

RSC Advances



This is an *Accepted Manuscript*, which has been through the Royal Society of Chemistry peer review process and has been accepted for publication.

Accepted Manuscripts are published online shortly after acceptance, before technical editing, formatting and proof reading. Using this free service, authors can make their results available to the community, in citable form, before we publish the edited article. This *Accepted Manuscript* will be replaced by the edited, formatted and paginated article as soon as this is available.

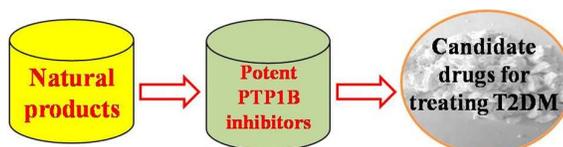
You can find more information about *Accepted Manuscripts* in the [Information for Authors](#).

Please note that technical editing may introduce minor changes to the text and/or graphics, which may alter content. The journal's standard [Terms & Conditions](#) and the [Ethical guidelines](#) still apply. In no event shall the Royal Society of Chemistry be held responsible for any errors or omissions in this *Accepted Manuscript* or any consequences arising from the use of any information it contains.

Natural and Semisynthetic Protein Tyrosine Phosphatase 1B (PTP1B)

Inhibitors as Anti-Diabetic Agents

Li-Jun Wang, Bo Jiang, Ning Wu, Shuai-Yu Wang, and Da-Yong Shi*



Natural products offered more opportunities to develop new drugs and leading compounds as

potent PTP1B inhibitors for treating T2DM.

Cite this: DOI: 10.1039/c0xx00000x

www.rsc.org/npr

REVIEW

Natural and Semisynthetic Protein Tyrosine Phosphatase 1B (PTP1B) Inhibitors as Anti-Diabetic Agents

Li-Jun Wang^a, Bo Jiang^a, NingWu^a, Shuai-Yu Wang^a and Da-Yong Shi^{*a}

Received (in XXX, XXX) XthXXXXXXXXX 20XX, Accepted Xth XXXXXXXXXXXX 20XX

5 DOI: 10.1039/b000000x

Diabetes mellitus, which includes both type 1 and type 2 diabetes mellitus (T2DM), is a major disease that threatens human health worldwide. Protein tyrosine phosphatase 1B (PTP1B) is a promising molecular-level therapeutic target that is effective in the management of T2DM. Natural products with various skeletons and diverse bioactivities offer opportunities for the development of new drugs and lead compounds with potent inhibitory activity against PTP1B *in vitro* and *in vivo*. Recently, a number of potent PTP1B inhibitors have been obtained from natural sources or prepared by synthesis/semi-synthesis, and they exhibit potential for the treatment of T2DM. In this review, we discuss the development of potent natural and semisynthetic PTP1B inhibitors with IC₅₀ values under 10 μM over the past six years (2009-2014), including their structural features, biological features, structure-activity relationships (SARs). We also discuss strategies for identifying potent PTP1B inhibitors from natural products to provide useful information for use by medicinal chemists in developing potent PTP1B inhibitors as T2DM treatments.

1 Introduction

2 Natural PTP1B inhibitors

2.1 Terpenoids

2.2 Flavonoids

30 2.3 Phenolic compounds

2.4 Bromophenols

2.5 Alkaloids

2.6 Steroids

3 Semi-synthetic PTP1B inhibitors

35 3.1 Maslinicacidderivates

3.2 Oleanolicacid derivatives

3.3 Triterpenesaponin derivatives

3.4 Lithocholicacid derivatives

3.5 Bromophenolderivates

40 4 Conclusion and perspective

5 Acknowledgement

6 Abbreviations

7 References

1 INTRODUCTION

45 Diabetes mellitus is a chronic disease that results from insulin deficiency or insulin resistance. Diabetes mellitus is the main threat to human health worldwide.^{1,2} The prevalence of the

disease is constantly increasing, with approximately 347 million individuals suffering from diabetes and more than 3 million deaths per year from consequences of uncontrolled hyperglycaemia, according to World Health Organization (WHO) reports.³ There are two main types of diabetes: type 1 and type 2 diabetes mellitus (T2DM). T2DM is noninsulin-dependent diabetes mellitus and is characterized by defects in signalling by the insulin receptor (IR) protein tyrosine kinase, and accounts for 90 % of cases of diabetes.^{4,5} Women, particularly those with a history of gestational diabetes, usually acquire this disease. Accumulating evidence has shown a strong association between childhood obesity and the onset of diabetes.⁶ Moreover, obesity is a major risk factor, and approximately 75 % of obese individuals develop T2DM.⁷ In addition, this disease causes many severe secondary complications, including atherosclerosis, microangiopathy, renal dysfunction and failure, cardiac abnormalities, diabetic retinopathy, and ocular disorders.^{8,9} Thus, developing novel therapies to treat these diseases is essential.

