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Simple approach for the immobilization of horseradish peroxidase on Poly-L-histidine modified reduced graphene oxide for amperometric determination of Dopamine and H₂O₂

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Abstract

In this work, immobilization of horseradish peroxidase (HRP) on poly-L-histidine (P-L-His) modified reduced graphene oxide (RGO) was demonstrated. The HRP/P-L-His-RGO bionanocomposite film was prepared through layer-by-layer (LBL) assembly. Scanning electron microscopy, Fourier transform infrared spectroscopy, electrochemical impedance spectroscopy, and UV-Vis spectroscopy were adopted to monitor the uniformity of the LBL assembly of the HRP/P-L-His-RGO bionanocomposite film. The immobilized HRP exhibited excellent electrocatalytic activity towards the reduction of hydrogen peroxide (H₂O₂). The catalysis currents showed linear relationships over the H₂O₂ concentrations, ranging from 0.2 to 5000 μM, respectively. The detection limits (S/N = 3) of H₂O₂ were 0.05 μM, respectively. The apparent Michaelis-Menten constant (K_m) was calculated to be 1.2 mM. Moreover, the modified electrode displayed a rapid response (5 s) to H₂O₂ with good stability and reproducibility. Based on the HRP/P-L-His-RGO bionanocomposite film, a third-generation reagentless biosensor was constructed for the determination of H₂O₂.

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1. Introduction

The study of the electrochemical behaviour of Dopamine (DA) has received considerable attention from researchers and has been extensively studied since the 1950s.¹ DA is a crucial catecholamine neurotransmitter existing in the brain and the peripheral nervous system of mammals.^{2,3} It has been found that a loss of DA-containing neurons may occur in some serious diseases such as Parkinson's, schizophrenia, epilepsy, and HIV infection to name a few.^{4,5} However, DA has a close oxidation potential with some interference, which can cause poor selectivity and sensitivity in the electrochemical determination of DA in brain fluids if it coexists with many interfering compounds.⁶ It is necessary to develop a simple and rapid analytical method for the determination of DA that will overcome these problems.⁷ In the past few years, several methods have been applied for the analytical determination of DA in pharmaceutical and clinical preparations, including, spectrophotometry,⁸ chemiluminescence,⁹ high performance liquid chromatography,¹⁰ and electrochemical methods. However, most of the above methods have some

disadvantages, such as time consuming, high costs, low sensitivity and complicated pretreatments.¹¹ On the other hand, electrochemical methods have been actively developed owing to their high sensitivity, rapid response time, low cost and simplicity of use.¹² Recently, much attention has been focused on the development of biosensors due to their advantages of easy preparation, fast detection, rapidity and high sensitivity.¹³

Hydrogen peroxide (H_2O_2) is a well-known oxidizing agent, which plays an important role in the production of textiles, cleaning products, organic compounds, and food industry.¹⁴ Owing to the increased usage of H_2O_2 , it has been released into the environment in large amounts. Thus, developing a simple, rapid, reliable and highly sensitive method for the determination of trace amounts of H_2O_2 at physiological pH has become a necessity.¹⁵ Furthermore, H_2O_2 is also a vital intermediate that is formed in the oxidase enzymes and its accurate and rapid determination is a key factor for the development of efficient biosensors.¹⁶ In recent years there have been numerous attempts to develop enzyme-based electrochemical biosensors for the detection of H_2O_2 .

However efforts have been limited by the problems of spatial control during the deposition process. The poor stability, the high cost of enzymes, the complicated immobilization procedure and the critical operational conditions due to sensitivity of the enzyme to pH, temperature, humidity, ionic detergents and toxic chemicals. It has become necessary to develop a simple enzyme immobilization strategy to achieve a device capable of sensing H_2O_2 with high sensitivity.¹⁷ Many techniques such as chemiluminescence,¹⁸ spectrophotometry,¹⁹ titrimetry,²⁰ and electroanalytical methods^{21,22} have been employed for the detection and determination of H_2O_2 . Among these methods, the enzyme-based electro analytical methods are more attractive because of their low cost, operational simplicity, high sensitivity and real-time detection.²³ Based on the efficient catalyzing ability of horseradish peroxidase (HRP) to decompose H_2O_2 , various kinds of biosensors have been developed.²⁴

HRP is a common enzyme widely used in H_2O_2 biosensors due to its easy availability, and low costs.²⁵ HRP contains a heme prosthetic group, which catalyzes the oxidation of different substrates as a consequence of the protein active site with the resting state of the heme-iron Fe (II).²⁶ However, some of the methods for enzyme immobilization are prone to problems such as low reproducibility, poor enzymatic activity, and poor

spatial control during the deposition process. The immobilization of HRP on the electrode surface is difficult.²⁷ In recent years, immobilization of HRP through several methods including absorption,²⁸ cross linking,²⁹ layer by layer assembly,³⁰ sol-gel entrapment,³¹ and electro polymerization have been proposed. Among these methods, the physical entrapment of enzymes is needed for the production of a more stable sensor with a decrease in the loss of the enzyme during measurement as well as a simpler and less-time consuming fabrication process, while still ensuring high stability and sensitivity, and biocompatibility, which can affect the activity of the enzyme.³²

Specifically, graphene is a two-dimensional nanomaterial that has received intense attention due to its unique physical and chemical properties, such as low resistivity, outstanding mechanical properties and large specific surface area.^{33,34} With the advantages of graphene substrate and active metal nanoparticle components, a new class of hybrid materials such as graphene supported metal nanocomposite has been developed.³⁵ They have shown great prospects for extensive application in many advanced fields such as memory electronics, optoelectronic transistors,³⁶

catalysis,³⁷ energy storage,³⁸ and electrochemical sensors and biosensors.^{39,40}

To the best of our knowledge, there has been no report on the fabrication of HRP/P-L-His-RGO nanocomposites on a glassy carbon electrode (GCE) surface for the determination of DA and H₂O₂. In this work, a novel amperometric biosensor based on the immobilization of HRP on the electrochemically polymerized L-His modified RGO has been proposed. The amine groups in the P-L-His provide positive surface charges on P-L-His-RGO, enabling a favourable microenvironment for the immobilization of HRP. This simple immobilization strategy demonstrates the great potential of P-L-His-RGO composite films to be used as a biomatrix for proteins in new generation biosensors. The proposed HRP/P-L-His-RGO biosensor has excellent sensitivity, wide linearity, favourable stability, good reproducibility, low detection limit and low Michaelis-Menten constant. In addition, the HRP/P-L-His-RGO composite film was applied for the determination of H₂O₂ and DA in real samples.

