ChemComm

Accepted Manuscript

This is an *Accepted Manuscript*, which has been through the Royal Society of Chemistry peer review process and has been accepted for publication.

Accepted Manuscripts are published online shortly after acceptance, before technical editing, formatting and proof reading. Using this free service, authors can make their results available to the community, in citable form, before we publish the edited article. We will replace this *Accepted Manuscript* with the edited and formatted *Advance Article* as soon as it is available.

You can find more information about *Accepted Manuscripts* in the [Information for Authors](http://www.rsc.org/Publishing/Journals/guidelines/AuthorGuidelines/JournalPolicy/accepted_manuscripts.asp).

Please note that technical editing may introduce minor changes to the text and/or graphics, which may alter content. The journal's standard [Terms & Conditions](http://www.rsc.org/help/termsconditions.asp) and the Ethical quidelines still apply. In no event shall the Royal Society of Chemistry be held responsible for any errors or omissions in this *Accepted Manuscript* or any consequences arising from the use of any information it contains.

www.rsc.org/chemcomm

Cite this: DOI: 10.1039/c0xx00000x

www.rsc.org/xxxxxx

ARTICLE TYPE

Direct Detection of the Mercury–**Nitrogen Bond in the Thymine**–**HgII** – **Thymine Base-pair with ¹⁹⁹Hg NMR Spectroscopy†**

Takenori Dairaku,‡^a Kyoko Furuita,‡^b Hajime Sato,‡^c Jakub Šebera,‡d,e Daichi Yamanaka,^a Hiroyuki Otaki,^a Shoko Kikkawa,^a Yoshinori Kondo,^a Ritsuko Katahira,^b F. Matthias Bickelhaupt,f,g Célia Fonseca Guerra,^f Akira Ono,^h Vladimír Sychrovský,*^d Chojiro Kojima,*^b and Yoshiyuki Tanaka*^a 5

Received (in XXX, XXX) Xth XXXXXXXXX 20XX, Accepted Xth XXXXXXXXX 20XX **DOI: 10.1039/b000000x**

We have observed the 1-bond ¹⁹⁹Hg– **¹⁵N** *J***-coupling** $({}^{1}J({}^{199}Hg,{}^{15}N) = 1050$ Hz) within the Hg^{II}-mediated thymine– ¹⁰ thymine base pair (T–Hg^{II}–T). This strikingly large $^{1}J(^{199}\text{Hg},^{15}\text{N})$ is the first one for canonical sp²-nitrogen atoms, **which can be a sensitive structure-probe of N-mercurated compounds and a direct evidence for N-mercuration.**

Mercury-199 NMR spectroscopy is used to probe coordination ¹⁵modes, coordinating elements, and the nature of metals in biomolecules.¹ Within the 199 Hg NMR data, those for N-Hg bonds are of particular importance as metals in proteins and in DNA/RNA molecules frequently interact with nitrogen atoms. Moreover, the $N-Hg^{II}$ bond formation in the Hg^{II} -mediated 20 thymine–thymine base pair $(T-Hg^{II}-T)$ corresponds to an irregular "deprotonative" N–mercuration in water of a bulk proton source. $2-7$ In addition, extraordinary thermal stability with a positive reaction entropy was observed for N-Hg^{II}-N bonding in a DNA duplex. $8-12$ The stability of the Hg-DNA complex can

²⁵be explained partly owing to the metallophilic attraction between Hg atoms in consecutive $T-Hg^{II}-T$ base pairs, and the metallophilic attraction itself is a recent hot topic of inorganic chemistry.¹³⁻¹⁷

