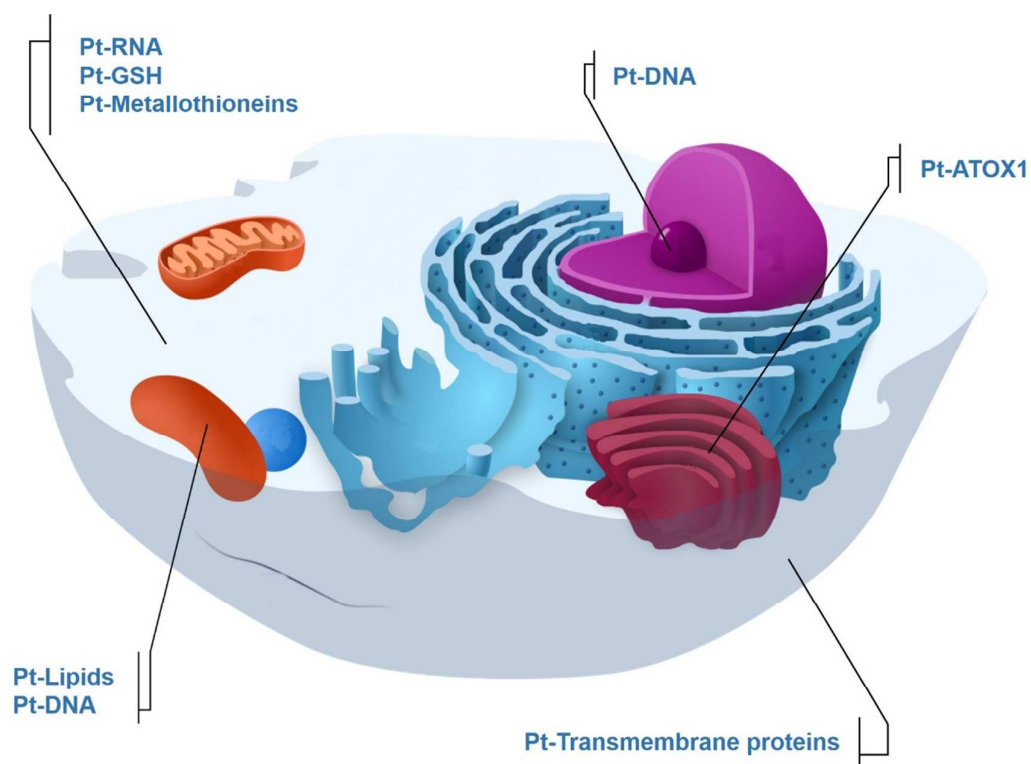


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The growing importance of biomarkers in platinum-based chemotherapy schemes are foreseen to play important role in medical decision-making. This mini review points out some targets for metalloomics to help them to reach this goal sooner.

Biomarkers to Assess the Efficiency of Treatment with Platinum-Based Drugs: What Can Metallomics Add to it?

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Abstract

Since the approval of cisplatin as an antineoplastic drug, the medical and the scientific communities are concerned about the side effects of platinum based drugs, which have been the dose limiting factor that leads to reduced treatment efficiency. Other important issue is the intrinsic or acquired resistance of some patients to treatment. Identifying proper biomarkers is crucial to evaluate the efficiency of the treatment, assisting physicians to determine, at early stages, whether the patient presents or not resistance to the drug, minimizing severe side effects, and allowing them to redirect the established course of chemotherapy. A great effort is being made to identify biomarkers that can be used to predict the outcome of the treatment of cancer patients with platinum-based drugs. In this context, the metallomic approach is not yet being used to its full potential. Since the basis of these drugs is platinum, the monitoring of biomarkers containing this metal should be the natural approach to evaluate the treatment progress. This review intends to show where the research in this field stands and points out some gaps that can be filled by metallomics.

Introduction

Since the late 1970's, platinum based drugs are being successfully used against a wide variety of tumours and they are still some of the most important agents for cancer treatment¹. The severe side effects concern the medical and scientific communities since the first trials and have been the dose limiting factor, which reduces treatment efficiency². Another important limitation of such treatment is the intrinsic or acquired resistance to these drugs. The mechanisms of such resistance have been reviewed recently by Galluzzi *et al.*³ and this is still a hot topic of scientific investigation^{3, 4, 5}. Once the treatment is not being effective or the prognostic is not favourable, there is no point to expose patients to the severe side effects caused by platinum based drugs^{6, 7}.

Biomarkers that can securely indicate the resistance of tumours or help to determine a prognostic to patients under treatment may play an important role, improving treatment efficiency through the adjustment of the chemotherapy strategy, and minimizing patients exposure to the drug side effects^{6, 8}. According to the National Cancer Institute, on its Dictionary of Cancer Terms⁹, a biomarker is defined as a “*biological molecule found in blood, other body fluids, or tissues that is a sign of a normal or abnormal process, or of a condition or disease. A biomarker may be used to see how well the body responds to a treatment for a disease or condition*”. However, the distance between the first attempts to identify a biomarker and its acceptance and use in the clinic is huge, and despite that, their scientific, social and economic potentials makes such endeavour worth⁸.

The importance of biomarkers can be inferred from the number of publications about the topic and by the growing number of specific databases condensing information about them, such as GOBIOM, BiomarkersBase, CancerDrive among others. A simple

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2
3 search for “cisplatin” at GVK BIO’s Online Biomarker Database (GOBIOM)¹⁰ returns
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5 around nine hundred molecules, however, the United States Food and Drug
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7 Administration (FDA) has approved only ten of them. Within this database, the
8
9 biomarkers are classified as biochemical, genomic, cellular, scoring scale, imaging, or
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11 physiological. These classes are related to the molecule characteristics and/or to the
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13 method applied to measure them. The biomarkers classified as the biochemical ones
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15 accounts for 59.7% of the outcome of the search, while the genomic ones represent
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17 29.3% of the total. Other classes of biomarkers are also represented: cellular (4.9%) and
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19 imaging (1.7%). Scoring scale and physiological types of biomarkers to access the
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21 effectiveness of platinum-based chemotherapy were not considered due to the scope of
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23 this review. The number of “under investigation” biomarkers reflects Drucker *et al.*
24
25 recent statement, that after the development of the “omics” the search for molecular
26
27 indicators increased substantially⁸. Most biochemical and genomic biomarkers were
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29 conceived using genomic or proteomic techniques. Metalloomics, as the newest *omic*
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31 approach, could be useful to identify biomarkers, especially to diagnose the action of
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33 metal-containing drugs, but, so far, the authors were not able to find any reported
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35 biomarker in clinical use identified through this approach.
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44 Many biochemical and genomic biomarkers have been suggested for the detection of
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46 different diseases, however, most of them is not specific and does not allow the
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48 identification of diseases in their earlier stages. Most of potential biomarkers did not
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50 jump from the laboratory stage to the clinical studies⁸. Despite that difficulty, the
51
52 research effort put on the identification of molecular predictors is huge. By
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54 understanding the biochemistry of platinum-based drugs in the cellular environment,
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56 their mechanisms of action or tumour resistance towards them, potential biomarkers can
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58 be foreseen and lead to reliable indications of the efficacy of treatment. In a near future,
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3 as the platinum pathway in human organism will be determined, other biological
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5 processes can be understood and reliable predicted. Platinum-containing biomarkers are
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8 post treatment indicators, which can bring information about individual's metabolism of
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10 the drug, making possible to evaluate and predict the outcome of the treatment.
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12 Platinum-based antineoplastic drugs allow the search for specific biomarkers to go into
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14 this direction, as platinum is a very rare element and can be detected in a sensitive way.
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16 In this work, selected biomarkers being used as clinical assistants in cancer prognostic,
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18 in the evaluation of the effectiveness of platinum chemotherapy or in the determination
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20 of the tumour resistance towards platinum-based drugs are reviewed. In addition, brief
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22 descriptions of the mechanism of action of these drugs, tumour resistance and the
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24 technologies used to investigate the biomarkers are presented, aiming to indicate the
25
26 potential of metalloomics in identifying reliable Pt-containing biomarkers.
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32 **Mechanisms of Platinum Based Drugs Activity**

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35 A complete comprehension of the mechanisms of action of platinum-based drugs has
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37 not yet been established, and conflicting data is observed in the literature¹¹. What seems
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39 as consensus is that the main cytotoxic mechanism of platinum-based drugs is the
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41 formation of covalent bond with the DNA, preventing cell division and inducing cell
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43 apoptosis. However, several other factors corroborate to their efficiency. Undoubtedly,
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45 cisplatin is the more studied drug of this group and serves as a reference in terms of
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47 activity and pharmacological principle to other drugs, thus its pharmacokinetics is
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49 briefly presented, and main differences among the drugs are highlighted.
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56 After application of cisplatin intravenously, the molecule undergoes hydrolysis due to
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58 the chloride concentration in the blood plasma and is carried by the blood in its original
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60 form. About 90% of cisplatin physically interacts with plasma proteins such as albumin,

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3 and the remainder is solubilized. Carboplatin is more stable than cisplatin in the blood;
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5 on the other hand oxaliplatin is hydrolysed even at high chloride concentrations. In the
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7 blood stream oxaliplatin interacts strongly with erythrocytes and blood proteins and
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9 only 12 % is available to enter other cells¹². Cisplatin is rapidly distributed in organs
10
11 and tissues and is found mainly in the liver and kidney¹³. The drug concentrations in
12
13 plasma rapidly decreases, with a half-life of ultrafilterable platinum in plasma ranging
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15 from 20 to 45 min. Approximately 25% of the drug is excreted in the urine over the first
16
17 24 h, up to 90% is eliminated by up to five days¹⁴.

