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ARTICLE TYPE

Hydroxytyrosol and tyrosol sulfate metabolites protect against oxidized cholesterol pro-oxidant effect in Caco-2 human enterocyte-like cells.

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Abstract

The aim of this study was to investigate the ability of sulfate metabolites of hydroxytyrosol (HT) and tyrosol (TYR) to act as antioxidants counteracting the pro-oxidant effect of oxidized cholesterol in
10 intestinal cells. For this purpose, we synthesized sulfate metabolites of HT and TYR using chemical methodology and examined their antioxidant activity in Caco-2 monolayers in comparison with the parent compounds. Exposure to oxidized cholesterol led to ROS production, oxidative damage, as indicated by MDA increase, a decrease of reduced glutathione concentration and an enhancement of glutathione peroxidase activity. All the tested compounds were able to counteract the oxidizing action of oxidized
15 cholesterol; HT and TYR sulfate metabolites showed an efficiency in protecting intestinal cells comparable to that of the parent compounds, strengthening the assumption that the potential beneficial effect of the parent compounds is retained, although extensive metabolisation occurs, being the resulting metabolites able to exert a biological action themselves.

²⁰ Keywords:

sulfate metabolites, oxidative stress, oxidized cholesterol, olive oil phenolic compounds

Introduction

Polyphenols present in extra virgin olive oil, the principal fat component of the Mediterranean diet, have been demonstrated to exert many potentially beneficial biological effects due to, at least in part, their ability to protect against oxidative stress mediated damage¹⁻². Hydroxytyrosol (3,4-dihydroxyphenylethanol; HT) and tyrosol (4-hydroxyphenylethanol; TYR) (Fig. 1) are the most biologically active phenolic alcohols present in the oil, where they exist in simple forms or as conjugates (such as oleuropein); they are structurally identical except that HT has an extra hydroxyl group in the meta position.

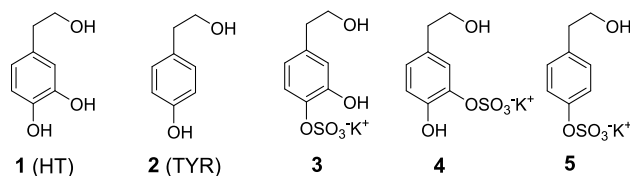


Fig. 1 Structure of tyrosol (TYR), hydroxytyrosol (HT) and their sulfate metabolites 3-5

The literature on phenolic absorption and metabolism after olive oil ingestion is extensive³⁻⁴; however, there are still several issues to be clarified. Some complex olive oil polyphenols, among the secoiridoids, are relatively stable under gastric conditions and reach the intestine where they may be directly absorbed or metabolised under absorption⁵; most of the complex olive oil polyphenols however seems to undergo gastrointestinal biotransformation (see de Bock for an excellent review)⁶, increasing the relative amount of simple phenols, mainly TYR and HT, entering the small and large intestine⁷. At this location the concentration of simple phenols may be quite high, in the high μM range⁷, and they might exert direct protective effects by scavenging reactive species and/or preventing their formation⁸. However, in the process of crossing enterocytes olive oil phenolic compounds are subjected to classical phase I/II biotransformation and to an important first pass metabolism, resulting in almost undetectable concentrations of free HT and TYR in body fluids⁹.

Sulfated and glucuronidated HT and TYR are the predominant metabolites found in human plasma and urine¹⁰⁻¹¹, and they have also been shown to concentrate in the intestinal epithelium, since glucuronidation and sulfation are the major pathways of phase II xenobiotic metabolism in the human intestine¹²⁻¹³. The potential health benefits of HT, TYR and their derivatives are likely to be due to both parental compounds and their phase I and phase II major metabolites. Glucuronidated metabolites of HT showed a more efficient radical scavenging potency than HT itself¹¹ and the ability to protect renal cells¹⁴ and erythrocytes from oxidative injury was also better than the parent compound¹⁵. However no data are available up-to-date concerning the antioxidant activity of sulfate metabolites of HT and TYR.

In this study we investigated the possible protective effect of the sulfate metabolites of TYR and HT, in comparison with the parent compounds, against the oxidative damage to intestinal mucosa due to oxidized cholesterol exposure in the human colon adenocarcinoma cell line, Caco-2. After confluence, these cells spontaneously undergo full differentiation in vitro with enterocyte-like features¹⁶. Caco-2 cells have been recognized as a suitable model for evaluating the effect of nutrient components, for both normal dietary constituents and toxicants, as oxidizing agents¹⁷. Large amounts of lipid oxidation products become available, of both exogenous and endogenous origin, at the level of the intestinal mucosa. Dietary oxysterols, derived from cholesterol degradation and oxidation after prolonged storage or cooking of foods rich in cholesterol, have recently been shown to contribute to the onset and further development of oxidative stress and inflammation related intestinal diseases¹⁸⁻¹⁹. The major oxysterols found in food may contribute to oxidative unbalance of the intestinal epithelium by inducing the generation of reactive oxygen species (ROS)^{18, 20}. Our purpose was to investigate the ability of sulfate metabolites of HT and TYR to act as antioxidants counteracting the oxysterols induced changes of the cellular redox state and compare them with their parent compounds. For this purpose, we synthesized sulfate metabolites of HT and TYR using chemical methodology and evaluate their uptake/stability in Caco-2 monolayers. Cytotoxicity, production of MDA and ROS species, levels of glutathione and glutathione peroxidase activity were measured to assess the protecting effect of the sulfate metabolites in comparison with their parent phenolic compounds.

Materials and methods

Preparation of oxidized cholesterol

Oxidized cholesterol was prepared essentially as previously described²¹. In brief, 2ml aliquots of pure cholesterol (Sigma Aldrich, St. Louis, MO) solution (5 mg/ml in EtOH) were dried down into round-bottomed glass test tubes under vacuum and heated in a 140 °C oil bath in air for 3 h. The oxidized mixture was separated and analysed by GC-MS for the determination of the relative oxysterols. Briefly, 2 µL of the mixture was diluted in 2 mL of EtOH (1: 1000 dilution). After evaporating under a nitrogen stream, 50 µL of pyridine (Sigma Aldrich) and 50 µL of N,O-Bis(trimethylsilyl)trifluoroacetamide with trimethylchlorosilane (BSTFA:TMCS; 99: 1) (Sigma Aldrich) were added, mixed and kept at 60 °C for 45 min. The derivatized sample (1.5 µL) was injected into an Agilent (Waldbronn, DE) GC-MS in splitless mode and the column temperature was programmed starting from 70 °C to 250 °C (10 °C/minute), after up to 290 °C (5 °C/min) maintained for 10 min, ending at 300 °C (2 °C/min) for 5 min (duration about 45 min total). The helium flow was set at 1.3 mL/min. The specific parameters of the instrument were as published by Calderon-Santiago et al.²² Spectra were acquired in scan mode.

