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ARTICLE TYPE

A Facile Naphthalene-Based Fluorescent Chemodosimeter for Mercury Ions in Aqueous Solution

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A facile naphthalene-based fluorescence "turn-on" chemodosimeter, 2-((2-(vinyloxy)naphthalen-1-yl)methylene) malononitrile (MS1), for rapid, selective and sensitive detection of Hg^{2+} by mercury-promoted hydrolysis of 10 vinylether group has been reported. The probe displayed a fast response time, and sensitive fluorescence response (100-fold fluorescence enhancement) to the detection of Hg^{2+} in aqueous solution.

Mercury, which is widely distributed in the environment such ¹⁵ as the air, soil, and water due to its use in batteries, dental amalgam, electrical apparatus, and industrial chemicals, is one of the most ubiquitous and poisonous heavy metals.¹ Mercury ions are not biodegradable, and hence can concentrate through the food chain in the tissues of fish and marine mammals.

²⁰ Excess mercury accumulation may induce strong damage to the central nervous system, various cognitive and motor disorders, and Minamata disease.² Due to the toxicity of Hg²⁺, the determination of mercury in biological and environmental samples is crucial both to the monitoring of environmental ²⁵ pollution and to the diagnosis of clinical disorders.

In the past several years, considerable efforts have been made to develop fluorescent chemosensors for Hg²⁺ based on the coordination of Hg²⁺ to heteroatom-based ligands, Hg²⁺ catalyzed desulfurization, and Hg²⁺ promoted hydrolysis of ³⁰ the vinyl ether group and β -alkynyl ether group.³ However,

³⁰ the vinyi ether group and *p*-arkynyi ether group. However, most of them still have limitations such as interference from other coexisting metal ions, poor water-solubility, and laborious synthesis processes expensive chemicals.⁴ Therefore, for practical applications, it is still desirable to ³⁵ develop simple Hg²⁺ sensors with good water solubility and

high selectivity and sensitivity.

Compared with the typically-developed chemosensors,⁵ fluorescent chemodosimeters, based-on highly specific chemical reactions between the dosimeters and the analytes,

- ⁴⁰ have received much research attention due to their relatively higher selectivity.⁶ Recently, Peng, Talukdar, Wu, and Ahn's groups have reported fluorescent chemodosimeters based on "deprotection-cyclization strategy" for the detection of fluoride ions,⁶ while the development of chemodosimeters for
- ⁴⁵ the specific determination of Hg²⁺ is drawing increasing research efforts. However, among the few available Hg²⁺ chemodosimeters reported,³ most employ the pH-sensitive fluorescein or 7-amino coumarin as the fluorophore and their



Scheme 1 Hydrolysis of MS1 by mercury ions.

pH-dependence may pose detection errors to the results. It is therefore strongly desirable to develop simple yet specific fluorescent chemodosimeters for Hg^{2+} that is immune to pH turbulence.

⁵⁵ It is known that Hg²⁺ catalyzes hydrolysis of vinylether to form the corresponding hydroxyl group.⁷ We proposed that the Hg²⁺ ion promoted hydrolysis of the vinyl enol ether group in **MS1** would generate the hydroxy intermediate, which will readily spontaneous cyclize to form a highly ⁶⁰ fluorescent chemodosimeter (Scheme 1).

Our research group is actively engaged in the development of novel selective and sensitive fluorescent probes for heavy



Scheme 2 Synthesis of MS1: (a) 1, 2-dibromoethane/K₂CO₃, acetone, 65 reflux, 3 h, 62%; (b) *t*-BuOK/DMSO, rt, 12 h, 46%; (c) CH₂(CN)₂/ piperidine, enthanol, rt, 1h, 48%.



Fig. 1 Absorption spectra of **MS1** (20 μ M) in PBS buffer solution (10 mM, pH 7.4, containing 1% CH₃CN) in the presence of different concentrations of Hg²⁺ (0-2.0 equiv.).

⁵ metal ions.⁸ Herein, we report the synthesis and properties of a deprotection-cyclization reaction based fluorescent chemodosimeter (**MS1**) that shows high selectivity and sensitivity for Hg²⁺.