22
23 Spatial distribution of platinum in lymphocytes of patients treated with cisplatin and in
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25 cultured cell lines are shown to be nearly the same: about 20% remain in the cell
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27 membrane; about 60% are found in the cytosolic fraction; 10% at the cytoskeleton and
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29 10% in the nucleus¹⁵. After crossing the cell membrane by passive or active transport¹⁶,
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31 cisplatin is in a medium with a chloride concentration of about 3 to 20 mmol L⁻¹ and
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33 undergoes hydrolysis converting into the most active forms of the drug³. Upon entering
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35 the cell, cisplatin interacts with the plasma membrane disrupting lipid-lipid and lipid-
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37 protein interactions¹⁷. It is believed that this interaction may lead to recruitment and
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39 activation of caspase 8 and, consequently, in the other caspase cascade leading to
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41 caspase-dependent cell apoptosis. Cisplatin in the cytoplasmic fraction interacts with a
42
43 number of nucleophilic species such as endogenous reduced glutathione (GSH),
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45 methionine, metallothionein, and other cytoplasmic proteins^{3,18,19,20}. The action of
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47 cisplatin in the cytoplasm has a depletive character over reduced species, providing a
48
49 favourable environment to oxidative stress, which enhance the action of the drug in the
50
51 cell nucleus³, at the endoplasmic reticulum and at the mitochondria^{17,21,22}. In the
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53 cytoplasmic fraction of the cell, cisplatin interacts with RNA, pledging to cell signalling
54
55 and gene expression²³. Considering what is known today, these are the major cytotoxic
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3 mechanisms of cisplatin. Most of other drug interactions in the cytoplasm are
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5 considered to be sources of resistance.
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9 In the cell nucleus, cisplatin binds to the DNA modifying its structure and preventing
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11 cell replication. About 1% of the total cisplatin absorbed by the cell is connected to the
12
13 nuclear DNA²⁴. Cisplatin interacts with DNA to form a series of adducts, preferably
14
15 linking the N7 position of the purine bases and can form double adducts with
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17 connections to a single strand or between the DNA strands. Cisplatin may also form
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19 adducts with DNA bases or DNA-protein adducts. If DNA damage is too extensive the
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21 cell progresses to apoptosis, the most important signalling pathway linking the DNA
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23 damage caused by cisplatin to apoptotic cell death involves the activation, in sequence,
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25 of a series of proteins in the nucleus and cytoplasm. The process starts by activating
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27 damage checkpoint 1 kinase (CHEK 1) proteins, and in turn phosphorylates the tumour
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29 suppressor protein p53^{25,26,27}. Once activated p53 triggers a number of lethal functions
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31 in the nucleus and cytoplasm, which lead to apoptosis of the cell^{28,22,29}.
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38 **Current biomarker investigation tools**

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42 In the last 20 years, the possibility of biomarkers investigation has immensely grown
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44 due to the advent of new technologies and tools made available to researchers. A myriad
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46 of approaches making use of these tools and the possible combinations of them are
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48 described on the literature. The advances in data treatment are also very important once
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50 scientists need to combine numerous results emerging from different sources in order to
51
52 extract the information about the responses to cancer treatment. These advances
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54 represent the basis for the implementation of new reliable indicators that will be used by
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56 physicians in a near future. Some of the main advances present in the literature are
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58 briefly discussed.
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At genomic field, the Next-Generation Sequencing (NGS) has been extensively used to identify somatic mutations and to determine the expression of many genes involved in crucial pathways such as EGFR and BRAF³⁰. It has also been used to determine the expression of KRAS gene, an relevant potential biomarker³¹. NGS platforms are based on sequencing-by-synthesis technology, with a DNA polymerase or ligase as the key component. Roche 454, Illumina, Helicos, and PacBio (Pacific Biosciences) use a DNA polymerase to drive their sequencing reaction, while SOLiD (Life Technologies) and Complete Genomics use a DNA ligase. The sequencing platforms can be further categorized as either single molecule-based (sequencing a single molecule) or ensemble-based (sequencing of multiple identical copies of a DNA molecule)³². Due to the huge amount of data produced with one single sequencing experiment (up to 200 million 100-nucleotides reads), careful experimental delimitation and a clear definition of aimed data are mandatory. Data treatment for identification of biomarkers through differential expression, for example, must be sought with the aid of specialized and validated software³².

Besides NGS, other powerful tools to investigate biomarkers are DNA microarrays. They enable high-throughput gene expression profiling through specific reaction of complementary DNA (cDNA) fixed on a rigid support with target RNA present in the sample³³. Through the overall RNA determination, the genic expression can be inferred. Many examples of the use of DNA microarray platform to study differences in genetic expressions, that could be used as biomarkers, are found in the literature for different malignancies such as lung adenocarcinoma³⁴, prostate cancer³⁵, among others, the approach was also used to evaluate DNA methylation changes³⁶.

Proteomic approach makes use a group of techniques that enable large-scale identification, characterization, and quantification of proteins in complex biological

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2
3 samples³⁷. Classical techniques making use immunoassays such Enzyme-Linked
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5 Immunosorbent Assay (ELISA) on research and are the most common approach in
6
7 clinical exams already prescribed by clinicians³⁸. Although ELISA is reliable and well
8
9 established, the need for high throughput and great specificity bring other techniques to
10
11 the spotlight. The analysis of proteome using 2D electrophoresis followed by mass
12
13 spectrometry (MS) is currently one of the most used approaches in comparative
14
15 proteomics. Samples are prepared in parallel and proteins are separated by 2D high
16
17 resolution electrophoresis, the different spots on the gel are usually characterised by
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19 Electro spray ionization MS (ESI-MS) or matrix-assisted laser desorption/ionisation
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21 (MALDI) time-of-flight (TOF) MS (MALDI-TOF) either in top-down or in bottom-up
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23 approach. MALDI-MS is the most common mass analyser employed due to its ability to
24
25 acquire peptide mass fingerprinting (PMF) with high throughput³⁹. PMF is a very useful
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27 approach for the rapid identification of a well-separated isolated protein. Tandem mass
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29 spectrometry (MS/MS) analysis exploits the fragmentation of selected precursor peptide
30
31 ions, showing greater confidence in protein identification³⁷.

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34 Protein microarrays are technical platforms for target proteomics based on quantitative
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36 protein expression focused on high throughput analysis. Similar to DNA microarrays
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38 these techniques count on a solid surface where hundreds to thousands of probing
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40 molecules are immobilized by robotic printing and the liquid sample is incubated on its
41
42 surface⁴⁰. The application of protein microarray to identify new biomarkers are
43
44 enormous and examples present in the literature account for the assessment of breast
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46 cancer recurrence, the evaluation of risk of developing bone metastasis from breast
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48 cancer⁴¹, as well as the evaluation of expression profile of colon cancer⁴², NSCLC⁴³,
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50 pancreatic cancer⁴⁴ and acute myelogenous leukemia⁴⁵.

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3 Regarding metallomics, the main characteristic of its approach is to be able to detect the
4 elements associated with biomolecules. This characteristic is, in general fulfilled by
5 inductively coupled plasma mass spectrometry (ICP-MS) hyphenated to a separating
6 technique such as HPLC. The separation techniques are employed to determine the
7 metallome because the detectors used are selective only for the differential elements,
8 (like Pt in the case of Pt based drugs). The target molecules must be separated prior to
9 the detection event. Electrophoresis has been used along with laser ablation (LA) in the
10 hyphenated technique LA-ICP-MS for the identification of metal containing molecules.
11 Most recently, LA-ICP-MS has been reported to be used in conjunction with protein
12 microarrays making possible to probe metal containing proteins from tissue lysates⁴⁶,
13 representing a huge step in metallomic development. The ability to probe a great
14 number of species with known activity is very promising for the investigation of
15 biomarkers and of biological processes in general.
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34 The association of metallomic techniques with proteomic allow information about the
35 general protein expression and which of them contain a specific element. In the imaging
36 field, Bianga *et al* used LA-ICP-MS and MALDI imaging to study the penetration and
37 distribution of two Pt-based metallodrugs (cisplatin and oxaliplatin) in human tumour
38 samples removed from patients diagnosed with colorectal or ovarian peritoneal
39 carcinomatosis⁴⁷.
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49 The extensive databases used both by genomics and proteomics require powerful
50 algorithms to assemble the pieces of the produced information, organise and manage
51 them in a systematic way so that useful information can be obtained. Bioinformatics is
52 fundamental for treating the huge amount of data produced by the techniques mentioned
53 above and to extract from them valid and reliable information. Great efforts are being
54 made on this field and databases such as Protein Data Base (PDB), Database of metal-
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3 binding sites in protein structures (MDB), Database of information on the catalytic
4 mechanisms of metal-dependent enzymes (Metal-MACiE), and Database of literature-
5 based annotation of metalloproteins (PROMISE)⁴⁸ are available. Software for data
6 treatment such as MITICS⁴⁹, OmniSpect⁵⁰, BioMap, MATLAB and Origin among
7 others are making possible to organize the data in a suitable way.
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16 It is important to point out that all the mentioned technologies are analytical tools, thus
17 standardization, validation and quality control are mandatory for good quality of
18 results⁵¹.
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23 24 **Gene expression based biomarkers**

