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A novel approach to discriminate Lycium barbarum from Zhongning area using FT-IR spectroscopy and chemometrics

Yuan Gao, Xiuzhu Yu *, Lirong Xu, Ning Wang, Rui Zhang

College of Food Science and Engineering, Northwest A&F University, 28 Xinong Road Yangling, 712100, Shaanxi, P. R. China

*Corresponding author. Tel.: +86-29-87092206; fax: +86-29-87092486. E-mail address: xiuzhuyu1004@hotmail.com.

Abstract: An increasing number of fake and inferior Lycium barbarum fruits from Zhongning production area appear in the market due to large demand of consumers. Present study was focused on the classification and identification of Zhongning Lycium barbarum (ZNL) from different production areas using Fourier transform infrared (FT-IR) spectroscopy coupled with chemometrics. Results revealed that the spectral region between 1909-1311 cm⁻¹ was found to be feasible for both classification and identification of ZNL. FT-IR in combination with discriminant analysis (DA) and soft independent modeling of class analogy were used for classification and identification of ZNL among different Lycium production areas in China, and FT-IR coupled with DA can classify and identify ZNL from the other production areas successfully. The recognition rates of the calibration and validation sets were 94.0 % and 100 %, respectively. In conclusion, the proposed method is a useful tool to identify ZNL among different Lycium production areas in China.

Introduction

Lycium barbarum L., known as Chinese wolfberry or Goji, is a kind of multi-branched shrub that belongs to Solanaceae. Its chemical constituents, which are commonly used in herbal medicine and tonic applications, have been extensively studied and found to contain polysaccharide-protein, polysaccharides, flavonoids, vitamins and zeaxanthin¹⁻⁴. Therefore, L. barbarum has numerous biological activities for potential pharmaceutical interest, including immune-regulatory, antitumor, anti-fatigue, and antioxidant properties; it also reduces neuronal damage and blood-retinal barrier disruption⁵⁻⁸. Geographical origin is one of the most important quality parameters for many foods, because climate, soil, and cultivation methods cause differences in the chemical composition of plants⁹. In China, the most popular Lycium ecotype currently growing is in

Zhongning County, Ningxia Hui Autonomous Region, China. Zhongning Lycium barbarum (ZNL) with its unique geography and suitable climate features, always produces high quality. The main features of Lycium are its large fruit size, nice color and high contents of polysaccharide, flavonoid, and trace elements, resulting in its high price in traditional medicine and international markets. Moreover, given the aforementioned features, ZNL is attracting increasing attention, which leads to its great need in the field of pharmacy and functional foods. At present, with the increasing demand for Lycium, the ZNL market is saturated with fake and inferior Lycium, and the specific differential detection method of Lycium is rarely reported. Therefore, simple and efficient methods to meet the demand are of urgent need.

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combined with chemometrics has been widely used in many scientific studies. It is a direct, reliable, and fast method that makes it possible to simultaneously obtain specific information about different parameters, mainly in the 3000-400 cm⁻¹ region, because bands are associated with the vibrations of functional groups of the molecules¹⁰⁻¹². The associated bands proteins, fats, lactose, and lactic acid are well known and of have been described in milk and cheese¹³⁻¹⁶. Currently, in the field of Lycium classification, different Lycium species are usually distinguished through micro-section, observation, and analysis by professionals. Such methods are highly subjective and undeterministic. Some works used FT-IR, second derivative IR spectra and 2D correlation infrared spectroscopy for the distinction of eight Lycium species and obtained acceptable results¹⁷⁻¹⁸. The discrimination of Goji's geographical origin through non-targeted liquid to quadruple time-of-flight mass spectrometry also obtained good results¹⁹. However, such methods had some disadvantages, such as complex, tedious and hard operation. Moreover, FT-IR spectroscopy combined with multivariate analysis has been used as a rapid and reliable method to determine the cultivation ages and cultivars of ginseng²⁰⁻²². Some researchers also conducted FT-IR spectroscopy with multivariate techniques to discriminate and classify red, blue, and green spray paints, and results proved the method's effectivity²³⁻²⁵. FT-IR spectroscopy has also been successfully used in the classification and discrimination of living matters²⁶⁻³⁰.