¹⁰ Fig. 2 (a) Fluorescence spectra of MS1 (10 μ M) in PBS buffer solution (pH 7.4, containing 1% CH₃CN) in the presence of different concentrations of Hg²⁺ (0-50 μ M) ($\lambda_{ex} = 395$ nm). Inset: fluorescence intensity changes as a function of Hg²⁺ concentration. (b) Emission spectra of MS1 (10 μ M) in PBS buffer solution (pH 7.4, containing 1% ¹⁵ CH₃CN) in the presence of various metal loss ($\lambda_{ex} = 395$ nm, 5.0 eq. of H₂²⁺ C₃²⁺ C₃





Fig. 3 Time-dependent fluorescence intensity changes of **MS1** (10 μ M) ²⁰ upon addition of various concentration of Hg²⁺ (0, 0.2, 1.0, 5.0 equiv. each) in PBS buffer solution (pH 7.4, containing 1% CH₃CN) (λ_{ex} = 395 nm).

As shown in Scheme 2, **MS1** can be readily prepared in three convenient steps under facile conditions with high yield ²⁵ starting with commercially available 2-hydroxy-1naphthaldehyde. The product (**MS1**) was well characterized by ¹H, ¹³C NMR, and HR-MS (ESI[†]).

We firstly assessed the UV-vis spectroscopic properties of **MS1** in PBS buffer solution (10 mM, pH = 7.4, containing 1% 30 CH₃CN). **MS1** (20 μ M) displayed a moderate UV-vis absorption around 538 nm. Upon addition of Hg²⁺ (0-2 equiv.), the absorption band at 538 nm decreased and a new band at 399 nm appeared instantly with an isosbestic point at 439 nm, which is owing to the loss of vinyl enol ether group ³⁵ and the formation of cyclic compound (Fig. 1).

As expected, **MS1** alone is almost non-fluorescent ($\lambda_{ex} = 395 \text{ nm}$, $\Phi = 0.002$, Table S1, ESI[†]) in neutral aqueous solution (10 mM PBS buffer, pH 7.4, containing 1% CH₃CN), while the addition of increasing concentrations of Hg²⁺ ⁴⁰ gradually enhanced the fluorescent signal and *ca*. 100-fold increasing was observed when 5.0 equiv. of Hg²⁺ was added (Fig. 2a, Table S1, ESI[†]), which was attributed to the cleavage of vinyl enol group by mercury ion promoted hydrolysis reaction and the formation of a highly fluorescent ⁴⁵ cyclic compound (Scheme 1). Moreover, a blue-green fluorescent compound **5** have been isolated from **MS1**-Hg²⁺ system (ESI[†]), which was agreed well with the proposed mercury induced deprotection-cyclization mechanism.

Subsequently, the time-dependence of **MS1** fluorescence ⁵⁰ was also evaluated in the presence of different concentration of Hg^{2+} . The result shows that the fluorescence of all tested solutions remarkably increased to their maximum value within the 10 minutes. No changes in fluorescense were detected in the absence of Hg^{2+} (Fig. 3).

Further, the fluorescence titration of MS1 with various metal ions was conducted to examine the selectivity (Fig.2b). Much to our delight, the turn-on response of MS1 is highly specific for Hg²⁺ and no obvious change of fluorescent emission was observed when it is treated with Co²⁺, Cr³⁺, ⁶⁰ Cu²⁺, Fe²⁺, Fe³⁺, K⁺, Mg²⁺, Mn²⁺, Na⁺, Ni²⁺, Pb²⁺, Sn⁴⁺, Ag⁺, Ca²⁺, and Zn²⁺. It should be mentioned that MS1 still responds to Hg²⁺ sensitively even in the presence of other relevant



Fig. 4 Fluorescence responses of **MS1** to various metal ions. Black bars represent the addition of 5.0 equiv. of Hg^{2+} and 10.0 equiv. of the other appropriate metal ion to a 10 μ M solution of **MS1**. Red bars represent the s addition of 5.0 equiv. of Hg^{2+} to the solutions containing **MS1** (10 μ M) and the appropriated metals (10.0 equiv.).

competing ions (Fig. 4). Therefore, these results suggest that **MS1** displays high selectivity toward Hg^{2+} in neutral aqueous solution.

- ¹⁰ Moreover, the Hg²⁺-sensing ability of **MS1** at a wide range of pH values was investigated. As depicted in Fig. 5, **MS1** alone is inert to pH in the range of 4.0-11.0. But in the presence of Hg²⁺, **MS1** have no fluorescence response in the highly basic environment (pH \ge 9) due to the reaction rate of
- ¹⁵ mercury ion-promoted hydrolysis of vinyl enol ether becomes slow at high pH value.⁶ However, satisfactory Hg²⁺-sensing abilities were exhibited in the range of pH from 4.0 to 8.0, indicating that **MS1** could be used in neutral natural systems, or a mildly acidic or basic environment.
- For practical purposes, the detection limit of **MS1** for the analysis of Hg^{2+} was also an important parameter. The fluorescence titration curve revealed that the fluorescence intensity of **MS1** at 470 nm increased linearly with the amount of Hg^{2+} in the range of 0-5.0 μ M ($R^2 = 0.994$) (Fig. S1, ESI†).
- ²⁵ Thus, the detection limit of **MS1** for Hg^{2+} was calculated to be 4.31×10^{-8} M (Hg content = 8.8 ppb), which reveals the high sensitivity for the analysis of the mercury ions.