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27 The gene expression control is of fundamental importance in the cell behaviour. This
28 becomes very clear if one considers that the whole genome is present in every cell, and
29 the differentiation among these cells is due to the genic expression. Which, when and
30 how much of a molecule will be expressed is dependent on a series of proteins and
31 enzymes that control DNA transcription⁵². All cancerous processes are related to
32 changes in cell genic expression. Many of the typical cancer cells behaviour, for
33 instance increased cell proliferation, insufficient apoptosis and cell differentiation, are
34 related to the expression of proteins and RNA that regulate these processes⁵³. The genic
35 expression biomarkers are the most common clinically in use and are being structured to
36 become a standardized practice^{54, 55, 56}. These biomarkers are the first to be used in the
37 development of the personalized medicine. Sentence commenting techniques employed
38 removed.
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57 TPMT gene encodes some enzymes involved in cisplatin metabolism, it presents some
58 single nucleotide polymorphism (SNP) that have been associated to a predisposition to
59 ototoxicity during or after cisplatin treatment⁵⁷. There still is some discussion on the
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3 literature whether this genetic marker is reliable⁵⁸, but this is the only biomarker cited in
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5 cisplatin label with recommendation and approval of FDA. The gene mutations are
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8 screened with *Illumina GoldenGate* assay and quantified by RT-PCR⁵⁷. Other genes are
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10 used in platinum-based therapy, but they are not specific for the patient response to
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12 treatment with this drug. Some important selected examples are shown below.
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16 A germline mutation in let-7 complementary site 6 (LCS6) within the untranslated
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18 region of the KRAS gene is known to be associated with poor outcome and drug
19
20 resistance in various cancers compared to the wild type allele⁵⁹. KRAS gene is a proto-
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22 oncogene and a single nucleotide substitution is responsible for activating mutation. The
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24 translated protein that results is implicated in various malignancies, including lung,
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26 mucinous adenoma, adenocarcinoma, ductal carcinoma of the pancreas and colorectal
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28 carcinoma⁶⁰. Besides, KRAS-variant is a potentially promising biomarker of poor
29
30 prognosis and a predictive biomarker of cisplatin resistance in head and neck squamous
31
32 cell carcinoma (HNSCC)⁵⁹, for Non-Small Cell Lung Cancer (NSCLC)⁶¹, among
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34 others. The gene variation is determined with a PCR-based assay, and validation is being
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36 pursued⁵⁹.
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42 RRM1 gene encodes one subunit, which constitutes ribonucleoside-diphosphate
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44 reductase, an essential enzyme for the production of deoxyribonucleotides prior to DNA
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46 synthesis in S phase of dividing cells. Lower expression levels of this gene, and SNP
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48 mutations are associated to a better response to cisplatin treatment in NSCLC^{62, 63}. The
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50 gene is screened and quantified by multiplex RT-qPCR. Significance of this gene
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52 expression as biomarker is usually associated with ERCC1 gene expression. Down
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54 regulation of the last is also associated with better treatment outcome and prognostic.
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56 Overexpression of ERCC1 is directly related to poorer prognostics and treatment
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58 outcome⁶⁴, expression of this is being measured with different approaches like in
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3 peripheral blood⁶⁵, tumour tissue, among other matrices, which makes its clinical use
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5 easier. Although there are problems with the expression measurement⁶⁶ this appears to
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7 be the most promising biomarker to enter clinical use⁶⁴.
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10 MiRNA are small non-protein-coding RNA molecules that play an important role in
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12 different biological processes, such as proliferation, differentiation and apoptosis by
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14 regulating gene expression. MiRNA-21 transcription is correlated with resistance to
15
16 cisplatin chemotherapy regimen: the lower the expression the better the outcomes. The
17
18 expression level of miR-21 in tumour tissue and plasma was indicated by Gao *et al.*⁶⁷ as
19
20 a biomarker to predict adjuvant platinum based chemotherapy response and disease free
21
22 survival in patients with NSCLC and also with oesophageal cancer⁶⁸. The miRNAs are
23
24 measured with specific microarray and RT-qPCR, and It has also been proposed as a
25
26 circulating biomarker able to provide prognostic, diagnosis and therapy progress⁶⁹.
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29 Besides the evaluation of individual genes, the genetic signatures (which make use of
30
31 statistics-based analytical tools) have been proposed as indicators of treatment outcome.
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33 Zhu *et al.* propose a 15-gene expression signature that is considered independent
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35 prognostic marker, which can predict patients most likely to benefit from adjuvant
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37 chemotherapy with cisplatin/vinorelbine⁷⁰. Kratz and co-workers established a
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39 quantitative-PCR-based assay based on 14 genes that, according to their findings, is
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41 able to identify patients with early-stage non-squamous NSCLC at high risk for
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43 mortality after surgical resection⁷¹.
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50 **Protein expression Biomarker**

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52 The genetic expression of a cell can be inferred by the pool of proteins found in it.
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54 Changes in the protein expression pattern are often observed when the cell is exposed to
55
56 some level of stress. Based on these principles, a *proteomic* approach can evaluate the
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3 behaviour of a tumour based on the comparative protein expression pattern of
4 healthy/cancerous cells or treated/non-treated cell (using, for instance, patient tissue or
5 blood). The protein expression is believed to be the next hot spot of research to clarify
6 the metabolism of platinum based drugs⁷². (Comments about techniques removed)
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13 Kuang *et al.* presented a proteomic approach making use of Western blot and real-time
14 PCR to identify possible biomarkers of efficiency for cisplatin-based treatment. The
15 group has identified eight differentially expressed proteins in one pair of cisplatin
16 sensitive/cisplatin resistant NSCLC cell lines. Special attention is given to DDH2
17 protein. This protein was investigated in serum of patients with NSCLC being treated
18 with cisplatin. They observed significantly different levels of the protein in the blood of
19 patients who presented disease progression, stability or amelioration⁷³.
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31 Fitzpatrick *et al.* used LC-MS to identify and quantify over 2000 proteins from two
32 pairs of cisplatin sensitive/cisplatin resistant cell lines. Among these, 760 proteins
33 showed significant expression changes. Based on the results, several potential pathways
34 that may be involved in cisplatin resistance in human ovarian cancer can be suggested.
35 This study provides a proteomic platform for large-scale quantitative protein analysis,
36 besides important information for investigation of new biomarkers of cisplatin
37 resistance in ovarian cancer⁷⁴.
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49 The efficacy of cisplatin treatment of tumour tissue from osteosarcoma has been
50 evaluated through protein expression profiling. A 2-D difference gel electrophoresis
51 was applied and 33 spots were found to differ significantly, allowing the classification
52 of patients in good or poor responders groups. These spots were later identified by
53 ESI-MS. Identification of the higher expression of peroxiredoxin 2 (PRDX2) in poor
54 responders was confirmed using Western blotting⁷⁵.
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3 The transmembrane 205 (TMEM205, previously known as MBC3205), a predicted
4 transmembrane protein, has shown expression profiles in normal human tissues
5 indicating a differential expression pattern with higher expression levels in the liver,
6 pancreas, and adrenal glands. Overexpression of TMEM205 in cells may be valuable as
7 a biomarker in cancer chemotherapy. Stable transfection of the TMEM205 gene confers
8 resistance to cisplatin by approximately 2.5 fold⁷⁶.
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18 Secreted proteome has been also evaluated as a biomarker. Such approach focus on the
19 possibility of collecting samples in the microenvironment of the tumour and/or around
20 the tumour. Sixteen proteins were shown to be differentially expressed by three
21 different epithelial ovarian carcinoma (EOC) lines, and the protein Collagen, type XI,
22 alpha 1 (COL11A1) was proposed as a biomarker for bad response to cisplatin
23 treatment. The authors evaluated the proteome of these cell lines by numerous
24 techniques⁷⁷.
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36 **Proposed biomarkers based on resistance mechanisms**

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39 The molecular basis of platinum-based drugs resistance can be described following their
40 molecular mode of action that involves several steps until the final response of the
41 tumour cell towards the drug. Problems to achieve DNA-damage response and
42 mitochondrial apoptosis^{78, 79} are the main factors impairing these drugs efficiency,
43 leading to poor treatment response. Herein, the goal is discuss, in the molecular level,
44 some points of the resistance mechanisms known so far in order to identify possible
45 biomarkers that may allow clinicians to predict and monitor clinical response to
46 platinum-based drugs chemotherapy. A better understanding of the mechanism by
47 which those molecules are regulated, as well as their roles in drug sensitivity and
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3 resistance, may indicate crucial prognostic markers and therapeutic targets for cancer
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5 treatment.
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9 Several molecules have been implicated in resistance to cisplatin, Methyl-CpG binding
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11 domain protein 1 (MBD1), which plays an important role in disease progression, is one
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13 of these. It is recruited to DNA damage sites under DNA damage conditions induced by
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15 cisplatin. Silencing of MBD1 significantly impaired activation of the DNA damage
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17 checkpoint response and inhibited DNA repair capacity⁸⁰. MBD1 binds mediator of
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19 DNA damage checkpoint protein 1 (MDC1), which is induced by radiation and
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21 regulates NBS1 activation in the presence of DNA damage repair⁸⁰.
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26 Another modulator of cisplatin activity in tumour recently described is Jab1 (a c-Jun
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28 coactivator), a multifunctional protein that participates in controlling cell proliferation
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30 and the stability of multiple proteins, plays an important role in the cellular response to
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32 cisplatin and irradiation by regulating DNA damage and repair pathways⁸¹. Jab1
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34 positively regulated Rad51 through p53-dependent pathway, and increased ectopic
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36 expression of Rad51 conferred cellular resistance to cisplatin, infrared light (IR) and
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38 UV radiation in Jab1-deficient cells. They showed that Jab1 is overexpressed in two
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40 relatively cisplatin-resistant, IR-resistant and UV-resistant nasopharyngeal carcinoma
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42 cells (NPC) cell lines.
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47 Furthermore, cisplatin activity can be modulate by pathways that, depending of the
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49 stimulus and cell type, enhance or reduce cisplatin efficiency. Activation of autophagy
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51 in the early stages of apoptosis, by BO-1051 (an N-mustard linked with a DNA-affinity
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53 molecule) acted as a defense system against cell death⁸². Inhibition of autophagy in its
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55 early or late stages resulted in an increase in the number of annexin V-positive cells.
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60 BO-1051-induced autophagy has a cytoprotective role and is connected to the ATM

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3 signaling pathway. This study revealed autophagy as a cytoprotective response against
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5 DNA damage-inducing chemotherapeutic agents, including BO-1051, cisplatin, and
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7 doxorubicin, in hepatocellular carcinoma cell lines⁸². On the other hand, autophagy is
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9 reported to enhance apoptosis induced by cisplatin. In lung cancer, treatment with
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11 cisplatin and radiation induced overexpression of autophagy-related genes, so as for the
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13 apoptosis signaling genes and a marked up-regulation of p21 expression, offering
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15 evidence that autophagy may enhance cisplatin efficiency⁸³.
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21 FOXO transcription factors, functioning downstream of the PI3K-PTEN-AKT (PKB)
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23 signaling cascade, are essential for cell proliferation, differentiation, DNA damage
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25 repair and apoptosis⁸⁴. Recent research indicates that the related transcription factor
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27 FOXM1 is a direct target of repression by FOXO proteins. Inactivation of FOXO or
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29 overexpression of FOXM1 is associated with tumourigenesis and cancer progression. In
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31 addition, the cytostatic and cytotoxic effects of a diverse spectrum of anti-cancer drugs,
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33 such as paclitaxel, doxorubicin, lapatinib, gefitinib, imatinib and cisplatin, are mediated
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35 through the activation of FOXO3a and/or the inhibition of its target FOXM1.
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37 Paradoxically, FOXO proteins also contribute to drug resistance by driving the
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39 expression of important genes for drug efflux as well as DNA repair and cell survival
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41 pathways in drug resistant cancers⁸⁴.
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48 One mechanism of cisplatin-resistant cells is through reduced intracellular platinum
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50 accumulation. This may result from reduced uptake, increased drug export or
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52 intracellular sequestration. While cisplatin uptake is mediated through the copper
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54 transporter protein Ctr1, efflux is performed by two other copper transporting p-type
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56 adenosine triphosphatases (ATP7A and ATP7B). Samimi *et al.* described that changes
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58 in the expression of these proteins implicated in cisplatin resistance and poor patient
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60 survival in some types of cancer, most notably ovarian cancer⁸⁵. The ATOX1 chaperone

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3 has been recently described as the transporter that brings Pt from CTR1 to ATP7B, the
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5 interaction of Pt with this protein appears to be γ -L-glutamyl-L-cysteinyl-glycine
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7 (reduced glutathione or GSH) competitive and stable ATOX1-Pt adducts have been
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9 observed⁸⁶.