2. Experimental Procedure

2.1. Reagents

The graphite powder (<20 μm) used in the experiments was purchased from Sigma-Aldrich. The L-

histidine, H₂O₂ (30%), Dopamine hydrochloride, NaH₂PO₄ and Na₂HPO₄ were supplied by Sigma-Aldrich. The horseradish peroxidase (E.C. 1.11.1.7 type-VI-A --S/5 mg, *Amoracia rusticana* source, 1840 U/mg) was provided by Sigma. The phosphate buffer solution (PBS) was prepared by mixing standard stock solutions of Na₂HPO₄ and NaH₂PO₄. The supporting electrolyte was a 0.05 M phosphate buffer solution (PBS). All experiments were performed using the PBS at room temperature, approximately 25 °C. All other chemicals were of analytical grade and were used as received. The water used throughout the experiment was purified by a Milli-Q water system (resistivity 18.25 MU cm, Millipore).

2.2. Apparatus

All electrochemical experiments were performed on a CHI 405A electrochemical work station with a conventional three-electrode system comprised of a platinum wire as the auxiliary electrode, an Ag/AgCl electrode as the reference electrode, and a modified glassy carbon electrode (GCE) as the working electrode. Rotating disc electrode (RDE) and amperometric measurements were performed using a CHI 405A electrochemical workstation with an analytical rotator AFMSRX (PINE Instruments, USA, Japan). Scanning electron microscopy (SEM) was performed using a

Hitachi S-3000 H instrument (Hitachi, Japan). Energy dispersive X-ray analysis (EDX) was recorded using the HORIBA EMAX X-ACT (Model 51-ADD0009). The UV absorption spectra were collected using a Model UV-3300 spectrophotometer (Hitachi, Japan). Fourier transform infrared (FT-IR) measurements were carried out using a Perkin Elmer spectrophotometer RXI. X-Ray photoelectron spectroscopy (XPS) was performed using a PHI 5000 Versa Probe equipped with an Al Kalpha X-ray source (1486.6 eV). Electrochemical impedance spectroscopy (EIS) measurements were carried out in the frequency range from 0.1 Hz to 1 MHz (IM6ex ZAHNER Kroanch, Germany).

2.3. Synthesis of GO

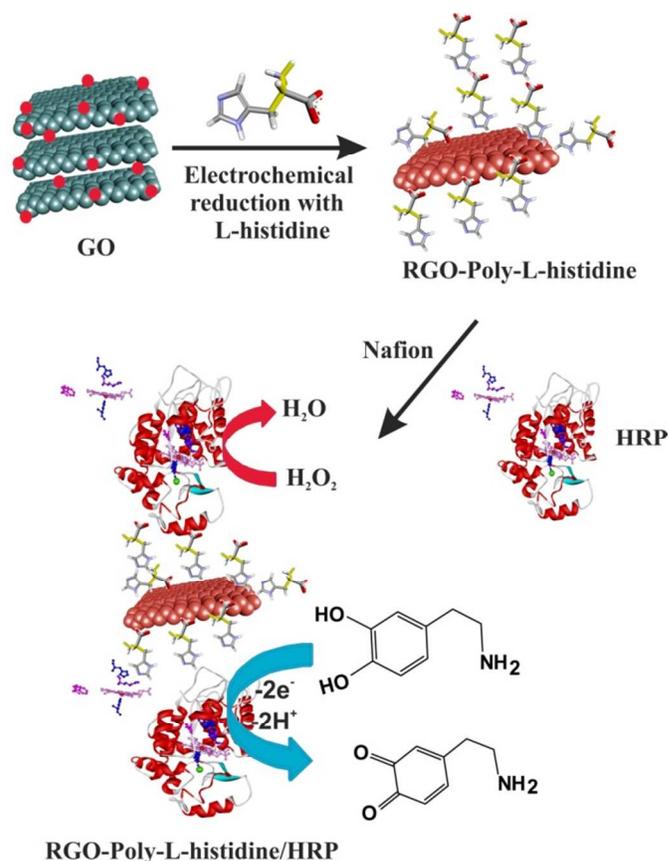
The graphene oxide (GO) used in this work was prepared using the modified Hummers method.⁴¹ Specifically, graphite powder (5.0 g) and 25 g of KMnO_4 were sequentially added to concentrated H_2SO_4 (150 mL) with constant stirring under ice-cold conditions. The resulting mixture was then continuously stirred at 35 °C for 5 h. Then it was diluted with 600 mL of deionized water and the temperature was kept under 50 °C. Subsequently, 30 mL of 30% H_2O_2 was added drop wise into this mixture. The obtained solid product was repeatedly filtered and washed with a 1 : 10 HCl aqueous solution (1000 mL) to remove the metal ions,

and then further washed with 1000 mL of deionized water until the solution pH became neutral (6.5). Finally, the obtained dark-yellow solid was dried in a vacuum oven at 40 °C for 48 h and then ground to fine powder. The as-obtained powder was dispersed in water and sonicated for 1 hour to obtain well-exfoliated GO.

2.4. Fabrication of HRP/P-L-His-RGO /GCE

Before modification, the GCE electrode (3mm) was first polished carefully with 0.05 μm of alumina slurry, and washed ultrasonically in water. Then it was electrochemically cleaned in 0.1 M H_2SO_4 by cyclic potential scanning between -0.2 and 1.2 V until a standard cyclic voltammogram was obtained. Finally, the GCE electrode was sonicated in water and absolute ethanol before being dried in a nitrogen stream before use. Schematic illustration 1 provides an overview of the fabrication process of the HRP biosensor. Prior to use, the as-synthesized GO (1mg) was dissolved by sonication in a mixture of 1mL water for 1 h. Then, 5 μL of the GO suspension was dropped onto the GCE surface, followed by evaporation of the solvent under an infrared lamp. The GO modified GCE was transferred to an electrochemical cell containing 0.05 M of the pH 5 PBS electrolyte with 5 mM of L-His. 30 cycles were recorded over the potential range of 0.0 to -1.5 V vs. Ag/AgCl as displayed in Fig. S1. During the first

cathodic potential scan, a large cathodic peak appears at -1.0 V with an onset potential of -0.75 V that is attributed to the reduction of the oxygen containing functional groups of GO. The cathodic current decreased continuously for each cycle, suggesting the effective reduction of functional groups of GO, the deposition of highly conducting material onto the GCE electrode and the electrochemical generation of RGO+P-L-His film on GCE. Then, a fresh HRP solution was prepared by dissolving 10 mg of HRP in 1 mL of 0.05 M, pH 7 PBS. 5 μ L of HRP solution was drop casted onto the surface of the RGO+P-L-His modified GCE electrode using a microsyringe and allowed to dry at 4 °C for 24 h. HRP was immobilized onto the RGO+P-L-His modified GCE through the electrostatic interactions between the positively charged P-L-His and negatively charged HRP molecules. After each modification step the electrode surface was rinsed with PBS. Finally, 5 μ L of 0.1% Nafion was coated on the modified electrode surface to form a protective film. The as-prepared enzyme modified electrode was dried at 4 °C until use.