³⁰ Fig. 5 Effect of the pH on the fluorescence emission of MS1 (10 μ M) alone and MS1 (10 μ M) reacted with Hg²⁺ (3.0 equiv.).

In conclusion, we have successfully developed a simple naphthalene-based fluorescense probe for Hg²⁺ based on mercury triggered cleavage reaction under mild conditions. ³⁵ The probe has the unique advantage of easy-preparation, good water solubility, and excellent selectivity and sensitivity response towards Hg²⁺ in aqueous solution. We anticipate that the experimental results of this study will inspire the future design of metal-ion sensors in water for a variety of chemical ⁴⁰ and biological applications.

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Notes and references

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† Electronic Supplementary Information (ESI) available: Experimental 55 details, synthetic details of MS1, additional spectroscopic data, and copies of NMR spectra. See DOI: 10.1039/b000000x/

(a) L. Magos, *Met. Ions Biol. Syst.*, 1997, 34, 321-370; (b) M. F. Wolfe, S. Schwarzbach and R. A. Sulaiman, *Environ. Toxicol. Chem.*, 1998, 17, 146-160; (c) P. B. Tchounwou, W. K. Ayensu, N. Ninashvili and D. Sutton, *Environ. Toxicol.*, 2003, 18, 149-175; (d) P. Grandjean, P. Weihe, R. F. White and F. Debes, *Environ. Res.*, 1998, 77, 165-172.

(a) C. R. Baum, *Curr. Opin. Pediatr.*, 1999, 11, 265-268; (b) E. K.
 Silbergeld, I. A. Silva and J. F. Nyland, *Toxicol. Appl. Pharmacol.*, 2005, 207, S282-S292; (c) R. K. Zalups and S. Ahmad, *J. Am. Soc. Nephrol.*, 2004, 15, 2023-2031; (d) Z. Zhang, X. Guo, X. Qian, Z. Lu and F. Liu, *Kidney Int.*, 2004, 66, 2279-2282; (e) J. Huang, X. Ma, B. Liu, L. Cai, Q. Li, Y. Zhang, K. Jiang and S. Yin, *J. Lumin.*, 2013, 141, 130-140.

(a) M. Y. Berezin and S. Achilefu, Chem. Rev., 2010, 110, 2641-2684; (b) K. P. Carter, A. M. Young and A. E. Palmer, Chem. Rev., 2014, 114, 4564-4601; (c) E. M. Nolan and S. J. Lippard, Chem. Rev., 2008, 108, 3443-3480; (d) X. Chen, T. Pradhan, F.

Wang, J. S. Kim and J. Yoon, *Chem. Rev.*, 2012, **112**, 1910-1956;
(e) M. Kaur and D. H. Choi, *Chem. Soc. Rev.*, 2015, **44**, 58-77; (f)
Z. Guo, S. Park, J. Yoon and I. Shin, *Chem. Soc. Rev.*, 2014, **43**, 16-29; (g) L. Yuan, W. Lin, K. Zheng, L. He and W. Huang, *Chem. Soc. Rev.*, 2013, **42**, 622-661; (h) J. Fan, M. Hu, P. Zhan and X.

Peng, *Chem. Soc. Rev.*, 2013, **42**, 29-43; (*i*) J. Du, M. Hu, J. Fan and X. Peng, *Chem. Soc. Rev.*, 2012, **41**, 4511-4535; (*j*) W. Xuan, C. Chen, Y. Cao, W. He, W. Jiang, K. Liu and W. Wang, *Chem. Commun.*, 2012, **48**, 7292-7294; (*k*) M. Vedamalai and S. P. Wu, *Org. Biomol. Chem.*, 2012, **10**, 5410-5416; (*l*) J. Liu, Y. Q. Sun, P. Wang, J. Zhang and W. Guo, *Analyst*, 2013, **138**, 2654-2660; (*m*) F. Song, S. Watanabe, P. E. Floreancig, and K. Koide, *J. Am. Chem.*