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12 The cMOAT/MRP2, which is another important efflux system, have its expression
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14 increased in tumour cells and several works have shown that this membrane transporter
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16 may contribute to cisplatin resistance⁸⁷. MRP2 requires GSH as a cofactor and its role in
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18 protection from the cytotoxic effects of cisplatin may be a result of its ability to
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20 transport GSH/cisplatin conjugates across the cell membrane⁸⁸. GSH can work in
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22 concert with cMOAT/MRP2 to pump GSH-cisplatin conjugates out of cells in an ATP-
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24 dependent manner. Resistance to cisplatin through this mechanism is entirely GSH-
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26 dependent⁸⁹. Increased levels of intracellular GSH are frequently observed in cisplatin-
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28 resistant tumours⁹⁰.

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31 The capacity of cisplatin and other drugs to form bonds with thiol groups, make it
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33 suitable to form bonds with metallothioneins (MTs), which are intracellular proteins
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35 containing the highest amount of thiol groups within the cytoplasm. These thiol groups
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37 are able to bind several cytotoxic agents, such as platinum compounds. The increase of
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39 the level of MT is one mechanism of resistance to these anticancer drugs, once
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41 intracytoplasmic binding of MT prevents the active molecules from reaching their
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43 target, the intranuclear DNA of tumour cells^{91,92}.

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46 Thioredoxin reductase (TrxR) is the major cellular protein disulfide reductase
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48 containing selenium. TrxR catalyzes NADPH-dependent reduction of the redox-active
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50 disulfide in thioredoxin (Trx), the so-called thioredoxin reductase/thioredoxin system,
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52 which serves a wide range of functions in cell proliferation and redox homeostasis⁹³.

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3 TrxR is overexpressed in many cancer cells and TrxR and Trx can enhance tumour
4 development and drug resistance, thus been validated as therapeutical target in several
5 studies⁹⁴, furthermore, TrxR/Trx system has a strong impact on tumour resistance to
6 cisplatin⁹⁵. As shown several reports, the cisplatin-resistant variants exhibited an
7 increased expression and activity of TRXR as well as TRX compared with the parental
8 cells, additionally, the inhibition of the TrxR/Trx system restored cell sensitivity to
9 cisplatin^{96,97}.

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21 The excision repair cross-complementation group 1 (ERCC1) is a key component of the
22 platinum-DNA repair machinery responsible for nucleotide excision repair (NER). Its
23 expression protein were markedly higher in cisplatin-resistant derivatives of several
24 tumour cell lines^{98, 99, 100, 101, 102}. The action of ERCC1 generate excised single-stranded
25 DNA of approximately 30 nucleotides containing the Pt adduct and attached NER
26 proteins. DNA polymerases and ligases fill in the gap using the normal strand as a
27 template¹⁰³.

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38 Altogether, both overexpression of proteins that regulates DNA damage response and
39 apoptosis, so as the over activation of antioxidant mechanisms will converge to the
40 reduced efficiency of cisplatin. Thus, an important issue for metalloomics is how to
41 efficiently monitor the outcome of metal-based treatments, so that this approach can
42 positively improve patient survival rate.

50 51 **Platinum containing biomarkers**

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54 The determination of biomolecules through the platinum atom bind to them is a unique
55 characteristic of the metallomic approach in cancer research. This characteristic is
56 fundamental for the investigation of a new generation of cancer biomarkers, once it
57 allows the targeting of chemical species containing Pt that are products of the
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3 metabolism of the drug itself. Those species might provide information about processes
4 that each patient goes through during treatment, enabling physicians the ability to
5 perform personalized medicine. Although metallomics present such a potential, up to
6 now there are no biomarkers containing platinum approved by FDA or with clinical use
7 reported. Many studies making use of metallomics have been conducted to clarify the
8 pharmacokinetics of platinum-based drugs but none of the identified species have been
9 reported as biomarkers so far. The information acquired through metallomics, together
10 with information about protein content, drugs metabolites and genetic expression
11 obtained through the different approaches may provide comprehensive information on
12 the fate of the drug (target, metabolism and resistance).
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28 Once the interaction of Pt with DNA is believed to be responsible for the cytotoxicity Pt
29 based drugs, great effort have been put on the determination of Pt-DNA adducts. These
30 studies showed also that there are between 1 to 5 Pt atoms per 10^6 nucleotides. Zayed
31 and coworkers developed a very sensitive method for the determination of Pt-DNA
32 adducts by liquid chromatography (LC) coupled to sector field (SF) ICP-MS (LC-SF-
33 ICP-MS) that could be used for *in vivo* tests with a detection limit of $0,14 \text{ ng mL}^{-1}$ of Pt
34 ¹⁰⁴. Nevertheless, so far, these molecules have not been reported as biomarker for
35 resistance, effectiveness or prognostic of treatment outcome.
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48 Wexselblatt and coworkers has reviewed the action of platinum compounds in the cell
49 and the DNA adducts generation. They emphasize not only the complexity and dynamic
50 nature of cells that prevent us from monitoring the fate of platinum complexes in cells,
51 but also they showed the inability of the current analytical techniques to provide non-
52 invasively and in real time direct information on the speciation of the platinum
53 complexes in cells. (Part about techniques Removed)
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3 As a modulator of gene expression at multiple levels, RNA is an important potential
4 biomarker. Pt-RNA adducts have the potential to impact cell fate by disrupting RNA
5 regulatory pathways. Hostetter *et al.* used *Saccharomyces cerevisiae* for in-cell analysis
6 of Pt adduct formation on mRNA, rRNA, and total RNA and DNA platinum adducts²³.
7
8 The estimated in-cell Pt concentrations and Pt accumulation on mRNA, rRNA, total
9 RNA, and DNA were determined using ICP-MS. It was described that similar Pt
10 accumulation was observed on rRNA and total RNA, but significantly less Pt
11 accumulated on mRNA. By using the mapping by reverse transcription, they
12 demonstrated specific Pt adduct formation on rRNA sequences conserved between yeast
13 and humans. Taken together, these data highlight important differences in the relative
14 accumulation of Pt on different RNA species and provide insight into the accessibility
15 of cellular RNA to small, cationic molecules²³.
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32 Due to a thiol group presence, numerous electron rich sites and, especially, due to its
33 high abundance in every cell compartments GSH have been associated with resistance
34 to Pt based drugs since the early 1990's. There are many reports associating the
35 superexpression of GSH with resistance to platinum based drugs, Shoeib and Sharp
36 showed many possible isomers for Pt-GSH structures⁵. Although there is controversial
37 data about the binding of Pt to GSH many techniques to determine this biomolecule are
38 present in the literature. In Table 1, a sample of biomarkers currently approved by FDA,
39 in use or under investigation is listed.
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52 Cisplatin and oxaliplatin interact to a high extent with blood biomolecules, a small
53 fraction of these drugs remains free 24 h after administration and the levels of Pt in the
54 blood of patients treated with these drugs remain high for decades^{105,106}. On the other
55 hand, most of infused carboplatin is excreted intact, most of the drug incubated with rat
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3 ultra-filtrated plasma is recovered intact¹⁰⁷, adducts with proteins having molecular
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5 weights similar to human serum albumin (HAS) and g-globulin were reported^{108,109}.
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10 11 **Conclusions**

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16 Molecular biomarkers are the most mature and are already in use to evaluate patients'
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18 prognostic, treatment outcome and overall survival rate. These biomarkers have solid
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20 and validated sample preparation and analysis kits are available in the market, which
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22 makes its application easier, less expensive and widespread among clinical analysis
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24 laboratories.
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28 The proteomic approaches appear as a very promising field to be explored. The
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30 complexity of the interactions of platinum-based drugs with proteins (in the blood
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32 stream, entering the cell, being distributed and actually exerting its cytotoxic effect) is
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34 large and many authors recognize the lack of complete information about it^{72, 86, 111}.
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39 A new biomarker generation might be identified based on the metallomic approach. The
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41 ability of atomic spectrometric methods to detect the Pt atoms bind to a diversity of
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43 biomolecules, previously separated by chromatography, electrophoresis end even with
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45 microarrays, can help to fill some of the gaps in the comprehension of the mechanisms
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47 of action of platinum-based drugs. This ability can reveal molecular markers that can
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49 help physicians to take decisions and foresee the treatment outcome and patients
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51 prognostics. Platinum containing molecules known for a long time such as Pt-GSH and
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53 metallothioneins can bring information about availability of the active compound inside
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55 the cell. However, the specific studies for them to become clinical biomarkers are still to
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60 be done. Pt-DNA adducts can bring information about activity of ERCC1 and NER

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3 system, consequently about resistance to Pt treatment. In contrast, in all works revised
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5 in this text, the digestion of the DNA is performed prior to platinum adduct
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7 determination. In such experimental design, the biological information contained in the
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9 specific fragments generated by the NER is mixed with the other digested nucleotides,
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11 losing the biological signature of the origin of adducts. Pt-ATOX1 and Pt-RNA adducts
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13 apparently can clarify the fate of platinum-based drugs and indicate treatment outcome.
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15 Besides Pt-containing molecules, it was shown that sulphur containing ones, such as
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17 TrxR can play important role as biomarkers of resistance.
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23 Other metallo-biomarkers, not included in the present revision, can be sought by
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25 applying cutting edge metalloomics tools shown above to investigate specific cellular
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27 processes like drug efflux; repair of DNA damage; secreted Pt-containing molecules;
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29 mitochondrial induced apoptosis; outside the cell apoptosis signaling (membrane
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31 protein).
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35 Important barriers must be overcome to reduce the apparent distance between the
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37 metalloomic tools and the medical community. Once the increasing interaction of
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39 analytical scientists, biologists and medical doctors matures, the advances in the field
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41 will certainly occur. In addition, evolution in sample treatment, simplification of
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43 analytical process and instrumentation, proper analytical validation, dissemination of
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45 the approach with consequent cost reduction are also important steps to be taken to
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47 bring metalloomics to examination prescription.
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53
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57
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Table 1: Examples of biomarkers related to Pt-based drugs.