Scheme. 1 Schematic illustration of the procedure used for the preparation of HRP/P-L-His-RGO film modified glassy carbon electrode.

3. Results and discussion

3.1 Surface studies and elemental analysis

Fig. 1 illustrates the SEM images of the as-prepared (A) P-L-His-RGO and (C) HRP/P-L-His-RGO bionanocomposites. The P-L-His-RGO contains flat, wrinkled sheets that are stacked with each other. Small plate and rod-like P-L-histidine molecules are also formed on the RGO sheets. Upon modification with HRP, several bead-like, globular structures are formed on the P-L-His coated RGO sheets, which can be

attributed to the electrostatic interaction between the negatively charged HRP and the positively charged P-L-His coated RGO. The EDX spectrum of P-L-His-RGO sheets (c) reveals the presence of C (50%), O (40%) and N (10%) related to the amino groups of histidine (See Fig. 1B). SEM results confirm the formation of HRP/P-L-His-RGO on the ITO surface, suggesting that the interaction between RGO/P-L-His and HRP indeed occurs, and may even influence the morphology of the dry films. Furthermore, the terminal functional groups of the positively charged P-L-His-RGO provide more active sites for the immobilization of the HRP biocomposite, leading to facile and efficient electron transfer.

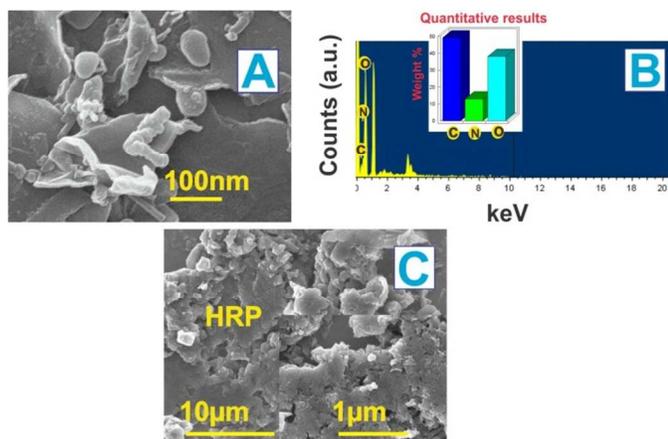


Fig. 1 SEM images of (A) P-L-His-RGO, (B) EDX spectra of P-L-His-RGO nanocomposite, (C) HRP/P-L-His-RGO bionanocomposite films.

Fig. 1

Fig 2(A&B) shows the TEM images of as prepared (A) RGO (B) P-L-His-RGO nanocomposite films. Fig 2A shows the as prepared reduced graphene was thin, transparent which consists of few layers of sheet like structures and B shows the P-L-His particles has been deposited as distorted shaped thin sheet like structure, which were uniformly dispersed in the RGO sheets. The presence of poly-L-His is clearly evident in the EDX analysis as shown in Fig 1 B.

The elemental composition of the P-L-His-RGO film after various stages of surface modification was ascertained by XPS (See Fig 2 C). It is well known that the bands centered at 284.8 and 531.0 eV are associated with C1s and O1s, respectively. It can be clearly seen that there is a significant decrease in the intensity of the O1s peaks of RGO compared with that of the P-L-His/RGO, which can be attributed to the loss of oxygen in GO as a result of electrochemical reduction (See Fig. 2D). A new peak observed at 404.2 eV corresponds to N1s, which can be assigned to the nitrogen in the amino groups of the P-L-His (See Fig. 2E). In addition, the new peak observed at 286 eV can be attributed to C-N of P-L-His.⁴² XPS results thus validate the electro deposition of the nitrogen containing P-L-His on RGO.

3.2 Characterization of HRP/P-L-His-RGO modified film.

Fig. S2 (a) shows the FT-IR spectra of (a) GO, and (b) P-L-His-RGO. FT-IR spectrum of GO shows the characteristic peaks of -OH ($\nu = 3440 \text{ cm}^{-1}$, 1398 cm^{-1} respectively), aromatic -C=C ($\nu = 1623 \text{ cm}^{-1}$), epoxy -C-O ($\nu = 1288 \text{ cm}^{-1}$), carboxyl -C=O and -C-O ($\nu = 1741 \text{ cm}^{-1}$ and alkoxy -C-O ($\nu = 1076 \text{ cm}^{-1}$) groups, revealing the existence of oxygen-containing functional groups on the GO nanosheets.⁴³ For P-L-His-RGO (Fig. S2 (b)), a new band appears at 1656 cm^{-1} , which corresponds to amide I (the -C=O stretch of P-L-His). The band at 1536 cm^{-1} is assigned to amide II (the -C-N stretch and -C-N-H deformation). Compared with RGO, the C=O stretch at 1623 cm^{-1} for carboxyl group disappeared, indicating the binding of P-L-His with the RGO surface.

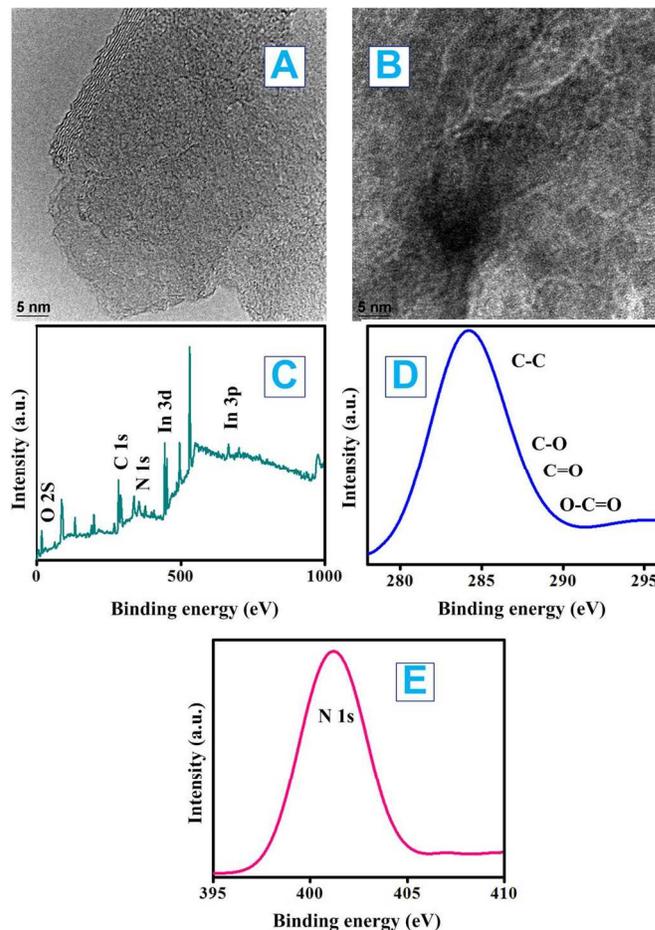


Fig.2 TEM images of (A) RGO (B) P-L-His-RGO nanocomposite films, and (C) XPS Survey of P-L-His-RGO. (D) C1s XPS spectra of P-L-His-RGO nanocomposite film. (E) N1s XPS spectra of P-L-His-RGO nanocomposite film.

