

Analytical Methods

Accepted Manuscript



This is an *Accepted Manuscript*, which has been through the Royal Society of Chemistry peer review process and has been accepted for publication.

Accepted Manuscripts are published online shortly after acceptance, before technical editing, formatting and proof reading. Using this free service, authors can make their results available to the community, in citable form, before we publish the edited article. We will replace this *Accepted Manuscript* with the edited and formatted *Advance Article* as soon as it is available.

You can find more information about *Accepted Manuscripts* in the [Information for Authors](#).

Please note that technical editing may introduce minor changes to the text and/or graphics, which may alter content. The journal's standard [Terms & Conditions](#) and the [Ethical guidelines](#) still apply. In no event shall the Royal Society of Chemistry be held responsible for any errors or omissions in this *Accepted Manuscript* or any consequences arising from the use of any information it contains.

1

2

3

4

5

6

7

8

9

10

11

12

13

14

15

16

17

18

19

20

21

22

23

24

**Direct determination of cadmium in geological samples by
slurry sampling electrothermal atomic absorption
spectrometry**

Hao Cui ^a, Wei Guo ^{a,*}, Mengting Cheng ^a, Ping Zhang ^b, Lanlan Jin ^a,

Qinghai Guo ^a, Shenghong Hu ^{a,**}

^a *State Key Laboratory of Biogeology and Environmental Geology, China University*

of Geosciences, Wuhan, 430074, P. R. China

^b *Beijing CKC, PerkinElmer, Inc. 100015, China*

*Corresponding Author. Tel and Fax: +86-27-67883495;

Email address: Wei.Guo@cug.edu.cn (Wei Guo); Shhu@cug.edu.cn (Shenghong Hu).

† Electronic supplementary information (ESI) available, see DOI: 10.1039/c5ay

XXXXX.

1
2
3
4 21 **ABSTRACT**
5
6

7 22 Cadmium (Cd) is an important element for the assessment of environmental pollution.
8
9
10 23 An accurate and high throughput method involving slurry sampling electrothermal
11
12 24 atomic absorption spectrometry (ETAAS) was developed to detect trace Cd content in
13
14
15 25 geological samples. After sonication of the sample with 0.5% v/v HNO₃ and 0.5% v/v
16
17
18 26 Triton X-100 solution, the slurry was directly introduced into a graphite atomizer and
19
20 27 detected by AAS. This simple method was shown to take 80% less time, and resulted
21
22
23 28 in 90% less reagent waste than the conventional acid digestion method. Under
24
25
26 29 optimized conditions (sample size $\leq 75\mu\text{m}$, sonication time: 20 min, pyrolysis
27
28 30 temperature: 450 °C, and atomization temperature: 1600 °C), the characteristic mass
29
30
31 31 and limit of detection for Cd were 0.8 pg and 0.002 $\mu\text{g g}^{-1}$, respectively. The proposed
32
33
34 32 method was applied to the determination of Cd in 86 soil, sediment, and rock standard
35
36
37 33 reference materials (SRMs). The results for 78 of these materials were in good
38
39
40 34 agreement with the reference values. The Cd levels in five limestone SRMs (GUI-1,
41
42 35 GUI-2, DIAN-1, DIAN-2, and DIAN-3) and one clay SRM (GBW03103) were
43
44
45 36 reported for the first time. The proposed method shows great potential for the direct
46
47
48 37 determination of trace Cd in various geological samples.

49
50 38
51
52
53
54
55
56
57
58
59
60

39 Introduction

40 The assessment of cadmium (Cd) in the environment or geological is important in the
41 study of environmental pollution because of the toxic effect of Cd on human health
42 and the fact that Cd-contaminated crops can grow from polluted soil.¹⁻³ The
43 abundance of Cd in upper continental crust is only $0.09 \mu\text{g g}^{-1}$. Therefore, the
44 concentration of Cd is at trace or ultra-trace levels in natural soils, sediments, and
45 rocks (except for Cd mineral deposits).⁴⁻⁶ Consequently, highly sensitive and
46 high-throughput analytical methods are needed for Cd determination of solid
47 environmental samples. In addition, accurate determination of Cd is imperative for Cd
48 isotopic analysis, which is vital for the study of cosmology, the tracing of
49 anthropogenic sources, the study of micronutrient cycling, and oceanography.⁷

50 Two main techniques have been proposed that can be used for the accurate
51 determination of trace Cd in soils, sediments, or rocks in typical geological
52 laboratories. One method is inductively coupled plasma mass spectrometry (ICP-MS),
53 which has high sensitivity, low detection limits, and is capable of simultaneous
54 multi-element analysis.⁸⁻¹⁹ However, isobaric and/or polyatomic interferences derived
55 from the presence of high concentrations of matrix elements are significant limitations
56 of this technology.²⁰ All isotopes of Cd are subject to interferences from various
57 isobaric ions and/or oxide or hydroxide ions of Pd, Sn, In, Zr, Mo, Ru, Nb, and Y.²¹
58 Complicated separation procedures, such as chromatography, precipitation, extraction,

1
2
3
4 59 and volatile species generation, are usually required to eliminate these interferences
5
6 22-25. Isotope dilution ICP-MS (ID ICP-MS) ^{26,27} and dynamic reaction cell ICP-MS
7
8
9 61 (DRC ICP-MS) ^{28,29} have been demonstrated as valid methods to accurately detect
10
11 62 trace Cd in environmental or geological samples. However, the high cost and complex
12
13
14 63 operating procedures associated with these methods prevent their routine use in a
15
16
17 64 high-throughput geological laboratory. The other method is electrothermal atomic
18
19 65 absorption spectrometry (ETAAS), which has emerged as a better technique than
20
21
22 66 ICP-MS for trace Cd determination as it is less subject to interference, more
23
24
25 67 economical, relatively simple, and is capable of direct determination of Cd in complex
26
27 68 matrices. ³⁰⁻³³