Currently, eight oral hypoglycaemic drug classes are available to treat patients suffering from T2DM. Classified according to their mechanisms of action, they are (i) α-glucosidase inhibitors, (ii) insulin secretagogues, (iii) biguanides, (iv) thiazolidinediones, (v) glucagon-like peptide-1 (GLP-1) receptor agonists, (vi) dipeptidyl peptidase-4 (DPP-4) inhibitors, (vii) sodium-glucose cotransporter-2 (SGLT2) inhibitors and (viii) amylin analogs.¹⁰ Although hypoglycaemic agents are widely available, these agents show many limitations, including adverse effects (e.g., gastrointestinal complaints, weight gain, peripheral oedema, headache, hypotension) and high rates of secondary failure, because they are not mimetic of insulin signaling.¹¹ Thus, novel drugs that mimic only the desired properties of insulin signalling are urgently needed.¹² Protein tyrosine phosphatases (PTPs) are enzymes that catalyze protein tyrosine dephosphorylation, which plays a key role in the regulation of insulin action by dephosphorylation of the activated autophosphorylated IR and downstream substrate proteins.¹³ Animal models, clinical studies and cell line studies have shown that a cytosolic non-receptor PTP, protein tyrosine phosphatase 1B (PTP1B), is a negative regulator of insulin signal transduction, and it has proven to be an important molecular target for potential treatment of T2DM.¹⁴⁻¹⁸

A number of PTP1B inhibitors have been synthesized as promising candidates for the treatment of T2DM since the late 1990s, including phosphonic acid, phosphonodifluoromethyl phenylalanine derivatives, carboxylic acids, imides, sulphonic

acids and vanadium compounds. These efforts focused on designing non-hydrolyzable p-Tyr surrogates that target the catalytic site, the second allosteric pocket or both catalytic and allosteric sites.^{19, 20} However, these molecules still lack efficacy *in vivo* because they have weak oral bioavailability, poor membrane permeability and weak selectivity over other PTPs.

It is well known that natural products with various skeletons and diverse bioactivities offer more opportunities for the development of new drugs and lead compounds.²¹ It has been proven that natural products are the most important sources for potential and novel PTP1B agents based on different *in vitro* and *in vivo* approaches. Many potent and specific natural PTP1B inhibitors with the potential to improve insulin resistance and normalize plasma glucose and insulin without inducing hypoglycaemia have been reported.²²⁻²⁴ Trodusquemine (MSI-1436, **1**, Fig. 1), which has an IC_{50} value of 1.3 μ M against PTP1B, is a natural novel aminosterol originally isolated from the liver of the dog fish shark (*Squalusacanthias*), and it showed excellent efficacy in preclinical studies and a good pharmacokinetic profile in a phase I clinical trial for the treatment of T2DM and obesity.²⁵⁻²⁸ These results indicate that potent natural inhibitors may provide a major advance in the treatment of T2DM in the future.

There are several reviews summarizing the development of PTP1B inhibitors focused on synthetic PTP1B inhibitors, and the literature before 2009 has been well summarized.¹⁹⁻²⁴ Since then, many potent and specific PTP1B inhibitors from natural products have been reported. Based on the above, we discuss the development of a diversity of natural PTP1B inhibitors reported during the past six years (2009-2014) with IC_{50} values under 10 μ M. Their structural features, biological features and structure-activity relationships (SARs) are reviewed, as are the strategies used to identify potent PTP1B inhibitors from natural products, which should provide useful information for medicinal chemists seeking to develop potent PTP1B inhibitors to treat T2DM.

35 2 NATURAL PTP1B INHIBITORS

More and more natural metabolites with anti-PTP1B activity are being reported with the progress in natural product chemistry. In this section, natural PTP1B inhibitors are categorized according to their chemical structures, including terpenes, flavonoids, phenolic compounds, bromophenols, and alkaloids.