To examine the nature of the interaction between the HRP and P-L-His-RGO we recorded the FT-IR spectra ((Fig.3A (a)). The two characteristic peaks centered at 1630 and 1567 cm^{-1} are assigned to the typical amide I and II absorption bands of HRP. The peak around 1082 cm^{-1} corresponds to the stretching vibration of C-O of HRP.⁴⁴ The biocompatibility of the

P-L-His-RGO film gives it great potential to be used as a matrix for the immobilization of HRP enzyme for biosensor fabrication. In addition, as can be seen from Fig.3A (b)), the positions of the amide I and II bands of HRP and that of the P-L-His-RGO film appeared at the same place, suggesting that the HRP retains its native structure in the HRP/P-L-His-RGO film. In short there has been no denaturation of HRP as it was stabilized through the electrostatic interactions.

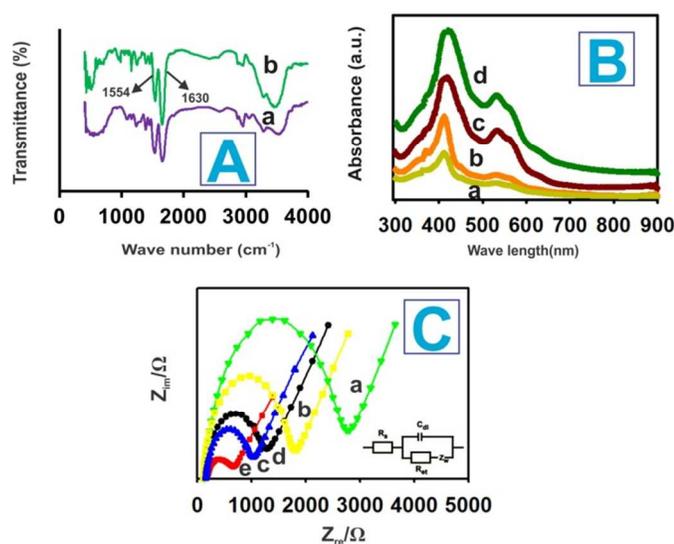


Fig. 3A FTIR spectra of (a) HRP and (b) HRP-P-L-His-RGO films. (B) UV-Vis spectra of (a) HRP and (b) HRP immobilized onto different bilayers (n = b-d) of the P-L-His-RGO films. (C) Nyquist plots for (a) HRP, (b) HRP/P-L-His, (c) RGO/GCE, (d) bare GCE, and (e) P-L-His-RGO/GCE electrodes for 5 mM [Fe(CN)₆]^{3-/4-} in a 0.05 M PBS (pH 7.0) solution

containing 0.05 M KCl. Amplitude: 5 mV, frequency: 0.1 Hz to 1 MHz. The bottom right inset shows Randles equivalence circuit for the above-mentioned electrodes.

The Soret absorption bands associated with the heme groups may provide information about the conformational integrity of the heme proteins and the possible denaturation of conformational change in the heme group region.^{45,46} As can be seen in Fig. 3(B), the Soret band of the HRP molecules appears at 408.8 nm (curve a). HRP immobilized onto different bilayers (n = b-d) of the P-L-His-RGO films exhibits the Soret band at 408.8 nm without any shifts, indicating that no significant denaturation of the protein occurred even after being immobilized onto the P-L-His-RGO composite with increased film thickness. Thus, HRP retained its native structure in the P-L-His-RGO film due to its good film-forming ability and excellent biocompatibility.

Fig. 2

3.3 Electrochemical characteristics and direct electrochemistry of HRP on the P-L-His-RGO modified film electrode

EIS measurement is a sensitive tool for studying the interface properties of surfaces after different steps of modification. The Nyquist plot of the EIS has a

semicircular portion at higher frequencies corresponding to the electron transfer limited process and a linear part at lower frequency ranges representing the diffusion limited process. Typical Nyquist plots of (a) HRP, (b) HRP/P-L-His, (c) RGO/ GCE, (d) bare GCE, and (e) P-L-His-RGO/GCE electrodes recorded in $[\text{Fe}(\text{CN})_6]^{3-/4-}$ solution over a frequency range of 0.1 Hz to 1 MHz are shown in Fig. 3C. The inset to Fig. 3C shows the equivalent circuit comprised of the ohmic resistance of the electrolyte solution (R_s), the Warburg impedance (Z_w), the capacitance (C), and the apparent charge transfer resistance (R_{et}). As depicted in Fig. 3C (d), the bare GCE electrode exhibits an R_{et} value of 1200 Ω . After modified with RGO, the R_{et} value of the RGO/GCE electrode decreased to 1120 Ω (See Fig. 3C(c)). With further modification of RGO/GCE with P-L-His decreased the R_{et} value to 730 Ω , likely due to the improved electron transport at the electrode interphase as a result of the electrostatic attraction between the positive charges in the amino groups of the L-His-RGO modified GCE electrode and the negative charged $[\text{Fe}(\text{CN})_6]^{3-/4-}$ redox probe (See Fig. 3C (e)). When the HRP enzyme was immobilized on the L-His-RGO modified GCE electrode, the R_{et} value increased to 1900 Ω (see Fig. 3C (b)), owing to the electrostatic repulsion between the negatively charged HRP and the negatively charged $[\text{Fe}(\text{CN})_6]^{3-/4-}$ redox electrochemical

probe. The EIS data indicates that HRP was successfully assembled on the P-L-His-RGO surface.

Fig. 4A shows the cyclic voltammetric responses of the (a) Bare GCE, (b) P-L-His-RGO and (c) HRP-L-His-RGO modified GCE electrodes in the 5mM $\text{K}_3[\text{Fe}(\text{CN})_6]$ containing 0.1 M KCl solution. Fig. 4A shows the cyclic voltammetry (CV) curve of the (a) The bare GCE witnessed a pair of well-defined redox peaks with anodic (E_{pa}) and cathodic (E_{pc}) peak potentials of 0.201 and 0.110 V, respectively, and redox peaks with a potential difference of 91 mV, and are attributed to the redox behavior of $[\text{Fe}(\text{CN})_6]^{3-/4-}$. Fig. 4A (b) shows an obvious increase in the redox peak current after the deposition of P-L-His onto the RGO/GCE surface. The positively charged P-L-His-RGO film modified GCE surface attracted the negatively charged $[\text{Fe}(\text{CN})_6]^{4-/3-}$ probe due to a stronger electrostatic interaction. The peak-to-peak separation (ΔE_p) was found to be 63 mV. When HRP was immobilized on the P-L-His-RGO modified GCE electrode, the peak currents decreased, while the peak-to-peak separation between the cathodic and the anodic waves increased. Fig. 4A (c) shows the CV curves of HRP-L-His-RGO modified GCE. There is a significant decrease in the peak currents, which can be attributed to the increase in the layer thickness and the electrostatic

repulsion between the negatively charged HRP enzyme on the L-His-RGO electrode surface and the negatively charged $\text{Fe}(\text{CN})_6^{4-/3-}$.