29
30 69 Sample preparation is the most time-consuming step for solid environmental
31
32
33 70 sample analysis, and microwave-assisted or high-pressure closed acid digestion has
34
35
36 71 been extensively employed for sample dissolution which avoids analyte losses and
37
38 72 contamination. ³⁴ The main drawbacks are the high cost of vessels and ovens, the risk
39
40
41 73 of explosions, low sample throughput, and the long time required for the cooling step.
42
43 74 ³⁵ Direct solid sampling ETAAS (DSS-ETAAS) may be used to simplify sample
44
45
46 75 preparation procedures, and to avoid sample contamination. ³⁶⁻⁴¹ However, the
47
48
49 76 DSS-ETAAS method was not widely accepted until recently, owing to the difficulty in
50
51
52 77 handling and introducing small sample masses, the high imprecision of the results due
53
54 78 to the heterogeneity of some natural samples, the difficulty in calibration due to the
55
56
57
58
59
60

1
2
3
4 79 requirement of solid standards with similar matrix composition and structure, the
5
6 80 limited linear working range of AAS, and the difficulty in diluting solid samples.³⁵
7
8
9 81 Another attractive alternative is the use of slurry sample introduction in ETAAS,
10
11 82 which combines the advantages of liquid and solid sampling and avoids many
12
13 83 problems associated with direct solid sampling. Here, simple aqueous standards are
14
15 84 used instead of solid-sample matrix calibration, the sample concentration can easily
16
17 85 be changed by dilution, it can avoid weighted errors and inhomogeneity problems are
18
19 86 eliminated, and the analyte concentration falls within the linear range. Furthermore,
20
21 87 slurry can be directly sampled using the conventional autosampler used in ETAAS,
22
23 88 avoiding the need for specialized expensive equipment such as a microbalance and
24
25 89 solid sampling accessories.^{42, 43} Slurry sampling ETAAS techniques have been
26
27 90 proposed for trace Cd determination in various biological and food materials, such as
28
29 91 plant, honey, wheat, and rice.^{32, 33, 44, 45} However, limited literature is available
30
31 92 regarding Cd determination in environmental soil or sediment samples.^{46, 47} Most
32
33 93 literature relates to acid digestion procedures but not direct slurry sampling. Therefore,
34
35 94 to investigate whether the slurry sampling method is effective for non-soft matrices,
36
37 95 such as soil, sediment, and rock, it is necessary to carefully optimize various operating
38
39 96 conditions and comprehensively evaluate the accuracy and precision of this method
40
41 97 for Cd determination.
42
43
44
45
46
47
48
49
50
51
52
53
54
55
56
57
58
59
60

1
2
3
4 98 The aim of this work was to develop an easy, accurate, reliable, green, and
5
6 99 routine method for determination of trace Cd in solid geological samples using slurry
7
8
9 100 sampling ETAAS. The optimization of the technique and its analytical performance,
10
11 101 as well as its application to the determination of trace Cd in 86 soil, sediment, and
12
13
14 102 rock standard reference materials (SRMs) are discussed in detail.
15
16
17 103

19 104 **Experimental**

22 105 **Instrumentation**

23
24
25
26 106 All measurements were performed with a PinAAcle™ 900T atomic absorption
27
28 107 spectrometer (PerkinElmer, Inc., Shelton, USA), which was equipped with a
29
30
31 108 longitudinal AC Zeeman background correction system, a transversely heated graphite
32
33 109 atomizer (THGA), PerkinElmer Cd electrodeless discharge lamps (EDL), and an AS
34
35
36 110 900 autosampler. A TubeView™ color furnace camera was used to monitor the
37
38
39 111 process of slurry sampling in the graphite tube (Fig. S1, see ESI†).

42 112 **Materials**

43
44
45
46 113 De-ionized water used for the preparation of all blank, standard, and sample solutions
47
48 114 was obtained from a water purification system (Merck Millipore, Fontenay-sous-Bois,
49
50
51 115 France). The Cd standard solution (1000 $\mu\text{g L}^{-1}$) was purchased from the National
52
53
54 116 Center for Analysis and Testing of Steel Materials, China. Nitric acid (65–70%, w/w,
55
56
57
58
59
60

1
2
3
4 117 99.999%) and Triton X-100 (> 99%) were purchased from Alfa Aesar Ltd. (Tianjin,
5
6 118 China). Eighty-six geological SRMs were used to evaluate the accuracy of the
7
8
9 119 proposed method, which covers 26 soil SRMs, 25 sediment SRMs, and 35 rock SRMs.
10
11 120 Five of these SRMs were purchased from the United States Geological Survey (USGS)
12
13
14 121 and the remainder from the Institute of Geophysical and Geochemical Exploration of
15
16
17 122 China (IGGE). A detailed description of these SRMs is given in Table S1 (see ESI†).

123 **Slurry sampling procedures**

124 Slurries were prepared by weighing 0.1000 to 0.3500 g of the sample in a 50 mL
125 conical polypropylene tube and diluting to 50 mL with 0.5% v/v HNO₃ containing
126 0.6% v/v Triton X-100. After homogenizing in an ultrasonic bath (Branson, USA) for
127 20 min, the uniform slurries were transferred into acid-cleaned propylene autosampler
128 cups. Then, 20 μL of slurry was taken up and delivered to the graphite tube for
129 ETAAS analysis.

130 For comparison, a conventional digestion method employing closed acid digestion
131 with a mixture of HF + HNO₃ was also used (see ESI Table 2S†)⁴⁸⁻⁵⁰

132

133 **Results and discussion**

134 **Optimization of slurry sampling ETAAS conditions**

135 To obtain accurate results with the slurry sampling ETAAS techniques, the slurry

1
2
3
4 136 sampling procedures and graphite furnace heating parameters were optimized in detail.
5
6 137 AGV-2, which is an andesite SRM provided by USGS with a certified Cd content of
7
8
9 138 $0.069 \mu\text{g g}^{-1}$, was used to optimize all of the parameters. As shown in Fig. 1a, the
10
11 139 recovery of Cd from the AGV-2 with HNO_3 was investigated by varying the HNO_3
12
13 140 concentrations from 0.1% and 0.6% (v/v), while other parameters, such as particle
14
15 141 size (62 μm), sonication duration (20 min), Triton X-100 concentration (0.5% v/v),
16
17 142 and slurry concentration (0.5% m/v), pyrolysis temperature (450 $^\circ\text{C}$), and atomization
18
19 143 temperature (1600 $^\circ\text{C}$), were kept constant. As is evident from the figure, the best
20
21 144 recovery of Cd is obtained when 0.5% v/v HNO_3 is used.
22
23
24
25
26

