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1 **Modification of lignin with dodecyl glycidyl ether and chlorosulfonic acid**
2 **for preparation of anionic surfactant**

3 Chang-Zhou Chen ^a, Ming-Fei Li ^{a,*}, Yu-Ying Wu ^a, Run-Cang Sun ^{a,b,*}

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5 ^a Beijing Key Laboratory of Lignocellulosic Chemistry, Beijing Forestry University, Beijing 100083,
6 China;

7 ^b State Key Laboratory of Pulp and Paper Engineering, South China University of Technology,
8 Guangzhou 510640, China

9 * Author to whom correspondence should be addressed:

10 Ming-Fei Li (Tel. +8610 62336592; E-mail address: limingfei@bjfu.edu.cn)

11 Run-Cang Sun (Tel. /Fax: +8610 62336972; E-mail address: rcsun3@bjfu.edu.cn)

12

13 **Abstract** Lignin was modified through incorporation of lipophilic and hydrophilic groups
14 for the preparation of surfactant. In this case, alkaline lignin reacted with dodecyl glycidyl
15 ether in the presence of dimethyl benzyl amine to incorporate lipophilic long alkyl chains,
16 and then sulfonated with chlorosulfonic acid for the introduction of hydrophilic
17 sulfonic acid group. Results showed that the reaction between dodecyl glycidyl ether and
18 carboxy group in lignin was the predominant reaction at 95 °C-110 °C. It was found that
19 the surface tension of the synthesized lignin surfactant solution was lower than that of
20 commercial surfactant sodium dodecylbenzenesulphonate when concentration was below
21 0.4 %, indicating that the surfactant prepared from alkaline lignin had a good surface
22 activity. A lowest critical micelle concentration of 0.50 g L⁻¹ and the corresponding surface
23 tension at 29.17 mN/m were achieved when the surfactant was derived from the lignin
24 grafted with dodecyl glycidyl ether at 110 °C. The anionic lignin surfactants prepared in
25 this study is a promising feedstock as detergents or to enhance oil recovery.

26

27

28 1. Introduction

29 The use of sustainable resources for energy and advanced eco-friendly material has
30 drawn much attention due to the increased consumption of fossil resources associated with
31 energy crisis and climate change.^{1,2} Lignocellulose, mainly composed of cellulose,
32 hemicellulose and lignin, is a promising and renewable resource for biofuel, chemicals,
33 and biomaterials without competition with human's food.³ There are some conundrums of
34 the development of bio-ethanol for making biomass-to-biofuels economic⁴, but it has great
35 potential economic benefits to develop new materials and chemicals derived from
36 lignocellulose. After cellulose and hemicelluloses, lignin is the third most abundant
37 biopolymer from biomass and considered as the most abundant aromatic renewable
38 resource. Lignin mainly consists of *p*-hydroxyphenyl (H), guaiacyl (G), and syringyl (S)
39 units derived from three major monolignols, namely, *p*-coumaryl alcohol, coniferyl
40 alcohol, and sinapyl alcohol. These units are connected by various inter-unit linkages (e.g.,
41 β -*O*-4', β -5', β - β ', 5-5', 5-*O*-4', and β -1', etc.) through an enzyme-initiated dehydrogenative
42 polymerization^{5,6} Lignin can be isolated from wood, annual plants (such as wheat straw)
43 and agricultural residues (such as corncobs) by different processes.^{7,8} However, lignin is
44 considered as low-value and underutilized by-product from pulping and bio-refinery
45 industries. Most of lignin fraction resulted from these processes is burned as fuel, and only
46 a small portion of lignin is converted to commercial products. The effective utilization of
47 lignin to produce value-added products through chemical modification will make the
48 biomass industry economically feasible with respect to the replacement of chemicals and
49 materials derived from fossil sources. Lignin is an excellent candidate for chemical
50 reactions due to its highly functional characteristics (rich in phenolic and aliphatic
51 hydroxyl groups) for the development of new chemicals.

52 Surfactants are extremely important chemicals, and numberless types of surfactants
53 are used for a myriad of applications, thus the development of lignin surfactant has a good
54 industrial prospect. Lignin itself is not very useful as a surfactant because of the nature
55 character of lignin, for example, the complex macromolecular structure, the large
56 molecular weight, the low hydrophilicity and lipophilicity.^{9,10} To work effectively, most

57 surfactant require water soluble and oil soluble characteristics. Lignin must be changed
58 into the fraction which has the hydrophilic or amphiphilic property. To date, the
59 introduction of hydrophilic groups into lignin by sulfonation^{11, 12}, carboxylation^{13, 14}, and
60 Mannich reaction¹⁵ has been investigated by many researchers. However, these methods
61 can only enhance hydrophilicity but to lipophilicity of lignin, and the surface activity of
62 these lignin surfactants is not satisfactory. In fact, a good lignin surfactant requires water
63 soluble and oil soluble characteristics by the incorporation of both lipophilic and
64 hydrophilic groups. *Morrow et al.*¹⁶ have described a method of producing surfactants for
65 enhancing oil recovery from lignin by two reactions of lignin, i.e., alkylation with
66 different alkyl halides at phenolic sites, sufficient oxidation to break the lignin into smaller
67 polymeric and monomeric units, and sulfonation to increase their water solubility. *Naae et*
68 *al.*¹⁷ reduced lignin in the presence of CO and H₂, the resulted products were alkoxyated
69 with α -olefin epoxide introducing a long alkyl chain, and these compounds were changed
70 into oil soluble lignin surfactant by sulfonation, sulfation and alkoxy-sulfation. *Zeng et al.*
71¹⁸ have reported that hydrophilic sulfonic group and lipophilic adbietic group were
72 introduced into the molecular structure of Kraft lignin, and the rosin-modified
73 lignosulfonate lowered the surface tension of a 10 g L⁻¹ aqueous solution to 49.3 mN/m.