Biomarker	Determination	Use/drug	Cancer type	Reference
TPMT*	Enzyme assay and RT-PCR	Predictive (Ear injury)/cisplatin	Solid pediatric tumours.	55-58
ATP7A and ATP7B	Microarray	Resistance/Multidrug	Diverse Malignancies	85, 110
BO-1051	qRT-PCR, immunoassay	Resistance/cisplatin	Lung	82-83
cMOAT/MRP2	HPLC-ICP-MS, qRT-PCR	Resistance/ cisplatin	Nasopharyngeal, HNSCC	87-90
COL11A1	LC-MS/MS	Resistance/ cisplatin	Ovarian carcinoma	77
ERCC1	Microarray, qRT-PCR	Predictive (treatment outcome)/Multidrug	Lung, Bladder, HNSCC, Ovarian.	6, 63-64, 98-103
FOXO	ELISA; RT-PCR	Resistance/Cisplatin	Breast	84
Jab1	RPPA, qRT-PCR, immunoassay.	Resistance/cisplatin	Nasopharyngeal carcinoma	81
KRAS	RT-PCR	Treatment outcome/multidrug	Gastric, HNSCC, Lung, Pancreas	30-31, 59-61
MBD1	Immunoblot analysis	Resistance/multidrug	Pancreas	80
MiRNA-21	Specific microarray and qRT-PCR	Predict resistance/multidrug	Oesophageal, Lung	67-69
RRM1	qRT-PCR	Treatment outcome/cisplatin	bladder, lung	62-63
TMEM205	RT-PCR/ qRT-PCR	Resistance/cisplatin	Liver, pancreas, adrenal gland	76
TrxR	2D GE,MS HPLC-ICP-MS	Prognostic and Resistance/Cisplatin	Diverse malignancies	93-97
Possible Metallomic target biomarkers				
Metallothionein	2D PAGE; SEC-ICP-MS	Response indicator/multidrug	Diverse Malignancies	91-92
Pt-DNA Adduct	HPLC-ICP-MS	Many studies about these molecules/multidrug	Diverse Malignancies	15, 104
Pt-GSH	HPLC-ICP-MS	Resistance/multidrug	Diverse malignancies	5, 88-90
Pt-RNA Adduct	Microarray, GE-ICP-MS, HPLC-ICP-MS	Resistance/cisplatin	Diverse malignancies	<u>23, 32</u>

* This is the only biomarker approved by FDA specifically to cisplatin.

References

1. July 20–26, 2013. *The Lancet* 382. i, DOI: [http://dx.doi.org/10.1016/S0140-6736\(13\)61581-0](http://dx.doi.org/10.1016/S0140-6736(13)61581-0).
2. G. Cavaletti, G. Tredici, G. Pizzini, A. Minoia, Tissue platinum concentrations and cisplatin schedules. *The Lancet* 1990, 336. 1003, DOI: [http://dx.doi.org/10.1016/0140-6736\(90\)92462-Q](http://dx.doi.org/10.1016/0140-6736(90)92462-Q).
3. L. Galluzzi, L. Senovilla, I. Vitale, J. Michels, I. Martins, O. Kepp, M. Castedo, G. Kroemer, Molecular mechanisms of cisplatin resistance. *Oncogene* 2012, 31. 1869-83, DOI: 10.1038/onc.2011.384
4. G. T. a. I. Diaz-Padilla, in *OVARIAN CANCER - A CLINICAL AND TRANSLATIONAL UPDATE*, ed. I. Diaz-Padilla. In Tech: Crocia, 2013, pp 205-224.
5. T. Shoeib, B. L. Sharp, Interactions of oxaliplatin with the cytoplasmic thiol containing ligand glutathione. *Metallomics* 2012, 4. 1308-20, DOI: 10.1039/c2mt20127e.
6. K. A. Olaussen, A. Dunant, P. Fouret, E. Brambilla, F. Andre, V. Haddad, E. Taranchon, M. Filipits, R. Pirker, H. H. Popper, R. Stahel, L. Sabatier, J. P. Pignon, T. Tursz, T. Le Chevalier, J. C. Soria, DNA repair by ERCC1 in non-small-cell lung cancer and cisplatin-based adjuvant chemotherapy. *N Engl J Med* 2006, 355. 983-91, DOI: 355/10/983 [pii]
7. I. Díaz-Padilla, A. Poveda, DNA Repair–Based Mechanisms of Platinum Resistance in Epithelial Ovarian Cancer: From Bench to Bedside. *Clinical Ovarian Cancer* 2010, 3. 29-35, DOI: 10.3816/COC.2010.n.005.
8. E. Drucker, K. Krapfenbauer, Pitfalls and limitations in translation from biomarker discovery to clinical utility in predictive and personalised medicine. *EPMA J* 2013, 4. 7, DOI: 10.1186/1878-5085-4-7
9. <http://www.cancer.gov/dictionary?CdrID=45618>; accessed 07/17/2014.
10. <https://gobiomdb.com/gobiom/>; Accessed 04/17/2014.
11. A. V. Klein, T. W. Hambley, Platinum drug distribution in cancer cells and tumors. *Chem Rev* 2009, 109. 4911-20, DOI: 10.1021/cr9001066.
12. B. Desoize, C. Madoulet, Particular aspects of platinum compounds used at present in cancer treatment. *Crit Rev Oncol Hematol* 2002, 42. 317-25, DOI: S1040842801002190 [pii].

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59
60
13. P. J. O'Dwyer, J. P. Stevenson, S. W. Johnson, Clinical pharmacokinetics and administration of established platinum drugs. *Drugs* 2000, *59*. 19-27, Doi 10.2165/00003495-200059004-00003.
14. R. S. Go, A. A. Adjei, Review of the comparative pharmacology and clinical activity of cisplatin and carboplatin. *J Clin Oncol* 1999, *17*. 409-22.
15. A. Zayed, T. Shoeib, S. E. Taylor, G. D. D. Jones, A. L. Thomas, J. P. Wood, H. J. Reid, B. L. Sharp, Determination of Pt-DNA adducts and the sub-cellular distribution of Pt in human cancer cell lines and the leukocytes of cancer patients, following mono- or combination treatments, by inductively-coupled plasma mass spectrometry. *Int J Mass Spectrom* 2011, *307*. 70-78, DOI 10.1016/j.ijms.2010.11.012.
16. B. G. Blair, C. A. Larson, R. Safaei, S. B. Howell, Copper transporter 2 regulates the cellular accumulation and cytotoxicity of Cisplatin and Carboplatin. *Clin Cancer Res* 2009, *15*. 4312-21, DOI: 10.1158/1078-0432.CCR-09-0311
17. N. M. Martins, N. A. G. Santos, C. Curti, M. L. P. Bianchi, A. C. Santos, Cisplatin induces mitochondrial oxidative stress with resultant energetic metabolism impairment, membrane rigidification and apoptosis in rat liver. *Journal of Applied Toxicology* 2008, *28*. 337-344, DOI: 10.1002/jat.1284.
18. R. R. Barefoot, Speciation of platinum compounds: a review of recent applications in studies of platinum anticancer drugs. *J Chromatogr B Biomed Sci Appl* 2001, *751*. 205-11.
19. J. Calderón, D. Ortiz-Pérez, L. Yáñez, F. Díaz-Barriga, Human exposure to metals. Pathways of exposure, biomarkers of effect, and host factors. *Ecotoxicology and Environmental Safety* 2003, *56*. 93-103, DOI: 10.1016/s0147-6513(03)00053-8.
20. L. A. Finney, T. V. O'Halloran, Transition metal speciation in the cell: insights from the chemistry of metal ion receptors. *Science* 2003, *300*. 931-6, DOI: 10.1126/science.1085049
21. J. L. Podratz, A. M. Knight, L. E. Ta, N. P. Staff, J. M. Gass, K. Genelin, A. Schlattau, L. Lathroum, A. J. Windebank, Cisplatin induced Mitochondrial DNA damage in dorsal root ganglion neurons. *Neurobiology of Disease* 2011, *41*. 661-668, DOI: 10.1016/j.nbd.2010.11.017.
22. N. Pabla, Z. Dong, Cisplatin nephrotoxicity: Mechanisms and renoprotective strategies. *Kidney International* 2008, *73*. 994-1007, DOI: 10.1038/sj.ki.5002786.
23. A. A. Hostetter, M. F. Osborn, V. J. DeRose, RNA-Pt adducts following cisplatin treatment of *Saccharomyces cerevisiae*. *ACS Chem Biol* 2012, *7*. 218-25, DOI: 10.1021/cb200279p.
24. V. M. Gonzalez, M. A. Fuertes, C. Alonso, J. M. Perez, Is cisplatin-induced cell death always produced by apoptosis? *Mol Pharmacol* 2001, *59*. 657-63.