2.1 Terpenoids

Terpenoids, as a large and varied class of hydrocarbons, have received considerable attention for their wide range of pharmacological properties, including anticancer, antimicrobial, antifungal, antiviral, anti-hyperglycaemic, anti-inflammatory and antiparasitic activities.²⁹ Recently, many terpenoids with effective anti-PTP1B activity have been reported.

Oleanolic acid (OA, **2**, Fig. 1), apentacyclic triterpenoid, has been in active clinical use as an antihepatitis drug in China, and it displays hypoglycaemic, anti-inflammatory, and antitumorigenic effects, protecting the liver against toxic injury. OA shows high inhibitory activity against PTP1B ($IC_{50} = 9.5 \pm 0.5 \mu$ M) that can be described by a linear mixed-type inhibition model based on the kinetic parameters (K_m and V_{max}), and it is usually used as a positive control drug in anti-PTP1B assays.^{30, 31}

Hung *et al.* investigated the PTP1B inhibitory constituents of *Gynostemma pentaphyllum* (Thunb.), which is used as an anti-diabetic agent in Vietnamese folk medicine. A $CHCl_3$ -soluble

fraction showed dose-dependent inhibitory activity against PTP1B enzyme with an IC_{50} value of 30.5 μ g/mL, and seven PTP1B inhibitors were found. Among these inhibitory compounds, (2*S*)-3 β , 20, 23 ξ -trihydroxydammarane-24-en-21-oic acid-21, 23 lactone (**3**, Fig. 1) showed the most potent PTP1B inhibitory activity with an IC_{50} value of $5.3 \pm 0.4 \mu$ M, and the inhibition mode was competitive toward p-NPP, with a K_i value of 2.8 M. It was found that the stereochemistry at C-20 could affect the inhibitory activity, and the presence of hydroxyl groups could decrease the affinity for a hydrophobic site of the enzyme.³² Seven PTP1B inhibitory triterpenoids were yielded by bioassay-guided fractionation of the leaves of *Rhododendron brachycarpum* G. Don (Ericaceae). Among them, ursolic acid (**4**, $IC_{50} = 3.1 \pm 0.3 \mu$ M, Fig. 1), corosolic acid (**5**, $IC_{50} = 7.0 \pm 0.6 \mu$ M, Fig. 1), 23-hydroxyursolic acid (**6**, $IC_{50} = 7.4 \pm 0.6 \mu$ M, Fig. 1), and rhododendric acid A (**7**, $IC_{50} = 6.3 \pm 0.6 \mu$ M, log P = 5.19, Fig. 1) exhibited strong inhibitory activity against PTP1B. This study also indicated that the hydroxyl group at C-3 in ursane-type triterpenoids is necessary for the inhibitory activity.³³ Xue *et al.* reported that an isomalabaricane triterpene isolated from the Hainan sponge *Stelletta* sp., stelletin G (**8**, Fig. 1) showed strong PTP1B inhibitory activity, with an IC_{50} value of $4.1 \pm 0.9 \mu$ M [34]. A 24-norursane triterpene, ilekudinol B (**9**, Fig. 1), was isolated as an active metabolite with IC_{50} value of $5.3 \pm 0.5 \mu$ M by means of *in vitro* bioassay-guided fractionation of MeOH extract of the leaves and stems of *Weigela subsessilis* (Caprifoliaceae).³⁵ These results suggest that the lactonization of a carboxyl group at C-28 in this type of triterpene may result in a loss of activity and that a free carboxyl group at C-28 could enhance the inhibition of PTP1B.