74 The aim of the present study was to prepare lignin surfactant by simultaneous
75 incorporation of lipophilic and hydrophilic groups into lignin. Alkaline lignin was
76 modified with dodecyl glycidyl ether in the presence of dimethyl benzyl amine as a
77 catalyst, and then was converted into anionic surfactant by sulfonation with
78 chlorosulfonic acid. The structure of lignin after reaction with dodecyl glycidyl ether was
79 examined by FT-IR, GPC, ³¹P-NMR and ¹³C-NMR, and the surface activity of the lignin
80 surfactant was investigated through determining the surface tension of the aqueous
81 solution.

82

83 **2. Experimental**

84 **2.1. Materials**

85 Alkaline lignin (AL) from corncob was obtained from Shandong Longlive
86 Bio-technology Co., Ltd, China. Lauryl alcohol, epichlorohydrin (ECH), tetrabutyl

87 ammonium bromide (TBAB), N,N-dimethylbenzylamine (BDMA), sodium dodecyl
88 benzene sulfonate (SDBS) and dimethyl sulfoxide (DMSO) were purchased from
89 Sinopharm Chemical Reagent Beijing Co., Ltd, China. Chlorosulfonic acid (99%) was
90 provided by Xiya reagent (Chengdu, China).

91 **2.2. Preparation of dodecyl glycidyl ether (DGE)**

92 To a vigorously stirred mixture containing 37.2 g (0.2 mol) of lauryl alcohol, 200 mL
93 of toluene, 50.0 g of 48% aqueous sodium hydroxide solution and 3.2 g (0.01 mol) of
94 TBAB, 37.0 g (0.4 mol) of ECH were added dropwise for 15 min at ambient temperature,
95 and the reaction was run for 6 h at 50 °C. When the resulting solution was cooled to room
96 temperature, the organic layer was separated and washed 4 times with distilled water (200
97 mL, 60 °C). Then the organic layer was evaporated at 60 °C under reduced pressure to
98 obtain DGE. The epoxy value of DGE was 0.33 mol epoxy / 100 g DGE, determined
99 according to the method described by Hou.¹⁹

100 **2.3. Reaction of AL with DGE**

101 A 250 mL round-bottom flask containing 8.0 g lignin, 1.8 g BDMA, 8.0 g DGE and
102 160 mL DMSO was heated for 5.0 h at various temperatures (95-110 °C). The reaction
103 was quenched by adding 20 mL of 2.0 M HCl under stirring for 30 min at room
104 temperature. The solvent was removed by rotary evaporation, and the residue was washed
105 with distilled water and petroleum ether for several times, and then dried overnight at
106 60 °C to give grafted lignin, named as DGE-AL.

107 **2.4. Sulfonation of DGE-AL**

108 The sulfonation of DGE-AL was conducted according to the procedure in a previous
109 report with minor modifications.¹¹ A total of 2.0 g chlorosulfonic acid in 20 mL anhydrous
110 dichloromethane (CH₂Cl₂) was added dropwise into 1.0 g of DGE-AL in 10 mL
111 dichloromethane. The mixture was mechanically stirred for 2 h in an ice bath. Next, 30
112 mL of 2 M NaOH was added and the mixture was stirred for 1 h in an ice bath, and then it
113 was adjusted with 1 M HCl to pH 8.0. The resulting mixture was dialyzed with a cellulose
114 tube (cut off Mw-3000) against water and lyophilized to yield sulfonated DGE-AL, named
115 as SD-AL.

116

117 **2.5. Sample characterization**

118 FT-IR spectra were recorded in a Thermo Scientific Nicolet iN10 FT-IR Microscope
119 (Thermo Nicolet Corporation, Madison, WI, USA) equipped with a liquid nitrogen cooled
120 MCT detector. The spectra were collected in the range of 4000-650 cm^{-1} at 4 cm^{-1}
121 resolution. NMR spectra were recorded on a Bruker AVIII 400 MHz spectrometer at 25 °C.
122 For ^{13}C -NMR spectra, 80 mg samples were dissolved in 1 mL of $\text{DMSO-}d_6$. The sample
123 was modified following the procedure below for ^{31}P -NMR determination. Approximate 30
124 mg of oven-dried sample was dissolved in pyridine/chloroform (1.6:1, v/v) in a small vial
125 and stirred continuously. N-hydroxyl naphthalimide was used as internal standard, and
126 chromium acetylacetonate as the relaxation reagent. Then,
127 2-chloro-4,4,5,5-tetramethyl-1,3,2-dioxaphosphalane (TMDP) was used for
128 phosphorylation of hydroxyl groups. The molecular weight of the sample was determined
129 by gel permeation chromatography (GPC). The lignin sample was dissolved in
130 tetrahydrofuran (2 mg/mL), and then the solution was analyzed on a high performance
131 liquid chromatography system (Agilent 1200 series, Agilent technologies, USA) with a
132 DAD detector. The molecular weights were calibrated via monodisperse polystyrene
133 standards. The sulfur content of the lignin surfactant was examined by elemental analysis
134 using a Vario EL III Elemental analyzer instrument (Elementar, Germany) according to a
135 previous report.²⁰

136 **2.6. Measurement of surface tension**

137 An automatic surface tensiometer (JK99B, Shanghai Zhongchen Digital Technic
138 Apparatus Co., Ltd) was used to measure the surface tension of the surfactant solution at
139 different concentrations at room temperature (18 °C). Deionized water (surface tension
140 74.950 mN/m) was employed for reference, and commercial surfactant SDBS was used as
141 a control. Critical micelle concentration (CMC) values of lignin surfactants were
142 identified from the plot of the surface tension versus the surfactant concentration of water
143 solution.