This study was conducted to identify the cultivation region and quality of ZNL. A total of 149 samples were obtained, and different spectral pretreatments were conducted. Various spectral regions were chosen, and different chemometric methods were applied to achieve ZNL classification from other regions. This study aimed to provide a new trend and perspective for ZNL discrimination and quality evaluation.

67 Samples

- 68 Mature wolfberries (L. barbarum) were obtained from Ningxia
- 69 Chinese Lycium Group Company, including ZNL (Ningxia,
- 70 China) and non-Zhongning Lycium (NZNL), which came from
- 71 other regions of Ningxia, Xinjiang, Qinghai, Gansu, China
- Reference samples were prepared by removing all foreign
 matters, cut into pieces with scissors, and desiccated by heated
 air combined with vacuum drying for 12 and 8 h, respectively.
 Finally, the dried samples were evenly milled and passed
 through a 60 mesh sieve to obtain the final samples.

77 Analytical protocol

A Bruker VERTEX 70 series FT-IR spectrometer equipped with a deuterated triglycine sulfate (DTGS) was used for this study. Moreover, high-speed universal grinder (Tianjin Taisite Instrument Co., Ltd.) and vacuum drying oven (Dalian Eilite Instrument Co., Ltd.) were used in this work. Transmission FT-IR spectra were taken though KBr pellets, in which the ratio of KBr to sample was 100:1 (w/w). The measurement parameters were as follows: 150 × 150mm window size, 4.0cm⁻¹ resolution, 32 co-added spectra, and 4000-400cm⁻¹ range. Each sample was measured three times. The average of the three spectra obtained from the same sample was used in subsequent analyses.

90 Data processing

Data covering the FT-IR wavelength region (4000–400cm⁻¹) were collected, and models were developed and validated using this complete range along with a number of subsets. Each spectrum was an ensemble average of three scans collected from the cell. The spectra of 149 samples were pretreated through normalization to correct the measurement variation. Each spectrum was aligned to the baseline using an instrumentation software before converting the spectra into JCAMP-DX format. The most accurate models were developed using the wavelength range of 1909-1311cm⁻¹, and only these

66 Material and method

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101 models were discussed further in this study.

102 Software

All statistical treatments were performed using
Unscrambler X10.1(CAMO, Norway) and TQ Analyst 8.3.125
(Thermo Scientific, USA). The commercial database software
used included OMNIC 7.3 (Thermo Scientific, USA) and
OPUS 5.5(Bruker, Germany).

108 Data analysis

109 Visual grouping

110 The samples were randomized and divided into two groups: a
111 calibration group that consisted of 111 of the samples and a
112 validation group that consisted of the remaining sample of 38.
113 Sample assignment to each group was performed by selecting
114 every sample as a member of the calibration group and the
115 remaining samples were assigned to the validation group.
116 Random selection did not generate equal-sized groups.

117 Soft independent modeling of class analogy (SIMCA)

SIMCA is based on making a PCA model for each class in the training set. Unknown samples were then compared with the class models and assigned to classes according to their proximity to the training samples. It is also known as a supervised pattern recognition method as the individual PCA models define classification rules. In the case of all classification methods, making a SIMCA model needed a training stage and a test stage. SIMCA modeling requires building one PCA model for each class which describes the structure of that class as well as possible. The optimal number of PCs should be chosen for each model separately, according to a suitable validation procedure. Before developing a SIMCA model, it is helpful to determine if the data being considered exhibit any tendency to cluster by the classes. Before using the models to predict class membership for new samples, one should also evaluate the model specificity, i.e. whether the