GS-MS analysis demonstrated that the mixture contained 58.9% of cholesterol, 13.5% of 7-ketocholesterol, 9.54% of 7β-hydroxycholesterol, 5.6% of 6β-hydroxycholesterol, 5.87% of 7α-hydroxycholesterol, 4.5% of cholesta-4,6-dien-3-ol and 2.1% of 3-keto-4-cholestene.

Synthesis of sulfate metabolites

General methods.

HT and HT acetate were obtained from Seprox Biotech (Madrid, Spain), TYR from Sigma Aldrich. All other chemicals obtained from commercial sources were used without further purification, unless otherwise noted. All reactions were monitored by TLC on precoated Silica-Gel 60 plates F254, and detected by heating with Mostain (500 mL of 10% H₂SO₄, 25g of (NH₄)₆Mo₇O₂₄·4H₂O, 1g Ce(SO₄)₂·4H₂O). Products were purified by flash chromatography with Merck Silica gel 60 (200-400 mesh). High resolution mass spectra were obtained on an ESI/quadrupole AutoSpec-Q mass spectrometer. NMR spectra were recorded on 300 or 500 MHz spectrometers, at room temperature for solutions in CDCl₃, or D₂O. Chemical shifts are referred to the solvent signal. Metabolites were purified by chromatography with Reverse Phase-C18 Silica gel. Data were processed using manufacturer software, raw data were multiplied by shifted exponential window function prior to Fourier transform, and the baseline was corrected using polynomial fitting.

General procedure for the microwave-assisted O-sulfation.

Microwave based sulfation reactions were performed using a microwave synthesizer in sealed reaction vessels. Phenolic derivatives (1.0 equiv), sulfur trioxide-trimethylamine complex, SO₃NMe₃, (5 equiv per OH; being this complex previously washed with H₂O, MeOH, and CH₂Cl₂ and dried under high vacuum) and a magnetic stirrer bar were placed in a 2-5 mL microwave reaction vial and fitted with a septum, which was then pierced with a needle. The closed vial was then evacuated in high vacuum for 2 h. The mixture was dissolved in dry CH₃CN (2.0 mL) and NEt₃ (0.3-1.0 mL) was then added. Reaction mixture was subjected to microwave radiation for 20–40 min (depending on the compound) at 100 °C (50-60W average power). MeOH (1 mL) and CH₂Cl₂ (1 mL) were added, and the solution was layered on the top of a Sephadex LH-20 chromatography column which was eluted with CH₂Cl₂/MeOH (1:1) to obtain the corresponding triethylammonium salt as a white powder (94–98% yield).

Triethylamine, 4-(2-(butyryloxy)ethyl)phenyl sulfate salt (7). Tyrosol butyrate **6**²³⁻²⁴ (70 mg, 0.33 mmol) and SO₃NMe₃ (233 mg, 1.7 mmol) were submitted under sulfation conditions for 20 min. TLC (Ethyl acetate: MeOH, 10:1) showed the formation of a major product and complete consumption of the initial material. Solvents were removed and the crude extract was purified by sephadex LH-20 (CH₂Cl₂:MeOH, 1:1) to afford **7** (92 mg, 98%) as a white powder. ¹H-NMR (300 MHz, D₂O) δ: 6.72, 7.18 (2d, 4H, J = 8.7 Hz, Harom), 4.24 (t, 2H, CH₂OAc), 3.10 (q, 6H, -CH₂CH₃), 2.88 (t, 2H, CH₂Ar), 2.19 (t, 2H, J = 7.5Hz, CH₂), 1.45 (m, 2H, CH₂), 1.18 (t, 9H, CH₂CH₃), 0.74 (t, 3H, CH₃); ¹³C-NMR (125 MHz, D₂O) δ: 177.1 (CO), 150.0, 136.2, 130.3 (2x CHarom), 121.3 (2x CHarom), 65.4 (CH₂OAc), 46.5, 36.0 (CH₂Ar), 34.0, 18.3, 12.6, 8.0 (CH₃). ESI-HRMS (ES⁻) Calcd. for C₁₂H₁₅O₆S (M-H) 287.0589, Found: 287.0594.

Potassium 4-(2-hydroxyethyl)phenyl sulfate (5). Compound **7** (97 mg, 0.32 mmol) was dissolved in MeOH (10 mL) and K₂CO₃ (90 mg, 0.66 mmol) was added. The reaction mixture was stirred at room temperature for 24 h and then neutralized with IR 120 H⁺ resin. Solvent was then removed in vacuum and the crude extract was purified by an RP-C18 column eluted with H₂O: MeOH (from 100:0 to 70:30). Fractions containing the desired product were concentrated and freeze-dried affording compound **5** as a white solid (68 mg, 94%). ¹H-NMR (400 MHz, D₂O) δ: 7.18, 6.86 (2d, 4H, J = 8.4 Hz, Harom), 3.78, 2.78 (2t, 4H, J = 6.7 Hz, CH₂OH, CH₂Ar); ¹³C-NMR (75 MHz, D₂O) δ: 157 (Cq), 130.3 (2x CHarom), 129.8 (Cq), 117.1 (2x CHarom), 63.0 (CH₂OH), 36.9 (CH₂Ar). ESI-HRMS (ES⁻) Calcd. for C₈H₉O₅S (M-H) 217.0171, Found: 217.0171.

3-((tert-butyldimethylsilyloxy)-4-hydroxyphenethyl acetate (9) and 4-((tert-butyldimethylsilyloxy)-3-hydroxyphenethyl acetate (10). To a solution of hydroxytyrosol acetate **8**²³⁻²⁴ (223 mg, 1.13 mmol) in DMF (anhydrous, 3 mL) cooled in an ice-water bath under argon were added sequentially tert-butyldimethylsilyl-trifluoromethanesulfonate (TBDMSOTf, 287 µL, 1.25 mmol, 1.10 equiv) and diisopropylethylamine (i-Pr₂Net, 265 µL, 1.52 mmol, 1.35 equiv). The mixture was allowed to stir for 30 min at 0 °C, and TLC (hexane:

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ethyl acetate 3:1) at that point indicated that the reaction was complete. The pale yellow reaction mixture was diluted with EtOAc (100 mL), cast into a separatory funnel, and washed with water (2x50 mL), brine (50 mL), and the organic phase was dried (Na₂SO₄). Filtration and concentration in vacuum afforded the crude extract that was purified by flash column chromatography (hexane:ethyl acetate from 15:1 to 10:1) to afford **9** and **10** (314 mg, 90%, powder) like a regioisomeric mixture in ratio ~ 1:1. ¹H NMR (400 MHz, CDCl₃) δ 6.88 (d, 1H, *J* = 8.1 Hz, Harom), 6.83 (s, 1H, Harom), 6.77 (d, 1H, *J* = 8.4 Hz, Harom), 6.73 (d, 1H, *J* = 8.4 Hz, Harom), 6.71 (s, 1H, Harom), 6.62 (d, 1H, *J* = 8.1 Hz, Harom), 5.52, 5.45 (2s, 2H, 2xOH), 4.26 (t, 2H, *J* = 6.7 Hz, CH₂OAc), 4.24 (t, 2H, *J* = 6.4 Hz, CH₂OAc), 2.86 (t, 2H, *J* = 6.7 Hz, CH₂Ar), 2.84 (t, 2H, *J* = 6.4 Hz, CH₂Ar), 2.07-2.05 (2s, 6H, CH₃C=O), 1.05, 1.03 (2s, 18H, C(CH₃)₃ x2), 0.30, 0.29 (2s, 12H, -Si(CH₃)₂ x2); ¹³C-NMR (125 MHz, CDCl₃) δ: 171.1, 171.0 (C=O), 147.1, 145.9, 142.3, 141.0, 131.8, 129.6 (Cqarom), 122.4, 120.2, 118.6, 117.7, 115.4, 114.8 (CHarom), 65.2, 65.0 (CH₂OAc), 34.5 (2x CH₂Ar), 25.7 (C(CH₃)₃), 21.0, 20.9 (CH₃C=O), 18.2 (C(CH₃)₃), -4.2 (Si(CH₃)₂); HRMS (ES⁺) Calcd. for C₁₆H₂₆O₄NaSi (M+Na) 333.1498, Found: 333.1508.

Triethylamine, 5-(2-acetoxyethyl)-2-((tert-butyl dimethylsilyl)oxy)phenyl sulfate salt (11) and **triethylamine, 4-(2-acetoxyethyl)-2-((tert-butyl dimethylsilyl)oxy)phenyl sulfate salt (12)**. Regioisomeric mixture of compounds **9** and **10** (157 mg, 0.506 mmol) and SO₃NMe₃ (351 mg, 2.52 mmol) were submitted under sulfation conditions for 2x20 min. TLC (Ethyl acetate:MeOH 10:1) showed the formation of a major product and complete consumption of the starting material. Solvents were removed and crude was purified by sephadex LH-20 in a solvent mixture of CH₂Cl₂: MeOH 1:1 to afford **11** and **12** (231 mg, 94%, powder) like a regioisomeric mixture in ratio ~ 1:1. ¹H-NMR (400 MHz, CDCl₃) δ: 7.50 (d, 1H, *J* = 7.9 Hz, Harom), 7.46 (s, 1H, Harom), 6.82 (d, 1H, *J* = 8.2 Hz, Harom), 6.78 (d, 1H, *J* = 8.2 Hz, Harom), 6.73 (d, 1H, *J* = 7.9 Hz, Harom), 6.72 (s, 1H, Harom), 4.20 (t, 4H, *J* = 7.08 Hz, CH₂OAc), 3.10-3.00 (dq, 12H, CH₂CH₃), 2.83 (t, 2H, *J* = 7.1 Hz, CH₂Ar), 2.82 (t, 2H, *J* = 7.05 Hz, CH₂Ar), 2.04, 2.03 (2s, 6H, CH₃C=O), 1.26 (t, 18H, CH₂CH₃), 1.00, 0.99 (2s, 18H, C(CH₃)₃ x2), 0.21, 0.20 (2s, 12H, -Si(CH₃)₂ x2); ¹³C-NMR (125 MHz, CDCl₃) δ: 171.0, 170.9 (C=O), 146.8, 145.6, 143.9, 142.8, 134.0, 130.7 (Cqarom), 124.5, 122.4, 121.9, 121.8, 121.6, 121.0 (CHarom), 65.1 (CH₂OAc), 46.3 (CH₂CH₃), 34.5, (CH₂Ar), 25.7 (C(CH₃)₃), 21.0 (CH₃C=O), 18.8 (C(CH₃)₃), 8.8 (CH₂CH₃), -4.2 (Si(CH₃)₂); ESI-HRMS (ES⁻) Calcd. for C₁₆H₂₅O₇SiS (M-H) 389.1090, Found: 389.1092.

Potassium 2-hydroxy-4-(2-hydroxyethyl)phenyl sulfate (3) and **potassium 2-hydroxy-5-(2-hydroxyethyl)phenyl sulfate (4)**. Regioisomeric mixture of **11** and **12** (231 mg, 0.47 mmol), potassium fluoride (KF, 55 mg, 0.94 mmol) and potassium carbonate (K₂CO₃, 130 mg, 0.94 mmol) were dissolved in MeOH (10 mL). The reaction mixture was stirred at room temperature for 18 h and solvent was then removed in vacuum. The crude extract was purified by column chromatography with RP-C18 silica gel eluting with H₂O: MeOH (from 100:0 to 70:30). Fractions containing the desired product were concentrated and freeze-dried affording compounds **3** and **4** (115 mg, 90%, white powder) like a regioisomeric mixture in ratio ~ 1:1. ¹H-NMR (300 MHz, D₂O) δ: 7.21 (d, 2H, *J* = 8.0 Hz, Harom), 7.17 (s, 1H, Harom), 6.98 (d, 1H, *J* = 8.5 Hz, Harom), 6.88 (d, 1H, *J* = 8.5 Hz, Harom), 6.80 (s, 1H, Harom), 6.70 (d, 1H, *J* = 8.0 Hz, Harom), 3.76-3.69 (m, 4H, CH₂OAc), 2.73-2.70 (m, 4H, CH₂Ar); ¹³C-NMR (125 MHz, D₂O) δ: 149.4, 147.4, 139.0, 138.4, 137.8, 130.7, 127.6, 123.0, 122.6, 120.0, 118.2, 117.6, 62.6, 62.4 (CH₂OAc), 37.4, 36.9 (CH₂Ar). HRMS-ESI (ES⁻) Calcd. for C₈H₉O₆S (M-H) 233.0120, Found: 233.0126.