Fig. 4B shows the CVs of the HRP/L-His-RGO modified GCE electrode in PBS at different scan rates (10-100 mV s^{-1}). The anodic and cathodic currents increase linearly as the scan rate increases from 10 to 100 mV s^{-1} , revealing a surface confined redox process. The linear regression equation can be expressed as follows: $I_{pa} = 1.909v - 0.228$, $R = 0.9938$; $I_{pc} = -1.312x + 0.524$, $R = 0.9962$. The effect of the scan rate is shown in Fig.4B. The reduction (I_{pc}) and oxidation (I_{pa}) peak currents exhibit a linear relationship with the scan rate in the range from 10 to 100 mV s^{-1} . The obtained results indicate that HRP immobilized on the L-His-RGO bio composite modified GCE electrode undergoes a surface confined electrochemical process. The surface average concentration of electro active HRP immobilized on the L-His-RGO bio composite modified GCE electrode can be calculated from the charge integration of the cathodic peak in the cyclic voltammogram according to the formula, $Q = nFA\Gamma$,⁴⁶ where Q is the charge consumed in C, A is the electrode area (cm^2), n is the number of electrons transferred and F is the Faraday constant. The surface concentration (Γ) of electro active HRP/L-His-RGO modified GCE

electrode is calculated to be $2.14 \times 10^{-10} \text{ mol cm}^{-2}$, which is much larger than for the HRP immobilized onto the PTMSPA@GNR electrode ($4.51 \times 10^{-11} \text{ mol cm}^{-2}$),⁴⁷ HRP adsorbed on the graphite electrode ($4 \times 10^{-11} \text{ mol cm}^{-2}$),⁴⁸ and HRP immobilized onto the Silica-hydroxyapatite modified GCE electrode ($2.13 \times 10^{-11} \text{ mol cm}^{-2}$).⁴⁹ The obtained result indicates the high loading of HRP in multi-layers of the L-His-RGO biocomposite film.

The heterogeneous electron transfer rate constant (k_s) of HRP adsorbed on the L-His-RGO modified GCE electrode is estimated (using Laviron's equation) to be about 4.82 s^{-1} , where $\Delta E_p < 200 \text{ mV}$.⁵⁰

$$\log K_s = \alpha \log(1-\alpha) + (1-\alpha) \log \alpha - \log(RT/nFv) - \alpha(1-\alpha)nF\Delta E_p/2.3RT, \text{---(2)}$$

where R is the gas constant ($8.314 \text{ J mol}^{-1} \text{ K}^{-1}$), T is the room temperature (298.15 K) and ΔE_p is the peak separation of $\text{Fe}^{3+/2+}$ in the active sites of HRP. Here, the α value is assumed to be 0.5 and the number of electrons (n) transferred is considered to be 2. The K_s value of the HRP immobilized on the L-His-RGO bio composite modified GCE electrode is higher than that of 1.13 s^{-1} for the HRP/DNA film electrode,⁵¹ $3.52 \pm 0.34 \text{ s}^{-1}$ for the HRP/chitosan/sol-gel/CNT film electrode,⁵² HRP immobilized onto graphite electrode ($1.3 \pm 0.2 \text{ s}^{-1}$)

film electrode,⁵³ HRP/PTMSPA@GNR film electrode (3.23± 0.1 s⁻¹),⁴⁷ HRP immobilized onto (2×10⁻⁴ s⁻¹) pyrolytic graphite.⁵⁴ These findings suggest that the L-His-RGO biocomposite modified GCE electrode provides a good microenvironment and high conductivity that facilitates the direct electron transfer between the HRP and the electrode surface.

electrode, (c) HRP-P-L-His-RGO/GCE modified electrode. Fig. 4B CVs of the HRP/P-L-His-RGO modified GCEs in 0.05 M deoxygenated PBS (pH 7) containing 1 mM DA obtained at different scan rates (from inner to outer: 10 to 100 mV s⁻¹). Inset: (*I*) linear dependence of *I*_{pa} and *I*_{pc} on the scan rates.

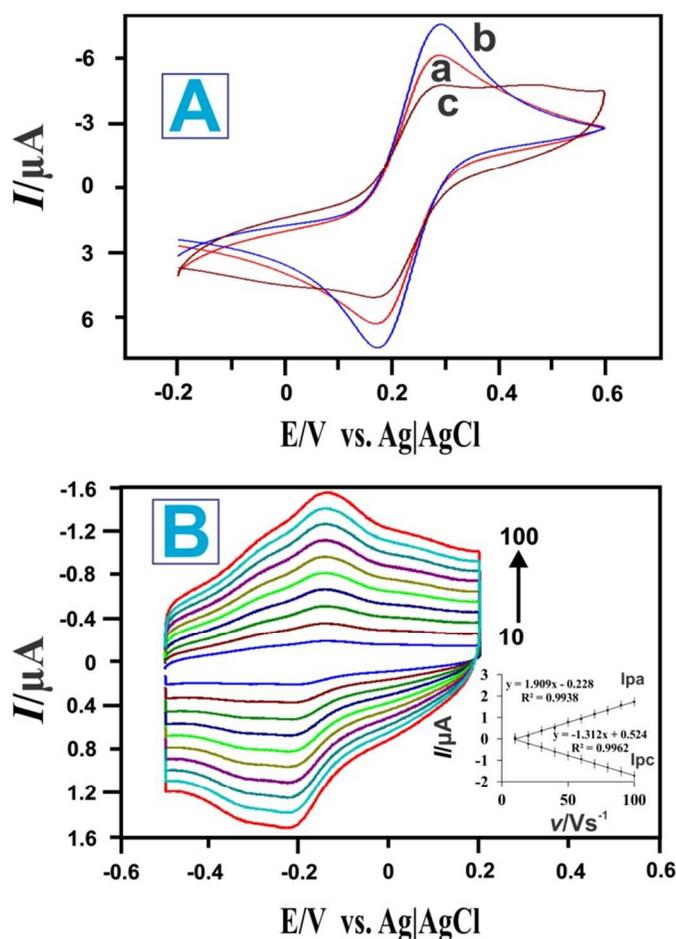


Fig. 4A CVs of the different modified electrodes recorded in the 0.05 M PBS (pH 7.0) solution containing 0.05 M KCl and 5mM [Fe(CN)₆]^{3-/4-}: (a) Bare GCE electrode, (b) P-L-His-RGO modified GCE

