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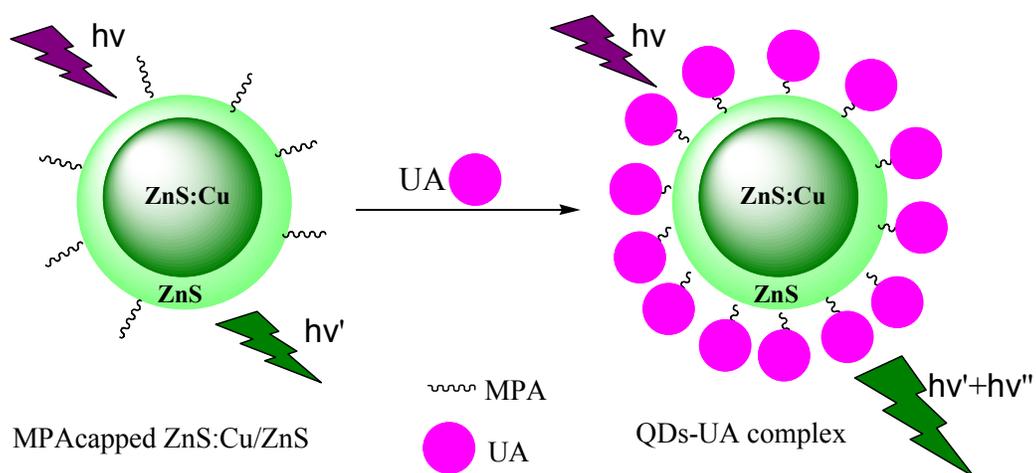
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Graphical Abstract

A simple core/shell ZnS:Cu/ZnS QDs based fluorescence enhancement system is constructed for detecting uric acid. The method with high sensitive, selective and lower toxicity features has applied to determination of UA in real samples.



1 **Fluorescence enhancement detection of uric acid based on**
2 **water-soluble 3-mercaptopropionic acid-capped core/shell**
3 **ZnS:Cu/ZnS**

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8 **Abstract**

9 3-Mercaptopropionic acid (MPA)-capped ZnS:Cu/ZnS core-shell quantum dots
10 (QDs) have been synthesized via a facile aqueous coprecipitation method and
11 characterized with fluorescence, UV-Vis absorption, infrared spectroscopy and
12 transmission electron microscopy. The fluorescence of ZnS:Cu/ZnS could be
13 increased in the presence of uric acid (UA). The affecting factors for the fluorescence
14 of ZnS:Cu/ZnS were examined including pH, temperature and reacting time. Under
15 the optimized conditions, the fluorescence intensity of the ZnS:Cu/ZnS QDs against
16 the UA concentration showed a linear response in the range of 0.66 μM to 3.3 μM
17 with the correlation coefficient (R^2) 0.9973 and the limit of detection 0.044 μM . Most
18 relevant molecules and physiological ions had no effect on the detection of UA. The
19 feasibility of developed method was further demonstrated by determining the
20 concentration of uric acid in human urine samples and the recoveries were
21 95%~103%. Our work provides a sensitive, selective and convenient fluorescence
22 method to determine UA in real samples.

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23 Keywords: Core/shell ZnS:Cu/ZnS; Fluorescence Detection; Uric acid

24

25 **1 Introduction**

26 Semiconductor nanoparticles or quantum dots (QDs) have emerged as the
27 attractive fluorescent probes for biomolecules detection due to their high fluorescence
28 quantum efficiency, size-dependent broad absorption and readily size-tunable narrow
29 emission as well as photochemical stability. Cadmium chalcogenide QDs hold a
30 special position by virtue of the ease preparation of high-quality samples and the
31 tunability of their luminescence over most of the visible spectrum. Nevertheless, the
32 main concern about the use of fluorophores based on cadmium chalcogenide QDs in
33 biological applications is represented by their inherent toxicity. [1] Hence critical
34 investigations on fabrication of low poisonous or nontoxic QDs are desirable. ZnS
35 QDs are extremely promising candidate with stable luminescence and high quantum
36 efficiency and particularly suitable for use as luminescent host materials for a very
37 large variety of dopants due to their wide band gap energy at room temperature [2].
38 Numerous metal ions, such as Mn^{2+} [3], Cu^+ [4], Ag^+ [5], and Eu^{2+} [6], have been
39 successfully doped into ZnS to produce photoluminescence in different wavelength
40 regions. These doped QDs (d-dots) with minimized self-absorption and higher
41 quantum efficiency can be used as cadmium-free emitters for biolabeling [7], efficient
42 donors for the Förster resonance energy transfer [8] or sensors for biomolecules[9]. On
43 the other hand, significant progress has been made on the surface capping ligands of
44 QDs and currently phosphines [10], amines[11], and carboxylic acids [12] are the