Cell culture

Cell culture materials were purchased from Invitrogen (Milano, Italy). Caco-2 cells were obtained from the European Collection of Cell Cultures (ECACC, Salisbury UK) and grown in Dulbecco's modified Eagle's medium (DMEM), supplemented with 2.5% of heat-inactivated bovine serum and 100 U/mL penicillin and 100 μg/mL streptomycin, at 37 °C under a humidified atmosphere of 95% air and 5% CO₂. For experimental studies Caco-2 cells, at passage 45–60, were plated at a density of about 1 × 10⁵/mL and used 21 days post seeding.

Cytotoxic activity

The cytotoxic effect caused by the exposure to increasing concentrations of oxidized cholesterol and the protective effect of HT, TYR, and sulfate metabolites was assessed on Caco-2 cells, seeded in 24-well plates, by the Neutral Red method²⁵. Cells were exposed to oxidized cholesterol (0–175 μg/mL) in complete medium and incubated for 24 h. In order to assess the protective effect of the phenolic compounds cells were pretreated with the tested compounds (2.5–10 μM in water solution, 30 min) prior to oxidized cholesterol exposure (100 μg/mL for 24 h). After incubation, the medium was removed; a Neutral Red Solution (Sigma Aldrich) (0.033% in medium) was then added to the wells to determine cell viability. After 30 min of incubation, the Neutral red solution was carefully removed and cells quickly rinsed with PBS. The incorporated dye was then solubilised in Neutral Red Solubilisation Solution, acetic acid/ethanol/water

(1/45/50, v/v/v), and the absorbance was measured at 540 nm.

Determination of MDA

MDA levels were determined in the medium of treated cells by the TBARS test with HPLC quantification, using the method described by Templar et al.²⁶, with some modifications. Briefly, 100 μ L of 10% trichloroacetic acid (TCA, Sigma Aldrich) was added to 400 μ L of the medium, samples were mixed and left at room temperature. After 20 min, 200 μ L of 2-thiobarbituric acid (TBA, Sigma Aldrich) (0.6%) were added; samples were incubated at 90 °C for 45 min and then centrifuged at 5000 x g for 15 min at 4 °C. Aliquots of the supernatant were injected into an Agilent 1100 HPLC system (Agilent Technologies, Santa Clara, CA) equipped with a diode-array detector (HPLC–DAD) and separation was achieved using a Varian (Middelburg, The Netherlands) Inertsil 5 ODS-2, 150 \times 4.6 mm column; the mobile phase was a mixture of KH₂PO₄ 50 mM pH 7/MeOH (65/35, v/v) at a constant flow rate of 1 mL/min. The formation of the MDA–TBA adduct was revealed measuring its absorbance at 532 nm. A standard curve was prepared using a 1,1,3,3,-tetraethoxypropane (TEP, Sigma Aldrich) solution in PBS (0.05–10 μ M).

ROS production in Caco-2 cells

Intracellular ROS production was monitored by adding the 2',7'-dichlorodihydrofluorescein diacetate (H₂-DCF-DA, Sigma Aldrich), according to Dinicola et al.²⁷, in Caco-2 cells exposed to oxidized cholesterol (75 μ g/mL) in complete medium and incubated for 0-180 min and in cells pretreated with the tested phenolic compounds (5-25 μ M in water solution, 30 min), prior to exposure to oxidized cholesterol (75 μ g/mL for 30 min), with some modifications. After incubation the culture medium was replaced with PBS and cells were loaded with 10 μ M H₂-DCF-DA for 30 min. After incubation, PBS and H₂-DCF-DA were removed and the cells were washed twice. Increase in cell fluorescence was measured at excitation and emission wavelengths of 490 and 520 nm, respectively, using an Infinite 200 auto microplate reader (Tecan, Salzburg, Austria), at 25 °C.

Glutathione (GSH) level and Glutathione peroxidase (GPx) activity

To assess the changes in the levels of GSH and GPx activity, cells were seeded in plate dishes and exposed to oxidized cholesterol (75 μ g/mL) in complete medium for 0-24 h. In order to assess the effect of HT, TYR, HT-S and TYR-S, cells were pretreated with the tested compounds (5-25 μ M in water solution, 30 min) prior to exposure to oxidized cholesterol 75 μ g/mL for 30 min to determine GSH level and for 18 h to measure GPx activity. At the end of incubation time, cells were washed with PBS, and subsequently scraped into 500 μ L of 5% metaphosphoric acid. Samples were then sonicated and centrifuged at 10000 \times g for 20 min at 4 °C; supernatants were collected and used to determine GSH level and GPx activity, using Glutathione Assay Kit and Glutathione peroxidase Assay Kit (Cayman Chemical Company, Ann Arbor, USA) according to the manufacturer's instructions.

Statistical analysis

Data are expressed as means \pm S.D (n=12 for each sample/condition). The statistical evaluation of the results was performed by analysis of variance (ANOVA) followed by a Bonferroni post-hoc test using GraphPad InStat version 3.05 (GraphPad Software, San Diego, CA, USA).

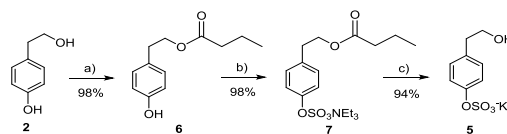
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Results

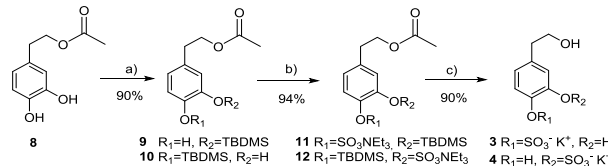
Synthesis of HT and TYR sulfate metabolites has been carried out using a protection-deprotection strategy together with the use of microwaves in the critical sulfation step. In the case of TYR, the primary alcohol was acyl-protected using immobilized lipase Novozym 435® and vinyl butyrate in *tert*-butyl methyl ether (98 % yield) (Scheme 1). Next, microwave-assisted sulfation was performed by treatment with $\text{SO}_3\cdot\text{NMe}_3$ complex and triethylamine in acetonitrile at 100 °C (98% yield). Final deprotection with K_2CO_3 in MeOH afforded tyrosol sulfate **5** in good yield (94%).



a) Novozym435, vinyl butyrate, TBME; b) $\text{SO}_3\cdot\text{NMe}_3$, TEA, CH_3CN , 100°C, 20 min, MW; c) K_2CO_3 , MeOH;

Scheme 1 Preparation of tyrosol sulfate **5**

Mono-sulfated hydroxytyrosol derivatives **3** and **4** were prepared as a regioisomeric mixture using a synthetic strategy similar to that of TYR. In this case, hydroxytyrosol acetate **8** was mono-silyl protected to avoid disulfated products difficult to separate from the mono-sulfated derivatives (Scheme 2).



a) TBDMSTf, DIPEA, CH_2Cl_2 ; b) $\text{SO}_3\cdot\text{NMe}_3$, TEA, CH_3CN , 100°C, 20 min, MW; c) KF, K_2CO_3 , MeOH.