- 1
2
3 25. S. Y. Shieh, J. Ahn, K. Tamai, Y. Taya, C. Prives, The human homologs of
4 checkpoint kinases Chk1 and Cds1 (Chk2) phosphorylate p53 at multiple DNA damage-
5 inducible sites. *Genes Dev* 2000, *14*. 289-300.
6
7
8 26. E. Appella, C. W. Anderson, Post-translational modifications and activation of
9 p53 by genotoxic stresses. *Eur J Biochem* 2001, *268*. 2764-72, DOI: ejb2225 [pii].
10
11
12 27. H. Zhao, H. Piwnica-Worms, ATR-mediated checkpoint pathways regulate
13 phosphorylation and activation of human Chk1. *Mol Cell Biol* 2001, *21*. 4129-39, DOI:
14 10.1128/MCB.21.13.4129-4139.2001.
15
16
17 28. J. Wang, N. Pabla, C. Y. Wang, W. Wang, P. V. Schoenlein, Z. Dong, Caspase-
18 mediated cleavage of ATM during cisplatin-induced tubular cell apoptosis: inactivation
19 of its kinase activity toward p53. *Am J Physiol Renal Physiol* 2006, *291*. F1300-7, DOI:
20 00509.2005 [pii]
21
22
23 29. L. Galluzzi, E. Morselli, O. Kepp, I. Vitale, M. Pinti, G. Kroemer, Mitochondrial
24 liaisons of p53. *Antioxid Redox Signal* 2011, *15*. 1691-714, DOI:
25 10.1089/ars.2010.3504.
26
27
28 30. M. T. Lin, S. L. Mosier, M. Thiess, K. F. Beierl, M. Debeljak, L. H. Tseng, G.
29 Chen, S. Yegnasubramanian, H. Ho, L. Cope, S. J. Wheelan, C. D. Gocke, J. R.
30 Eshleman, Clinical validation of KRAS, BRAF, and EGFR mutation detection using
31 next-generation sequencing. *Am J Clin Pathol* 2014, *141*. 856-66, DOI:
32 10.1309/AJCPMWGWGO34EGOD
33
34
35 31. N. Kothari, M. J. Schell, J. K. Teer, T. Yeatman, D. Shibata, R. Kim,
36 Comparison of KRAS mutation analysis of colorectal cancer samples by standard
37 testing and next-generation sequencing. *J Clin Pathol* 2014. DOI: jclinpath-2014-
38 202405 [pii]
39
40
41 32. Y. Chu, D. R. Corey, RNA sequencing: platform selection, experimental design,
42 and data interpretation. *Nucleic Acid Ther* 2012, *22*. 271-4, DOI:
43 10.1089/nat.2012.0367.
44
45
46 33. M. Schena, D. Shalon, R. W. Davis, P. O. Brown, Quantitative monitoring of
47 gene expression patterns with a complementary DNA microarray. *Science* 1995, *270*.
48 467-70.
49
50
51 34. L. C. Lai, M. H. Tsai, P. C. Chen, L. H. Chen, J. H. Hsiao, S. K. Chen, T. P. Lu,
52 J. M. Lee, C. P. Hsu, C. K. Hsiao, E. Y. Chuang, SNP rs10248565 in HDAC9 as a
53 novel genomic aberration biomarker of lung adenocarcinoma in non-smoking women. *J*
54 *Biomed Sci* 2014, *21*. 24, DOI: 10.1186/1423-0127-21-24
55
56
57 35. P. Perot, V. Cheynet, M. Decaussin-Petrucci, G. Oriol, N. Mugnier, C.
58 Rodriguez-Lafrasse, A. Ruffion, F. Mallet, Microarray-based identification of
59 individual HERV loci expression: application to biomarker discovery in prostate cancer.
60 *J Vis Exp* 2013. e50713, DOI: 10.3791/50713.

- 1
2
3 36. C. J. Lee, J. Evans, K. Kim, H. Chae, S. Kim, Determining the effect of DNA
4 methylation on gene expression in cancer cells. *Methods Mol Biol* 2014, *1101*. 161-78,
5 DOI: 10.1007/978-1-62703-721-1_9.
6
7
8 37. B. Flatley, P. Malone, R. Cramer, MALDI mass spectrometry in prostate cancer
9 biomarker discovery. *Biochim Biophys Acta* 2014, *1844*. 940-9, DOI:
10 10.1016/j.bbapap.2013.06.015
11
12 38. R. Morgan, A. Boxall, A. Bhatt, M. Bailey, R. Hindley, S. Langley, H. C.
13 Whitaker, D. E. Neal, M. Ismail, H. Whitaker, N. Annels, A. Michael, H. Pandha,
14 Engrailed-2 (EN2): A Tumor Specific Urinary Biomarker for the Early Diagnosis of
15 Prostate Cancer. *Clinical Cancer Research* 2011, *17*. 1090-1098, DOI:10.1158/1078-
16 0432.Ccr-10-2410.
17
18
19 39. A. D. Weston, L. Hood, Systems biology, proteomics, and the future of health
20 care: toward predictive, preventative, and personalized medicine. *J Proteome Res* 2004,
21 *3*. 179-96.
22
23
24 40. D. A. Hall, J. Ptacek, M. Snyder, Protein microarray technology. *Mech Ageing*
25 *Dev* 2007, *128*. 161-7, DOI: S0047-6374(06)00254-5 [pii]
26
27 41. N. Hayashi, G. C. Manyam, A. M. Gonzalez-Angulo, N. Niikura, H. Yamauchi,
28 S. Nakamura, G. N. Hortobagyi, K. A. Baggerly, N. T. Ueno, Reverse-phase protein
29 array for prediction of patients at low risk of developing bone metastasis from breast
30 cancer. *Oncologist* 2014, *19*. 909-14, DOI: 10.1634/theoncologist.2014-0099
31
32 42. K. Malinowsky, U. Nitsche, K. P. Janssen, F. G. Bader, C. Spath, E. Drecoll, G.
33 Keller, H. Hofler, J. Slotta-Huspenina, K. F. Becker, Activation of the PI3K/AKT
34 pathway correlates with prognosis in stage II colon cancer. *Br J Cancer* 2014, *110*.
35 2081-9, DOI: 10.1038/bjc.2014.100
36
37 43. R. Ummanni, H. A. Mannsperger, J. Sonntag, M. Oswald, A. K. Sharma, R.
38 Konig, U. Korf, Evaluation of reverse phase protein array (RPPA)-based pathway-
39 activation profiling in 84 non-small cell lung cancer (NSCLC) cell lines as platform for
40 cancer proteomics and biomarker discovery. *Biochim Biophys Acta* 2014, *1844*. 950-9,
41 DOI: 10.1016/j.bbapap.2013.11.017
42
43 44. Y. J. Huang, M. L. Frazier, N. Zhang, Q. Liu, C. Wei, Reverse-phase protein
44 array analysis to identify biomarker proteins in human pancreatic cancer. *Dig Dis Sci*
45 2014, *59*. 968-75, DOI: 10.1007/s10620-013-2938-9.
46
47 45. S. M. Kornblau, A. Qutub, H. Yao, H. York, Y. H. Qiu, D. Graber, F. Ravandi,
48 J. Cortes, M. Andreeff, N. Zhang, K. R. Coombes, Proteomic profiling identifies
49 distinct protein patterns in acute myelogenous leukemia CD34+CD38- stem-like cells.
50 *PLoS One* 2013, *8*. e78453, DOI: 10.1371/journal.pone.0078453
51
52 46. L. Waentig, S. Techritz, N. Jakubowski, P. H. Roos, A multi-parametric
53 microarray for protein profiling: simultaneous analysis of 8 different cytochromes via
54 differentially element tagged antibodies and laser ablation ICP-MS. *Analyst* 2013, *138*.
55 6309-6315, DOI:10.1039/C3an00468f.
56
57
58
59
60

- 1
2
3 47. J. Bianga, A. Bouslimani, N. Bec, F. Quenet, S. Mounicou, J. Szpunar, B.
4 Bouyssiere, R. Lobinski, C. Larroque, Complementarity of MALDI and LA ICP mass
5 spectrometry for platinum anticancer imaging in human tumor. *Metallomics* 2014, 6.
6 1382-1386, DOI: 10.1039/C4mt00131a.
7
8
9
10 48. I. Bertini, G. Cavallaro, Bioinformatics in bioinorganic chemistry. *Metallomics*
11 2010, 2. 39-51, DOI: 10.1039/b912156k.
12
13 49. O. Jardin-Mathe, D. Bonnel, J. Franck, M. Wisztorski, E. Macagno, I. Fournier,
14 M. Salzert, MITICS (MALDI Imaging Team Imaging Computing System): a new open
15 source mass spectrometry imaging software. *J Proteomics* 2008, 71. 332-45, DOI:
16 10.1016/j.jprot.2008.07.004
17
18 50. R. M. Parry, A. S. Galhena, C. M. Gamage, R. V. Bennett, M. D. Wang, F. M.
19 Fernandez, omniSpect: an open MATLAB-based tool for visualization and analysis of
20 matrix-assisted laser desorption/ionization and desorption electrospray ionization mass
21 spectrometry images. *J Am Soc Mass Spectrom* 2013, 24. 646-9, DOI: 10.1007/s13361-
22 012-0572-y.
23
24 51. L. J. Kricka, S. R. Master, Validation and quality control of protein microarray-
25 based analytical methods. *Mol Biotechnol* 2008, 38. 19-31, DOI: 10.1007/s12033-007-
26 0066-5.
27
28 52. S. Clancy, DNA transcription. *Nature Education* 2008, 1.
29
30 53. W. A. SCHULZ, *MOLECULAR BIOLOGY OF HUMAN CANCERS*. 1 ed.;
31 Springer: Dordrecht, 2005.
32
33 54. V. A. Fusaro, C. Brownstein, W. Wolf, C. Clinton, S. Savage, K. D. Mandl, D.
34 Margulies, S. Manzi, Development of a scalable pharmacogenomic clinical decision
35 support service. *AMIA Jt Summits Transl Sci Proc* 2013, 2013. 60.
36
37 55. A. J. Thompson, W. G. Newman, R. A. Elliott, S. A. Roberts, K. Tricker, K.
38 Payne, The cost-effectiveness of a pharmacogenetic test: a trial-based evaluation of
39 TPMT genotyping for azathioprine. *Value Health* 2014, 17. 22-33, DOI:
40 10.1016/j.jval.2013.10.007
41
42 56. C. A. Brownstein, D. M. Margulies, S. F. Manzi, Misinterpretation of TPMT by
43 a DTC genetic testing company. *Clin Pharmacol Ther* 2014, 95. 598-600, DOI:
44 10.1038/clpt.2014.60
45
46 57. C. J. D. Ross, H. Katzov-Eckert, M.-P. Dubé, B. Brooks, S. R. Rassekh, A.
47 Barhdadi, Y. Feroz-Zada, H. Visscher, A. M. K. Brown, M. J. Rieder, P. C. Rogers, M.
48 S. Phillips, B. C. Carleton, M. R. Hayden, Genetic variants in TPMT and COMT are
49 associated with hearing loss in children receiving cisplatin chemotherapy. *Nature*
50 *Genetics* 2009, 41. 1345-1349, DOI: 10.1038/ng.478.
51
52 58. J. J. Yang, J. Y. Lim, J. Huang, J. Bass, J. Wu, C. Wang, J. Fang, E. Stewart, E.
53 H. Harstead, S. E. G. W. Robinson, W. E. Evans, A. Pappo, J. Zuo, M. V. Relling, A.
54
55
56
57
58
59
60

1
2
3 Onar-Thomas, A. Gajjar, C. F. Stewart, The role of inherited TPMT and COMT genetic
4 variation in cisplatin-induced ototoxicity in children with cancer. *Clin Pharmacol Ther*
5 2013, *94*. 252-9, DOI: 10.1038/clpt.2013.121
6