Despite such biological/chemical importance, N-Hg^{II} bonds 30 remained uncharacterized. Particularly, the measurements of 1 *J*(199 Hg, 15 N) is challenging, owing to the large chemical shift anisotropy (CSA) of 199 Hg and low natural abundance of 15 N.¹⁸ The only $|^{1}J(^{199}Hg,^{15}N)|$ value of a linear two-coordinate complex was recorded for $(Me_3Si)_2N-Hg^{II}-N(SiMe_3)_2$.¹⁹ The $_{35}$ |¹ $J(^{199}Hg,^{15}N)$ | values for other coordination modes of ^{199}Hg are also limited to Hg^{II}-CyDTA (*trans*-1,2-diaminocyclohexane- NNN '-tetraacetate)²⁰ and Hg^{II} —(NHMe₂)₂Cl₂²¹ complexes (Tables S1 and S2 in ESI†). However, in all cases, some of important parameters such as structure, ¹⁵N or ¹⁹⁹Hg NMR ⁴⁰ chemical shifts $(\delta \Box^{15} N)$ or $\delta \Box^{199} Hg$)), 2-bond $^{15}N^{-15}N$ *J*couplings across Hg^{II}, $(^{2}J(^{15}N,^{15}N))$ or hybridization state of nitrogen atoms always remained unknown. Therefore, a complete ¹⁹⁹Hg/¹⁵N NMR *J*/ δ dataset for a structurally well-defined compound has never been recorded so far.

 45 In this sense, the T-Hg^{II}-T base pair (Figure 1) provides an excellent platform for studying ${}^{1}J(^{199}Hg, {}^{15}N)$, as its chemical and 3-dimensional (3D) structures have been solidly determined^{3,6,7,12,22} and historically accumulated data^{2-4,23,24} are

available. Regarding the NMR parameters of the $T-Hg^{II}-T$ base ⁵⁰ pair, the ¹⁹⁹Hg chemical shift $\delta(^{199}Hg)^{23}$, the 2-bond ¹⁵N-¹⁵N Jcoupling across Hg^{II}, ${}^2J(^{15}N,{}^{15}N)^6$ and $\delta(^{15}N)^6$ were previously determined. Hence, the only missing NMR parameter for characterizing the unique physicochemical properties of the N– Hg^H bond is ¹ $J(^{199}Hg¹⁵N)$. Once it is measured for T-Hg^{II}-T, the 55 T-Hg^{II}-T system will provide a complete J/δ dataset for ¹⁹⁹Hg^{/15}N with a reliable structure, and the ${}^{1}J(^{199}Hg, {}^{15}N)$ value may provide a key concept for constructing molecular devices^{8,25-} 44 from Hg^{II}-DNA complexes.

To measure ${}^{1}J({}^{199}Hg,{}^{15}N)$ in T-Hg^{II}-T, its highly soluble ¹⁵N-60 labeled complex is crucial for 199 Hg^{$/15$}N signal detection. In addition, Hg^{II}-ligand exchanges must be suppressed to avoid the disappearance of ${}^{1}J(^{199}Hg, {}^{15}N)$ owing to exchange broadening. Considering these facts, we determined the $^1J(^{199}Hg, ^{15}N)$ value by using a thymidine- Hg^{II} -thymidine complex (T- Hg^{II} -T). To 65 confirm if the splitting of the ¹⁹⁹Hg resonance is ¹ $J(^{199}$ Hg,¹⁵N), we monitored the disappearance of the splitting upon ¹⁵N-decoupling using a special NMR probe for detecting $15N$ -heteronucleus correlations. Lastly, the derived ${}^{1}J(^{199}Hg, {}^{15}N)$ value was also investigated theoretically with relativistic density functional 70 theory (DFT) including spin-orbit coupling effects.

In this study, we used $15N$ -labeled thymidine to produce $15N$ labeled T-Hg^{II}-T. To suppress the exchange of Hg^{II} ligands, we prepared a sample that contained $T-Hg^{II}-T$ exclusively, without any anion (competitive Hg^{II}-ligands against thymine). Such 75 sample was prepared by the reaction [thymidine + HgO \rightarrow T– Hg^{II} -T + H₂O] followed by H₂O evaporation.²⁴ The resulting pure 15 N-labeled T-Hg^{II}-T was subjected to 199 Hg NMR measurements in dimethyl sulfoxide-d6 (DMSO-d6) (Figure 1). The ¹⁹⁹Hg NMR signal was successfully observed as a triplet so resonance at $\delta(^{199}Hg) = -1784$ ppm, with the absolute ¹J-value $|{}^{1}J({}^{199}\text{Hg},{}^{15}\text{N})| = 1050$ Hz (Figure 1 and Table 1). The observed δ^{199} Hg) value was the same as that observed previously in T- Hg^{II} –T,²³ which ensured successful sampling.