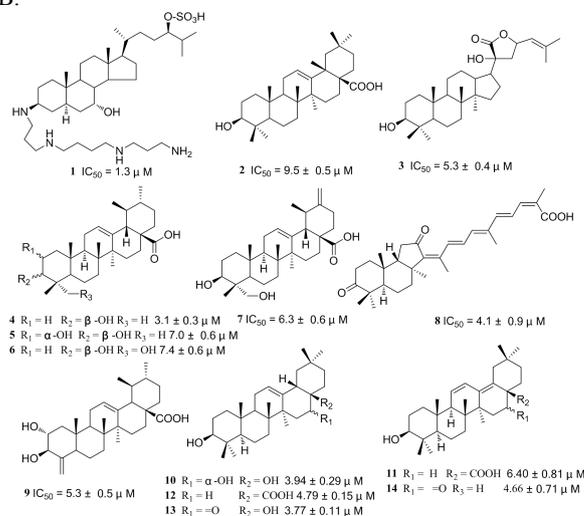


Fig. 1 Structures of compounds 1-14

Six oleanane-type triterpenes (**2**, **10-14**, Fig. 1) were isolated from an EtOAc-soluble extract of fruit peels of *Camellia japonica* (Theaceae), and they showed strong PTP1B inhibitory activity (IC_{50} values ranging from 3.77 ± 0.11 to $6.40 \pm 0.81 \mu$ M) as well as significant cytotoxicity (IC_{50} values ranging from 0.51 ± 0.05 to $13.55 \pm 1.44 \mu$ M).³⁶ From the above results, the SARs of these triterpene compounds suggest that the hydroxyl group at C-3 and carbonyl group at C-28 or C-27 of the oleanane-type triterpenoids are essential structural features for the PTP1B inhibitory activity,

and the number of hydroxyl groups is inversely proportional to the PTP1B inhibitory potency.³²⁻³⁷

Three potently active constituents of terpenoids, betulinic acid (**15**, Fig. 2), betulinic acid methyl ester (**16**, Fig. 2) and mokko lactone (**17**, Fig. 2), along with nine inactive compounds were identified through activity-guided fractionation of the MeOH extract of roots of *Saussurea lappa* C.B. Clarke (Compositae), which resulted in potent inhibition of PTP1B with IC₅₀ values of 1.54 ± 0.07 μM, 1.91 ± 0.15 μM and 6.03 ± 0.08 μM, respectively.³⁸

Cedrodorol B (**18**, Fig. 2), a new apotirucallane-type triterpenoid isolated from the twigs and leaves of *C. odorata*, showed significant inhibitory activity against PTP1B, with an IC₅₀ value of 8.05 μM.³⁹ Fang et al. investigated inhibitory activities against PTP1B from *Schisandra chinensis* (Turcz.) Baill. (SCTB). Compounds **19** and **20** (Fig. 2) were isolated from petroleum ether (PE) extract of this functional and medicinal herb, and they showed potent inhibitory activity, with IC₅₀ values of 2.36 ± 0.20 μM and 9.78 ± 0.10 μM that were comparable with the positive controls NaVO₄ (IC₅₀ = 22 ± 0.20 μg/mL) and acarbose (2.10 ± 0.10 μg/mL).⁴⁰

Four novel sesquiterpene quinones with an unprecedented "dysidavarane" carbon skeleton were isolated from the South China Sea sponge *Dysideaavara* by the Lin group.⁴¹ In these compounds, dysidavarone A (**21**, Fig. 2) showed significant inhibitory activity against PTP1B (IC₅₀ = 9.98 μM).

A sesquiterpene quinone, dysidine (**22**, Fig. 2), which was isolated from the Hainan sponge *Dysidea villosa*, shows the strongest PTP1B inhibitory activity with an IC₅₀ value of 6.70 μM.⁴² Further studies revealed that compound (**22**) is a novel slow-binding PTP1B inhibitor with moderate inhibition selectivity over other PTPs. It strongly activates the insulin signalling pathway, promotes membrane translocation of glucose transporter 4 (GLUT4) in CHO-K1 and 3T3-L1 cells and greatly promotes glucose uptake in 3T3-L1 cells.⁴³ Yamazaki *et al.* reported that a sesquiterpene, dehydroeuryspongins A (**23**, Fig. 2), from a dehydro product was formed from euryspongins A (a unique sesquiterpene isolated from the marine sponge *Euryspongia* sp. collected at Iriomote Island, Okinawa, Japan) with CDCl₃ in an NMR tube. It is very interesting that compound **23** has potent inhibitory activity against PTP1B, with an IC₅₀ value of 3.6 μM, whereas euryspongins A shows no inhibitory effect against PTP1B.⁴⁴ One new dinorremophilane derivative and four new eremophilanolactones were isolated from 95% EtOH extract of *Ligularia fischeri*, which is used as a traditional Chinese medicine (TCM) for treating cough, inflammation, jaundice, scarlet fever, rheumatoid arthritis, and hepatic diseases. Among these derivatives, (3β, 6β, 8α, 10β)-3-acetyl-6, 8, 10-trihydroxyeremophil-7 (11)-eno-12, 8-lactone (**24**, Fig. 2) exhibited inhibitory activity against PTP1B with an IC₅₀ value of 1.34 μM.⁴⁵