- 134 classes overlap or are sufficiently distant from each other.
- 135 Specific tools, such as model distance and modeling power are
- 136 available for this purpose. The discrimination power of SIMCA
- 137 was based on the largest possible distance among classes.
- 138 Discriminant analysis (DA)
- DA is a classification method of TQ Analyst software. Different spectral pretreatment methods, including a constant, peak or normalize ratio (A/b=k*c), multiplicative signal correction (MSC), Standard Normal Variate (SNV), Norris derivative filter, and Savitzky-Golay (SG) filter) were selected to produce better results. The Savitzky-Golay filter is a type of filter. The method essentially performs a local polynomial regression (of degree k) on a distribution (of at least k+1 equally spaced points) to determine the smoothed value for each point. Methods are also provided for calculating the first up to the fifth derivatives. The samples were classified, and the spectral information of the samples was imported. To make a robust and available discriminate model, different spectral pretreatment method were applied, and the analysis spectral regions and various principal component (PC) numbers were changed. The value was either decided by the highest recognition rate or the lowest error rate.

156 Results and discussion

157 Spectral analysis

- 158 The chemical compounds of Lycium are polysaccharides,
- 159 organic acids, steroids, peptides, flavonoids, and trace elements.
- 160 Fig. 1 shows the FT-IR spectra of the two kinds of typical
- *Lycium* samples at room temperature.



As shown in Fig. 1, several characteristics could be extracted, such as the strongest peak at 3390cm⁻¹ belonging to the stretching vibration of O-H groups, and the peak at 2925 cm⁻¹ assigned to the stretching vibration of -CH₂ groups. Furthermore, the peak at 1626 cm⁻¹ was mostly ascribed to the stretching vibration of C=O groups in the volatile oils and other compounds embodying carbonyl group, and the stronger peaks in 1600-1000 cm⁻¹ were mainly attributed to the stretching vibration of C-O and C=O, which displayed the characteristic absorption bands of polysaccharides and glycosides.

Although the spectra of the two kinds of *Lycium* species
were rather similar, some differences in the shape or intensity
were observed. The absorbance peak of *Lycium* was extremely
different under the range of 1800–1000 cm⁻¹. In particular, the

- 182 characteristic band at 1108 cm⁻¹ was from the ZNL sample,
- 183 while characteristic band at 1098 cm⁻¹ was from the NZNL
- 184 sample. Therefore, we could create a robust classification
- **185** model of *Lycium* under the spectra range of $1800-1000 \text{ cm}^{-1}$.

186 Effective wavenumber (EW) selection

187 EW selection can simplify the model, and improve the
188 forecasting capacity and robustness of the calibration models
189 to eliminate of collinear variables³¹. The value of X-loading
190 weights was employed in the EW selection method and it was
191 estimated using the PCA model.





Fig. 2 Effective wavenumber selection using X-loading weight

The X-loading weights of the first four PCs by PCA are shown in Fig. 2. The loading weights showed how much each wavenumber contributed to the response variation. Wave numbers with large loading weight values are important for the PCA model. The spectral regions of X-loading weights with absolute values of over 0.02 were used for modeling. As shown in Fig. 2, the spectral regions of 3700-3100 and 1800-900 cm⁻¹showed higher loading weight and contributed more to the robustness of the identification model. Compared with the PCA model obtained with full spectra, the X-loading weight methods considerably improved the performance of the model. Excellent performance was obtained from the PCA model with EWs selected using the X-loading weight method. PCA is a