The 199 Hg NMR spectrum under 15 N-decoupling and 15 N NMR ⁸⁵spectrum were recorded to exclude the possibility that the observed splitting of the ¹⁹⁹Hg signal might arise from a structural polymorphism. Notably, the splitting disappeared upon the 15 Ndecoupling (Figure 1b). It should be further noted that this $15N$ decoupled ¹⁹⁹Hg NMR spectrum can't be recorded with ⁹⁰conventionally available probes. This measurement became

possible only by using the special probe, which can perform a $15N-199$ Hg double resonance spectroscopy. In addition, the splitting of the $15N$ resonance (1050 Hz) was observed as satellite peaks at $\delta(^{15}N)$ = 184 ppm in the 1-dimensional ¹⁵N NMR s spectrum (Figure S1 in ESI†). Thus, the splitting of the 199 Hg resonance shown in Figure 1a should be interpreted as 1 *J*(199 Hg, 15 N).

Figure 1. One-dimensional ¹⁹⁹Hg NMR spectrum (71.667 MHz ¹⁰ for 199 Hg frequency) of the thymidine-Hg^{II}-thymidine complex (25 mM) in DMSO-d6 under natural abundance 199 Hg (16.84%). (a) The $1D^{199}$ Hg NMR spectrum without ¹⁵N-decoupling. (b) The $1D^{199}$ Hg NMR spectrum with ¹⁵N-decoupling. The ¹⁹⁹Hg NMR chemical shifts are displayed with respect to dimethylmercury (0 μ ₁₅ ppm) using 1 M HgCl₂ in DMSO-d6 as a secondary reference (– 1501 ppm).⁵⁵ The chemical structure of the $T-Hg^{II}-T$ is depicted above the spectrum, with "R" denoting ribose.

The $|^{1}J(^{199}Hg,^{15}N)|$ value of 1050 Hz for T-Hg^{II}-T was strikingly larger than the ¹J-coupling of $(Me_3Si)_2N-Hg^{II}$ -20 N(SiMe₃)₂ (316.2 Hz),¹⁹ Hg^{II}–CyDTA complexes (365.7–395.5 Hz),²⁰ and Hg^{II}-(NHMe₂)₂Cl₂ (14.7 Hz)²¹ (Table 1 and Tables S1, and S2 in ESI[†]). Thus, the observed $|^{1}J(^{199}Hg,^{15}N)|$ value for T $-Hg^{II}-T$ is the largest of all ¹J-values reported to date.

Here we investigate the correlation between $|^{1}J(^{199}Hg^{15}N)|$ 25 value and N-hybridization state. Within the compounds whose $|{}^{1}J({}^{199}Hg, {}^{15}N)|$ were reported, T-Hg^{II}-T and $(Me_3Si)_2N-Hg^{II}$ -N(SiMe³)2 possess the linear two-coordinate structure, and their $|^{1}J(^{199}Hg,^{15}N)|$ values can be compared. Regarding the Nhybridization state of $(Me_3Si)_2N-Hg^{II}-N(SiMe_3)_2$, an sp²-like

30 planar structure of the nitrogen atoms was suggested from the electron diffraction study,¹⁹ which is further supported by Bent's rule⁴⁵ (see Supporting Discussion in ESI† for Bent's rule). Therefore, the Hg^{II}-bound nitrogen atoms in both samples belong to the $sp²$ category basically, and the current data of $_{35}|^{1}J(^{199}Hg,^{15}N)|$ are insufficient for us to correlate between $|^{1}J(^{199}Hg,^{15}N)|$ and N-hybridization, due to the lack of the ^{1}J values for N(sp)- Hg^{II} and N(sp³)- Hg^{II} bonds.