Scheme 2 Preparation of hydroxytyrosol sulfate metabolites **3** and **4**

Random TBDMS-protection of HT acetate **8** and subsequent chromatographic separation afforded a 1:1 regioisomeric mixture of the two possible mono-phenolic compounds **9** and **10**. The same sulfation reaction conditions used before were applied to give 94% yield of a 1:1 mixture of isomers **11** and **12**. Finally, acetyl and silyl deprotection was carried out in one step using KF and K_2CO_3 in MeOH to obtain a mixture of mono-sulfated hydroxytyrosol derivatives **3** and **4** (90% yield).

To investigate the potential protective effect of the HT and TYR sulfate metabolites, in comparison with their parent compounds, against the pro-oxidant effect of oxidized cholesterol in intestinal cells, Caco-2 monolayers were treated for 24 h with a mixture obtained from the oxidation of cholesterol at 140 °C for 3 h. In these oxidizing conditions half of the initial cholesterol was turned into oxidation products (reported in the methods section). Treatment with growing concentration of the mixture induced production of MDA and cell death (Fig. 2), indicating the presence of oxidative cell injury.

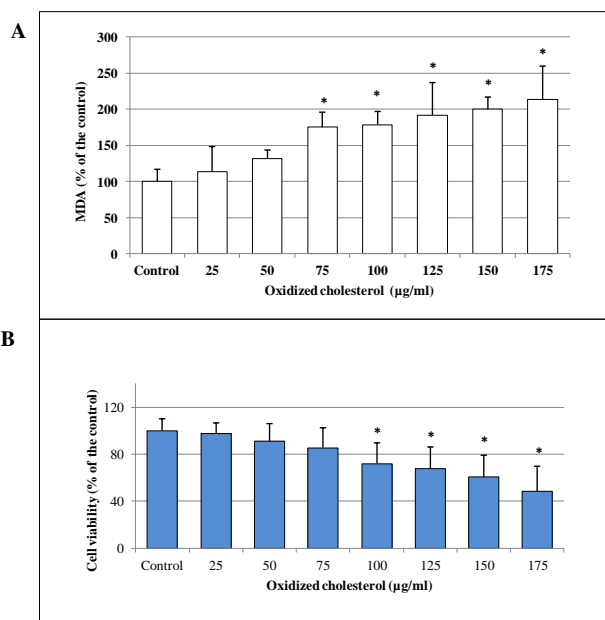


Fig.2 Values of MDA (2A) and cell viability (2B) measured in Caco-2 cells after 24 h incubation with different concentrations of oxidized cholesterol.

* = $p < 0.05$ versus control

Pretreatment with the phenolic compounds significantly inhibited the increase of MDA (Fig. 3) in Caco-2 cells treated with 75 µg/mL of the oxidizing mixture, the highest amount able to induce a significant production of MDA but not cell death. MDA reduction was significant from 2.5 µM for HT-S and from 5 µM for HT; TYR and its sulfate metabolite exerted the same efficacy starting from 2.5 µM.

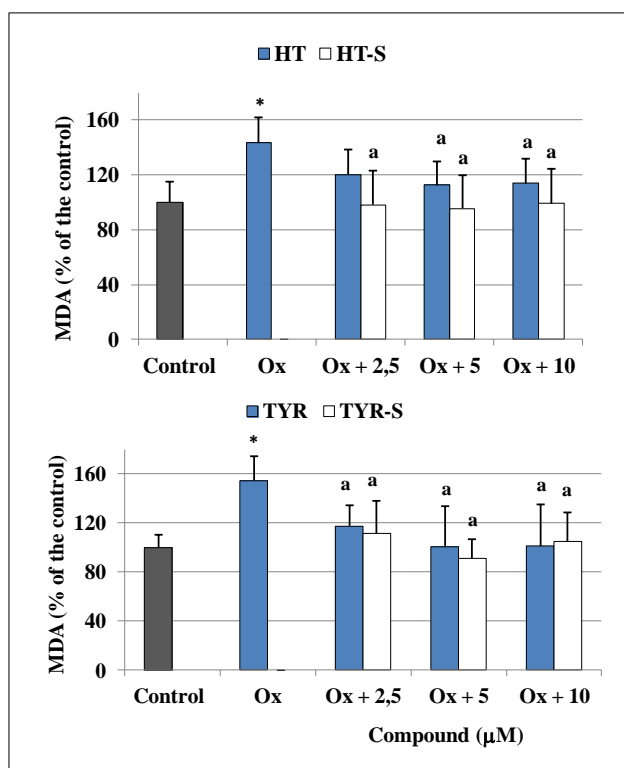


Fig.3 Values of MDA measured in Caco-2 cells after 24 h incubation with 75 µg/mL oxidized cholesterol and pretreated (30 min) with TYR, HT, HT sulfate metabolites 3-4 (HT-S) and TYR sulfate metabolite 5 (TYR-S) (2.5-10 µM). * = $p < 0.05$ versus control, a = $p < 0.05$ versus oxidized cholesterol treated (Ox).

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Next we examined the ability of HT, TYR and their corresponding sulfate metabolites to protect cells against death in Caco-2 cells treated with 100 $\mu\text{g/mL}$ of oxidized cholesterol, the lowest amount able to induce a significant cell death (Fig. 4). We observed that HT preserved cell viability from the concentration of 5 μM whereas HT sulfates 3-4 showed protection at the 10 μM concentration. In the case of TYR and its sulfate derivative, both improved cell viability at 10 μM concentration.

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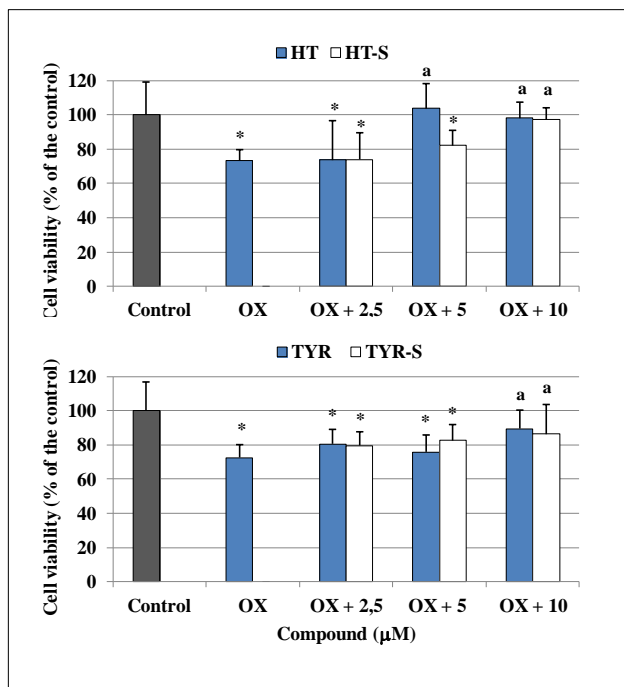


Fig.4 Cell viability measured in Caco-2 cells after 24 h incubation with 100 $\mu\text{g/mL}$ oxidized cholesterol and pretreated (30 min) with TYR, HT or their sulfate metabolites (2.5-10 μM). * = $p < 0.05$ versus control, a= $p < 0.05$ versus oxidized cholesterol treated (Ox).