7
8 59. J. B. Weidhaas, J. W. Lee, R. Slebos, J. Howard, J. Perez, J. Gilbert, S. Nallur,
9 T. Paranjape, J. J. Garcia, B. Burtness, A. A. Forastiere, C. H. Chung, Association of
10 the 3'-untranslated region KRAS-variant with cisplatin resistance in patients with
11 recurrent and/or metastatic head and neck squamous cell carcinoma. *Journal of Clinical*
12 *Oncology* 2013, *31*.
13

14
15 60. C. Almoguera, D. Shibata, K. Forrester, J. Martin, N. Arnheim, M. Perucho,
16 Most human carcinomas of the exocrine pancreas contain mutant c-K-ras genes. *Cell*
17 1988, *53*. 549-54, DOI: 0092-8674(88)90571-5 [pii].
18

19
20 61. N. Karachaliou, C. Mayo, C. Costa, I. Magri, A. Gimenez-Capitan, M. A.
21 Molina-Vila, R. Rosell, KRAS Mutations in Lung Cancer. *Clin Lung Cancer* 2013, *14*.
22 205-214, DOI: 10.1016/j.clcc.2012.09.007.
23

24
25 62. G. Q. Wu, N. N. Liu, X. L. Xue, L. T. Cai, C. Zhang, Q. R. Qu, X. J. Yan,
26 Multiplex Real-time PCR for RRM1, XRCC1, TUBB3 and TS mRNA for Prediction of
27 Response of Non-small Cell Lung Cancer to Chemoradiotherapy. *Asian Pac J Cancer*
28 *Prev* 2014, *15*. 4153-8.
29

30
31 63. F. Mazzoni, F. L. Cecere, G. Meoni, C. Giuliani, L. Boni, A. Camerini, S.
32 Lucchesi, F. Martella, D. Amoroso, E. Lucherini, F. Torricelli, F. Di Costanzo, Phase II
33 trial of customized first line chemotherapy according to ERCC1 and RRM1 SNPs in
34 patients with advanced non-small-cell lung cancer. *Lung Cancer* 2013, *82*. 288-93,
35 DOI: 10.1016/j.lungcan.2013.08.018
36

37
38 64. K. K. Wei, L. Jiang, Y. Y. Wei, Y. F. Wang, X. K. Qian, Q. Dai, Q. L. Guan,
39 The prognostic value of ERCC1 expression in gastric cancer patients treated with
40 platinum-based chemotherapy: a meta-analysis. *Tumour Biol* 2014. DOI:
41 10.1007/s13277-014-2128-1.
42

43
44 65. M. Schena, S. Guarrera, L. Buffoni, A. Salvadori, F. Voglino, A. Allione, G.
45 Pecorari, E. Ruffini, P. Garzino-Demo, S. Bustreo, L. Consito, P. Bironzo, G. Matullo,
46 DNA repair gene expression level in peripheral blood and tumour tissue from non-small
47 cell lung cancer and head and neck squamous cell cancer patients. *DNA Repair* 2012,
48 *11*. 374-380, DOI: DOI 10.1016/j.dnarep.2012.01.003.
49

50
51 66. J. G. Schneider, N. Farhadfar, A. Sivapiragasam, M. Geller, S. Islam, E. Selbs,
52 Commercial laboratory testing of excision repair cross-complementation group 1
53 expression in non-small cell lung cancer. *Oncologist* 2014, *19*. 459-65, DOI:
54 10.1634/theoncologist.2013-0311
55

56
57 67. W. Gao, X. Lu, L. Liu, J. Xu, D. Feng, Y. Shu, MiRNA-21: a biomarker
58 predictive for platinum-based adjuvant chemotherapy response in patients with non-
59 small cell lung cancer. *Cancer Biol Ther* 2012, *13*. 330-40, DOI: 10.4161/cbt.19073
60

- 1
2
3 68. Y. Hu, A. M. Correa, A. Hoque, B. Guan, F. Ye, J. Huang, S. G. Swisher, T. T.
4 Wu, J. A. Ajani, X. C. Xu, Prognostic significance of differentially expressed miRNAs
5 in esophageal cancer. *Int J Cancer* 2011, *128*. 132-43, DOI: 10.1002/ijc.25330.
6
7
8 69. G. Cheng, Circulating miRNAs: Roles in cancer diagnosis, prognosis and
9 therapy. *Adv Drug Deliv Rev* 2014. DOI: S0169-409X(14)00199-9 [pii]
10
11 70. C. Q. Zhu, K. Ding, D. Strumpf, B. A. Weir, M. Meyerson, N. Pennell, R. K.
12 Thomas, K. Naoki, C. Ladd-Acosta, N. Liu, M. Pintilie, S. Der, L. Seymour, I. Jurisica,
13 F. A. Shepherd, M. S. Tsao, Prognostic and Predictive Gene Signature for Adjuvant
14 Chemotherapy in Resected Non-Small-Cell Lung Cancer. *Journal of Clinical Oncology*
15 2010, *28*. 4417-4424, DOI: 10.1200/jco.2009.26.4325.
16
17
18 71. J. R. Kratz, J. He, S. K. Van Den Eeden, Z. H. Zhu, W. Gao, P. T. Pham, M. S.
19 Mulvihill, F. Ziaei, H. Zhang, B. Su, X. Zhi, C. P. Quesenberry, L. A. Habel, Q. Deng,
20 Z. Wang, J. Zhou, H. Li, M. C. Huang, C. C. Yeh, M. R. Segal, M. R. Ray, K. D. Jones,
21 D. J. Raz, Z. Xu, T. M. Jahan, D. Berryman, B. He, M. J. Mann, D. M. Jablons, A
22 practical molecular assay to predict survival in resected non-squamous, non-small-cell
23 lung cancer: development and international validation studies. *Lancet* 2012, *379*. 823-
24 32, DOI: 10.1016/S0140-6736(11)61941-7
25
26
27 72. O. Pinato, C. Musetti, C. Sissi, Pt-based drugs: the spotlight will be on proteins.
28 *Metallomics* 2014, *6*. 380-95, DOI: 10.1039/c3mt00357d.
29
30
31 73. P. Kuang, C. Zhou, X. Li, S. Ren, B. Li, Y. Wang, J. Li, L. Tang, J. Zhang, Y.
32 Zhao, Proteomics-based identification of secreted protein dihydrodiol dehydrogenase 2
33 as a potential biomarker for predicting cisplatin efficacy in advanced NSCLC patients.
34 *Lung Cancer* 2012, *77*. 427-432, DOI: 10.1016/j.lungcan.2012.03.016.
35
36
37 74. D. P. G. Fitzpatrick, J.-S. You, K. G. Bemis, J.-P. Wery, J. R. Ludwig, M.
38 Wang, Searching for potential biomarkers of cisplatin resistance in human ovarian
39 cancer using a label-free LC/MS-based protein quantification method. *PROTEOMICS –*
40 *Clinical Applications* 2007, *1*. 246-263, DOI: 10.1002/prca.200600768.
41
42
43 75. D. Kubota, K. Mukaihara, A. Yoshida, H. Tsuda, A. Kawai, T. Kondo,
44 Proteomics study of open biopsy samples identifies peroxiredoxin 2 as a predictive
45 biomarker of response to induction chemotherapy in osteosarcoma. *J Proteomics* 2013,
46 *91*. 393-404, DOI: 10.1016/j.jprot.2013.07.022
47
48
49 76. D. W. Shen, J. Ma, M. Okabe, G. Zhang, D. Xia, M. M. Gottesman, Elevated
50 expression of TMEM205, a hypothetical membrane protein, is associated with cisplatin
51 resistance. *J Cell Physiol* 2010, *225*. 822-8, DOI: 10.1002/jcp.22287.
52
53
54 77. P. N. Teng, G. Wang, B. L. Hood, K. A. Conrads, C. A. Hamilton, G. L.
55 Maxwell, K. M. Darcy, T. P. Conrads, Identification of candidate circulating cisplatin-
56 resistant biomarkers from epithelial ovarian carcinoma cell secretomes. *Br J Cancer*
57 2014, *110*. 123-32, DOI: 10.1038/bjc.2013.687
58
59
60

- 1
2
3 78. E. R. Jamieson, M. P. Jacobson, C. M. Barnes, C. S. Chow, S. J. Lippard,
4 Structural and kinetic studies of a cisplatin-modified DNA icosamer binding to HMG1
5 domain B. *J Biol Chem* 1999, 274. 12346-54.
6
7
8 79. S. M. Cohen, S. J. Lippard, Cisplatin: from DNA damage to cancer
9 chemotherapy. *Prog Nucleic Acid Res Mol Biol* 2001, 67. 93-130.
10
11
12 80. J. Xu, W. Zhu, W. Xu, X. Cui, L. Chen, S. Ji, Y. Qin, W. Yao, L. Liu, C. Liu, J.
13 Long, M. Li, X. Yu, Silencing of MBD1 reverses pancreatic cancer therapy resistance
14 through inhibition of DNA damage repair. *Int J Oncol* 2013, 42. 2046-52, DOI:
15 10.3892/ijo.2013.1901.
16
17
18 81. Y. Pan, Q. Zhang, V. Atsaves, H. Yang, F. X. Claret, Suppression of Jab1/CSN5
19 induces radio- and chemo-sensitivity in nasopharyngeal carcinoma through changes to
20 the DNA damage and repair pathways. *Oncogene* 2013, 32. 2756-66, DOI:
21 10.1038/onc.2012.294
22
23
24 82. L. H. Chen, C. C. Loong, T. L. Su, Y. J. Lee, P. M. Chu, M. L. Tsai, P. H. Tsai,
25 P. H. Tu, C. W. Chi, H. C. Lee, S. H. Chiou, Autophagy inhibition enhances apoptosis
26 triggered by BO-1051, an N-mustard derivative, and involves the ATM signaling
27 pathway. *Biochem Pharmacol* 2011, 81. 594-605, DOI: 10.1016/j.bcp.2010.12.011
28
29
30 83. M. Liu, S. Ma, Y. Hou, B. Liang, X. Su, X. Liu, Synergistic killing of lung
31 cancer cells by cisplatin and radiation via autophagy and apoptosis. *Oncol Lett* 2014, 7.
32 1903-1910, DOI: 10.3892/ol.2014.2049
33
34
35 84. J. M. Kwok, B. Peck, L. J. Monteiro, H. D. Schwenen, J. Millour, R. C.
36 Coombes, S. S. Myatt, E. W. Lam, FOXM1 confers acquired cisplatin resistance in
37 breast cancer cells. *Mol Cancer Res* 2010, 8. 24-34, DOI: 10.1158/1541-7786.MCR-09-
38 0432
39
40 85. G. Samimi, N. M. Varki, S. Wilczynski, R. Safaei, D. S. Alberts, S. B. Howell,
41 Increase in expression of the copper transporter ATP7A during platinum drug-based
42 treatment is associated with poor survival in ovarian cancer patients. *Clin Cancer Res*
43 2003, 9. 5853-9.
44
45
46 86. A. Galliani, M. Losacco, A. Lasorsa, G. Natile, F. Arnesano, Cisplatin handover
47 between copper transporters: the effect of reducing agents. *J Biol Inorg Chem* 2014, 19.
48 705-14, DOI: 10.1007/s00775-014-1138-1.
49
50
51 87. S. M. Xie, W. Y. Fang, T. F. Liu, K. T. Yao, X. Y. Zhong, Association of
52 ABCC2 and CDDP-Resistance in Two Sublines Resistant to CDDP Derived from a
53 Human Nasopharyngeal Carcinoma Cell Line. *J Oncol* 2010, 2010. 915046, DOI:
54 10.1155/2010/915046.
55
56
57 88. M. Tonigold, A. Rossmann, M. Meinold, M. Bette, M. Marken, K. Henkenius,
58 A. C. Bretz, G. Giel, C. Cai, F. R. Rodepeter, V. Benes, R. Grenman, T. E. Carey, H.
59 Lage, T. Stiewe, A. Neubauer, J. A. Werner, C. Brendel, R. Mandic, A cisplatin-
60 resistant head and neck cancer cell line with cytoplasmic p53 exhibits ATP-binding