As a further investigation, $|^{1}J(^{199}Hg, ^{15}N)|$ values for the "sp² nitrogen" in T-Hg^{II}-T and "sp²-like nitrogen" in $(Me_3Si)_2N-$ ⁴⁰ Hg^{II}-N(SiMe₃)₂ were strikingly different (Table 1). However, this may be because the sp²-like N-hybridization in $(Me_3Si)_2N$ - Hg^{II} -N(SiMe₃)₂ might be different from the "canonical sp² nitrogen" in T-Hg^{II}-T. This possibility was also inferred from

 $14/15$ N NMR spectroscopic data⁴⁶, where the ¹⁴N NMR chemical 45 shift for the $Si₂N-Hg^{II}-NSi₂$ linkage showed a rather sp³-like value $(\delta^{14}N) = 66.2$ ppm⁴⁶, Table S1 in ESI†). By contrast, δ ¹⁵N) for Hg^{II}-linked N3 in T-Hg^{II}-T is 184 ppm, and the value is located within the empirical range for an sp^2 -hybridized nitrogen (Figure S1 and Table S1 in ESI†). From these facts, the 50 N-hybridization state of $(Me_3Si)_2N-Hg^{II}-N(SiMe_3)_2$ can't be unambiguously assigned (see also Supporting Discussion in ESI† for details). However, on the basis of the investigations mentioned above, the $|^{1}J(^{199}Hg,^{15}N)|$ value might be a sensitive NMR parameter for detecting differences in the fine electronic ⁵⁵ structures of T-Hg^{II}-T and $(Me_3Si)_2N-Hg^{II}-N(SiMe_3)_2$.

Table 1. Experimental and theoretical ¹⁹⁹Hg NMR parameters.

Ligand	Method	$N-$ hybrid $^{[a]}$	$ {}^1J_{\text{HgN}} ^{[b]}$	$\delta\Box^{199}$ Hg) ^[c]
Thymine ^[d]	Experiment	sp ²	1050	-1784
	Theory ^[e]	sp ²	$931^{[f]}$	-1848
$N(SiMe3)2[g]$	Experiment	sp^2 -like ^[h]	$316.2^{[i]}$	-992 ^[j]
	Theory ^[e]	sp^2 -like ^[h]	278.4 ^[f]	–827

[a] Hybridization state of nitrogen atoms [b] The "absolute" 1bond ¹⁹⁹Hg⁻¹⁵N *J*-coupling, $|^{1}J(^{199}Hg^{15}N)|$, in Hz. [c] ¹⁹⁹Hg NMR chemical shift in ppm with respect to dimethylmercury (0 ppm). ω [d] The T-Hg^{II}-T complex. [e] The theoretical calculation (ZORA-SO-B3LYP/TZ2P) in this work. The average values of ${}^{1}J(^{199}Hg, {}^{15}N)$ and $\delta\Box$ ¹⁹⁹Hg) were calculated for rotational conformers of thymidine-Hg^{II}-thymidine, because the energy barrier for rotation around the N-Hg^{II}-N axis was smaller than 1.1 65 kcal.mol⁻¹. The calculated $\delta\Box^{199}$ Hg) and $^{1}J_{HgN}$ values were therefore averaged over respective rotamers (Table S5 in ESI†). [f] The "–" sign was calculated for *J*-coupling (Table S4 in ESI†). [g] The $(Me_3Si)_2N-Hg^H-N(SiMe_3)_2$ complex. [h] See Supporting Discussion for details. [i] Reference 19 [j] Reference 53. For 70 chemical shift referencing see the footnote to Table S1 in ESI†. It should be noted that ${}^{15}N^6$ and ${}^{1}H^{54}$ chemical shift perturbations for the thymidine-Hg^{II}-thymidine complexation were coherent with those observed for the formation of the T-Hg^{II}-T base-pairs in a DNA duplex (Table S1 in ESI†).