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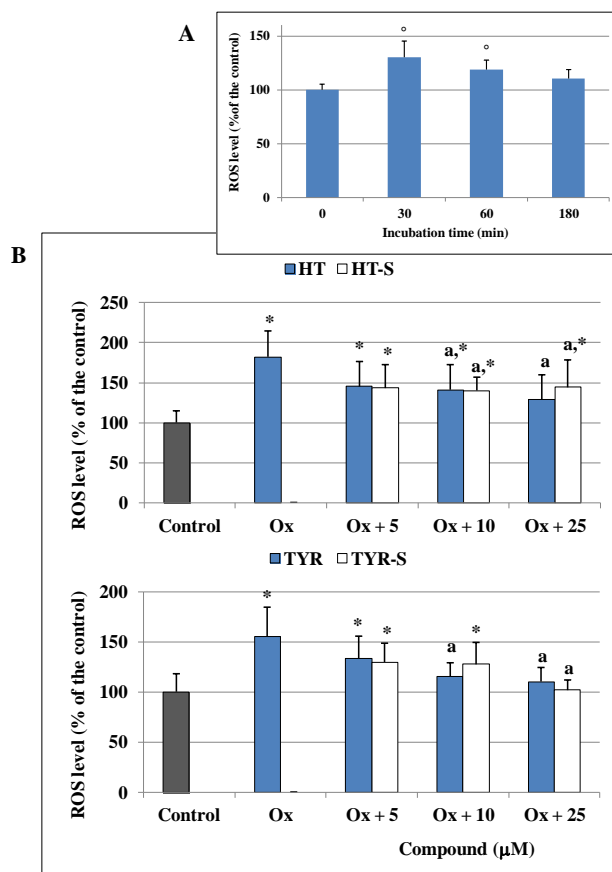


Fig.5 ROS level, expressed as % of the control samples, in Caco-2 cells, treated with 75 μg/ml oxidized cholesterol for different incubation times (5A) or pretreated (30 min) with TYR, HT or their sulfate metabolites (5-25 μM) and treated with oxidized cholesterol for 30 min (5B), exposed to 2',7'-dichlorodihydrofluorescein diacetate H₂-DCF-DA (10 mM) for 30 min. * = p<0.05 versus control, a = p<0.05 versus oxidized cholesterol treated (Ox), ° = p<0.05 versus 0 min

The oxidizing action of the mixture of oxidized cholesterol was then investigated monitoring the alteration of the cellular redox status with time: after 30 min of incubation a significant production of ROS was observed in the cells treated with the oxidized cholesterol in comparison with the control (Fig. 5A). Next, we measured ROS production at 30 min, after pretreatment with the different phenolic compounds (5, 10, 25 μM). We observed that ROS production was significantly lower from the concentration of 10 μM for all the tested compounds, except for TYR-S (Fig. 5B).

After 30 min of incubation with the oxidized cholesterol, ROS production was associated with a significant reduction of Caco-2 cellular GSH, around 40% of the initial value, as shown in Figure 6A. The level of GSH increased with time, reaching the control value at 3/6 h, to decrease again thereafter. Pretreatment with both HT and TYR preserved the control level of GSH at all the tested concentrations and their sulfate metabolites exerted a comparable efficacy (Fig. 6B). Interestingly, none of the tested compounds was able to alter GSH level when incubated alone (25 μM) in the experimental conditions used (p>0.5 versus control).

With respect to the considered parameters, we did not observe any concentration dependent protective effect of the tested phenolic compounds, nor significant differences among parent compounds and metabolites.

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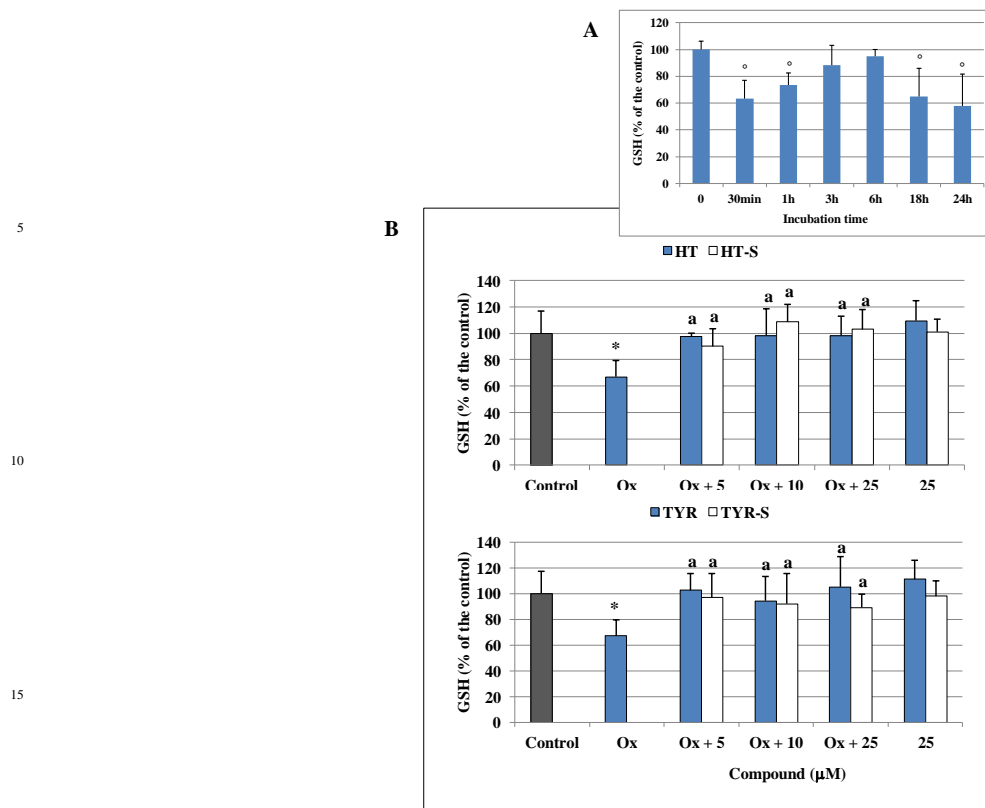