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kind of data dimension reduction technique of multivariate statistical analysis. PC is the result of the original variable reorganization, replacing the original variables involved in the modeling, reduced the workload of the analysis process. Selected a suitable PCs value, when the cumulative contribution rate of PC was greater than 85%, we can think it is enough to response the information of original variables, the corresponding value is the number of PCs. According to the PCA analysis, we found that the value of 3 PC is perfect, with the cumulative contribution rate of 90%, selected for further research. Most of the peaks that appeared in the regions of 3700-3100cm⁻¹ belonged to the vibration of O-H bond, including stretching vibration, deviational vibration and bending vibration. All those molecular movements made a small difference among various Lycium samples. Nevertheless, simple and effective identification pre-analysis was conducted. According to the pre-analysis results and the X-loading weight values, the spectra of 1909-1311cm⁻¹ was chosen for in-depth and detailed analysis. 0.30 0.25



(a) Characteristic spectrum of the ZNL sample





Fig. 3 compiles the characteristic FT-IR spectra of *Lycium* from
different production areas under room temperature. As shown
in Fig. 3, the typical part of 1900-1300cm⁻¹ was chosen.
Detailed peak positions of the typical samples are also
illustrated in Fig. 3.

These FT-IR spectra showed their macro-fingerprints. Comparative and separate analyses were performed in the spectra of two typical samples. For example, the strongest peak at 1638 and 1630cm⁻¹ was due to the bending of carbonyl. The absorbance values of these two sample types greatly differed, and various amounts of amino acid and polypeptide possibly exist in these samples. The peak at 1501 and 1509cm⁻¹ belonged to the bending of -C-H groups in Lycium, whereas that at 1458 and 1459cm⁻¹ was attributed to the bending vibration of methyl and methylene groups. Several kinds of amino acids might be present in these samples. The peak at 1386 and 1385cm⁻¹ was due to the plane vibration of O-H groups in Lycium, phenolic acid compounds and carbohydrates. In the spectrum of ZNL sample, an absorbance peak was observed at 1401cm⁻¹, which was attributed to the bending vibration of methyl and methylene groups. This finding indicated the typical absorption of amino acids. The typical

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peak of NZNL sample was at 1542cm⁻¹, which was mainly 265 result of the SIMCA classification, a suitable spectral region attributed to the bending vibration of N-H groups and tretching vibration of C-N groups in protein amide II band. This peak displayed the characteristic absorptions of fatty acid and 262 protein.

266	was selected. Model and data processing were performed under
267	this region. Various spectral pretreatment methods were
268	applied to identify the ZNL in the typical production areas. All
269	of the methods were verified by multivariate statistical
270	analysis.

263 SIMCA

Upon analysis of the spectral region and considering the

Table 1 Measurement of performance in the discrimination of Lycium samples of the calibration and validation groups using

272					SIMCA						
	Spectral regions/cm ⁻¹			Calibration			Validation				
	1909–1	311	No pretreatment	SNV	SG (9.4)	MSC	No pretreatment	SNV	SG (9.4)	MSC	
	P opognition rate	ZNL	55.56	90.60	83.76	52.99	58.12	85.47	82.91	52.99	
	Recognition rate	NZNL	53.13	87.5	84.37	46.88	53.13	81.25	81.25	56.25	
273	A suitable value of three PCs was chosen for each class. The				283	the ide	entification of ZNL	samples.	As we	all know,	an
274	spectral region of the model was 1909-1311cm ⁻¹ . Different				284	applicable model at least meet the demands of recognition r					rate
275	models were constructed using various data processing			285	over 90%, and the further study should be performed without				iout		
276	76 methods (No, SNV, SG 9.4, and MSC). Table 4 presents the 286 any hesitation.										
277	classification rate of each model. The type of ZNL sample										
278	obtained a better classification regardless of the technique										
279	used, in which the t	ype of NZN	IL sample was ext	tremely	288	Th	e DA method emplo	yed variou	is spectral	l regions un	Ider
280	confusing. The process	sing method	of SNV and SG she	owed a	289	the sa	me pretreatment	and vari	ous PCs	s or vari	ous
281	better classification th	an that of M	ISC and No pretrea	atment.	290	pretreat	ments on the typical	spectral	region. Ai	n identificat	tion
282	The SNV processing undoubtedly showed great significance in 291 method was achieved according to the results of the model.										