144

145

146 **3. Results and discussion**

147 **3.1. Synthesis and characterization of DGE-AL**

148 Typically, Lewis bases are used to accelerate the reaction between epoxides and the
 149 hydroxyls of phenols, alcohols and carboxyl.²¹ In this study, BDMA was chosen as a
 150 catalyst of the reaction between DGE and lignin, and DGE was introduced into lignin
 151 through lignin hydroxyl-epoxide reaction (see Scheme 1). Because of the low
 152 hydrophilicity and lipophilicity, lignin had a low solubility in neutral water and most of
 153 organic solvents. For a homogeneous reaction between lignin and DGE, DMSO was
 154 employed as a solvent for the reaction system. In order to avoid self-polymerization of
 155 DGE in the presence of tertiary amine²², the reaction temperature was set at 95, 100 and
 156 110 °C. After reaction, hydrochloric acid was used to deactivate the remnant DGE, and the
 157 inactivated DGE and DGE homopolymer were washed off with petroleum ether. As seen
 158 from Table 1, the DGE-AL yields were as high as 62.68-65.25%, and the molecular
 159 weights of the products derived from the reaction conducted at 95, 100, and 110 °C were
 160 1300, 1410, and 2050 g mol⁻¹, respectively.

161

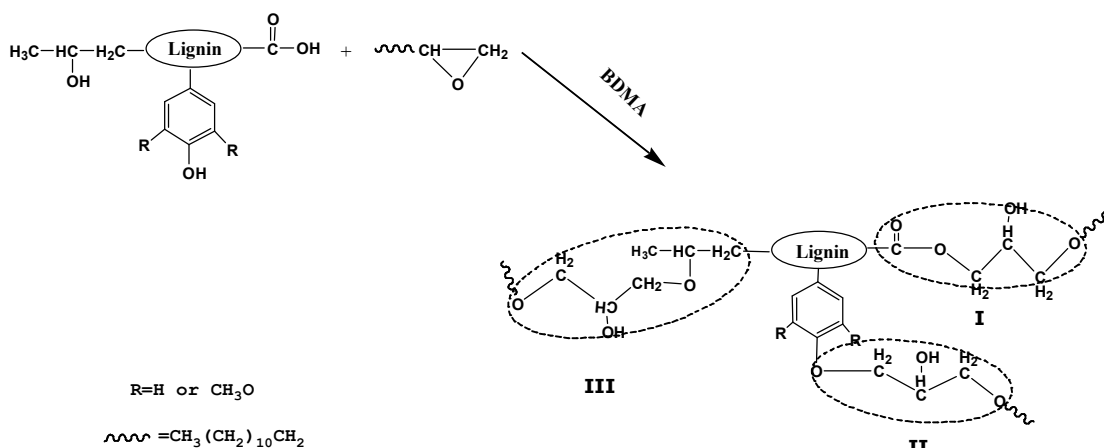
162 Table 1 The yields, weight average (M_w) and number average (M_n) molecular weight of
 163 the products from the reaction of DGE with alkaline lignin

Samples ^a	Yield (%) ^b	M_w (g mol ⁻¹)	M_n (g mol ⁻¹)	M_w/M_n
DGE-AL 1	65.25	2050	720	2.85
DGE-AL 2	64.31	1410	600	2.37
DGE-AL 3	62.68	1300	580	2.23
AL	-	850	540	1.59

164 ^a AL, raw material alkaline lignin; DGE-AL 1, DGE-AL 2 and DGE-AL 3 were obtained
 165 by the reaction of DGE with lignin in the presence of BDMA at 110, 100 and 95 °C for 5 h,
 166 respectively.

167 ^b the yield of DGE-AL was based on the total weight of reactant.

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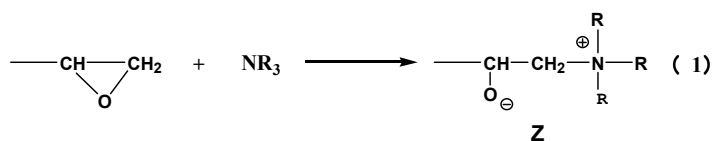


169

170 Scheme 1 The reaction of DGE with lignin in the presence of BDMA.

171

172 ³¹P-NMR technique has been widely applied in classification and quantitative
 173 determination of several different classes of hydroxyl groups in the native and modified
 174 lignin.²³⁻²⁵ In this study, phosphitylating agent TMDP was utilized to obtain the qualitative
 175 and quantitative information about hydroxyl groups in lignin before and after graft. The
 176 ³¹P-NMR spectra of the samples are illustrated in Figure 1 and the contents of different
 177 hydroxyl groups are listed in Table 2. For DGE-AL 1 and DGE-AL 3, the content of total
 178 OH was lower than that of AL, indicating the successful introduction of DGE into lignin
 179 in the presence of BDMA. As compared to DGE-AL 3, the dramatic reduction of the
 180 carboxylic OH content in DGE-AL 1 suggested that the reaction between epoxide and
 181 carboxyl was the predominant reaction when reaction temperature increased from 95 °C to
 182 110 °C. This can be explained by the mechanism of glycidyl ether reactions with alcohols,
 183 phenols and carboxylic acids in the presence of tertiary amine as a catalyst.



184

185 From Equation 1, a zwitterion (Z) was formed by the interaction between tertiary amine
 186 and epoxide which acted as an efficient catalyst due to its high basicity.²¹ Because of the
 187 relatively high acidity of carboxyl in lignin, the reaction between carboxyl and glycidyl
 188 ether occurred first. As see from Fig. 1, the chemical shifts of 150.0-145.4 ppm,
 189 144.5-137.0 ppm, and 136.0-133.6 ppm are assigned to aliphatic, phenolic, and carboxylic

190 acid OH, respectively.²⁶ With respect to the reaction mechanism of hydroxyl-glycidyl
 191 ether reactions, the following hypothesis was made: some new hydroxyl groups were
 192 derived from the open-ring reaction of epoxy group of DGE. Accordingly, three new kinds
 193 of signals were assigned at the chemical shifts of aliphatic OH, derived from the reactions
 194 of DGE with aliphatic, phenolic, and carboxylic acid OH in lignin. This was supported by
 195 the presence of three new peaks (147.9-147.4, 147.1-146.8, and 146.6-145.8 ppm) in the
 196 spectra of DGE-AL 1 and DGE-AL 3, which correspond to the chemical shifts of aliphatic
 197 OH. From Fig. 1, the signals at 146.6-145.8 ppm varied more noticeably as compared to
 198 those at 147.9-147.4 ppm and 147.1-146.8 ppm probably due to the large reduction of the
 199 content of carboxylic OH in DGE-AL 1 as compared to DGE-AL 3 (Table 2). The signals
 200 at 146.6-145.8 ppm were probably due to the hydroxyls derived from carboxyl-glycidyl
 201 ether reaction, whereas the absorptions at 147.9-147.4 ppm and 147.1-146.8 ppm probably
 202 correspond to the hydroxyl groups derived from the alcohol and phenol-glycidyl ether
 203 reactions, respectively.