Fig.6 GSH level, expressed as % of the control samples, in Caco-2 cells, treated with 75 μg/mL oxidized cholesterol for different incubation times (6A) or pretreated (30 min) with TYR, HT or their sulfate metabolites (5-25 μM) and treated with oxidized cholesterol for 30 min (6B). * = p<0.05 versus control, a = p<0.05 versus oxidized cholesterol treated (Ox), ° = p<0.05 versus 0min

A significant increase of GPx activity, around 50% above the control level, was also observed in Caco-2 cells treated with the oxidized cholesterol after 18 h of incubation (Fig. 7A). HT (25 μM) induced an increase, although not significant, of the basal GPx activity; however, once incubated together with the oxidized cholesterol HT reduced the increase of the enzyme activity from 10 μM with respect to the oxidized samples (Fig. 7B). HT sulphates **3** and **4** did not alter the control level but exerted the same activity of the parent compound for the oxidized samples.

In presence of TYR, GPx activity was significantly above the control level; a small effect was observed in pretreated samples with respect to the oxidized ones. In contrast, Caco-2 cells pretreated with all tested concentrations of TYR sulphate **5** resulted in an enzyme activity significantly lower with respect to the oxidized samples, and the enzyme basal activity was not influenced (Fig. 7B).

Finally, we evaluated the stability and uptake of HT, TYR and their sulfate metabolites from 30 min to 24 h of incubation in the Caco-2 cultures. UPLC-MS analyses (see supporting information) revealed a partial loss of the compounds in the culture medium, maybe due to the action of the fetal bovine serum present or to the intrinsic stability to air oxidation of each compound. Analysing medium and supernatants (from lysed cells) we observed that all the tested compounds entered the cells from 30 min of incubation and undergo an extensive metabolisation, giving rise to a pool of metabolites, among which we detected mainly sulfate and methyl-sulfate metabolites.

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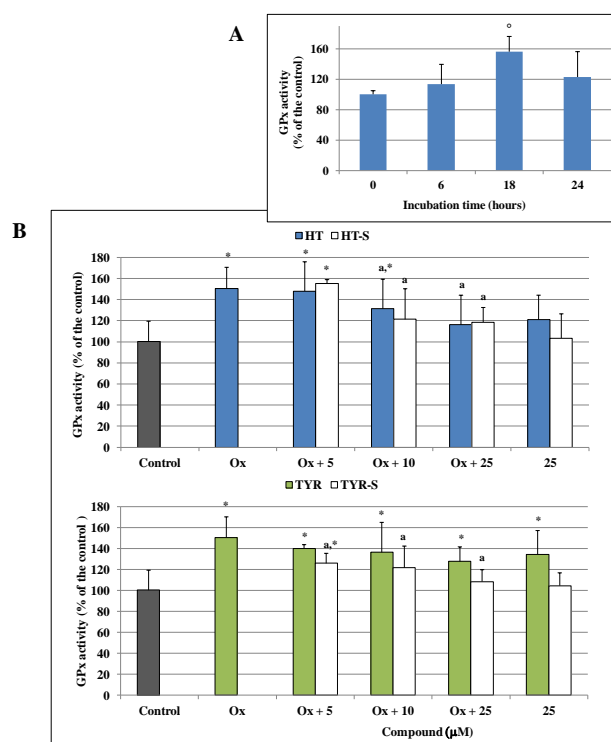


Fig.7 GPx activity, expressed as % of the control samples, in Caco-2 cells, treated with 75 $\mu\text{g/mL}$ oxidized cholesterol for different incubation times (7A) or pretreated (30min) with TYR, HT or their sulfate metabolites (5-25 μM) and treated with oxidized cholesterol for 18 h (7B). * = $p < 0.05$ versus control, a = $p < 0.05$ versus oxidized cholesterol treated (Ox), ° = $p < 0.05$ versus 0 h

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Discussion

It is known that polyphenols undergo strong phase I and phase II metabolism, in which they are hydrolyzed (phase I) and later conjugated (phase II) into their glucuronidated, methylated and sulfated forms in order to be absorbed and excreted. Recently, such metabolites are being taken into consideration to provide in vivo relevance for any activity.

Sulfate metabolites of phenolic compounds as resveratrol²⁸⁻³⁰, quercetin, epicatechin and catechin³¹⁻³³, have been shown to exert a biological activity comparable to that of the parent compound. Quercetin-3-O-sulfate has been shown to retain an antioxidant effect, significantly inhibiting the Cu²⁺-induced oxidation of human LDL³⁴. The free radical scavenging activity of a 3-sulfate metabolite of resveratrol was comparable to that of the parent compound, when measuring the ability to quench the DPPH radical²⁸.

In this work we have studied the potential protecting activity of HT and TYR sulfate metabolites in Caco-2 human endothelial intestinal cells where oxidative stress was provoked by oxidized cholesterol. Although the liver is the major organ for metabolism of many drugs and xenobiotics, the small intestine also contributes substantially to it by several pathways involving phase I and phase II reactions. At the same time, concentration of sulfate metabolites in the intestinal epithelium may be high, as glucuronidation and sulfation are major pathways of phase II xenobiotic metabolism in the human intestine¹²⁻¹³, and might contribute to preserve intestinal epithelium integrity against pro-oxidant dietary agents.

We selected oxidized cholesterol for our oxidative stress cell model since cholesterol and its oxidation products have recently been shown to potentially interfere with the homeostasis of the intestinal epithelium^{18, 20}, due to their pro-oxidant and pro-inflammatory properties. Oxysterols are more polar and more readily diffusible through cell membranes, and have consistently been shown to be more reactive than unoxidized cholesterol, possessing marked pro-inflammatory and cytotoxic effects in a number of cells and tissues³⁵⁻³⁶. In our oxidizing conditions, about 50% of the initial cholesterol was turned into oxidation products, mainly 7-ketocholesterol and some of the hydroxycholesterol isomers commonly found in cholesterol rich foodstuffs³⁷. It has been reported that dietary cholesterol and some oxysterols of dietary origin are able to lead differentiated Caco-2 cells to both necrotic and apoptotic death, depending on the experimental conditions^{18, 38-40}.