45 most widely used species, through which improved properties and functions can be
46 achieved, such as good biocompatibility and molecular recognition capabilities. The
47 surface modification of QDs with particular capping ligands as the platform makes
48 QDs available for interaction with various target analytes. [13] Therefore,
49 functionalized QDs have been progressed toward different chemical species including
50 ions [14], small molecules [15], and biological macromolecule [16].

51 Uric acid (UA) is the end product of purine nucleoside, adenosine and guanosine
52 metabolism and is a major antioxidant in humans. Fast and accurate determination of
53 UA level in human physiological fluids has been recognized as a vital clinical
54 indicator in the diagnosis of patients suffering from numerous metabolic disorders
55 such as gout, renal impairment, leukemia, diabetes, and high blood pressure [17].
56 Different approaches have been involved in the uric acid determination, such as
57 enzymatic-spectrophotometric method [18], electrochemical [20], or fluorescent
58 techniques [21] and capillary electrophoresis [22]. Among them, fluorescence analysis
59 has attracted substantial attention with their unique advantages of simplicity, rapidity,
60 high sensitivity and low cost of instrumentation and maintenance. Ferrer et al. [23]
61 proposed a fluorescent sol-gel biosensor and applied to determine UA in biological
62 fluids. Ren and co-workers [24] reported a new strategy for highly sensitive
63 determination of uric acid based on fluorescence resonance energy transfer (FRET)
64 using gold nanoparticles as energy acceptors. Dey's group [25] synthesized
65 naphthyridine based fluorescent receptors for the recognition of uric acid. To the best
66 of our knowledge, few references were reported based on QDs fluorescence detection

67 of UA. Zhang et al. [26] found that CdTe quantum dots can act as fluorescence probe
68 for detecting uric acid in biological fluid. We try to use ZnS QDs as the alternative to
69 construct UA detection system toward high sensitivity and low toxicity.

70 Recently inorganically passivated or core/shell structured QDs have emerged
71 because of their higher-emission quantum efficiency and thermal stability over the
72 single core nanocrystals [27]. A succession of core/shell QDs have been synthesized
73 in aqueous solution, such as ZnSe:Eu/ZnS, CdS:Mn/ZnS, ZnS:Mn/ZnS. However,
74 ZnS:Cu/ZnS core/shell QDs are seldom reported because the different crystal
75 structure between CuS and ZnS led to co-precipitation occur difficultly. Zhang's
76 group [28] firstly synthesized successfully and characterized ZnS:Cu/ZnS core/shell
77 nanocrystals with excellent water-solubility, stability and demonstrated that core/shell
78 structures can be used as a powerful strategy to enhance PL properties of doped
79 semiconductor NCs. This core/shell structure QDs has been employed to determine
80 folic acid in aqueous media [29]. Further extended analytical application of
81 ZnS:Cu/ZnS core-shell quantum dots is desirable.

82 In this paper, we synthesized ZnS:Cu/ZnS core-shell quantum dots in aqueous
83 solution by using 3-Mercaptopropionic acid (MPA) as the surface-ligand. The polar
84 carboxylic acid group of MPA assembling on the surface of QDs renders the QDs
85 water-soluble and available for interaction with specific analytes. The fluorescence of
86 ZnS:Cu/ZnS QDs were increased dramatically in the presence of UA. The fluorescent
87 detection system of UA was constructed based on core/shell ZnS:Cu/ZnS QDs with
88 high sensitive and selective features and applied to determination of UA in human

89 urine samples. The results revealed that the as-synthesized ZnS:Cu/ZnS core/shell
90 QDs might be a new promising fluorescence probe for biomedical or clinical analysis.