204

205 Table 2 Quantification of hydroxyl content of alkaline lignin before and after reaction with
 206 DGE by quantitative ³¹P-NMR method

Samples ^a	Hydroxyl content (mmol g ⁻¹ of lignin)						Total phenol OH	Total OH
	Aliphatic OH	Syringyl OH	Guaiacyl OH	<i>p</i> -Hydroxy phenyl OH	Carboxylic OH			
AL	2.26	0.84	1.67	1.02	2.24	1.52	8.03	
DGE-AL 1	2.77	0.64	1.16	0.55	0.39	1.04	5.51	
DGE-AL 3	2.70	0.78	1.32	0.40	0.88	1.20	6.27	

207 ^a AL, DGE-AL 1 and DGE-AL 3 corresponding to the lignin samples in Table 1.

208

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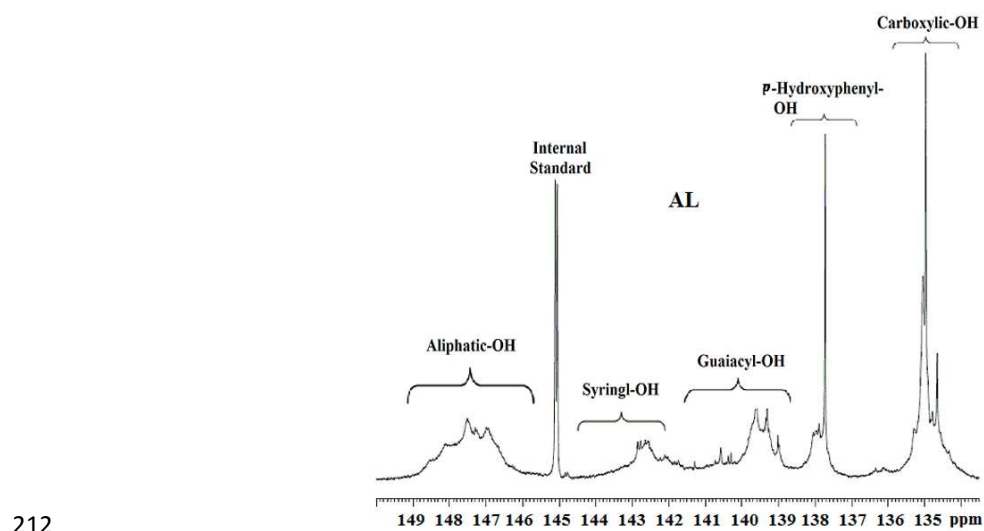
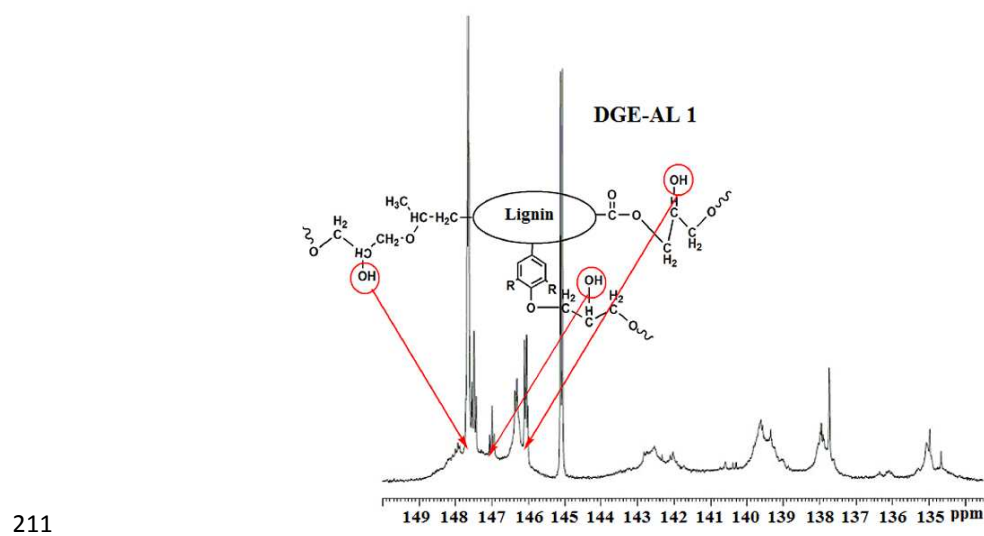
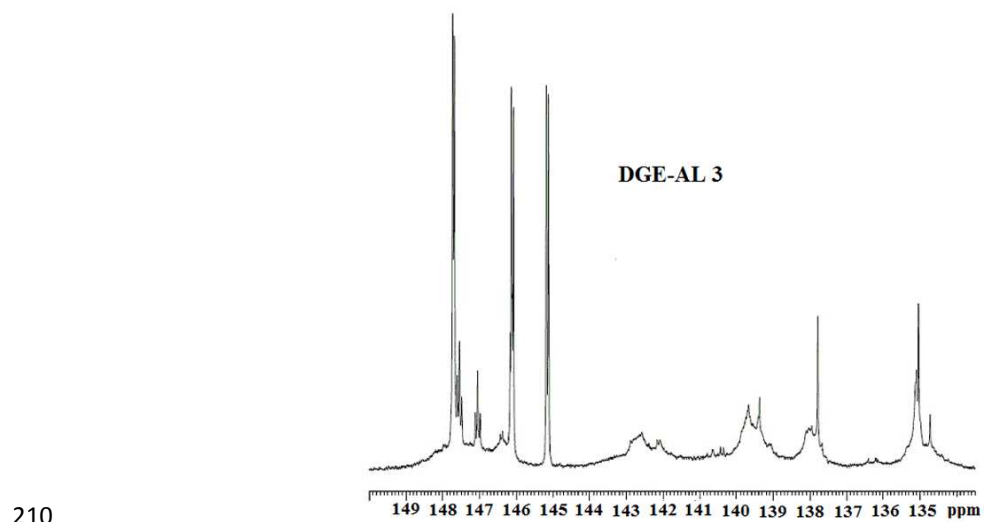
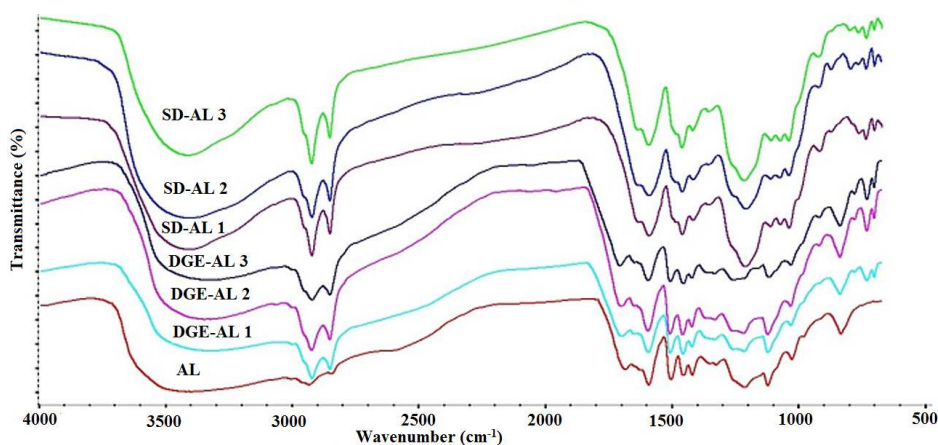