 Table 2 Effect of different spectra without pretreatments on recognition rate of calibration and validation samples

Spectral	PC Calibration set/% Val		Valida	alidation set/%	
regions/cm ⁻¹	numbers	ZNL	NZNL	ZNL	NZNL
3999–410	10	74.70	81.82	85.71	70
3780-2980	10	73.49	86.36	78.57	70
2980-2100	10	78.31	81.82	82.14	80
2560-1760	10	81.93	77.27	85.71	80
2100-1120	10	89.16	86.36	92.86	90
1300–960	10	86.75	81.82	85.71	80
1909–1311	10	93.98	90.91	100	100
920-410	10	86.75	86.36	92.86	80
1240–660	10	81.93	81.82	85.71	90
3999-410	10	74.70	81.82	85.71	70

293 The numbers of the ZNL calibration and validation sets were

83 and 28, respectively, and the remaining number of samples

was the number of NZNL. Different spectra were purposefully
selected without any pretreatments. Various spectral regions
obviously resulted in various recognition rates of the
calibration and validation sets. As shown in Table 2, in order to
find a suitable spectral region, a constant value of 10 PCs was
chosen for each treatment, different spectra exerted tiny effects
on the recognition rate of the validation set. A perfect result of

100 % was achieved in the range of 2560-1760 cm¹. Various

303	spectral regions exerted a great influence on the calibration set
304	compared with the validation set. Only one spectral region had
305	a recognition rate over 90% for both samples, and the
306	recognition rates under the range of 1909-1311 \mbox{cm}^1 were
307	93.98% and $90.91%.$ These results were due to the fact that the
308	two samples exhibited different feature absorbance peaks in the
309	range of 1909-1311 cm^{-1} .

Spectral	DC	Calibra	ation set/%	Validation set/%	
regions/cm ⁻¹	PC numbers –	ZNL	NZNL	ZNL	NZNL
1909-1311	4	51.81	78.57	68.42	90
1909-1311	5	66.26	71.43	78.95	80
1909-1311	6	69.88	78.57	78.95	100
1909-1311	7	79.52	82.14	86.84	100
1909-1311	8	77.11	82.14	84.21	100
1909-1311	9	75.9	82.14	84.21	100
1909-1311	10	93.98	90.91	100	100
1909-1311	11	87.95	89.29	100	100
1909-1311	12	87.95	89.29	100	100
1909-1311	13	87.95	89.29	97.37	100

311	Under the analysis of different spectral interval, considering
312	that the PC numbers contributed to the classification of the
313	samples, a constant value of 10 PCs was chosen for each
314	treatment. From the Table 3, it's tempting to conclude that the

315	PCs of 10 perfected for the deeper DA classification research.
316	Further analysis of the results showed that different spectral
317	interval had the same conclusion, the value of 10 PCs was an

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ble 3, it's tempting to conclude that the	318	appropriate identification conditions.
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0 319 0 1

Table 4 Effect of different spectral pretreatments on the recognition rate of calibration and validation samples

Spectral Spectral		Calibrati	Calibration set/%		Validation set/%	
regions/cm ⁻¹	pretreatment	ZNL	NZNL	ZNL	NZNL	
 1909-1311	SNV	87.95	86.36	92.86	80	
1909-1311	SNV+SG(7.3)	84.34	86.36	96.43	90	
1909-1311	SG. (7.3)	79.51	87.95	100	100	
1909-1311	MSC+SG. (7.3)	80.72	81.81	89.29	100	
1909-1311	MSC	84.34	86.36	96.43	90	
1909-1311	MSC+N (7.3)	81.93	81.81	85.71	90	
1909-1311	Norris (7.3)	83.13	86.36	85.71	100	
1909-1311	SNV+Norris (7.3)	85.54	77.27	92.86	90	
1909-1311	SG (9.4)	86.75	89.29	96.43	80	
1909-1311	Norris (9.4)	83.13	90.91	89.28	80	
1909-1311	SNV+Norris (9.4)	81.93	86.36	96.43	90	
1909-1311	MSC+Norris (9.4)	87.95	90.91	89.29	100	