27 145 Sonication and the addition of the stabilizing medium Triton X-100 were used to
28
29 146 improve the homogeneity of the sample slurries, and similar optimization procedures
30
31 147 as for HNO_3 concentration were performed for Triton X-100 concentration and
32
33 148 sonication time, as shown in Fig. 1b and Fig. 1c. Under the optimized Triton X-100
34
35 149 concentration (0.5% v/v) and sonication time (20 min), sample particles formed
36
37 150 homogenous and stable slurries (Fig. S2, see ESI†).
38
39
40
41
42

43 151 It is important to optimize particle size in order to attain quantitative recoveries
44
45 152 with the heat extraction procedure. Different particle sizes ranges of a soil sample
46
47 153 with a Cd content of $0.230 \pm 0.012 \mu\text{g g}^{-1}$ (determined by the closed acid digestion
48
49 154 ETAAS method) were studied. Particle sizes of 83 μm , 75 μm , 62 μm , and 38 μm
50
51 155 were investigated, as shown in Fig. 1d, and quantitative recoveries were attained for
52
53
54
55
56
57
58
59
60

1
2
3
4 156 particles both 38 μm (97%) and 62 μm (98%) in size. 10 min of sample grinding in a
5
6 157 ball mill is required to obtain a particle size of 38 μm . However, only 4 min is
7
8
9 158 required to obtain a particle size of 62 μm . Therefore, 62 μm particle sizes (240 mesh)
10
11
12 159 was employed for the rest of this work.

14 160 The concentration of slurry suspensions is an important factor, which can affect
15
16
17 161 the measurement precision. Here, a number of suspensions with different percentages
18
19
20 162 of SRM AGV-2 were prepared, and the precision (RSD, %) of the integrated
21
22 163 absorbance results was explored. Fig. 1e shows the relationship between precision and
23
24
25 164 slurry concentration in the range 0.1 – 1.0% (m/v). The optimum concentration ranges
26
27 165 from 0.2% to 0.7% (m/v). Although quantitative recovery in the slurry sampling
28
29
30 166 procedure is achieved for sample concentrations outside this range ($> 0.7\%$ m/v or $<$
31
32 167 0.2%), a high level of error (RSD $> 9.5\%$) is observed.

35 168 As it is a volatile element, Cd is lost from the graphite atomizer at temperatures
36
37
38 169 higher than 300 $^{\circ}\text{C}$ in the absence of a chemical modifier.^{51,52} To increase the thermal
39
40
41 170 stability of Cd and allow higher pyrolysis temperatures for the removal of higher
42
43 171 levels of concomitants, $\text{Pd}(\text{NO}_3)_2$ solution was assayed as a chemical modifier. The
44
45
46 172 effect of the pyrolysis and atomization temperature on the integrated absorbance were
47
48
49 173 studied in experiments on the AGV-2 SRM slurry with or without a $\text{Pd}(\text{NO}_3)_2$
50
51 174 chemical modifier. As shown in Fig. 1f, the addition of 5 μL 0.05% m/v $\text{Pd}(\text{NO}_3)_2$
52
53
54 175 modifier allowed the optimum pyrolysis temperature to increase from 350 $^{\circ}\text{C}$ (without

1
2
3
4 176 modifier) to 450 °C. The optimum atomization temperature was found to be 1600 °C
5
6 177 (Fig. 1f), which is similar to that of without a modifier. In addition to the graphite
7
8
9 178 furnace temperature programs described in Table 1, a pre-pyrolysis step (350 °C)
10
11 179 using air was added to avoid carbon residues on the platform of the graphite tube ⁵³.

15 180 **Analytical features of merit**

18 181 Calibrations were carried out using aqueous standard solutions with a linear range of
19
20 182 0.1 to 3.0 ng mL⁻¹ Cd. The characteristic mass (m_0), defined as the mass of analyte
21
22 183 corresponding to 0.0044 absorbance units, is 0.8 pg. The limit of detection (LOD) is
23
24 184 0.002 µg g⁻¹, which was calculated from 20 consecutive measurements of the blank
25
26 185 solutions, and a 200-fold sample dilution factor, i.e., a 0.5% m/v slurry concentration.
27
28 186 The tube lifetime using the proposed slurry sampling ETAAS method is 785 ± 28
29
30 187 analytical firings, which is only 15% shorter than that with the conventional closed
31
32 188 acid digestion ETAAS method. By considering the time taken the preparation of
33
34 189 sample slurries (10 samples per each batch spent 0.5 hour) and the determination by
35
36 190 ETAAS, the analytical throughput of the method is estimated to be 7 samples per hour.
37
38 191 For each sample analysis, only 0.25 mL HNO₃ and 0.30 mL Triton X-100 is required
39
40 192 with the proposed method. Conversely, conventional closed acid digestion ETAAS
41
42 193 requires 6.0 mL HNO₃ and 2.0 mL HF per sample, representing much higher cost and
43
44 194 more waste.
45
46
47
48
49
50
51
52
53
54
55
56
57
58
59
60

195 **Geological sample analysis**

196 Five USGS SRMs were assessed using the proposed method (Table 2). For
197 comparison, the USGS certified Cd values and those reported elsewhere in the
198 literature are also listed in Table 2. Cd contents determined for SRMs AGV-2, SBC-1,
199 SGR-1b, and BCR-2 are in agreement with the USGS reference values. The reference
200 value of BHVO-2 (USGS) is $0.06 \mu\text{g g}^{-1}$, which is lower than our reported value of
201 $0.116 \pm 0.011 \mu\text{g g}^{-1}$. However, our result is consistent with the value obtained by
202 isotope dilution ICP-MS ($0.112 \mu\text{g g}^{-1}$),²⁶ which is regarded as the most reliable
203 method for Cd detection. To further confirm that our reported values are reasonable,
204 ten separate aliquots of BHVO-2 were analyzed over a period of three months, and
205 consistent results ($0.093\text{-}0.123 \mu\text{g g}^{-1}$) were obtained (Fig. 2a). The Cd content of
206 BCR-2 obtained by ICP-MS ($0.227 \mu\text{g g}^{-1}$,⁵⁴ and $0.75 \mu\text{g g}^{-1}$,⁵⁵ etc.) is significantly
207 higher than the USGS reference value ($0.14 \mu\text{g g}^{-1}$) and the values obtained by
208 ID-ICP-MS ($0.136 \mu\text{g g}^{-1}$),⁵⁶ and our proposed ETAAS method ($0.142 \pm 0.008 \mu\text{g g}^{-1}$).
209 The large errors obtained using the ICP-MS method could be attributed to the
210 presence of large amounts of Zr ($\text{Zr}/\text{Cd} > 1880$) and Mo ($\text{Mo}/\text{Cd} > 2510$) in BCR-2.
211 Similar results were observed for AGV-2 and SGR-1b, which further confirms that
212 our slurry sampling ETAAS method is more reliable than ICP-MS.