91 **2 Experimental**

92 **2.1. Reagents**

93 All the chemicals were of reagent grade and used without further purification.
94 3-Mercaptopropionic acid (MPA), sodium sulfide nonahydrate ($\text{Na}_2\text{S}\cdot 9\text{H}_2\text{O}$),
95 Copper(II) chloride dihydrate (CuCl_2), sodium dihydrogen phosphate (NaH_2PO_4) and
96 disodium hydrogen phosphate (Na_2HPO_4), sodium hydroxide (NaOH), heptahydrate
97 zinc sulphate ($\text{ZnSO}_4\cdot 7\text{H}_2\text{O}$) and Uric acid (UA) were purchased from Aladdin. Water
98 used throughout was doubly deionized.

99 **2.2. Apparatus**

100 Fluorescence measurements were performed using a Cary Eclipse fluorescence
101 spectrophotometer. UV-Vis absorption spectra were collected using a TU-1901
102 Double beam UV-Vis spectrophotometer. JEOL JEM-2100 high-resolution
103 transmission electron microscope was adopted to examine the appearance and size of
104 nanoparticles. Infrared spectroscopy was carried out using a FTIR-8400S Infrared
105 spectrometer.

106 **2.3. Methods**

107 **2.3.1. Synthesis of ZnS:Cu QDs**

108 MPA-capped ZnS:Cu QDs were synthesized via an aqueous route similar to
109 Yang's method [30] for ZnS:Mn. For 1% copper doping, the synthesis is as follows. In
110 a three-neck round-bottomed flask, 5 mL of 0.1 M $\text{ZnSO}_4\cdot 7\text{H}_2\text{O}$, 0.5 mL of 0.01 M

111 CuCl_2 , and 20 mL of 0.1 M MPA were combined. This solution was diluted to 45 mL
112 with water, and the pH was adjusted to 11 using 1 M NaOH. Next the solution was
113 degassed by bubbling N_2 gas for 30 min, after which 4.5 mL of degassed 0.1 M
114 $\text{Na}_2\text{S}\cdot 9\text{H}_2\text{O}$ was quickly injected into the solution. It remained clear and was stirred
115 for 20 min. The crude solution showed nearly no luminescence. The reaction mixture
116 was then exposed to air and aged 2 h at 50 °C, then The luminescence was observed.
117 The MPA-capped ZnS:Cu QDs could be precipitated by ethanol, the precipitate was
118 centrifuged and washed with ethanol, then dried in vacuum. Stock solutions of
119 $\text{ZnSO}_4\cdot 7\text{H}_2\text{O}$ and CuCl_2 were prepared and used for many syntheses. MPA and
120 $\text{Na}_2\text{S}\cdot 9\text{H}_2\text{O}$ solutions were less stable and prepared fresh each time the synthesis was
121 performed.

122 **2.3.2 Synthesis of ZnS:Cu/ZnS QDs**

123 The preparation of ZnS:Cu/ZnS QDs was performed according to Zhang's
124 method [28] with some improvements. The solution after aging was used without
125 purification, and then 8 mL 0.1 M $\text{ZnSO}_4\cdot 7\text{H}_2\text{O}$ / 5 mL 0.1 M Na_2S / 17.2 mL 0.1 M
126 MPA/ 2 mL 1 M NaOH were added successively to the solution in 40 times injections
127 with a 30 s interval in between each injection. Then the mixture refluxed for 2 h. After
128 cooling to room temperature the reaction mixture was precipitated with ethanol and
129 centrifuged. The nanocrystals were further washed with ethanol and finally dried
130 under vacuum at room temperature. The obtained powder is white and highly soluble
131 in water.

132 **2.3.3 Analytical procedure**

133 1 mL of 5 mg mL⁻¹ ZnS:Cu/ZnS QDs, 2 mL 0.02 M PBS (pH 7.0) were mixed
134 into a 10 mL calibrated test tube, then a certain amount of UA were stepwise added
135 into the mixture and shaken thoroughly for 7 min. At an excitation wavelength of 332
136 nm, the fluorescence spectra of the mixture were determined in the 380-600 nm
137 emission wavelength range and the fluorescence intensity at 480 nm was used for
138 quantitative analysis.

139 **3. Results and discussion**

140 **3.1. Surface characters of ZnS:Cu/ZnS QDs**

141 To identify the conjugation mode between 3-mercaptopropionic acid and
142 ZnS:Cu/ZnS QDs, the IR spectra of the QDs coated by 3-mercaptopropionic acid are
143 measured and compared with pure 3-mercaptopropionic acid. Fig. 1 shows that the
144 spectral features and several peak positions are similar. However, the absorption of
145 S-H stretching vibration at 2600-2500 cm⁻¹ has not been found in the IR spectra of
146 ZnS:Cu/ZnS QDs, which demonstrate that the S-H bond in 3-mercaptopropionic acid
147 of the QDs is covalently bound to Zn on the QDs surface.