- ⁷⁵ We then quantum chemically computed $|{}^{1}J({}^{199}Hg,{}^{15}N)| = 931$ Hz and $\delta(^{199}Hg)$ = -1848 ppm for T-Hg^{II}-T using DFT including relativistic corrections from the zeroth-order regular approximation (ZORA) with spin-orbit (SO) coupling, as implemented in the ADF program $47-49$ (see Table 1 and Tables so S3-S5 in ESI[†]). The theoretical $|{}^{1}J(^{199}Hg, {}^{15}N)|$ and $\delta(^{199}Hg)$ values agree well with the experimental data (Table 1). With reference to the theoretical values given by Bagno and Saielli $(|^{1}J(^{199}Hg,^{15}N)| = 670$ Hz, $\delta(^{199}Hg) = -1727$ ppm),⁵⁰ the theoretical $|{}^{1}J(^{199}Hg, {}^{15}N)|$ value was refined by using the complex ⁸⁵where it was actually recorded (Tables S3 in ESI†). The $|{}^{1}J({}^{199}\text{Hg},{}^{15}\text{N})|$ value of 278.4 Hz calculated for $(\text{Me}_3\text{Si})_2\text{N-Hg}^{\text{II}}$ $N(SiMe₃)₂$ also agreed satisfactorily with experiment (316.2) Hz)¹⁹. The signs of ¹J(¹⁹⁹Hg,¹⁵N) for T-Hg^{II}-T and (Me₃Si)₂N- Hg^{II} –N(SiMe₃)₂ were both "–" theoretically (Table S4 in ESI†).
- ⁹⁰ In order to investigate the correlation between $|^{1}J(^{199}Hg,^{15}N)|$

values and N-hybridization states theoretically, we further analyzed the theoretical $\frac{1}{J}$ The calculated 1 *J*(199 Hg, 15 N) values were dependent on the "Fermi Contact" + "Spin Dipole coupling" (FC+SD) term (Table S4 in ESI†). With ⁵the dominance of this FC term, one may find the correlation

between the N-hybridization and $|^{1}J(^{199}Hg, ^{15}N)|$ in the future, although it should be experimentally explored.

Empirically, the δ^{199} Hg) values are clustered in terms of linked elements, hybridization states, and other factors of Hg^{II}-

- 10 linked atoms (Table S2 in ESI†). Such phenomena were explained on the basis of the empirical correlation of $\delta^{(199)}$ Hg) with the ionicity of the X–Hg^{II} bond (high ionicity \rightarrow up-field shift of $\delta^{(199)}$ Hg)),⁵¹ Unfortunately, owing to both a paucity of experimental $\delta(^{199}Hg)$ values for a linear two-coordinate N-Hg^{II}-
- $_{15}$ N linkage and the uncertain N-hybridization state in $(Me_3Si)_2N Hg^{II}$ –N(SiMe₃)₂, the correlation between δV^{199} Hg) and the Nhybridization state of Hg^{II}-linked nitrogen also remains obscure. Nevertheless, the highly up-field-shifted $\delta^{(199)}$ Hg) value for T-Hg^{II}-T among those of N-mercurated compounds suggests that
- 20 N(sp²)–Hg^{II}–N(sp²) covalent linkages possess significant ionic character, which agrees with our previous studies (Table 1 and Tables S1, and S2 in ESI†).^{7,52} This observation suggests that $\delta^{(199)}$ Hg) values can be used as a sensitive indicator for probing the Hg^{II} coordination environment not only in C-mercurated ²⁵complexes but also in N-mercurated complexes, including

metalloproteins¹ and metallo-DNA/RNA.

Accordingly, ¹⁹⁹Hg NMR parameters, especially ${}^{1}J(^{199}Hg, {}^{15}N)$, are sensitive parameters for characterizing the electronic structures of N-mercurated complexes and their N-Hg^{II} bonds as

³⁰ well as their Hg atoms. Hence, the ${}^{1}J(^{199}Hg, {}^{15}N)$ value could be a key parameter for predicting the physicochemical properties of N-mercurated complexes and making them into molecular devices, based on a bottom-up approach.

Conclusions

- ³⁵ The $|^{1}J(^{199}Hg,^{15}N)|$ value of 1050 Hz has been reported for canonical sp²-hybridized nitrogen for the first time. From this result, the $T-Hg^{II}-T$ system provides a comprehensive and reliable 199 Hg^{/15}N NMR dataset for probing the Hg^{II} environment in N-mercurated compounds. This newly observed ${}^{1}J(^{199}Hg, {}^{15}N)$
- ⁴⁰coupling can be used for detecting N–Hg bond formations and precisely characterizing these bonds.