213 The method we report herein was also used to determine Cd levels in 81 Chinese
214 geological SRMs, comprising 25 soil, 26 sediment, and 30 rocks. The obtained values

1
2
3
4 215 and their certified values compared in Fig. 3, except for those of one clay
5
6 216 (GBW03103), and one ultra-basic rock (GBW07101), and five limestone (GUI-1,
7
8
9 217 GUI-2, DIAN-1, DIAN-2, and DIAN-3) samples, which have no available certified or
10
11 218 literature values. As shown in Fig. 3, the Cd levels of 74 of the geological SRMs are
12
13
14 219 in good agreement with the certified values or reference values. The detailed results
15
16
17 220 for these SRMs are given in Table S1 (see ESI[†]). As shown in Table 3, the Cd content
18
19 221 found for the ultra-basic rock GBW07101 is $0.044 \pm 0.002 \mu\text{g g}^{-1}$, which is nearly
20
21 222 twice its reference value $0.024 \mu\text{g g}^{-1}$. This SRM was analyzed ten times over a
22
23
24 223 three-month period, and the results ranged from 0.043 to $0.047 \mu\text{g g}^{-1}$ (Fig. 2b). The
25
26
27 224 proposed method was also used to analyze one clay SRM GBW03103, and five
28
29
30 225 limestone SRMs (GUI-1, GUI-2, DIAN-1, DIAN-2, and DIAN-3), which have no
31
32
33 226 reported Cd value available (Table 3). Our recommended values are $0.108 \pm 0.009 \mu\text{g}$
34
35 227 g^{-1} (GBW03103), $0.056 \pm 0.005 \mu\text{g g}^{-1}$ (GUI-1), $0.176 \pm 0.012 \mu\text{g g}^{-1}$ (GUI-2), 0.134
36
37
38 228 $\pm 0.009 \mu\text{g g}^{-1}$ (DIAN-1), $0.335 \pm 0.005 \mu\text{g g}^{-1}$ (DIAN-2), and $0.244 \pm 0.004 \mu\text{g g}^{-1}$
39
40
41 229 (DIAN-3), respectively. The results of repeated analysis over a period of three months
42
43
44 230 are very stable (Fig. 4), which further confirms that our reported values are reasonable.
45
46 231 In addition, these values are in good agreement with the conventional closed acid
47
48
49 232 digestion ETAAS method (Table 3).

50
51 233

52 53 54 234 **Conclusion**

1
2
3
4 235 A method using slurry sampling ETAAS to accurately determine trace Cd in various
5
6 236 soils, sediments, and rocks was developed and validated. In our method, sample
7
8
9 237 digestion is not needed; thus, the problems related to conventional acid digestion,
10
11 238 such as the use of concentrated acids, time consumption, the risk of contamination,
12
13
14 239 and possible analyte loss are avoided. Using of the proposed method, Cd levels were
15
16
17 240 determined for 78 international geological SRMs, and the results were found to be in
18
19 241 good agreement with their certified or literature values of the samples. The Cd levels
20
21
22 242 in five limestone SRMs (GUI-1, GUI-2, DIAN-1, DIAN-2, and DIAN-3) and one clay
23
24
25 243 SRM (GBW03103) were reported for the first time in the literature, and the values for
26
27 244 one ultra-basic rock SRM (GBW07101) and one basalt SRM (BHVO-2) were updated.
28
29
30 245 In conclusion, our method has great potential for the direct determination of trace
31
32
33 246 levels of Cd in various solid geological samples.
34
35
36
37
38
39 247
40
41
42
43
44
45
46
47
48
49
50
51
52
53
54
55
56
57
58
59
60

248 **Acknowledgement**

249 This work was supported by the National Nature Science Foundation of China
250 (No. 21175120 and 21207120) and the National Key Scientific Instrument and
251 Equipment Development Projects of China (No. 2011YQ06010008), and the
252 Fundamental Research Funds for the Central Universities, China University of
253 Geosciences (Wuhan) (No. CUGL140411).

254

255 **References**

- 256 1. X. F. Mao, J. X. Liu, Y. T. Huang, L. Feng, L. H. Zhang, X. Y. Tang, J. Zhou, Y. Z.
257 Qian and M. Wang, *Journal of agricultural and food chemistry*, 2013, **61**, 848-853.
- 258 2. K. Ono, *Journal of hazardous materials*, 2013, **262**, 741-747.
- 259 3. S. Enamorado, J. M. Abril, A. Delgado, J. L. Mas, O. Polvillo and J. M. Quintero,
260 *Journal of hazardous materials*, 2014, **266**, 122-131.
- 261 4. R. J. Bian, S. Joseph, L. Q. Cui, G. X. Pan, L. Q. Li, X. Y. Liu, A. Zhang, H.
262 Rutledge, S. W. Wong, C. Chia, C. Marjo, B. Gong, P. Munroe and S. Donne, *Journal*
263 *of hazardous materials*, 2014, **272**, 121-128.
- 264 5. E. Galunin, J. Ferreti, I. Zapelini, I. Vieira, C. R. T. Tarley, T. Abrao and M. J.
265 Santos, *Journal of hazardous materials*, 2014, **265**, 280-287.
- 266 6. D. W. Xue, H. Jiang, X. X. Deng, X. Q. Zhang, H. Wang, X. B. Xu, J. Hu, D. L.
267 Zeng, L. B. Guo and Q. Qian, *Journal of hazardous materials*, 2014, **280**, 269-278.
- 268 7. A. D. Schmitt, S. J. G. Galer and W. Abouchami, *Earth Planet Sc Lett*, 2009, **277**,
269 262-272.
- 270 8. R. F. Wei, Q. J. Guo, H. J. Wen, J. X. Yang, M. Peters, C. W. Zhu, J. Ma, G. X.
271 Zhu, H. Z. Zhang, L. Y. Tian, C. Y. Wang and Y. X. Wan, *Analytical Methods*, 2015, **7**,
272 2479-2487.
- 273 9. Z. H. Wang, Q. Z. Xu, S. Y. Li, L. Y. Luan, J. Li, S. X. Zhang and H. H. Dong,
274 *Analytical Methods*, 2015, **7**, 1140-1146.