148

149 **3.2. HRTEM images of ZnS:Cu and ZnS:Cu/ZnS QDs**

150 High-resolution transmission electron microscopy (HRTEM) is performed to
151 study the morphology of the prepared QDs. Fig. 2 shows the image of the prepared
152 ZnS:Cu and ZnS:Cu/ZnS QDs. The shape of ZnS:Cu QDs is close to spherical and
153 aggregated. After the growth of the ZnS shell, they show no difference with ZnS:Cu
154 QDs except for their average size. Due to the growth of the ZnS shell on ZnS:Cu QDs,

155 the average size (4.0 nm) of ZnS:Cu/ZnS core/shell QDs is larger than that (2.3 nm)
156 of the ZnS:Cu core QDs. The difference between them means that the shell thickness
157 is around 1.7 nm.

158

159 **3.3. Spectral characteristics of ZnS:Cu and ZnS:Cu/ZnS QDs**

160 The UV-vis absorption and the fluorescence emission spectra of the ZnS:Cu
161 solution and the ZnS:Cu/ZnS solution are shown in Fig. 3 and Fig.4. Fig.3 shows that
162 the ZnS:Cu solution produces a UV absorption excitonic peak around 280 nm, after 2
163 h of reflux, the peak is red-shifted to 290 nm. This shift is due to shell growth, which
164 indicate that ZnS shell is coated around the ZnS:Cu core, instead of gathering into a
165 nuclear. Fig. 4 shows that the ZnS:Cu solution develops an emission peak at 474 nm.
166 After ZnS shell is coated around the ZnS:Cu core, the fluorescence intensity increases
167 by almost four times with a very slight shift in emission to 481 nm, which is likely
168 due to surface Cu sites that were not emissive originally become emissive upon
169 incorporation into the ZnS lattice with further ZnS shell growth.

170

171 **3.4. Optimization of the system**

172 Here, the effect of pH value, reaction temperature and reaction time on the
173 fluorescence intensity of the ZnS:Cu/ZnS QDs–UA mixture was studied in detail.

174 **3.4.1 Effect of pH**

175 The effect of pH on the fluorescence intensity of ZnS:Cu/ZnS QDs and
176 ZnS:Cu/ZnS QDs with 3.3 μ M UA was studied in the pH range of 5-10. As shown in

177 Fig.5, the fluorescence intensity of ZnS:Cu/ZnS QDs and ZnS:Cu/ZnS-UA mixture
178 solution both increased with the change of pH from 5.0 to 8.0. This is due to the
179 protonation of the stabilizer MPA in acid medium. With the increase of pH, the
180 deprotonation of the thiol group in the MPA molecule occurs. This deprotonation
181 could strengthen the covalent bond between Zn and MPA molecule, which could
182 enhance the fluorescence intensity. However, the fluorescence intensity begins to
183 decline with the further increase of pH value, which might be caused by the
184 disintegration of MPA in extreme basic medium. Considering the protonation of the
185 stabilizer MPA in acid medium, and the environment uric acid present, the
186 physiological condition of pH 7.0 PBS was used in further experiments.

187

188 **3.4.2 Effect of Reaction temperature**

189 The effect of the reaction temperature on the fluorescence intensity of
190 ZnS:Cu/ZnS QDs and ZnS:Cu/ZnS QDs with 3.3 μM UA was studied, and the results
191 were shown in Fig.6. It shows that the fluorescence intensity of the ZnS:Cu/ZnS QDs
192 with 3.3 μM UA is strongest at 25 $^{\circ}\text{C}$, then decreased gradually with the increase of
193 reaction temperature from 25 $^{\circ}\text{C}$ to 50 $^{\circ}\text{C}$, which might be caused by the
194 decomposition of UA at high temperature. The fluorescence intensity of the
195 ZnS:Cu/ZnS QDs decreased slightly gradually with the increase of reaction
196 temperature from 20 $^{\circ}\text{C}$ to 50 $^{\circ}\text{C}$. The fluorescence-enhanced extent of the
197 ZnS:Cu/ZnS QDs solution by UA reached a maximum at 25 $^{\circ}\text{C}$, so the reaction
198 temperature of 25 $^{\circ}\text{C}$ was chosen for the further experiments.