Song, S. Walanabe, P. E. Floreancig, and K. Kolde, J. Am. Chem.
 Soc., 2008, 130, 16460-16461; (n) H. Jiang, J. Jiang, J. Cheng, W.
 Dou, X. Tang, L. Yang, W. Liu and D. Bai, New J.Chem., 2014, 38, 109-114; (o) L. Chen, L. Yang, H. Li, Y. Gao, D. Deng, Y. Wu and L. Ma, Inorg. Chem., 2011, 50, 10028-10032; (p) M. Saleem, R.
 Abdullah, A. Ali, B. J. Park, E. H. Choi, I. S. Hong and K. H. Lee, Anal. Methods, 2014, 6, 3588-3597.

 (a) M. Tian and H. Ihmels, *Chem. Commun.*, 2009, 3175-3177; (b)
 N. Kumari, N. Dey, S. Jha and S. Bhattachary, *ACS Appl. Mater. Interfaces*, 2013, 5, 2438-2445; (c) S. Madhu, R. Kalaiyarasi, S. K. Basu, S. Jadhav and M. Ravikanth, *J. Mater. Chem. C*, 2014, 2,

95

2534-2544; (d) M. Tian, L. Liu, Y. Li, R. Hu, T. Liu, H. Liu, S. Wang and Y. Li, *Chem. Commun.*, 2014, **50**, 2055-2057.

- (a) P. Dinake, P. E. Prokhorova, V. S. Talanov, R. J. Butcher and G. G. Talanova, *Tetrahedron Lett.*, 2010, **51**, 5016-5019; (b) J. H. Kim, J. Y. Noh, I. H. Hwang, J. J. Lee and C. Kim, *Tetrahedron Lett.*, 2013, **54**, 4001-4005; (c) P. Srivastava, R. Ali, S. S. Razi, M. Shahid, S. Patnaik and A. Misra, *Tetrahedron Lett.*, 2013, **54**, 3688-3693; (d) L. N. Neupane, J. Y. Park, J. H. Park and K. H. Lee, *Org. Lett.*, 2013, **15**, 254-257; (e) M. Kumar, N. Kumar, V. Bhalla, H.
- Singh, P. R. Sharma and T. Kaur, Org. Lett., 2011, 13, 1422-1425; (f) M. Kumar, S. I. Reja and V. Bhalla, Org. Lett., 2012, 14, 6084-6087.
- 6 (a) Y. Peng, Y. M. Dong, M. Dong and Y. W. Wang, J. Org. Chem., 2012, 77, 9072-9080; (b) A. Roy, D. Kand, T. Saha and P. Talukdar,
- ¹⁵ Chem. Commun., 2014, **50**, 5510-5513; (c) D. Kim, S. Singha, T. Wang, E. Seo, J. H. Lee, S. J. Lee, K. H. Kim and K. H. Ahn, Chem. Commun., 2012, **48**, 10243-10245; (d) J. T. Yeh, P. Venkatesan and S. P. Wu, New J. Chem., 2014, **38**, 6198-6204.

7 (a) M. Santra, B. Roy and K. H. Ahn, Org. Lett., 2011, 13, 3422-

- 3425; (b) J. Jiang, W. Liu, J. Cheng, L. Yang, H. Jiang, D. bai and
 W. Liu, Chem. Commun., 2012, 48, 8371-8373; (c) Y. S. Cho, K. H.
 Ahn, Tetrahedron Lett., 2010, 51, 3852-3854; (d) S. Zhang, J. Geng,
 W. Yang and X. Zhang, RSC Adv., 2014, 4, 12596-12600.
- 8 (a) K. Wu, Y. Gao, Z. Yu, F. Yu, J. Jiang, J. Guo and Y. Han, Anal.
 25 Methods, 2014, 6, 3560-3563; (b) X. Li, Y. Gong, K. Wu, S. H. Liang, J. Cao, B. Yang, Y. Hu and Y. Han, RSC Adv., 2014, 4, 36106-36109; (c) X. Li, C. Yang, K. Wu, Y. Hu, Y. Han and S. H. Liang, Theranostics, 2014, 4, 1233-1238; (d) J. Liu, K. Wu, S. Li, T. Song, Y. Han and X. Li, Dalton Trans., 2013, 42, 3854-3859; (e)
- ³⁰ J. Liu, K. Wu, X. Li, Y. Han and M. Xia, *RSC Adv.*, 2013, **3**, 8924-8928.