1
2
3 cassette transporter upregulation and high glutathione levels. *J Cancer Res Clin Oncol*
4 2014. DOI: 10.1007/s00432-014-1727-y.
5
6

7 89. E. S. Arner, H. Nakamura, T. Sasada, J. Yodoi, A. Holmgren, G. Spyrou,
8 Analysis of the inhibition of mammalian thioredoxin, thioredoxin reductase, and
9 glutaredoxin by cis-diamminedichloroplatinum (II) and its major metabolite, the
10 glutathione-platinum complex. *Free Radic Biol Med* 2001, 31. 1170-8, DOI:
11 S0891584901006980 [pii].
12
13

14 90. S. Singh, T. Okamura, F. Ali-Osman, Serine phosphorylation of glutathione S-
15 transferase P1 (GSTP1) by PKC α enhances GSTP1-dependent cisplatin metabolism
16 and resistance in human glioma cells. *Biochem Pharmacol* 2010, 80. 1343-55, DOI:
17 10.1016/j.bcp.2010.07.019
18
19

20 91. J. Gumulec, M. Raudenska, V. Adam, R. Kizek, M. Masarik, Metallothionein -
21 immunohistochemical cancer biomarker: a meta-analysis. *PLoS One* 2014, 9. e85346,
22 DOI: 10.1371/journal.pone.0085346
23
24

25 92. J. Gumulec, J. Balvan, M. Sztalmachova, M. Raudenska, V. Dvorakova, L.
26 Knopfova, H. Polanska, K. Hudcova, B. Ruttkay-Nedecky, P. Babula, V. Adam, R.
27 Kizek, M. Stiborova, M. Masarik, Cisplatin-resistant prostate cancer model: Differences
28 in antioxidant system, apoptosis and cell cycle. *Int J Oncol* 2014, 44. 923-33, DOI:
29 10.3892/ijo.2013.2223.
30
31

32 93. G. Powis, D. L. Kirkpatrick, Thioredoxin signaling as a target for cancer
33 therapy. *Curr Opin Pharmacol* 2007, 7. 392-7, DOI: S1471-4892(07)00091-4 [pii]
34

35 94. S. Li, J. Zhang, J. Li, D. Chen, M. Matteucci, J. Curd, J. X. Duan, Inhibition of
36 both thioredoxin reductase and glutathione reductase may contribute to the anticancer
37 mechanism of TH-302. *Biol Trace Elem Res* 2010, 136. 294-301, DOI:
38 10.1007/s12011-009-8544-1.
39
40

41 95. B. Zhou, J. Huang, Y. Zuo, B. Li, Q. Guo, B. Cui, W. Shao, J. Du, X. Bu, 2a, a
42 novel curcumin analog, sensitizes cisplatin-resistant A549 cells to cisplatin by inhibiting
43 thioredoxin reductase concomitant oxidative stress damage. *Eur J Pharmacol* 2013,
44 707. 130-9, DOI: 10.1016/j.ejphar.2013.03.014
45
46

47 96. T. Sasada, H. Nakamura, S. Ueda, N. Sato, Y. Kitaoka, Y. Gon, A. Takabayashi,
48 G. Spyrou, A. Holmgren, J. Yodoi, Possible involvement of thioredoxin reductase as
49 well as thioredoxin in cellular sensitivity to cis-diamminedichloroplatinum (II). *Free*
50 *Radic Biol Med* 1999, 27. 504-14, DOI: S0891-5849(99)00101-X [pii].
51
52

53 97. A. B. Witte, K. Anestal, E. Jerremalm, H. Ehrsson, E. S. Arner, Inhibition of
54 thioredoxin reductase but not of glutathione reductase by the major classes of alkylating
55 and platinum-containing anticancer compounds. *Free Radic Biol Med* 2005, 39. 696-
56 703, DOI: S0891-5849(05)00231-5 [pii]
57
58

59 98. M. Z. Muallem, S. Marnitz, R. Richter, C. Kohler, J. Sehouli, R. Arsenic,
60 ERCC1 expression as a predictive marker of cervical cancer treated with cisplatin-based
chemoradiation. *Anticancer Res* 2014, 34. 401-6, DOI: 34/1/401 [pii].

- 1
2
3 99. M. Z. Muallem, I. Braicu, M. Nassir, R. Richter, J. Sehouli, R. Arsenic, ERCC1
4 expression as a predictor of resistance to platinum-based chemotherapy in primary
5 ovarian cancer. *Anticancer Res* 2014, *34*. 393-9, DOI: 34/1/393 [pii].
6
7
8
9 100. O. Ozdemir, P. Ozdemir, A. Veral, H. Uluer, M. H. Ozhan, ERCC1 expression
10 does not predict survival and treatment response in advanced stage non-small cell lung
11 cancer cases treated with platinum based chemotherapy. *Asian Pac J Cancer Prev* 2013,
12 *14*. 4679-83.
13
14 101. J. E. Bauman, M. C. Austin, R. Schmidt, B. F. Kurland, A. Vaezi, D. N. Hayes,
15 E. Mendez, U. Parvathaneni, X. Chai, S. Sampath, R. G. Martins, ERCC1 is a
16 prognostic biomarker in locally advanced head and neck cancer: results from a
17 randomised, phase II trial. *Br J Cancer* 2013, *109*. 2096-105, DOI:
18 10.1038/bjc.2013.576
19
20 102. Y. Torii, R. Kato, Y. Minami, K. Hasegawa, T. Fujii, Y. Udagawa, ERCC1
21 expression and chemosensitivity in uterine cervical adenocarcinoma cells. *Anticancer*
22 *Res* 2014, *34*. 107-15, DOI: 34/1/107 [pii].
23
24
25 103. E. C. Friedberg, How nucleotide excision repair protects against cancer. *Nat Rev*
26 *Cancer* 2001, *1*. 22-33, DOI: 10.1038/35094000.
27
28
29 104. A. Zayed, G. D. Jones, H. J. Reid, T. Shoeib, S. E. Taylor, A. L. Thomas, J. P.
30 Wood, B. L. Sharp, Speciation of oxaliplatin adducts with DNA nucleotides.
31 *Metallomics* 2011, *3*. 991-1000, DOI: 10.1039/c1mt00041a.
32
33
34 105. J. Szpunar, A. Makarov, T. Pieper, B. K. Keppler, R. Lobinski, Investigation of
35 metallo-drug-protein interactions by size-exclusion chromatography coupled with
36 inductively coupled plasma mass spectrometry (ICP-MS). *Anal. Chim. Acta* 1999, *387*.
37 135-144, DOI:10.1016/S0003-2670(99)00074-4.
38
39
40 106. P. Allain, O. Heudi, A. Cailleux, A. Le Bouil, F. Larra, M. Boisdron-Celle, E.
41 Gamelin, Early biotransformations of oxaliplatin after its intravenous administration to
42 cancer patients. *Drug Metab Dispos* 2000, *28*. 1379-84.
43
44
45 107. P. Guo, S. Li, J. M. Gallo, Determination of carboplatin in plasma and tumor by
46 high-performance liquid chromatography-mass spectrometry. *J Chromatogr B Analyt*
47 *Technol Biomed Life Sci* 2003, *783*. 43-52, DOI: S1570023202004890 [pii].
48
49
50 108. R. Xie, W. Johnson, L. Rodriguez, M. Gounder, G. S. Hall, B. Buckley, A study
51 of the interactions between carboplatin and blood plasma proteins using size exclusion
52 chromatography coupled to inductively coupled plasma mass spectrometry. *Anal*
53 *Bioanal Chem* 2007, *387*. 2815-22, DOI: 10.1007/s00216-007-1147-9.
54
55
56 109. M. Sooriyaarachchi, A. Narendran, J. Gailer, Comparative hydrolysis and
57 plasma protein binding of cis-platin and carboplatin in human plasma in vitro.
58 *Metallomics* 2011, *3*. 49-55, DOI: 10.1039/c0mt00058b.
59
60

1
2
3 110. S. Owatari, S. Akune, M. Komatsu, R. Ikeda, S. D. Firth, X. F. Che, M.
4 Yamamoto, K. Tsujikawa, M. Kitazono, T. Ishizawa, T. Takeuchi, T. Aikou, J. F.
5 Mercer, S. Akiyama, T. Furukawa, Copper-transporting P-type ATPase, ATP7A,
6 confers multidrug resistance and its expression is related to resistance to SN-38 in
7 clinical colon cancer. *Cancer Res* 2007, 67. 4860-8, DOI: 67/10/4860 [pii]
8
9

10 111. E. Wexselblatt, E. Yavin, D. Gibson, Cellular interactions of platinum drugs.
11 *Inorg Chim Acta* 2012, 393. 75-83, DOI 10.1016/j.ica.2012.07.013.
12
13
14
15
16
17
18
19
20
21
22
23
24
25
26
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