199

200 **3.4.3 Effect of Reaction time**

201 The effect of reaction time on the fluorescence intensity of ZnS:Cu/ZnS QDs
202 with 3.3 μM UA was studied and the results were shown in Fig.7. It can be seen that
203 the fluorescence of the QDs increased immediately after the addition of UA, and
204 reached equilibrium after a reaction time of 7min. So in the further study, the mixture
205 of ZnS: Cu/ZnS QDs and UA were shaken thoroughly for 7 min and then the
206 fluorescence intensity was recorded.

207

208 **4. Fluorescence enhancement of MPA-capped ZnS:Cu/ZnS QDs by UA**

209 **4.1. Analytical characteristics**

210 Fig. 8 shows the fluorescence spectra of ZnS:Cu/ZnS QDs upon the addition of
211 different concentrations of UA. It can be seen that the fluorescence intensity of
212 ZnS:Cu/ZnS QDs increased with the UA added. No obvious emission peak shift and
213 change of full width at half maximum were found. The fluorescence enhancing
214 phenomenon might be caused by that after adding uric acid to QDs solutions, the
215 surface of QDs might be changed and cause the decrease of surface defects. Through
216 the formation of hydrogen bond between secondary amine of uric acid and carboxyl
217 of MPA coated on the QDs[26], UA is conjugated to the MPA of nanocrystal surface,
218 such complex will result in the formation of a passivation shell around the nanocrystal,
219 which could remove the surface defects, and the luminescent efficiency will be
220 improved dramatically.

221 In Fig. 9, it can be seen that the fluorescence intensity of ZnS:Cu/ZnS QDs
222 gradually increased with the addition of UA until 13.2 μM . The insert in Fig.9
223 illustrates that the fluorescence intensity, $F-F_0$ exhibits a linear relationship with the
224 logarithm of the concentration of uric acid in the range from 0.66 μM to 3.3 μM ,
225 which is best described by the equation:

$$226 \quad F-F_0 = -239.30967 + 368.04448 \log C_{\text{UA}} (10^{-7}\text{M})$$

227 F_0 and F are the fluorescence intensity of the ZnS:Cu/ZnS QDs in the absence and
228 presence of uric acid, respectively. The correlation coefficient was 0.9973. The
229 relative standard deviation (RSD) was 1.3 %. The limit of detection (LOD) calculated
230 according to IUPAC definitions is to be 0.044 μM . Table.1 shows the comparison of
231 the established method and other fluorescence method of UA. The results revealed that
232 the as-conducted system is convenient and sensitive and the ZnS:Cu/ZnS QDs
233 material has lower toxicity over organic fluorescent substances.

234 **4.2 Selectivity study**

235 we studied the effect of a series of physiologically related ions such as K^+ , Na^+ ,
236 NH_4^+ , Ca^{2+} and Mg^{2+} and other small organic molecules such as ascorbic acid,
237 creatinine, dopamine, urea, glucose, d-lactose and sucrose and some amino acids such
238 as L-alanine, L-cysteine and L-tryptophan on the fluorescence intensity of the
239 ZnS:Cu/ZnS QDs under the same conditions, and the results were shown in Fig. 10. It
240 could be found that the fluorescence intensity of ZnS:Cu/ZnS increases to 2-fold in
241 solution containing 3.3 μM UA, and was almost unaffected by other molecules such
242 as ascorbic acid, dopamine, etc. Therefore, the method has a high selectivity and

243 could be applied to the direct determination of trace amounts of uric acid in urine
244 samples after dilution.

245

246 **4.3 Determination of uric acid in human urine**

247 In order to demonstrate the practical utility of the developed system, the prepared
248 method was applied to determine the concentration of uric acid in two human urine
249 samples. To get the appropriate concentration of sample solutions, urine samples are
250 directly diluted with ultrapure water. The results obtained by the standard addition
251 method were shown in Table1, and the accuracy of the proposed method was
252 evaluated by determining the average recoveries of UA in real samples. It can be seen
253 that the RSD was lower than 2.23 % and the average recoveries of UA in the real
254 samples were between 95 % and 103 %. The above results demonstrated the potential
255 applicability of the ZnS:Cu/ZnS QD-based fluorescence probe for the detection of UA
256 in human urine.