- 1
2
3
4 275 10. J. Sun, Z. G. Yang, H. W. Lee and L. Wang, *Analytical Methods*, 2015, 7,
5
6 276 2653-2658.
7
8
9 277 11. Y. X. Liu, T. Zhou, J. Y. Wang, Y. L. Wang, D. Zhao and Q. Zhang, *Analytical*
10
11 278 *Methods*, 2015, 7, 1872-1877.
12
13
14 279 12. D. P. Bishop, D. J. Hare, A. de Grazia, F. Fryer and P. A. Doble, *Analytical*
15
16 280 *Methods*, 2015, 7, 5012-5018.
17
18
19 281 13. A. Philippe, M. Gangloff, D. Rakcheev and G. E. Schaumann, *Analytical Methods*,
20
21 282 2014, 6, 8722-8728.
22
23
24 283 14. P. Li, X. Q. Zhang, Y. J. Chen, H. Z. Lian and X. Hu, *Analytical Methods*, 2014, 6,
25
26 284 4205-4211.
27
28
29 285 15. C. Han, J. N. Sun, H. Y. Cheng, J. H. Liu and Z. G. Xu, *Analytical Methods*, 2014,
30
31 286 6, 5369-5375.
32
33
34 287 16. S. Z. Chen, X. L. Guo, Y. Zhang and D. B. Lu, *Analytical Methods*, 2014, 6,
35
36 288 3668-3673.
37
38
39 289 17. M. Balski, H. Kipphardt, A. Berger, S. Meyer and U. Panne, *Analytical Methods*,
40
41 290 2014, 6, 77-85.
42
43
44 291 18. Z. Bao, H. L. Yuan, R. Wen and K. Y. Chen, *Analytical Methods*, 2015, 7,
45
46 292 5034-5040.
47
48
49 293 19. N. I. Rousis, L. N. Pasiadis and N. S. Thomaidis, *Analytical Methods*, 2014, 6,
50
51 294 5899-5908.
52
53
54
55
56
57
58
59
60

- 1
2
3
4 295 20. W. Guo, S. H. Hu, J. A. Zhao, S. S. Jin, W. J. Liu and H. F. Zhang, *Microchemical*
5
6 296 *Journal*, 2011, **97**, 154-159.
7
8
9 297 21. T. W. May and R. H. Wiedmeyer, *Atomic Spectroscopy*, 1998, **19**, 150-155.
10
11 298 22. T. C. Duan, X. J. Song, P. R. Guo, H. F. Li, L. H. Pan, H. T. Chen and J. W. Xu,
12
13 299 *Journal Of Analytical Atomic Spectrometry*, 2007, **22**, 403-406.
14
15
16
17 300 23. X. Y. Jia, Y. Han, X. L. Liu, T. C. Duan and H. T. Chen, *Microchim Acta*, 2010,
18
19 301 **171**, 49-56.
20
21
22 302 24. P. H. Liao, S. J. Jiang and A. C. Sahayam, *Journal Of Analytical Atomic*
23
24 303 *Spectrometry*, 2012, **27**, 1518-1524.
25
26
27 304 25. W. N. Chen, S. J. Jiang, Y. L. Chen and A. C. Sahayam, *Analytica chimica acta*,
28
29 305 2015, **860**, 8-14.
30
31
32 306 26. A. Makishima, H. Kitagawa and E. Nakamura, *Geostand Geoanal Res*, 2011, **35**,
33
34 307 57-67.
35
36
37 308 27. R. Q. Thompson and S. J. Christopher, *Analytical Methods*, 2013, **5**, 1346-1351.
38
39
40 309 28. W. Guo, S. H. Hu, Y. F. Xiao, H. F. Zhang and X. J. Xie, *Chemosphere*, 2010, **81**,
41
42 310 1463-1468.
43
44
45 311 29. J. M. Jarrett, G. Xiao, K. L. Caldwell, D. Henahan, G. Shakirova and R. L. Jones,
46
47 312 *Journal Of Analytical Atomic Spectrometry*, 2008, **23**, 962-967.
48
49
50 313 30. Naeemullah, T. G. Kazi, H. I. Afridi, F. Shah, S. S. Arain, A. H. Panhwar, S. A.
51
52 314 Arain and M. B. Arain, *Analytical Methods*, 2014, **6**, 6909-6915.
53
54
55
56
57
58
59
60

- 1
2
3
4 315 31. S. Mohajer, M. Chamsaz and M. H. Entezari, *Analytical Methods*, 2014, **6**,
5
6 316 9490-9496.
7
8
9 317 32. R. F. de Oliveira, C. C. Windmoller, W. B. Neto, C. C. de Souza, M. A. Beinner
10
11 318 and J. B. B. da Silva, *Analytical Methods*, 2013, **5**, 5746-5752.
12
13
14 319 33. I. C. F. Damin, A. V. Zmozinski, A. R. Borges, M. G. R. Vale and M. M. da Silva,
15
16 320 *Analytical Methods*, 2011, **3**, 1379-1385.
17
18
19 321 34. Y. Z. Yi, S. J. Jiang and A. C. Sahayam, *Journal Of Analytical Atomic*
20
21 322 *Spectrometry*, 2012, **27**, 426-431.
22
23
24 323 35. J. Sardans, F. Montes and J. Penuelas, *Spectrochim Acta B*, 2010, **65**, 97-112.
25
26
27 324 36. P. Torok and M. Zemberyova, *Spectrochim Acta B*, 2011, **66**, 93-97.
28
29
30 325 37. A. de Jesus, A. V. Zmozinski, I. C. F. Damin, M. M. Silva and M. G. R. Vale,
31
32 326 *Spectrochim Acta B*, 2012, **71-72**, 86-91.
33
34
35 327 38. P. Torok and M. Zemberyova, *Food chemistry*, 2012, **132**, 554-560.
36
37
38 328 39. A. V. Zmozinski, L. D. Passos, I. C. F. Damin, M. A. B. E. Santo, M. G. R. Vale
39
40 329 and M. M. Silva, *Analytical Methods*, 2013, **5**, 6416-6424.
41
42
43 330 40. A. V. Zmozinski, T. Pretto, A. R. Borges and M. G. R. Vale, *Analytical Methods*,
44
45 331 2015, **7**, 3735-3741.
46
47
48 332 41. M. Resano, A. C. Lapena and M. A. Belarra, *Analytical Methods*, 2013, **5**,
49
50 333 1130-1139.
51
52
53 334 42. M. C. Santos, J. A. Nobrega, N. Baccan and S. Cadore, *Talanta*, 2010, **81**,