Guo's group systematically investigated PTP1B inhibitors from marine biology resources. Sarsolilide A (**25**, Fig. 2), a capnosane diterpene isolated from the Hainan soft coral *Sarcophyton trocheliophorum* Marenzeller, exhibited inhibitory activity with an IC₅₀ value of 6.8 ± 0.9 μM.⁴⁶ Two unprecedented diterpenoids, methyl sarcotroates A and B (**26**, Fig. 2), possessing a tetradecahydrocyclopenta [3', 4'] cyclobuta [1', 2':4, 5]-cyclonona

[1, 2-b] oxirene ring system, were isolated from the Hainan soft coral *Sarcophyton trocheliophorum*. Of these, in the first report of a natural PTP1B inhibitor containing a hydro-peroxide group, compound **26** exhibited potent inhibitory activity, with an IC₅₀ value of 6.97 μM. In an ongoing investigation of *S. trocheliophorum*, eleven cembranoids were isolated and evaluated for their inhibitory activity against human PTP1B.⁴⁷ Most of these terpenoids show potent activity, particularly compounds **27** and **28** (Fig. 2), which exhibited significant activity and had IC₅₀ values of 5.95 and 6.33 μM, respectively. The preliminary SARs of these compounds were summarized as follows: (I) the structure of the α, β-unsaturated ε-lactone should not be essential for the activity; (II) the methyl ester group at C-18 could increase the inhibitory activity; (III) the conjugated diene/ester moiety should not be crucial for the activity; and (IV) the presence of an epoxide or diol group on the macrocyclic ring is responsible for a dramatic decrease in activity.

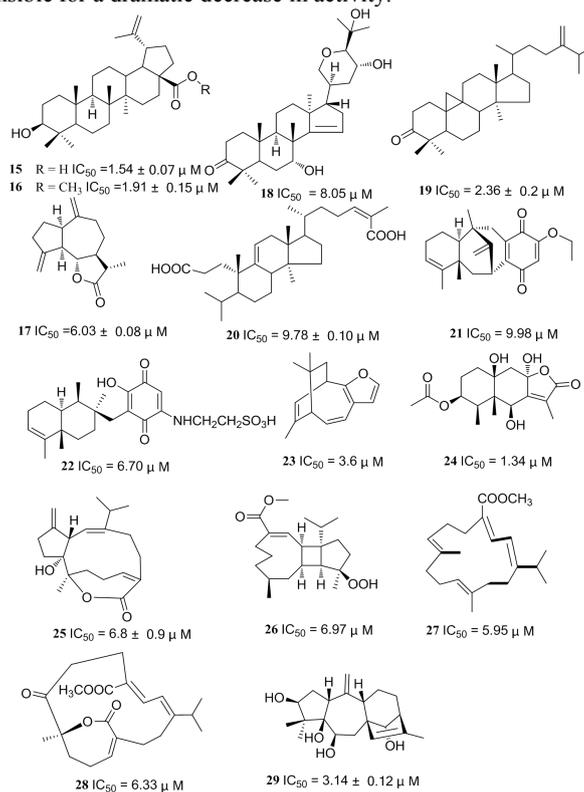


Fig. 2 Structures of compounds **15-29**

Liu *et al.* investigated the chemical constituents of *R. principis*, which afforded six new grayanane diterpenoids. Among them, principinol E (**29**, Fig. 2) exhibited significant inhibitory activity against PTP1B with an IC₅₀ of 3.14 ± 0.12 μM. A preliminary summary of the SARs was concluded based on their results as follows: (I) the presence of OH-5 and OH-6 is essential for the activity; (II) when OH-6 is epoxidized or acetylated, activity will decrease; (III) the Δ¹⁵ double bond could increase the activity.⁴⁸ *Aralia continentalis* Kitag. (syn. = *A. cordata* Thunb., Araliaceae) is a perennial herb whose sprouts are consumed in salads in Korea and whose roots are used as a TCM. Jung *et al.* investigated the most active fractions in detail to isolate active compounds, and they summarized their SARs in PTP1B inhibition.⁴⁹ Nine diterpenoids possessing excellent anti-PTP1B