Acknowledgements

This work was performed using the NMR spectrometer under the Cooperative Research Program of the Institute for Protein

- ⁴⁵Research, Osaka University. This work was supported by the Platform for Drug Discovery, Informatics, and Structural Life Science from the Ministry of Education, Culture, Sports, Science and Technology (MEXT), Japan. This work was supported by grants-in-aid for Scientific Research (A) (24245037 to A.O. and
- ⁵⁰Y.T), (B) (24310163 to Y.T and C.K.), (C) (18550146 to Y.T) from MEXT, Japan; a Human Frontier Science Program (HFSP) Young Investigator Grant from HFSPO, France (Y.T. and V.S.); and GAČR (P205/10/0228 and 15-21387S to V.S.) from the Czech Republic. T.D. and K.F. are the recipients of a Research
- ⁵⁵Fellowship for Young Scientists from the Japan Society for the

Promotion of Science (JSPS). Y.T. and V.S. were further supported by an Invitation Fellowship for Research in Japan (Short-Term) from JSPS. F.M.B. and C.F.G. were supported by the National Research School Combination - Catalysis (NRSC-C) 60 and the Netherlands Organization for Scientific Research (NWO-CW and NWO-EW).

Notes and references

- *a Graduate School of Pharmaceutical Sciences, Tohoku University, 6-3 Aza-Aoba, Aramaki, Aoba-ku, Sendai, Miyagi 980-8578, Japan. E-mail:*
- ⁶⁵*tanaka@mail.pharm.tohoku.ac.jp b Institute for Protein Research, Osaka University, 3-2 Yamadaoka, Suita,*
- *Osaka 565-0871, Japan. E-mail: kojima@protein.osaka-u.ac.jp c Application, Bruker BioSpin K.K., 3-9 Moriya-cho, Kanagawa-ku, Yokohama, Kanagawa 221-0022, Japan.*
- *d* ⁷⁰*Institute of Organic Chemistry and Biochemistry, Academy of Sciences of the Czech Republic, v.v.i., Flemingovo námestí 2, 16610, Praha 6, ̌ Czech Republic. E-mail: vladimir.sychrovsky@uochb.cas.cz e Institute of Physics, Academy of Sciences of the Czech Republic, v.v.i, Na Slovance 2, CZ-182 21 Prague 8, Czech Republic.*
- *f* ⁷⁵*Department of Theoretical Chemistry and Amsterdam Center for Multiscale Modeling (ACMM), VU University Amsterdam, De Boelelaan 1083, NL-1081 HV Amsterdam, The Netherlands.* ^g Institute for Molecules and Materials (IMM), Radboud University
- *Nijmegen, Heyendaalseweg 135, NL-6525 AJ Nijmegen, The Netherlands. h* ⁸⁰*Department of Material & Life Chemistry, Kanagawa University, 3-27-1 Rokkakubashi, Kanagawa-ku, Yokohama, Kanagawa 221-8686, Japan.* † Electronic Supplementary Information (ESI) available: Additional information as noted in text. See DOI: 10.1039/c0xx00000x/
- ‡ These authors contributed equally to this work.
- ⁸⁵1 L. M. Utschig, J. W. Bryson, T. V. O'Halloran, *Science* 1995, **268**, 380–385.
- 2 S. Katz, *Biochim. Biophys. Acta* 1963, **68**, 240–253; references cited therein.
- 3 L. D. Kosturko, C. Folzer, R. F. Stewart, *Biochemistry* 1974, **13**, 3949–3952