- 1
2
3
4 335 1781-1787.
5
6 336 43. A. R. Borges, E. M. Becker, C. Lequeux, M. G. R. Vale, S. L. C. Ferreira and B.
7
8 337 Welz, *Spectrochim Acta B*, 2011, **66**, 529-535.
9
10 338 44. R. A. Sanchez-Moreno, M. J. Gismera, M. T. Sevilla and J. R. Procopio,
11
12 339 *Phytochem Analysis*, 2010, **21**, 340-347.
13
14 340 45. C. K. de Andrade, V. E. dos Anjos, M. L. Felsner, Y. R. Torres and S. P. Quinaia,
15
16 341 *Food chemistry*, 2014, **146**, 166-173.
17
18 342 46. D. Baralkiewicz and H. Gramowska, *Analytica chimica acta*, 2004, **510**, 249-256.
19
20 343 47. A. L. Souza, F. F. da Silva, G. A. R. Kelmer and P. V. Oliveira, *Analytical*
21
22 344 *Methods*, 2013, **5**, 2059-2063.
23
24 345 48. W. Guo, S. H. Hu, X. J. Wang, J. Y. Zhang, L. L. Jin, Z. L. Zhu and H. F. Zhang,
25
26 346 *Journal Of Analytical Atomic Spectrometry*, 2011, **26**, 1198-1203.
27
28 347 49. W. Guo, S. H. Hu, J. Y. Zhang, L. L. Jin, X. J. Wang, Z. L. Zhu and H. F. Zhang,
29
30 348 *Journal Of Analytical Atomic Spectrometry*, 2011, **26**, 2076-2080.
31
32 349 50. W. Guo, S. H. Hu, J. Y. Zhang and H. F. Zhang, *Sci Total Environ*, 2011, **409**,
33
34 350 2981-2986.
35
36 351 51. E. C. Lima, F. Barbosa and F. J. Krug, *Fresen J Anal Chem*, 2001, **369**, 496-501.
37
38 352 52. P. Wu, C. H. Li, J. B. Chen, C. B. Zheng and X. D. Hou, *Appl Spectrosc Rev*,
39
40 353 2012, **47**, 327-370.
41
42 354 53. W. Guo, P. Zhang, L. L. Jin and S. H. Hu, *Journal Of Analytical Atomic*
43
44
45
46
47
48
49
50
51
52
53
54
55
56
57
58
59
60

1
2
3
4 355 *Spectrometry*, 2014, **29**, 1949-1954.
5

6 356 54. S. K. Marx and B. S. Kamber, *Applied Geochemistry*, 2010, **25**, 1221-1237.
7

8
9 357 55. Z. C. Hu and S. Gao, *Chem Geol*, 2008, **253**, 205-221.
10

11 358 56. R. D. Loss, K. J. R. Rosman and J. R. Delaeter, *Geochimica Et Cosmochimica*
12

13
14 359 *Acta*, 1984, **48**, 1677-1681.
15

16
17 360
18

19 361
20
21
22
23
24
25
26
27
28
29
30
31
32
33
34
35
36
37
38
39
40
41
42
43
44
45
46
47
48
49
50
51
52
53
54
55
56
57
58
59
60

1
2
3
4
5 362 **Figure captions**
6
7

8 363 **Fig. 1.** Effects of the slurry sampling ETAAS conditions on Cd recoveries from the
9
10 364 geological SRM AGV-2. (a) Effect of HNO₃ concentration, (b) effect of Triton X-100
11
12 365 concentraton, (c) effect of sonication time, (d) effect of sample particle size, (e) effect
13
14 366 of slurry concentration, and (f) effect of pyrolysis and atomization temperatures.
15
16

17
18 367 **Fig. 2.** Results of repeated analyses of (a) a basalt SRM BHVO-2, and (b) an
19
20 368 ultra-basic rock SRM GBW 07101. The average values of Cd in BHVO-2 and
21
22 369 GBW07101 are $0.116 \pm 0.011 \mu\text{g g}^{-1}$ and $0.044 \pm 0.002 \mu\text{g g}^{-1}$, respectively. Ten
23
24 370 separate aliquots of the samples were analyzed during a period of three months.
25
26

27
28 371 **Fig. 3.** Obtained Cd values for 74 Chinese geological SRMs by slurry sampling
29
30 372 ETAAS vs. their reference values.
31
32

33
34 373 **Fig. 4.** The stability of Cd values obtained by our method in six geological SRMs: (a)
35
36 374 clay SRM GBW 03103, (b) limestone SRM GUI-1, (c) limestone SRM GUI-2, (d)
37
38 375 limestone SRM DIAN-1, (e) limestone SRM DIAN-2, and (f) limestone SRM
39
40 376 DIAN-1. Ten separate aliquots of the samples were analyzed during a period of three
41
42 377 months.
43
44

45
46
47 378
48
49
50
51
52
53
54
55
56
57
58
59
60

379

380 **Table 1**

381 Graphite furnace temperature programs and spectrometer operating conditions

| Step | Temp. (°C) | Ramp. time (s) | Hold time (s) | Inter. flow (mL min ⁻¹) | Gas type |
|------------------------------|------------|-------------------|--|--|------------------|
| Drying | 110 | 5 | 30 | 250 | Ar |
| | 130 | 15 | 30 | 250 | Ar |
| Pyrolysis | 350 | 15 | 15 | 50 | Air ^a |
| | 450 | 10 | 20 | 250 | Ar |
| Atomization | 1600 | 0 | 4 | 0 | / |
| Cleaning | 2500 | 1 | 3 | 250 | Ar |
| Wavelength/nm: 228.8 | | | Measurement mode: Peak area | | |
| Spectral bandwidth/nm: 0.7 | | | Zeeman-effect background correction system | | |
| Cathode lamp intensity/mA: 8 | | | Pyrolytic graphite tubes with platform | | |
| Integration time/s: 3 | | | Injection volume/μL: 20 | | |

382 ^a Air gas come from the air compressor.