320 To reduce the scattering effects and compare the Lycium

321 samples, the spectral region of the FT-IR $(1909-1311 \text{ cm}^{-1})$

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was subjected to different pretreatments (i.e., normalization, first and second derivative, SG, and Norris. Only the best classification results for the spectral region are presented in the present study (Table 4). Different pretreatments of the spectral region led to a small fluctuation on the recognition rate of the calibration set. The original spectrum is available and valuable. The optimal reference set was obtained by iteratively adding and removing reference samples until the recognition error and the uncorrected number of external estimates were minimized or more precisely balanced. Finally, the no-pretreatment of the spectral region at 1909-1311 cm⁻¹ was regarded as the best choice. Under this condition, the recognition rates of the calibration set were 93.98% and 90.91%, whereas those of the validation set were 100% and 100%.

Conclusions

FT-IR combined with discrimination technique is a powerful tool for Lycium sample discrimination. Two methods were tested on different Lycium produces areas and good results were obtained. DA is an excellent methodology for ZNL discrimination, because it is a rapid technique that does not need a chemical reagent. The optimal identification results of the recognition rates of the ZNL and NZNL calibration set were 93.98% and 90.91%, respectively, whereas the ZNL and NZNL validation set were 100% and 100%, respectively. DA can meet the requirements of simple, feasible, and ordinary data processing with common software. Therefore, a simple, rapid, and reliable overall discrimination of ZNL cultivar was obtained at a low cost, which might be applied for rapid classification of ZNL Lycium cultivar. However, this study presented some limitations, and further work is necessary to obtain more robust classification rules with consideration for regional and time variability. Nevertheless, this study achieved 354 a powerful and practical identification method for ZNL and 355 provided a new trend and perspective for quality evaluation and breed discrimination.

Acknowledgement

- The authors would like to thank the Fundamental Research
- Funds for the Central Universities (QN 2013057) for the

360	financial support
361	Notes and references
362	1. D. Donno, G. L. Beccaro, M. G. Mellano, A. K. Cerutti and G. Bourous, Journal of Functional Foods, 2014
202	G. Bounous, Journal of Functional Foods, 2014.
204 265	2. N. He, A. Yang, Y. Jiao, L. Han and Y. Zhao, Food
202	2 B. I. Badawall, D. Curti, I. Wang, I.M. Dahrushawaka, C. I.
267	5. K. J. Kedgwell, D. Curu, J. wang, J. M. Dobruchowska, G. J.
260	Corbohydrata Dolymars, 2011, 84, 1244, 1240
260	A M Zhang E Wang P Lin X Tang O Zhang and Z Zhang
270	4. M. Zhang, F. Wang, K. Liu, A. Tang, Q. Zhang and Z. Zhang, LWT Food Science and Technology 2014, 59
271	LW 1 - Food Science and Technology, 2014, 58,
371	5 H Amagase and N R Farnsworth Food Research
372	International 2011 44 1702-1717
374	6 H Amagase B Sun and C Borek Nutrition research 2009
375	29 19-25
376	7 S P Gannasin N M Adzahan M Y Hamzah S Mustafa
377	and K. Muhammad. Food Chem. 2015, 182.
378	292-301
379	8. O. Zhao, B. Dong, J. Chen, B. Zhao, X. Wang, L. Wang, S.
380	Zha, Y. Wang, J. Zhang and Y. Wang, Carbohydr
381	Polym, 2015, 127, 176-181.
382	9. GQ. Zheng, ZY. Zheng, X. Xu and ZH. Hu, Biochemical
383	Systematics and Ecology, 2010, 38, 275-284.
384	10. A. S. Luna, A. P. da Silva, J. S. A. Pinho, J. Ferré and R.
385	Boqué, Food Research International, 2015, 67,
386	206-211.
387	11. D. L. Wetzel, Vibrational Spectroscopy, 2012, 60, 29-33.
388	12. YI. Zhang, JB. Chen, Y. Lei, Q. Zhou, SQ. Sun and I.
389	Noda, Journal of Molecular Structure, 2010, 974,
390	144-150.
391	13. R. Karoui, M. Hammami, H. Rouissi and C. Blecker, Food
392	Chem, 2011, 127, 743-748.
393	14. A. Kassouf, J. Maalouly, D. N. Rutledge, H. Chebib and V.
394	Ducruet, Waste Management, 2014, 34, 2131-2138.
395	15. S. Kucheryavskiy, Chemometrics and Intelligent
396	Laboratory Systems, 2013, 120, 126-135.
397	16. S. T. Martín-del-Campo, D. Picque, R. Cosío-Ramírez and
398	G. Corrieu, International Dairy Journal, 2007, 17,
399	835-845.
400	17. S. Wu, Y. Wang, G. Gong, F. Li, H. Ren and Y. Liu, Food