Fig. 1 ^{31}P -NMR of AL, DGE-AL 1 and DGE-AL 3.

215 More information of the structure of DGE-AL was acquired by FT-IR and ^{13}C -NMR
216 spectra. Fig. 2 shows the FT-IR spectra of the samples. As compared to AL, the intensity
217 of the absorption bands at 2924 and 2853 cm^{-1} (saturated C-H stretching vibration)
218 increased, the absorption at 1120 cm^{-1} corresponding to C-O-C stretching vibration
219 became clearer, and a new peak appeared at 731 cm^{-1} ($(\text{CH}_2)_n \geq 2$ rocking vibration) in
220 DGE-AL spectra. The results above suggested that long alkyl chain was introduced into
221 lignin through the reaction between DGE and lignin in the presence of BDMA, in
222 accordance with the result of ^{31}P -NMR analysis aforementioned.

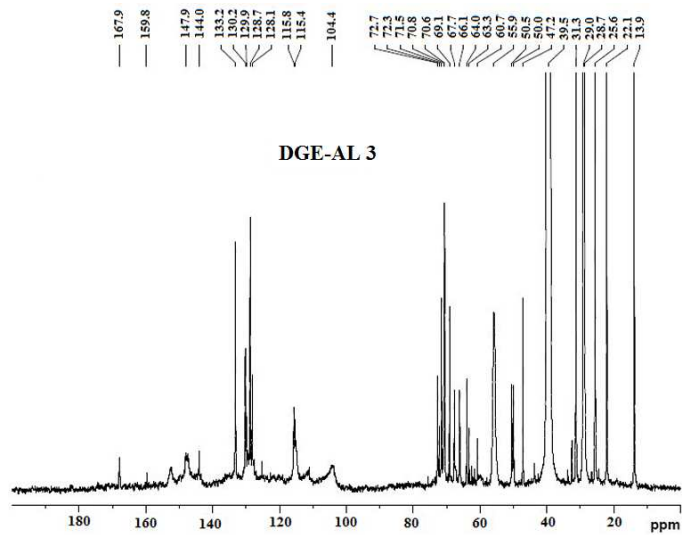


223 Fig. 2 FT-IR spectra of AL, DGE-AL and SD-AL samples.

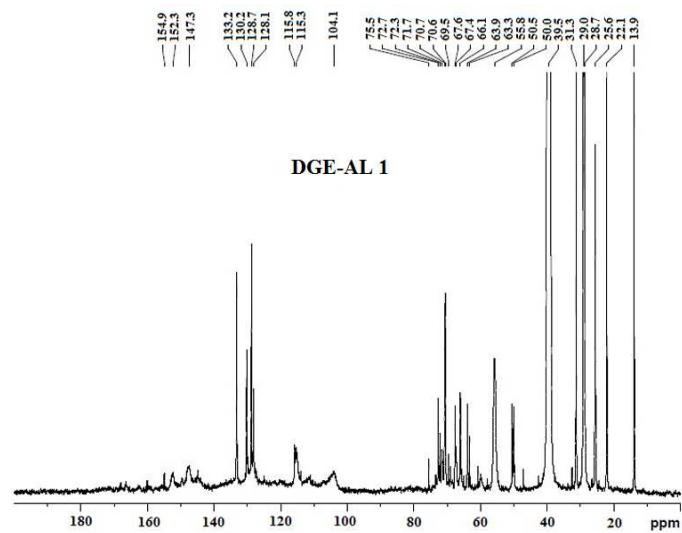
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225
226 Fig. 3 shows the ^{13}C -NMR spectra of AL and DGE-AL. In the aliphatic region, five
227 well-defined peaks were observed at 14.0, 25.6, 28.7, 29.0 and 31.3 ppm in DGE-AL
228 spectra, and these peaks are assigned to the methyl carbon and methylene carbon of
229 dodecyl chains. Generally, the ^{13}C -NMR chemical shift of the carbon atoms bonded to
230 oxygen (CH_2O , C-O-C, C-O-C=O) in glycerol ether or glycerides are assigned at the
231 bands of 60.0-70.0 ppm.²³⁻²⁹ As expected, the additional peaks at 60.0-72.7 ppm attributed
232 to the carbon atoms in glycerol ethers cross-linked structure (see Scheme 1, I, II, III) were
233 observed in DGE-AL spectra. Because of the complexity of the ring-opening reaction of
234 epoxy compound as well as the complicated structure of lignin macromolecule,³⁰ no effort
235 was made to assign these peaks. It was concluded that the expected compounds were
236 formed in the reaction of lignin and DGE under the catalysis of dimethylbenzylamine.