383

384

385 **Table 2**386 Cd detected results of USGS geological SRMs, $\mu\text{g g}^{-1}$

| USGS SRMs | This study (N=15) | USGS Ref. ^a | Literature values |
|-------------------|-------------------|------------------------|---|
| AGV-2, andesite | 0.070 ± 0.003 | (0.061) | 0.121^{b} , 0.082^{c} , |
| BHVO-2, basalt | 0.120 ± 0.012 | (0.06) | 0.112^{d} |
| SBC-1, shale | 0.400 ± 0.003 | (0.4) | / |
| SGR-1b, oil shale | 0.900 ± 0.040 | (0.9) | 1.14^{c} |
| BCR-2, basalt | 0.155 ± 0.013 | (0.14) | 0.227^{b} , 0.75^{c} , 0.136^{e} |

387 ^a Reference value by USGS388 ^b ICP-QMS value by Marx and Kamber ⁵⁴389 ^c ICP-QMS value by Hu and Gao ⁵⁵390 ^d ID-ICP-MS value by Makishim et al. ²⁶391 ^e ID-ICP-MS value by Loss et al. ⁵⁶

392

393

394 **Table 3**395 Updated Cd values for seven SRMs determined by this method, $\mu\text{g g}^{-1}$

| Geological SRMs | This work (N=10) | Reference values | Closed acid digestion ETAAS method (N=3) |
|---------------------------|---------------------|----------------------|---|
| GBW07101, Ultrabasic rock | 0.044 ± 0.004 | (0.024) ^a | 0.048 ± 0.012 |
| GBW03103, Clay | 0.108 ± 0.009 | / ^b | 0.108 ± 0.029 |
| GUI-1, Limestone | 0.056 ± 0.005 | / ^b | 0.060 ± 0.012 |
| GUI-1, Limestone | 0.176 ± 0.012 | / ^b | 0.185 ± 0.015 |
| DIAN-1, Limestone | 0.134 ± 0.009 | / ^b | 0.142 ± 0.022 |
| DIAN-2, Limestone | 0.335 ± 0.005 | / ^b | 0.342 ± 0.025 |
| DIAN-3, Limestone | 0.244 ± 0.004 | / ^b | 0.238 ± 0.018 |

396 ^a Reference value by the Institute of Geophysical and Geochemical Exploration of

397 China

398 ^b No reference or reported value

399

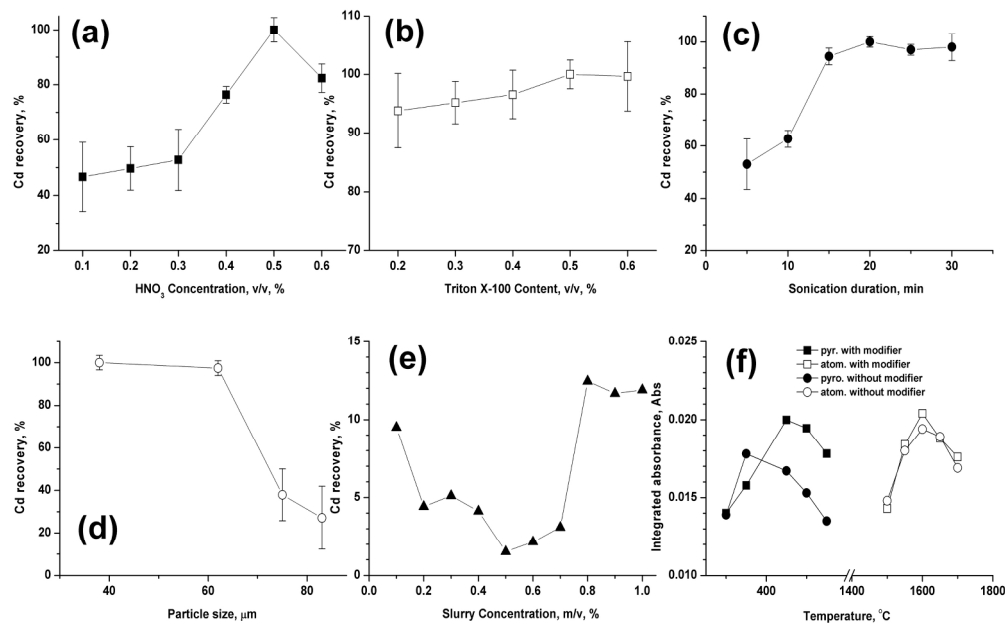


Fig. 1. Effects of the slurry sampling ETAAS conditions on Cd recoveries from the geological SRM AGV-2. (a) Effect of HNO₃ concentration, (b) effect of Triton X-100 concentration, (c) effect of sonication time, (d) effect of sample particle size, (e) effect of slurry concentration, and (f) effect of pyrolysis and atomization temperatures.

210x148mm (300 x 300 DPI)

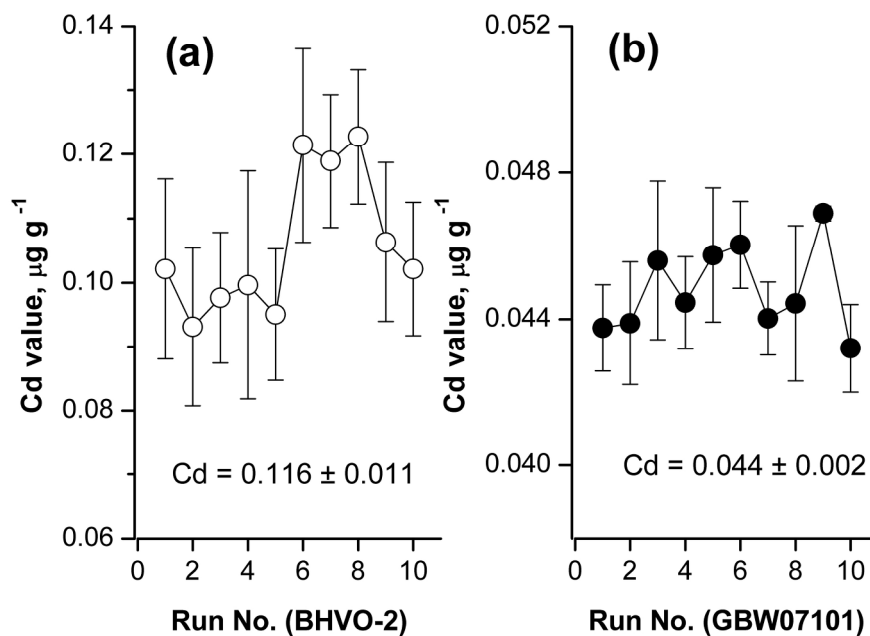


Fig. 2. Results of repeated analyses of (a) a basalt SRM BHVO-2, and (b) an ultra-basic rock SRM GBW 07101. The average values of Cd in BHVO-2 and GBW07101 are $0.116 \pm 0.011 \mu\text{g g}^{-1}$ and $0.044 \pm 0.002 \mu\text{g g}^{-1}$, respectively. Ten separate aliquots of the samples were analyzed during a period of three months. 210x148mm (300 x 300 DPI)