[Fig. 3]

3.4 Electro catalysis of the HRP/P-L-His-RGO modified film.

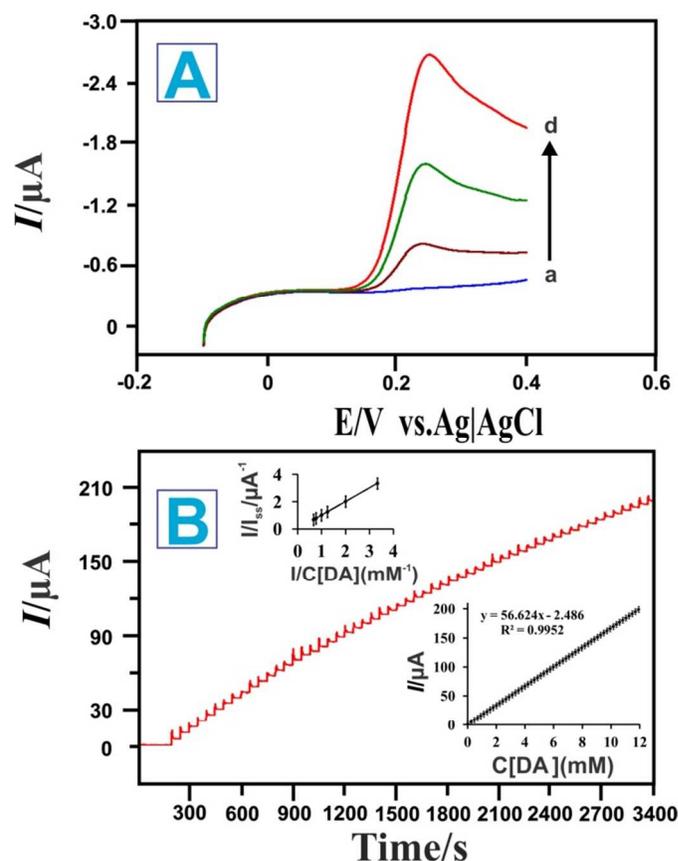


Fig. 5A Linear sweep voltammetry of the HRP-P-L-His-RGO modified GCEs electrode in PBS with DA concentrations of (a) blank, (b) 1, (c) 2, and (d) 3, mM.

(B) Amperometric response of the fabricated biosensor to successive additions of DA into the PBS solution (0.05 M, pH 7). Applied potential: +0.23 V. Inset: calibration curve and linear fitting curve between the current and the DA concentration.

The electrocatalytic performances of the (a) bare GCE, (b) RGO /GCE, (c) P-L-His-RGO/GCE, and (d) HRP/P-L-His-RGO bionanocomposite are evaluated by recording the linear sweep voltammetry in 0.05 M PBS (pH 7.0) containing 3mM DA at 50 mV s^{-1} . It can be seen that there is an electrocatalytic reduction towards DA on the bare GCE and RGO/GCE exhibited by the lower peak current and higher over potential than for the HRP/P-L-His-RGO bionanocomposite. Under the same conditions, the P-L-His-RGO/GCE modified electrode exhibits an E_{pa} of DA with a more negative potential with an enhanced peak current (I_{pa}) response greater than that of the bare GCE or RGO/GCE. This large decrease in oxidation over potential corresponds to the presence of high density arrays of P-L-His-RGO. Their electrocatalytic behaviour indicates that P-L-His-RGO greatly facilitates electron transfer with a negative potential shift. In addition, the catalytic activity of the HRP/P-L-His-RGO bionanocomposite (Fig. 7D) is evident from a decrease in the over potential as well as an increase in the response current for DA oxidation

compared with the other bare GCE, RGO/GCE, and P-L-His-RGO/GCE modified electrodes. There is a two-fold increase in the oxidation peak current (I_{pa}) of DA on the HRP/P-L-His-RGO bionanocomposite over that of the other electrodes (bare GCE, RGO/GCE, P-L-His-RGO/GCE). These experiments clearly indicate that the presence of P-L-His-RGO provided a favourable microenvironment that preserved HRP bioactivity and assisted the process of electron transfer between the protein and the GCE electrode surface. This enhanced the signal and reduced the over potential for DA electro-oxidation, both of which are very useful for practical applications for the fabrication of a sensitive and selective electrochemical DA sensor, with less risk of interference.

The electrochemical catalytic oxidation of DA at the HRP/P-L-His-RGO/GCE modified electrode was investigated by linear sweep voltammetry. When DA was added to the pH 7.0 PBS, a new oxidation peak appeared at about 0.2 V, as shown in Fig. 5A. The new peak originates from the oxidation of DA. The anodic peak current increased for each addition of DA into the buffer solution. The above experimental results demonstrate the good electro catalytic activity of the HRP/P-L-His-RGO/GCE modified electrode towards the oxidation of DA. Thus, the HRP/P-L-His-RGO/GCE

modified electrode showed greater sensitivity and faster catalytic activity towards DA. We only observed a 3% decrease in the initial response even after carrying out 50 continuous cyclic scans in the potential range of -0.1 to 0.4 V at a scan rate of 50 mV/s, showing its high stability. The mechanism of the catalytic Oxidation of the immobilized HRP on P-L-His-RGO/GCE modified electrode to DA was illustrates by the following schemes: ⁵⁵

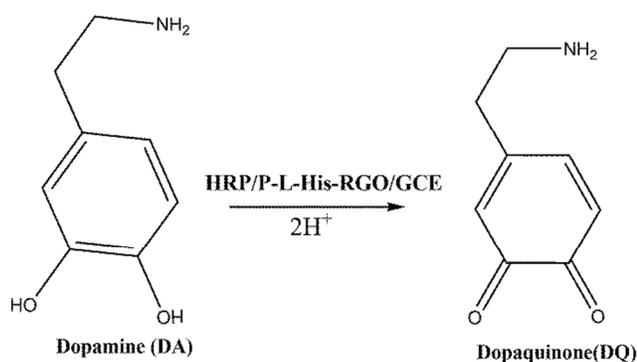


Fig. 5B illustrates a typical current-time plot obtained at the HRP/L-His-RGO modified rotating disc electrode for each successive addition of DA into the 0.05 M pH 7.0 PBS. The electrode potential was held at +0.23 V vs. Ag/AgCl. When the background current became stable, certain amount of DA was added into the solution and stirred at 1200 rpm. Fig. 5B shows that the amperometric peak current was linearly proportional to the concentration of DA. A stable and fast amperometric response could be observed for each successive injections of DA into the PBS solution. The

time required to reach the steady state was less than 5 s, which indicates the rapid response of this sensor for DA. The inset to Fig. 5B shows the calibration plot of the DA obtained over the concentration range of 0.2-12000 μM . The corresponding regression equations for the linear plot are expressed as follows: $I(\mu\text{A}) = 56.624 + 2.486 [\text{DA}]$, $R = 0.9952$. The sensitivity of the sensor to DA was determined to be $16.2\mu\text{A mM}^{-1}\text{cm}^{-2}$; while the detection limit was found to be $0.42 \mu\text{M}$ at the signal-to-noise ratio of 3. The further comparison of HRP/P-L-His-RGO/GCE modified electrode developed in this study with other modified electrodes for determinations of DA are listed in Table S1. Since, amperometry is more sensitive than cycle voltammetry, we used amperometric method to determine the affinity of DA at the HRP/P-L-His-RGO/GCE modified electrode.