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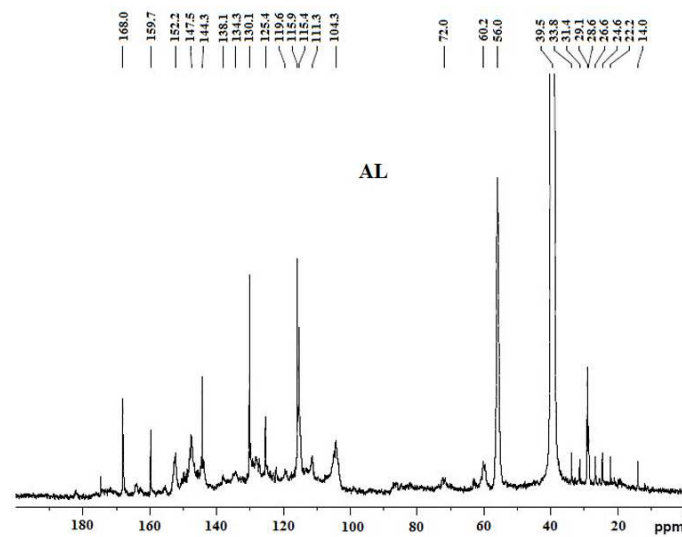
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Fig. 3 ^{13}C -NMR of AL, DGE-AL 1 and DGE-AL 3.

242

243 3.2. Preparation and characterization of SD-AL

244 Surfactants are amphiphilic materials containing both a polar long-chain hydrocarbon
245 “tail” and polar, usually ionic, “head” groups. Because of the lack of hydrophilic groups,
246 only the introduction of long alkyl chain into lignin cannot make lignin become a good
247 surfactant. In this case, lignin achieved both hydrophilic and hydrophobic characteristics
248 which were different from those of commercial surfactants. AL grafted with DGE under
249 different temperatures were sulfonated with chlorosulfonic acid, and then neutralized with
250 sodium hydroxide, yielding anionic surfactant SD-AL. The FT-IR spectra of SD-AL are
251 shown in Fig. 2. For the spectra of SD-AL, the intensity of the bands at 2924 and 2853
252 cm^{-1} correspond to methyl and methylene vibration, respectively. The new absorptions at
253 1073 and 1040 cm^{-1} correspond to the symmetric and asymmetric stretching vibrations of
254 S–O bond, respectively.³¹ The appearance of a strong peak at 1217 cm^{-1} is indicative of
255 the S=O stretch of the sulfatesalt as well as the sulfonic acid,³² indicating the existence of
256 SO_3 groups in SD-AL. As compared to DGE-AL, the intensity of the peak at 1120 cm^{-1}
257 corresponding to C-O-C stretching vibration in SD-AL became weak, indicating the
258 partial hydrolysis of ester groups during the preparation of SD-AL.

259 The effect of hydrolysis of ester groups during the preparation of SD-AL was
260 investigated through examining the content of the dichloromethane-insoluble components
261 of DGE-AL after treatment with 2 M NaOH for 1 h in an ice bath. Briefly, 1g of DGE-AL
262 was added to 30 mL of 2 M NaOH and the solution was stirred for 1h in an ice bath. After
263 the reaction, DGE-AL was regenerated by precipitation through adjusting the resulting
264 solution to pH=2 with 30% acetic acid. The result showed that the contents of
265 dichloromethane-insoluble components were 29.71%, 27.72 % and 26.43% of the weight
266 of regenerated DGE-AL 1, DGE-AL 2 and DGE-AL 3, respectively. This indicated that
267 some de-alkylation of DGE-AL occurred during the preparation of SD-AL. The
268 generation of dichloromethane-insoluble components should be due to the hydrolysis of
269 the ester groups in DGE-AL. Thus, optimization of the neutral condition for sulfonic acid
270 group should be conducted in a future study.

271

272

273 Table 3 Elemental analysis, yield, CMC and surface tension of water at CMC of all
 274 surfactant samples

Samples ^a	Elemental analysis (%)			Yield (%) ^b	CMC (g L ⁻¹)	Surface tension of CMC (mN/m)
	S	C	H			
AL	-	63.00	5.60	-	-	-
SD-AL 1	7.14	45.00	5.21	78.2	0.50	29.17
SD-AL 2	7.27	46.15	5.96	77.8	0.55	29.65
SD-AL 3	7.81	46.23	5.91	78.5	0.58	30.21
SDBS	-	-	-	-	0.97	30.50

275 ^a SDBS, commercial surfactant sodium dodecyl benzene sulfonate; SD-AL 1, SD-AL 2
 276 and SD-AL 3 derived from the sulfonation of DGE-AL 1 DGE-AL 2, and DGE-AL 3 at
 277 the same condition, respectively.