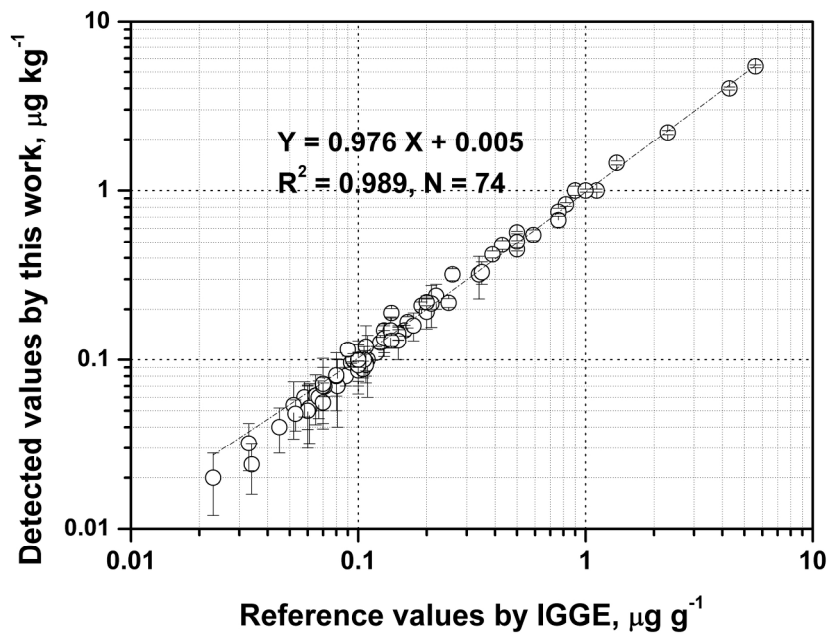


Fig. 3. Obtained Cd values for 74 Chinese geological SRMs by slurry sampling ETAAS vs. their reference values.
210x148mm (300 x 300 DPI)

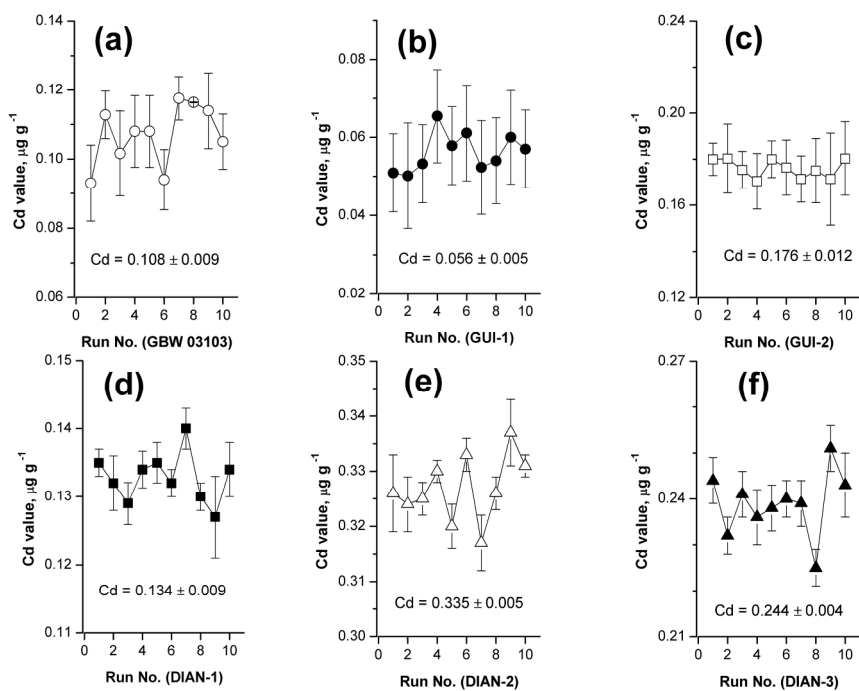
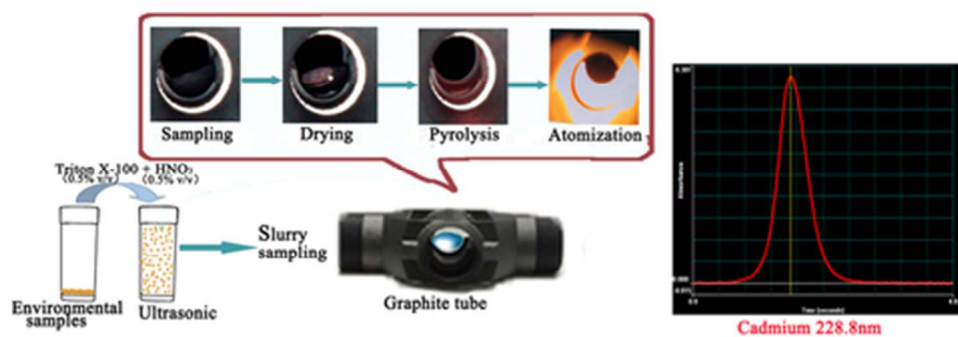


Fig.4 The stability of Cd values obtained by our method in six geological SRMs: (a) clay SRM GBW 03103, (b) limestone SRM GUI-1, (c) limestone SRM GUI-2, (d) limestone SRM DIAN-1, (e) limestone SRM DIAN-2, and (f) limestone SRM DIAN-3. Ten separate aliquots of the samples were analyzed during a period of three months.

209x148mm (300 x 300 DPI)



Graphical Abstract: An accurate and high throughput method involving slurry sampling ETAAS was developed to detect trace Cd content in various geological samples
41x14mm (300 x 300 DPI)