257

258 **5. Conclusion**

259 In summary, we have constructed a core/shell ZnS:Cu/ZnS QDs based
260 fluorescence system for UA and successfully applied to determination of UA levels in
261 human urine. The as-prepared QDs probes for UA detection exhibits highly sensitivity,
262 selectivity and lower toxicity over other QDs systems. To the best of our knowledge,
263 MPA-capped ZnS:Cu/ZnS core/shell QDs was firstly utilized to the determination of
264 uric acid. As a kind of novel fluorescence probe, it is the most favorable alternative.

265 The present approach extends a new applicable assay for developing economical,
266 sensitive, and selective QD-based fluorescent methods in biomedical or bioprocess.

267 **Acknowledgements**

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319 **Figure captions:**

320 **Table1. The comparison of the established method and other florescence method**

321 **Table2. The determination results of the Human urine samples**

322 **Fig. 1 Infrared spectra of pure 3-mercaptopropionic acid and MPA-coated**
323 **ZnS:Cu/ZnS QDs**

324 **Fig. 2 HRTEM images of ZnS:Cu and ZnS:Cu/ZnS QDs**

325 **Fig. 3 The UV-vis absorption spectra of the ZnS:Cu and the ZnS:Cu/ZnS**

326 **Fig. 4 The fluorescence emission spectra of the ZnS:Cu and the ZnS:Cu/ZnS**

327 **Fig. 5 The effect of pH on the fluorescence intensity of Zn:Cu/ZnS QDs and**
328 **ZnS:Cu/ZnS QDs with 3.3 μ M UA**

329 **Fig. 6 The effect of the reaction temperature on the fluorescence intensity of**
330 **ZnS:Cu/ZnS QDs and ZnS:Cu/ZnS QDs with 3.3 μ M UA**

331 **Fig. 7 The effect of reaction time on the fluorescence intensity of**

332 **ZnS:Cu/ZnS QDs with 3.3 μ M UA**

333 **Fig. 8 The fluorescence spectra of ZnS:Cu/ZnSQDs upon stepwise addition**

334 **(increments of 2 μ L) of a 0.5 mM UA solution in PBS (pH 7.0) buffer solution**

335 **Fig. 9 The fluorescence intensity of ZnS:Cu/ZnS QDs with different**

336 **concentrations of UA. Inset: plot of FL intensity versus logarithm of the**

337 **concentration of UA in the range of 0.66–3.3 μ M. PBS: 0.02 M phosphate buffer**

338 **solution (pH 7.0).**

339 **Fig. 10 Relative fluorescence intensity of ZnS:Cu/ZnS QDs in 0.02 M phosphate**

340 **buffer solution (pH 7.0) containing various substances (0.033 mM). F and F_0 are**

341 **the fluorescence intensity in the presence and absence of foreign substances,**

342 **respectively.**

343 **Table1.**

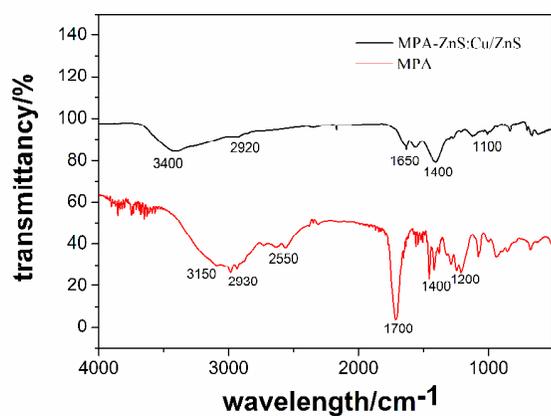
Methods	Material	Detection limit	Ref
fluorescent sol-gel biosensor	A coupled uricase–peroxidase system	20nM	[23]
FRET	Gold nanoparticle	25nM	[24]
fluorescence quenching	Naphthyrindine	90nM	[25]
fluorescence quenching	CdTe quantum dots	103nM	[26]
Fluorescence enhancement	ZnS:Cu/ZnS	44nM	This work