In the present study, oxidized cholesterol toxic effects were investigated in terms of alteration of cellular redox state and oxidative damage. Oxidized cholesterol exposure resulted in a significant production of ROS, in accordance with results reported by Biasi et al.^{18, 41}, being this likely the cause of oxidative damage and death. ROS production in Caco-2 cells treated with specific oxysterols has been reported to be due, at least in part, to the activation of NADPH oxidase¹⁸. In our experimental conditions ROS production was inversely correlated to a decrease of GSH level. GSH is the main non-enzymatic antioxidant defence within the cell, reducing different peroxides, hydroperoxides, and radicals (alkyl, alkoxy, peroxy, etc.). It is usually assumed that GSH depletion reflects intracellular oxidation whereas an increase in GSH concentration could be expected to prepare the cell against a potential oxidative insult⁴²⁻⁴³.

In our oxidative stress model, in cells pretreated with HT, we observed an inhibition in the formation of ROS and also prevention of GSH level decrease. HT protective effect could predominantly be attributed to the capacity of acting as free radical scavenger, as already documented in the same cells treated with other oxidizing agents, TBH⁸ and acrylamide⁴⁴. The well known free radical scavenging and metal chelating activities are mostly due to HT ortho-diphenolic structure, whose high antioxidant activity may be explained by the high electron donating effect of the second hydroxyl group.

Despite the lack of the catechol group in their structure, HT sulfate metabolites (**3-4**) were as effective as HT itself. It is interesting to note however that, as shown by our uptake studies (see supporting information), both HT and its sulfate metabolite enter the cells and are partially converted one into the other, thus what we observe is in part the resultant of the activity of both compounds.

At the same time, TYR and its sulfate metabolite **5** display very similar protecting ability against oxidation in our cell model, and TYR possesses only one OH phenolic group whereas TYR sulfate does not have any hydroxyl substituent in its aromatic ring.

In the case of TYR, it has been shown to have scavenging effects on ONOO⁻⁴⁵, O₂⁻⁴⁶ and peroxy radicals⁸.

It is important to remark that, in addition to the antioxidant capacity of natural polyphenols by directly scavenging extracellular/intracellular ROS, several phenolic compounds have been shown to provide a parallel protection by enhancing the level of GSH and the activity of a number of related protective enzymes⁴⁷. Among these enzymes, GPx catalyses the reduction of peroxides and is suggested to act as barrier against hydroperoxide attack⁴⁸. In our experimental conditions, none of the tested compounds altered the basal level of GSH, but TYR was able to significantly increase the activity of GPx after 18 h of incubation. At that time point, in Caco-2 cells exposed to the oxidized cholesterol we detected a significant increase of the GPx activity, indicating a positive response of the cell defence system to face an oxidative insult⁴²⁻⁴³. Thus, pre-treatment with HT and HT sulfates prevented the increase in the activity of GPx induced by oxidative stress, either by direct action on the enzyme activity or via the decreasing of ROS production and consequently in cell damaging peroxide species.

The same result was found in presence of TYR sulfate metabolite. GPx activity in Caco-2 cells was significantly enhanced with respect

to the control in all samples pretreated with TYR, in presence or not of oxidized cholesterol. This outcome is the first evidence of an inductive effect on GPx activity of TYR, which has been shown to preserve GSH level under oxidative stress conditions without altering GPx basal level in Caco-2⁴⁹ and macrophage-like (J774 A1)⁵⁰ cell lines. Our data are in agreement with that previously published with HT in HepG2^{43, 51} and in Caco-2⁴⁴ cells. Similarly, flavanols, such as epicatechin-3-gallate and procyanidin B2, induce a significant increase in several antioxidant-related enzymes activities, accompanied by a decrease in ROS production not affecting GSH content in Caco-2 cells⁵².

When Caco-2 cells were treated with oxidized cholesterol for 24 h, a significant increase of MDA production was observed, together with a decrease in cell viability, indicating a lipo-peroxidative injury to the cell. Pretreatment with all the phenolic compounds and the corresponding sulfate metabolites inhibited MDA formation at all tested concentrations. At 24h of incubation, regardless the phenolic compound initially present in the incubation mixture (parent compound or sulfate metabolite), we found a pool of compounds (see supporting information), mainly sulfate and methyl-sulfate metabolites, in agreement with Rubio et al.¹³, due to the intense metabolising activity of Caco-2 cells, all of which probably contribute to the final effect.

None of the compounds however was equally efficient in preventing cell death. This result suggests that the alteration of cellular redox status did not led only to a direct oxidative injury, but likely modulated signalling pathways involved in cell survival, as demonstrated in differentiated Caco-2 cells, where both unoxidized cholesterol as well as an oxysterol mixture were able to induce apoptosis as a consequence of enhanced ROS production¹⁸.

The actual contribution of phenolic metabolites to the biological activity of the parent compounds is still quite controversial. In fact, Khymenets et al.⁵³ reported the lack of antioxidant activity for HT glucuronate metabolites compared with that of the parent compounds, suggesting that they were not chief contributors to the antioxidant effects provided by olive oil consumption. In contrast, Paiva-Martins et al.¹⁵ recently described the protective effects of HT glucuronides on erythrocyte oxidative-induced hemolysis to a similar extent to that observed for HT itself. These results suggested that HT metabolites may play an important role in the protective activity of olive oil phenolic compounds.

Conclusions

In conclusion, our data provides evidence of the ability of sulfate metabolites of HT and TYR to protect intestinal cells against the pro-oxidant effect of oxidized cholesterol, with an efficiency comparable to that of the parent compounds, strengthening the assumption that the potential beneficial effect of the parent compounds is retained, if not enhanced, although extensive metabolisation occurs, being the resulting metabolites able to exert a biological action themselves. Sulfate metabolites may be present in the intestinal epithelium, as the result of deconjugation/conjugation processes; thus, they may significantly contribute to the protective activity exerted by olive oil simple phenols against the oxidation process triggered by lipid peroxides on the intestinal mucosa.

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Abbreviations

HT	hydroxytyrosol
TYR	tyrosol
5 HT-S	HT sulfate metabolites 3-4
TYR-S	TYR sulfate metabolite 5
MDA	malonyldialdehyde
NR	neutral red
ROS	reactive oxygen species
10 GSH	glutathione
GPx	glutathione peroxidase

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25 [†] Electronic Supplementary Information (ESI) available: [1H-NMR and 13C-NMR spectra of compounds 3-4, 5, 7, 9-10, 11-12 and TYR, HT and sulfate
metabolites stability/uptake in Caco-2 cultures]. See DOI: 10.1039/b000000x/

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Hydroxytyrosol and tyrosol sulfate metabolites counteract the oxidizing action of oxidized cholesterol in Caco-2 cells with an efficiency comparable to that of the parent compounds