activity with IC_{50} values under 10 μ M, including continental acid ($IC_{50} = 0.66 \pm 0.18 \mu$ M, **30**, Fig. 3); ent-pimara-8 (14), 15-diene-19-ol ($IC_{50} = 9.85 \pm 0.20 \mu$ M, **31**, Fig. 3); 7-oxo-ent-pimara-8 (14), 15-diene-19-oic acid ($IC_{50} = 0.09 \pm 0.06 \mu$ M, **32**, Fig. 3); kaurenoic acid ($IC_{50} = 4.64 \pm 0.82 \mu$ M, **33**, Fig. 3); 16 α -hydroxy-17-isovaleryloxy-ent-kauran-19-oic acid ($IC_{50} = 1.51 \pm 0.07 \mu$ M, **34**, Fig. 3); 16 α ,17-dihydroxy-ent-kauran-19-oic acid ($IC_{50} = 0.56 \pm 0.10 \mu$ M, **35**, Fig. 3); 8 α -hydroxy-ent-pimara-15-en-19-ol ($IC_{50} = 1.34 \pm 0.56 \mu$ M, **36**, Fig. 3); 17-hydroxy-ent-kaur-15-en-19-oic acid ($IC_{50} = 9.12 \pm 0.92 \mu$ M, **37**, Fig. 3); and 15 α ,16 α -epoxy-17-hydroxy-ent-kauran-19-oic acid ($IC_{50} = 1.96 \pm 0.06 \mu$ M, **38**, Fig. 3), were found in the active n-hexane and EtOAc soluble fractions. The results indicated that the SARs were as follows: (I) the molecular type of the diterpenoid and the types of substituent present in the molecule are important for strong interactions with the enzyme molecule and for the consequent inhibition of the enzyme; (II) an isovaleryloxy moiety at C-17, introduction of a hydroxyl group or reduction of a carboxyl group at C-19 would abolish the inhibitory effects towards PTP1B; (III) a C-7 ketone or C-19 carboxylic acid group could influence the inhibitory activity, and an oxo group at the C-7 position is important for the inhibition of PTP1B activity. Continuing research into the kinetics and molecular docking of these substances has indicated that compounds (**30**, **32**) and 7 β -hydroxy-ent-pimara-8(14), 15-diene-19-oic acid (**39**, Fig. 3) show negative binding energies of -5.3 to -6.1 kcal/mol and a high affinity for PTP1B residues (Phe182 and Asp181 in the WPD loop; Cys215 in the active sites; Tyr46, Arg47, Asp48, Val49, Ser216, Ala217, Gly218, Ile219, Gly220, Arg221, Gln262, and Gln266 in the pocket site), which indicates that they may stabilize the open form and generate tighter binding to the catalytic site of PTP1B.⁵⁰ Therefore, these diterpene PTP1B inhibitors show promise as therapeutic agents for the treatment of diabetes and related disorders.

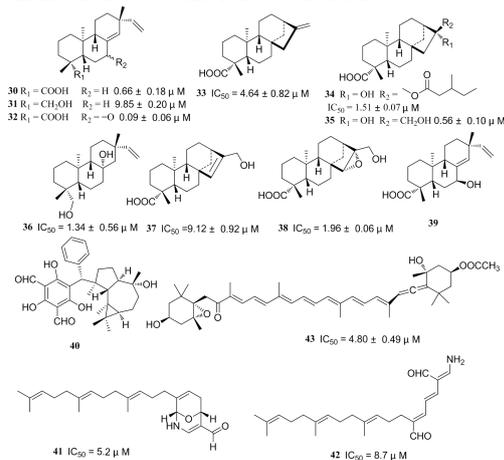


Fig. 3 Structures of compounds 30-43

Zhang's group reported that a novel sesquiterpenoid, psidial B (**40**, Fig. 3), was isolated from the leaves of *Psidium guajava* L. with an inhibition rate for PTP1B enzyme of 61.7% at 10 μ M.⁵¹ Piao *et al.* reported that two active compounds (**41**, **42**, Fig. 3) isolated from the sponge *Hippospongia lachne* off Yongxing Island in the South China Sea show potent PTP1B inhibitory activity with IC_{50} values of 5.2 μ M and 8.7 μ M.⁵²