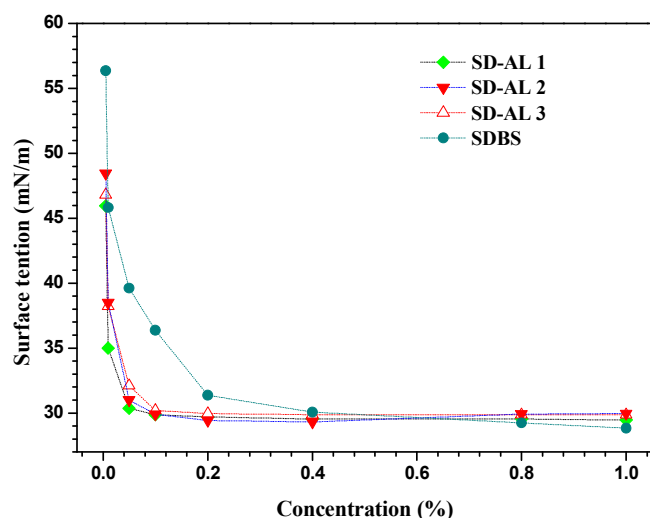
278 ^b the yield of SD-AL was based on the weight of DGE-AL.
 279 -, no measurement.

280

281 3.3. Characteristics of SD-AL as a surfactant

282 Fig. 4 shows the isotherms of the surface tension of aqueous solution plotted against
 283 concentrations of SD-AL as compared to commercial surfactant SDBS. By the addition of
 284 lignin surfactant, the surface tension of water decreased with the increase of the
 285 concentration. For SD-AL 1, SD-AL 2, and SD-AL 3, the surface tension was lower than
 286 that of SDBS when the concentration of aqueous surfactant solution was below 0.4 %.
 287 After sulfurization under the same conditions, the lignin surfactant derived from the
 288 reaction of lignin with DGE at a high temperature had a low surface tension at equivalent
 289 concentration of aqueous solution. It was consistent with the result reported by Košíková
 290 *et al.*, in which the higher amount of alkylhalide was used to modify the prehydrolysis
 291 lignin, the lower surface tension was observed.³³

292



293

294 Fig. 4 Isotherms of surface tension of water plotted against concentration of SD-AL
295 surfactant and SDBS.

296

297 Generally, the concentration of surfactant influences the physical and chemical
298 properties of surfactant solution. CMC is considered to be a fundamental property of
299 surfactants. A number of interfacial phenomena such as detergency can take place due to
300 the presence of micelles in solutions. In addition, other phenomena such as the surface
301 tension reduction is not directly influenced by the formation of micelles.³⁴ Therefore,
302 CMC and the corresponding surface tension are important indicators to measure the
303 surface activity of surfactants SD-AL. As seen from the data listed in Table 3, the CMC of
304 SD-AL decreased with the increase of reaction temperature between DGE and lignin, a
305 lowest CMC (0.50 g L^{-1}) and the corresponding surface tension (29.17 mN/m) were
306 achieved when the surfactant derived from the lignin grafted with DGE under $110 \text{ }^\circ\text{C}$. This
307 was because under a similar content of sulfur, surfactant derived from the lignin grafted
308 with DGE under high temperature has a high lipophilicity than that prepared under low
309 temperature. The CMC and surface tension at the corresponding CMC of all surfactants
310 derived from lignin were lower than the CMC (0.97 g L^{-1}) and the corresponding surface

311 tension (30.50 mN/m) of the commercial surfactant SDBS, indicating that SD-AL
312 surfactants had a good surface activity. Furthermore, the lignin surfactant prepared showed
313 a good ability to reduce the surface tension of aqueous solution, comparable to those of
314 the products prepared previously. In Košíková B's work,³³ a series of lignin-based surface
315 active agents derived from alkylation of beechwood prehydrolysis lignin with
316 laurylbromide decreased the surface tension of aqueous solution to 30-40 mN/m at the
317 concentration of 1 g L⁻¹. Homma *et al.*³⁵ reported that nonionic polymeric surfactants
318 derived from the modification of acetic acid lignin with polyethylene glycol
319 diglycidylethers lowered the surface tension of aqueous solution to 36 mN/m at the
320 corresponding CMC (50g L⁻¹). The results of the present study suggested that lignin
321 surfactants with high surface activity was prepared by modification of lignin with DGE
322 and chlorosulfonic acid.

323

324 4. Conclusion

325 A long alkyl chain was successfully introduced into lignin by the reaction between
326 lignin and DGE in the presence of BDMA as a catalyst. The reaction between DGE and
327 carboxy group was the predominant reaction at 95 °C-110 °C. The surface tension of the
328 lignin surfactant solution was lower than that of commercial surfactant sodium
329 dodecylbenzenesulphonate when the concentration of surfactant solution was below 0.4 %,
330 indicating that the surfactants prepared from modified alkaline lignin had a good surface
331 activity. A lowest CMC (0.50 g L⁻¹) and the corresponding surface tension (29.17 mN/m)
332 were achieved when the surfactant was derived from the lignin grafted with DGE at 110
333 °C. The anionic lignin surfactants prepared in this study could be utilized as detergents or
334 to enhance oil recovery.

335

336

337 **Acknowledgements**

338 We express our gratitude for the financial support from Beijing Forestry University Young
339 Scientist Fund (BLX2012025), the National Natural Science Foundation of China
340 (3110103902), the Major State Basic Research Projects of China (973-2010CB732204),
341 and the Open Foundation of the State Key Laboratory of Pulp and Paper Engineering
342 (201261).