and Bioproducts Processing, 2015, 93, 148-155.

- 18. X. Yao, Y. Peng, Q. Zhou, P. Xiao and S. Sun, Journal of Molecular Structure, 2010, 974, 161-164.
- 19. I. Bondia-Pons, O. Savolainen, R. Törrönen, J. A. Martinez, K. Poutanen and K. Hanhineva, Food Research International, 2014, 63, 132-138.
- 20. H.-Y. Fu, D.-C. Huang, T.-M. Yang, Y.-B. She and H. Zhang, Chinese Chemical Letters, 2013, 24,
- 639-642.
- 21. Q. Fan, C. Chen, Y. Lin, C. Zhang, B. Liu and S. Zhao,
- Journal of Molecular Structure, 2013, 1051, 66-71.

Analytical Methods

1		
2	412	22. Y. K. Kwon, M. S. Ahn, J. S. Park, J. R. Liu, D. S. In, B. W.
3	413	Min and S. W. Kim, Journal of ginseng research,
4	414	2014, 38, 52-58.
5	415	23. D. Bu, B. Wan and G. McGeorge, Chemometrics and
6	416	Intelligent Laboratory Systems, 2013, 120, 84-91.
7	417	24. D. Custers, T. Cauwenbergh, J. L. Bothy, P. Courselle, J. O.
γ Q	418	De Beer, S. Apers and E. Deconinck, Journal of
0	419	pharmaceutical and biomedical analysis, 2014.
9	420	25. C. Muehlethaler, G. Massonnet and P. Esseiva, Forensic
10	421	science international, 2014, 244, 170-178.
11	422	26. B. Aliakbarian, M. Casale, M. Paini, A. A. Casazza, S.
12	423	Lanteri and P. Perego, LWT - Food Science and
13	424	Technology, 2015, 62, 376-383.
14	425	27. W. Jin, L. Xibing, C. Tao and Y. Jinlin, Procedia
15	426	Engineering, 2011, 26, 374-381.
16	427	28. C. Alamprese and E. Casiraghi, LWT - Food Science and
17	428	Technology, 2015, 63, 720-725.
18	429	29. D. Cozzolino, M. Holdstock, R. G. Dambergs, W. U.
19	430	Cynkar and P. A. Smith, Food Chemistry, 2009, 116,
20	431	761-765.
21	432	30. LQ. Zhao, ZQ. Qiu, B. Narasimhamoorthy and J. A.
22	433	Greaves, Industrial Crops and Products, 2013, 47,
23	434	51-57.
24	435	31. F. Liu, Y. He and L. Wang, Analytica chimica acta, 2008,
25	436	615, 10-17.
26		
27		
28		
29		
30		
31		
32		
33		
34		
35		
55		