- 4 Z. Kuklenyik, L. G. Marzilli, *Inorg. Chem*., 1996, **35**, 5654-5662.
- 5 Y. Miyake, H. Togashi, M. Tashiro, H. Yamaguchi, S. Oda, M. Kudo, Y. Tanaka, Y. Kondo, R. Sawa, T. Fujimoto, T. Machinami, A. Ono, *J. Am. Chem. Soc.* 2006, **128**, 2172–2173.
- ⁹⁵6 Y. Tanaka, S. Oda, H. Yamaguchi, Y. Kondo, C. Kojima, A. Ono, *J. Am. Chem. Soc.* 2007, **129**, 244–245.
- 7 T. Uchiyama, T. Miura, H. Takeuchi, T. Dairaku, T. Komuro, T. Kawamura, Y. Kondo, L. Benda, V. Sychrovský, P. Bouř, I. Okamoto, A. Ono, Y. Tanaka, *Nucleic Acids Res.*, 2012, **40**, 5766– 100 5774.
	- 8 A. Ono, H. Togashi, *Angew. Chem.* 2004, **116**, 4400–4402; *Angw. Chem. Int. Ed.* 2004, *43*, 4300–4302.
	- 9 H. Torigoe, A. Ono, T. Kozasa, *Chem. Eur. J.*, 2010, **16**, 13218- 13225.
- ¹⁰⁵10 H. Torigoe, Y. Miyakawa, A. Ono, T. Kozasa, *Thermochimica Acta*, 2012, **532**, 28–35.
	- 11 J. Šebera, J. Burda, M. Straka, A. Ono, C. Kojima, Y. Tanaka, V. Sychrovský, *Chem. Eur. J.* 2013, **19**, 9884–9894.
- 12 H. Yamaguchi, J. Šebera, J. Kondo, S. Oda, T. Komuro, T. 110 Kawamura, T. Daraku, Y. Kondo, I. Okamoto, A. Ono, J. V. Burda, C. Kojima, V. Sychrovský, Y. Tanaka, *Nucleic Acid Res.* 2014, **42**, 4094–4099.
	- 13 P. Pyykkö, *Chem. Rev.* 1997, **97**, 597-636.
- 14 F.-A. Polonius, J. Müller, *Angew. Chem.* 2007, **119**, 5698–5701; ¹¹⁵*Angew. Chem. Int. Ed.* 2007, **46**, 5602–5604.
	- 15 L. Benda, M. Straka, Y. Tanaka; V. Sychrovský, *Phys. Chem. Chem. Phys.*, 2011, **13**, 100–103.
	- 16 L. Benda, M. Straka, V. Sychrovský, P. Bouř, Y. Tanaka, *J. Phys. Chem. A,* 2012, **116**, 8313-8320.
- ¹²⁰17 S. Kumbhar, S. Johannsen, R. K. O. Sigel, M. P. Waller, J. Müller, *J. Inorg. Biochem.* 2013, **127**, 203–210.
- 18 J. Mason (Chapters 12) and R. J. Goodfellow (Chapter 21) in *Multinuclear NMR* (Ed.: J. Mason) Plenum Press, New York 1987.
- 19 P. Bernatowicz, S. Szymański, B. Wrackmeyer, *J. Phys. Chem. A* 2001, **105**, 6414–6419.
- ⁵20 E. H. Curzon, N. Herron, P. Moore *J. Chem. Soc., Dalton Trans.* 1980, 721–725.
- 21 S. S. Al-Showiman, *Inorg. Chim. Acta* 1988, **141**, 263–274.
- 22 J. Kondo, T. Yamada, C. Hirose, I. Okamoto, Y. Tanaka, A. Ono, *Angew. Chem.* 2014, *126*, 2417–2420; *Angew. Chem. Int. Ed.* 2014, ¹⁰**53**, 2385–2388.
- 23 A. R. Norris, R. Kumar, *Inorg. Chim. Acta* 1984, **93**, 33–35.
- 24 E. Buncel, C. Boone, H. Joly, R. Kumar, A. R. Norris, *J. Inorg. Biochem.* 1985, **25**, 61–73.
- 25 K. Tanaka, M. Shionoya, *J. Org Chem.* 1999, **64**, 5002–5003.
- ¹⁵26 H. Weizman, Y. Tor, *J. Am. Chem. Soc.* 2001, **123**, *33*75–3376. 27 C. Switzer, S. Sinha, P. H. Kim, B. D. Heuberger, *Angew. Chem.*
	- 2005, **117**, 1553–1556; *Angew. Chem. Int. Ed.* 2005, **44**, 1529–1532.
	- 28 K. Tanaka, G. H. Clever, Y. Takezawa, Y. Yamada, C. Kaul, M. Shionoya, T. Carell, *Nat. Nanotechnology*, 2006, **1**, 190–U195.