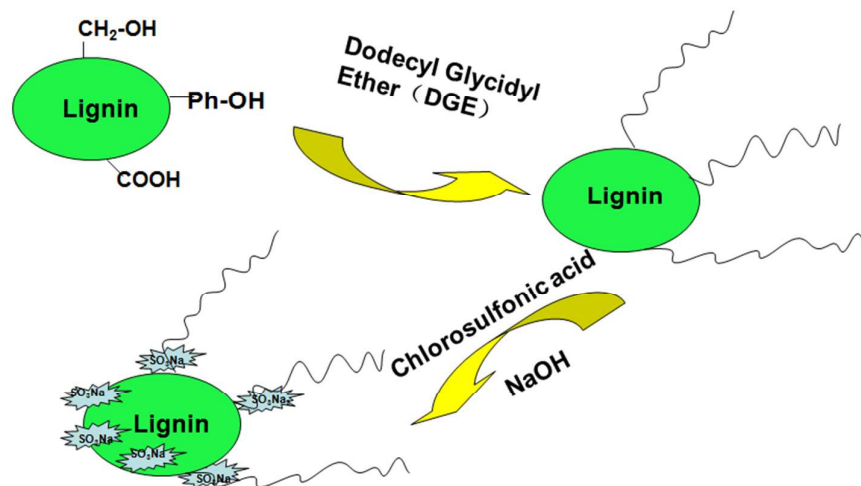
343

344 **References**

- 345 1. M. Enquist-Newman, A. M. E. Faust, D. D. Bravo, C. N. S. Santos, R. M. Raisner, A. Hanel,
346 P. Sarvabhowman, C. Le, D. D. Regitsky, S. R. Cooper, *Nature*, 2013, **505**, 239–243.
- 347 2. Y. Li, A. J. Ragauskas, *RSC Adv*, 2012, **2**, 3347-3351.
- 348 3. A. J. Ragauskas, C. K. Williams, B. H. Davison, G. Britovsek, J. Cairney, C. A. Eckert, W. J.
349 Frederick, J. P. Hallett, D. J. Leak, C. L. Liotta, *Science*, 2006, **311**, 484-489.
- 350 4. M. G. Adsul, D. V. Gokhale, *Biofuels*, 2012, **3**, 383-386.
- 351 5. W. Boerjan, J. Ralph, M. Baucher, *Annual review of plant biology*, 2003, **54**, 519-546.
- 352 6. R. Y. Nsimba, C. A. Mullen, N. M. West, A. A. Boateng, *ACS Sustainable Chemistry & Engineering*,
353 2013, **1**, 260-267.
- 354 7. M. Dashtban, A. Gilbert, P. Fatehi, *RSC Adv*, 2014, **4**, 2037-2050.
- 355 8. S. Laurichesse, L. Avérous, *Prog. Polym. Sci.*, 2013.
- 356 9. S. N. Fhlaithearta, S. Ernst, K. G. Nierop, G. de Lange, G. J. Reichart, *Mar. Micropaleontol.*, 2013,
357 **102**, 69–78.
- 358 10. Z. Liu, L. Zhao, S. Cao, S. Wang, P. Li, *BioResources*, 2013, **8**, 6111-6120.
- 359 11. Y. Matsushita, S. Yasuda, *Bioresour. Technol.*, 2005, **96**, 465-470.

- 360 12. X. Ouyang, L. Ke, X. Qiu, Y. Guo, Y. Pang, *J. Disper. Sci. Technol.*, 2009, **30**, 1-6.
- 361 13. S. Y. Lin, *US Pat.*, 3910873, 1975.
- 362 14. M. D. Matzinger, *US Pat.*, 6045606, 2000.
- 363 15. Y. Matsushita, S. Yasuda, *J. Wood. Sci.*, 2003, **49**, 166-171.
- 364 16. L. R. Morrow, M. G. DaGue, L. E. Whittington, *US Pat.*, 4790382, 1988.
- 365 17. D. G. Nae, M. G. DaGue, N. G. Dunn, *US Pat.*, 5230814, 1993.
- 366 18. L. L. Zhao, Z. G. Liu, X. E. Huang, Y. H. Liu, *Advanced Materials Research*, 2013, **634**, 456-460.
- 367 19. S. S. Hou, Y. P. Chung, C. K. Chan, P. L. Kuo, *Polymer*, 2000, **41**, 3263-3272.
- 368 20. D. Chen, J. Zhu, P. Yuan, S. Yang, T. H. Chen, H. He, *J. Therm. Anal. Calorim.*, 2008, **94**, 841-848.
- 369 21. W. Tänzer, S. Reinhardt, M. Fedtke, *Polymer*, 1993, **34**, 3520-3525.
- 370 22. L. Shechter, J. Wynstra, *Industrial & Engineering Chemistry*, 1956, **48**, 86-93.
- 371 23. B. Ahvazi, O. Wojciechowicz, T. M. Ton-That, J. Hawari, *J. Agric. Food Chem.*, 2011,
- 372 **59**, 10505-10516.
- 373 24. Y. Pu, S. Cao, A. J. Ragauskas, *Energy & Environmental Science*, 2011, **4**, 3154-3166.
- 374 25. J. Qin, M. Wolcott, J. Zhang, *ACS Sustainable Chemistry & Engineering*, 2013.
- 375 26. J. L. Wen, S. L. Sun, T. Q. Yuan, F. Xu, R. C. Sun, *J. Agric. Food Chem.*, 2013, **61**, 11067-11075.
- 376 27. R. Burns Jr, J. Friedman, M. Roberts, *Biochemistry*, 1981, **20**, 5945-5950.
- 377 28. F. Blainvaux, P. J. Madec, E. Marechal, *Polym. Bull.*, 1985, **13**, 237-244.
- 378 29. A. A. Bredikhin, Z. A. Bredikhina, V. G. Novikova, A. V. Pashagin, D. V. Zakharychev, A.
- 379 T. Gubaidullin, *Chirality*, 2008, **20**, 1092-1103.
- 380 30. M. Schömer, J. Seiwert, H. Frey, *ACS Macro Letters*, 2012, **1**, 888-891.
- 381 31. D. Chen, S. Wang, M. Xiao, Y. Meng, *Energy & Environmental Science*, 2010, **3**, 622-628.

- 382 32. T. Huang, J. M. Messman, K. Hong, J. W. Mays, *Journal of Polymer Science Part A: Polymer*
383 *Chemistry*, 2012, **50**, 338-345.
- 384 33. B. KošíWkova, M. Ďuriš, V. Demianova, *Eur. Polym. J.*, 2000, **36**, 1209-1212.
- 385 34. M. J. Rosen, J. T. Kunjappu, *Surfactants and interfacial phenomena*. John Wiley & Sons, 2012.
- 386 35. H. Homma, S. Kubo, T. Yamada, Y. Matsushita, Y. Uraki, *J. Wood Chem. Technol.*, 2008, **28**,
387 270-282.
- 388



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