344

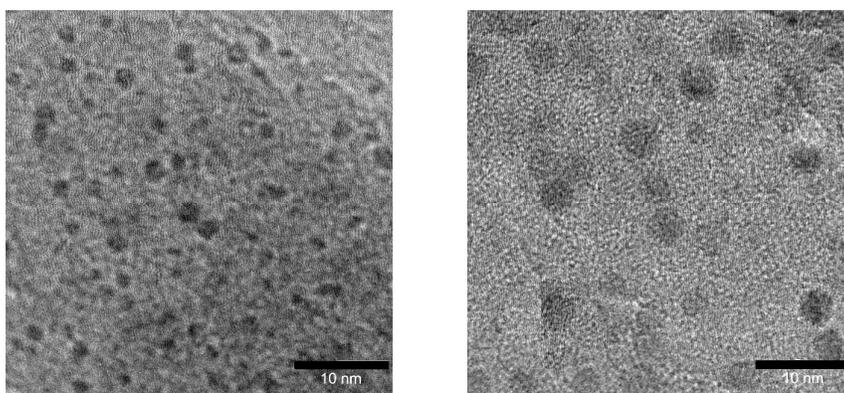
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346 **Table2.**

Samples	Average found / 10^{-7} molL $^{-1}$	Added / 10^{-7} molL $^{-1}$	Total found / 10^{-7} molL $^{-1}$	RSD/ (%) (n=5)	Recovery / (%)
Human urine1	6.81	5	11.97	1.36	103
Human urine2	6.34	5	11.41	0.64	101
		15	21.53	1.38	98
		15	20.58	2.23	95

347 **Fig. 1**

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349 **Fig. 2**

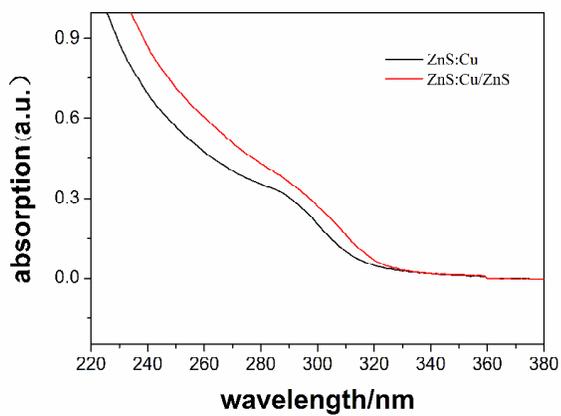
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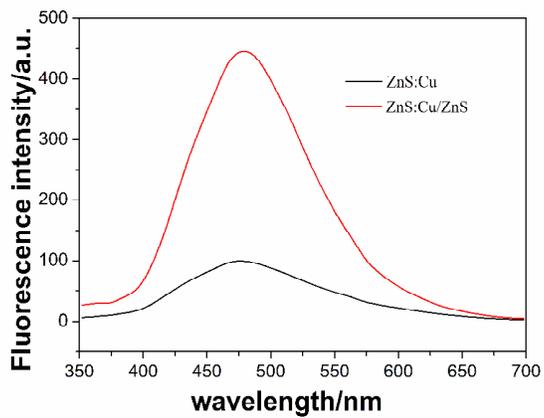
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355 **Fig. 3**

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358 **Fig. 4**

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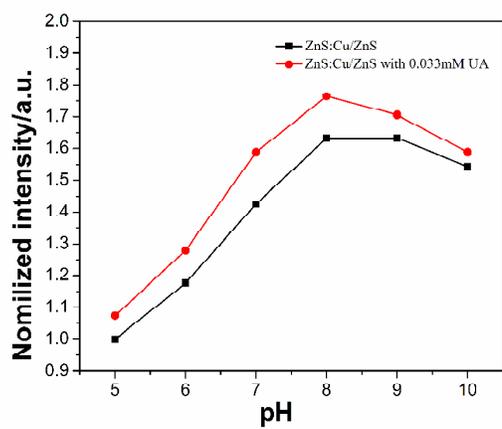
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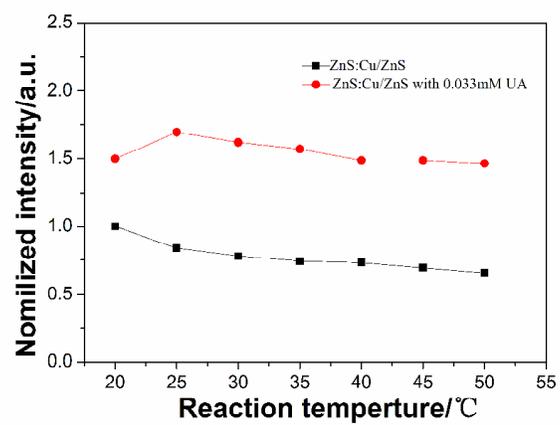
366 Fig. 5