$$\frac{1}{I_{ss}} = \frac{1}{I_{max}} + \frac{K_m}{I_{max}C} \quad (3)$$

As shown in Fig. 5B, the apparent Michaelis-Menten constant (K_m) can be obtained from the Lineweaver-Burk equation to evaluate the affinity of the immobilized enzyme towards DA.⁴⁵ In this work, the K_m of the HRP/P-L-His-RGO/GCE modified electrode

was determined to be 2.5 mM. Thus, the P-L-His-RGO modified electrode has an improved affinity for the immobilization of HRP to DA. Evidently, the small K_m value reveal the higher affinity of HRP/P-L-His-RGO/GCE for DA. Moreover, the selectivity of the DA biosensor was evaluated by using the amperometric method. The results show that the common interfering species including 100 μ M ascorbic acid, 100 μ M glucose, 100 μ M uric acid, and 100 μ M cysteine did not cause any observable interference with the amperometric response to 100 μ M DA at the HRP/P-L-His-RGO/GCE modified electrode. As shown in Fig. S5, the prepared HRP/P-L-His-RGO/GCE modified electrode allowed high sensitivity and selectivity for DA detection under normal physiological conditions.

3.5 Electrocatalytic properties towards the reduction of H_2O_2

In order to investigate the electrocatalytic activity of the different electrodes: (a) bare GCE, (b) RGO modified GCE, (c) P-L-His modified GCE, (d) P-L-His-RGO modified GCE, (e) HRP/P-L-His-RGO modified GCE towards the reduction of H_2O_2 , CV was carried out in the presence of H_2O_2 at the HRP/P-L-His-RGO bionanocomposite modified GCE electrodes. Fig.S4 shows the CVs of 3 mM H_2O_2 in a pH 7 PBS buffer solution for the unmodified (a) bare GCE, (b)

RGO modified GCE, (c) P-L-His modified GCE, (d) P-L-His-RGO modified GCE, and (e) HRP/P-L-His-RGO modified GCE electrodes. The results demonstrate the superior performance of the as-obtained HRP/P-L-His-RGO bionanocomposite. The HRP/P-L-His-RGO bionanocomposite modified GCE shows a greater background current than that of the other modified electrodes, which can be attributed to the large surface area of the P-L-His-RGO film. The nanostructures in this material give it a large surface area so that the composite P-L-His-RGO matrix has a higher capability for enzyme immobilization. Therefore, there was a two-fold increase in the reduction peak current of H_2O_2 at HRP/P-L-His-RGO modified GCE electrodes. These results show that the P-L-His-RGO modified GCE electrodes can improve the catalytic activity of HRP with more effective electrocatalytic activity toward H_2O_2 than the other modified electrodes.

Fig. 6A shows the CVs of the HRP/P-L-His-RGO/GCE recorded in the presence of different concentrations of H_2O_2 . The electrocatalytic responses of both the bare GCE and HRP/P-L-His-RGO/GCE towards the electrocatalytic reduction process of H_2O_2 have also been shown in Fig. 6A. However, no reduction peak was observed at the bare GCE. The intercalated HRP molecules produce good

electrocatalytic activity due to the interaction between the HRP and the P-L-His-RGO/GCE.

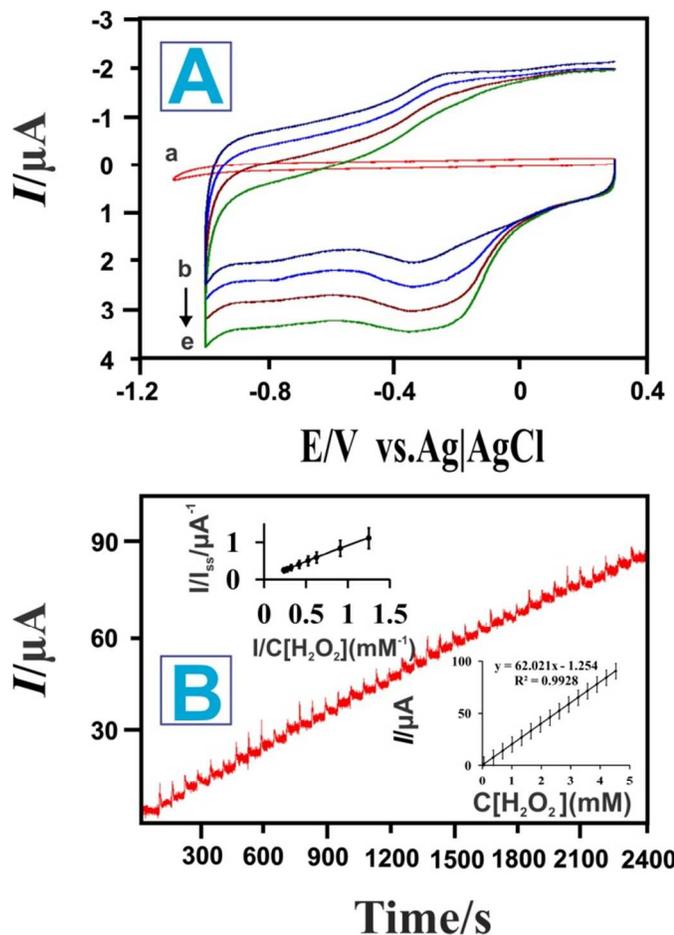
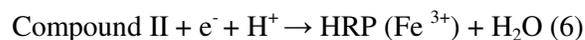
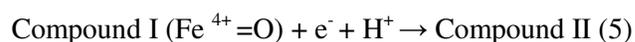
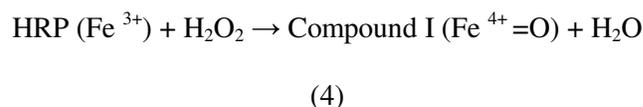


Fig. 6A Bioelectrocatalysis of the HRP/P-L-His-RGO modified GCEs towards H₂O₂ in PBS with a scan rate of 0.5 V s⁻¹ and H₂O₂ concentrations of (a) Bare GCE, (b) Blank, (c) 1, (d) 2, and (e) 3 mM. (B) Typical current-time response curves for the HRP/P-L-His-RGO modified GCEs after successive additions of H₂O₂ into pH 7 PBS. Applied potential: -0.2 V (vs. Ag|AgCl). Inset: calibration curve showing variation of the biosensor response with H₂O₂ concentration.