- ²⁰29 G. H. Clever, C. Kaul, T. Carell, *Angew. Chem.* 2007, *119*, 6340– 6350; *Angew. Chem. Int. Ed.* 2007, **46**, 6226–6236.
	- 30 J. Müller, *Eur. J. Inorg. Chem.* 2008, 3749–3763.
- 31 A. Ono, S. Cao, H. Togashi, M. Tashiro, T. Fujimoto, T. Machinami, S. Oda, Y. Miyake, I. Okamoto, Y. Tanaka, *Chem. Commun.* 2008, ²⁵4825–4827.
- 32 S. Johannsen, N. Megger, D. Böhme, R. K. O. Sigel, J. Müller, *Nat. Chem.* 2010, **2**, 229–234.
- 33 A. Ono, H. Torigoe, Y. Tanaka, I. Okamoto, *Chem. Soc. Rev.* 2011, **40**, 5855–5866.
- ³⁰34 E. Meggers, P. L. Holland, W. B. Tolman, F. E. Romesberg, P. G. Schultz, *J. Am. Chem. Soc.* 2000, **122**, 10714–10715.
	- 35 S. Atwell, E. Meggers, G. Spraggon, P. G. Schultz, *J. Am. Chem. Soc.* 2001, **123***,* 12364–12367.
	- 36 E. Meggers, *Curr. Opin. Chem. Biol.* 2007, **11**, 287–292.
- ³⁵37 M. K. Schlegel, L.-O. Essen, E. Meggers, *J. Am. Chem. Soc.* 2008, **130**, 8158–8159.
	- 38 K. Tanaka, A. Tengeiji, T. Kato, N. Toyama, M. Shionoya, *Science* 2003, **299**, 1212–1213.
- 39 G. H. Clever, S. J. Reitmeier, T. Carell, O. Schiemann, *Angew.* ⁴⁰*Chem.*, 2010, **122**, 5047–5049; *Angew. Chem. Int. Ed.*, 2010, **49**, 4927–4929.
- 40 T. Carell, C. Behrens, J. Gierlich, *Org. Biomol. Chem.* 2003, **1**, 2221–2228.
- 41 T. Ito, G. Nikaido, S. I. Nishimoto, *J. Inorg. Biochem.* 2007, **101**, 1090–1093
- 42 J. Joseph, G. B. Schuster, *Org. Lett.* 2007, **9**, 1843–1846.
- 43 L. Q.Guo, N. Yin, G. N. Chen, *J. Phys. Chem. C* 2011, **115**, 4837– 4842.
- 44 H. Isobe, N. Yamazaki, A. Asano, T. Fujino, W. Nakanishi, S. Seki, ⁵⁰*Chem. Lett.* 2011, **40**, 318–319.
- 45 H. A. Bent, *Chem. Re*v. 1961, **61**, 275–311.
- 46 O. Just, D. A. Gaul, W. S. Rees, Jr., *Polyhedron* 2001, **20**, 815–821.
- 47 G. te Velde, F. M. Bickelhaupt, E. J. Baerends, C. Fonseca Guerra, S. J. A. van Gisbergen, J. G. Snijders, T. Ziegler, *J. Comput. Chem.* ⁵⁵2001, **22**, 931–967.
- 48 M. Swart, C. Fonseca Guerra, F. M. Bickelhaupt, *J. Am. Chem. Soc.* 2004, **126**, 16718–16719.
- 49 J. M. Fonville, M. Swart, Z. Vokacova, V. Sychrovský, J. E. Šponer, J. Šponer, C. W. Hilbers, F. M. Bickelhaupt, S. S. Wijmenga, *Chem.* ⁶⁰*Eur. J.* 2012, **18**, 12372–12387.
- 50 A. Bagno, G. Saielli, *J. Am. Chem. Soc.* 2007, **129**, 11360–11361.
- 51 D. Rehder, *Coord. Chem. Rev.* 1991, **110**, 161–210.
- 52 Y. Tanaka, A. Ono, *Dalton Trans.* 2008, 4965–4974.
- 53 A. E. Wetherby, Jr. S. D. Benson, C. S. Weinert, *Inorg. Chim. Acta* ⁶⁵2007, **360**, 1977–1986.
- 54 Y. Tanaka, H. Yamaguchi, S. Oda, M. Nomura, C. Kojima, Y. Kondo and A. Ono, *Nucleosides Nucleotides Nucleic Acids*, 2006, **25**, 613–624.
- 55 S. S. Lemos, D. U. Martins, V. M. Deflon, J. Elena, *J. Organomet.*
- ⁷⁰*Chem.* 2009, **694**, 253–258.