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368 Fig. 6

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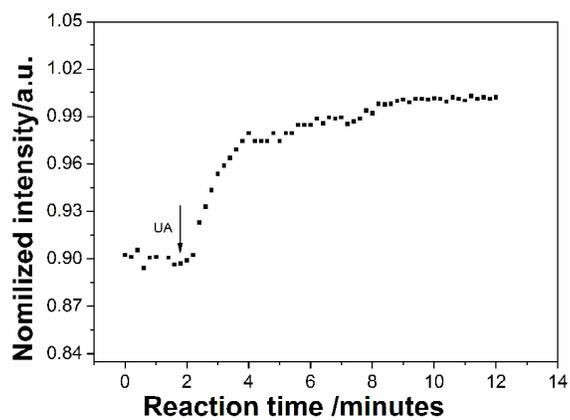
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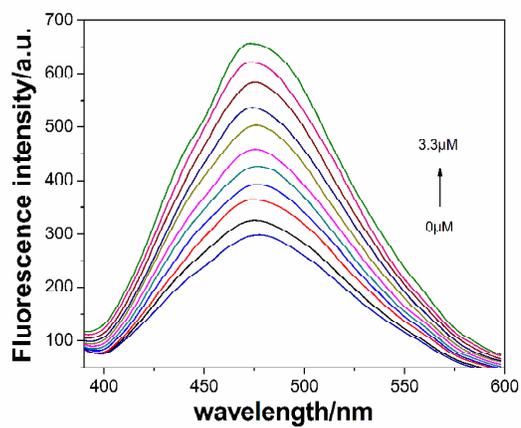
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377 **Fig. 7**

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379 **Fig. 8**

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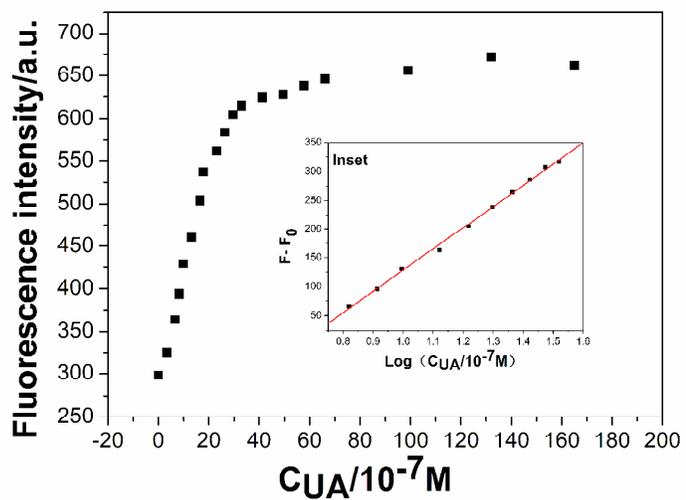
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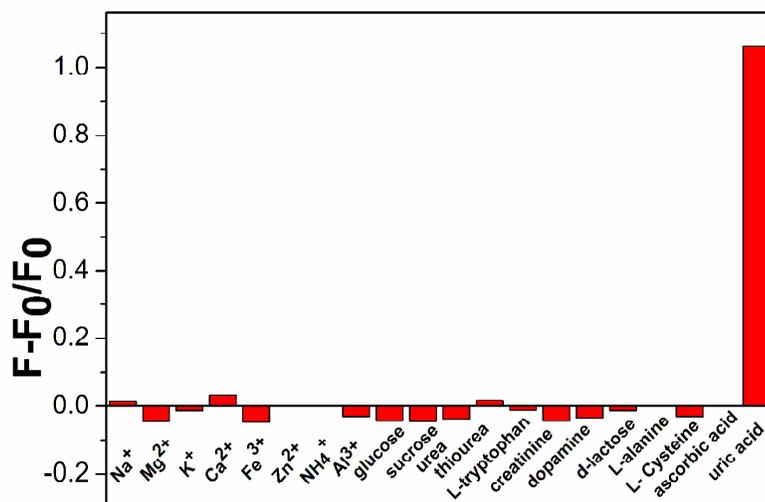
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388 Fig. 9



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390 Fig. 10



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