It is evident that the reduction current of H₂O₂ increased when increasing the H₂O₂ concentrations from 1 mM to 3 mM. These observations clearly indicate that the HRP/P-L-His-RGO/GCE exhibits excellent catalytic activity toward the reduction of H₂O₂. The electrocatalytic process can be expressed as follows:^{27, 46}



The overall reaction would be



We then investigated the amperometric response of the HRP/P-L-His-RGO/GCE modified rotating disc electrode towards each successive additions of H₂O₂ into 0.05 M pH 7.0 PBS at an applied potential of -0.2 V as shown in 6B. Upon addition of aliquots of H₂O₂ into the 0.05 M pH 7.0 PBS solution, the prepared HRP/P-L-His-RGO/GCE sensor responded rapidly and achieved the maximum steady-state current within 5 s. As shown in Fig. 6B a linear dependence of the peak current over the concentration of H₂O₂ was observed in the range of 0.2 to 5000 μM with a

sensitivity of $266.4 \text{ mA mM}^{-1} \text{ cm}^{-2}$. The detection limit was estimated to be $0.05 \text{ }\mu\text{M}$ at a signal-to-noise ratio of 3. The electrocatalytic efficiency of the proposed HRP/P-L-His-RGO/GCE electrode for the determination of H_2O_2 was compared with other kinds of HRP modified electrode as shown in Table 1. The apparent Michaelis-Menten constant (K_m), which is an indicator of the enzyme-substrate kinetics was calculated from the Lineweaver-Burk equation, where I_{ss} is the steady-state current attained after the addition of the substrate, I_{max} is the maximum current measured under saturated substrate conditions and C is the bulk concentration of the substrate. The K_m value was found to be 1.2 mM , which suggests an apparent affinity between immobilized HRP and L-His-RGO/GCE. Furthermore, the effect of possible interfering species such as uric acid (UA), glucose (Glu), and ascorbic acid (AA) on H_2O_2 detection was also investigated. The

amperometric responses of the HRP/L-His-RGO/GCE to the continuous addition of $100 \text{ }\mu\text{M}$ UA, $100 \text{ }\mu\text{M}$ AA, $100 \text{ }\mu\text{M}$ glucose and $100 \text{ }\mu\text{M}$ H_2O_2 at working potentials of -0.2 V in PBS (0.05 M , $\text{pH } 7.0$) were studied. As shown in Fig. S6 there is clear indication that the interference from UA or AA is negligible at an applied potential of -0.2 V .

Table 1

Comparison of the performance of the reported HRP modified electrodes for detection of H_2O_2 .

Modified electrodes	Linear range μM	Sensitivity $\text{mA mM}^{-1} \text{ cm}^{-2}$	LOD μM	Ref

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Au/graphene sheet/HRP/ chitosan	5-5130	-	1.7	[27]
Sonogel-carbon electrode /HRP	4.0-100	12.8	1.6	[45]
MWNT-silica-polyaniline -NW-HRP	1×10^{-6} - 1000	58.1	1×10^{-6}	[47]
GCE/Silica-hydroxyapatite /HRP	1-100	0.051	0.35	[56]
HRP-Nafion-screen printed electrode	5.98-35.36	0.143	0.48	[58]
ZnO/GNPs/Nafion/HRP	15-1100	-	9	[59]
Clay/HRP/Au/CS	39-3100	-	9	[60]
Au/CS/HRP	12.2-2430	0.013	6.3	[61]
CMCS/Au/HRP	5-1400	-	0.401	[62]
HRP-ZrO ₂ /Au	20-9450	-	2	[63]
HRP/P-L-His-RGO/ GCE	0.2 -5000	266.4	0.05	This work

Table 2

Determination of H₂O₂ in real eye drop samples obtained using the HRP/P-L-His-RGO modified glassy carbon electrode

Samples	Added [μM]	Found [μM]	Recovery rate (%)
1	5	4.8	101%
2	10	9.6	99.5%
3	20	18.2	98%

3.6 Real sample analysis

We investigated the response of the HRP/P-L-His-RGO/GCE biosensor for the measurement of H₂O₂ in real eye drop samples. The eye drop sample was obtained from a pharmacy. 1 mL of the sample was dissolved in the PBS (pH 7) prior to measurements. The analytical applicability of the HRP/P-L-His-RGO/GCE biosensor was evaluated by determining the recovery towards different concentrations of hydrogen peroxide using the standard addition method. The obtained results are given in Table 2, which indicate that this method is reliable and could be effectively used in future applications. We further explored the practicality of the proposed HRP/P-L-His-RGO/GCE biosensor for the determination of the DA in human urine and serum samples. Sampling was performed using the standard additional method. All the urine and serum samples (1 mL) were dissolved in PBS pH 7 prior to use. The

analytical results are shown in Table S2. These results indicate the effectiveness of this method for practical applications.

3.7 Stability and reproducibility of the sensor

The as-prepared HRP/P-L-His-RGO/GCE modified electrode showed an acceptable reproducibility with a relative standard deviation of 3.6 % for the current determined at an DA concentration of 50 μM . The reproducibility of different modified electrodes was also examined. Compared with the HRP/P-L-His-RGO/GCE, only RGO and P-L-His-RGO films have an RSD of 3.22% and 4.22%, respectively. In addition, the stability of the HRP/P-L-His-RGO/GCE modified electrode was evaluated after being stored in 0.05 M pH 7.0 PBS at 4 °C. The storage stability was evaluated by periodically checking its relative response current in PBS containing 50 μM of DA. After a storage period of 20 days, the HRP/P-L-His-RGO/GCE modified electrode showed an 8% loss of activity. Thus, the good repeatability, reproducibility and selectivity of the developed biosensor make it attractive for practical applications.

4. Conclusion

In summary, a biocompatible nanocomposite electrode based on HRP/P-L-His-RGO film was

proposed for the determination of H₂O₂ and DA. We have demonstrated the feasibility of this novel design for an enzymatic electrochemical H₂O₂ and DA sensor based on HRP/P-L-His-RGO/GCE modified electrode. The as-prepared HRP/P-L-His-RGO/GCE modified electrode exhibited very strong and sensitive amperometric responses to H₂O₂ and DA. Ease of fabrication, fast response and high sensitivity and long stability show the promise of the enzymatic H₂O₂ and DA sensor proposed in this study for the detection of H₂O₂ and DA in real clinical and waste water samples. The HRP/P-L-His-RGO/GCE modified electrode provides a promising platform for the design of reagent-less biosensors owing to with the advantages of simplicity, rapidity and ease of operation.

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