Fucoxanthin (**43**, Fig. 3), a marine carotenoid, is characteristically present in edible brown sea weeds such as *Eisenia bicyclis* (Arame), *Undaria pinnatifida* (Wakame), and *Hijikia fusiformis* (Hijiki), and it can enhance insulin resistance and decrease blood glucose levels. Jung *et al.* showed that fucoxanthin showed potent inhibitory activity against PTP1B, with an IC_{50} value of 4.80 \pm 0.49 μ M. Kinetic study revealed that fucoxanthin is a mixed-type inhibitor, indicating that fucoxanthin can bind to the allosteric site of the free enzyme or to the enzyme-substrate complex. A docking study indicated that fucoxanthin was stably positioned in the pocket of PTP1B through three hydrogen-bond interactions with the Phe30, Phe52, and Gly183 residues of the enzyme and the two hydroxyl groups of fucoxanthin as well as hydrophobic interactions between the long hydrocarbons of fucoxanthin harbouring conjugated double bonds and the Ile219, Tyr46, Val49, and Ala217 residues of PTP1B.⁵³

2.2 Flavonoids

Flavonoids are polyphenolic compounds that have two aromatic rings connected by a three-carbon bridge. They are ubiquitous in plants, and they include chalcone, dibenzylmethane, flavanone, dihydroflavonol, isoflavan, isoflavone, flavone, pterocarpan, coumestran, aurone, dihydrochalcone, biflavonoid, and flavonoid glycoside. They have a broad range of biological properties, including antioxidative, anti-allergic, anti-inflammatory, antidiabetic, antiproliferative, hepato- and gastro-protective, antiviral, and antineoplastic activities.⁵⁴ Flavonoids have also been found to display anti-PTP1B activity.

The MeOH extract of the stem bark of *E. lysistemon* Hutch. was found to inhibit PTP1B activity by more than 80 % at 30 μ g/mL.⁵⁵ By using bioassay-guided fractionation of the active extract, twelve pterocarpanes were isolated, including three new pterocarpanes along with nine known pterocarpanes. Most of them showed strong inhibition of PTP1B *in vitro*, especially erybraedin A (**44**, Fig. 4) with an IC_{50} value of 2.4 \pm 0.7 μ M. It was found that (I) prenylating the pterocarpanes could enhance their inhibitory activity; (II) the presence of a C-8 aldehyde and C-6 α hydroxyl group might be responsible for a loss in activity; (III) prenylation of the A ring and/or D ring could increase the PTP1B inhibitory activity. The same research group investigated the EtOAc extract of the stem bark of *Erythrina abyssinica* (Leguminosae) and isolated three new and twelve known pterocarpan derivatives, which were evaluated for their inhibitory effects on PTP1B. Of these, six compounds including neorautenol (**45**, Fig. 4), erybroadin D (**46**, Fig. 4), erybroadin B (**47**, Fig. 4), folitenol (**48**, Fig. 4), erysubin E (**49**, Fig. 4) and erybroadin C (**50**, Fig. 4) showed excellent anti-PTP1B activity with IC_{50} values of 7.6 \pm 0.9 μ M, 4.2 \pm 0.2 μ M, 7.8 \pm 0.5 μ M, 6.4 \pm 0.6 μ M, 8.8 \pm 0.5 μ M and 7.3 \pm 0.1 μ M, respectively.

Li *et al.* screened a compound library of 42 licorice flavonoids by assessing their PTP1B inhibitory activity and observed that licoagrone (**51**, Fig. 4), licoagrodin, licoagroaurone, and *isobavachalcone* showed potent PTP1B inhibitory activity. Among them, compound **51** ($IC_{50} = 6.0 \mu$ M) showed the most potent PTP1B inhibitory activity, while it showed very weak inhibition against T-cell protein tyrosine phosphatase (TCPTP, a protein tyrosine phosphatase homologous to PTP1B), indicating that compound **51** is a selective PTP1B inhibitor. Studies of the inhibition mode and cellular activities of this compound in the

insulin-signalling pathway revealed that it increased the pAkt levels, though not in a concentration-dependent manner.⁵⁶

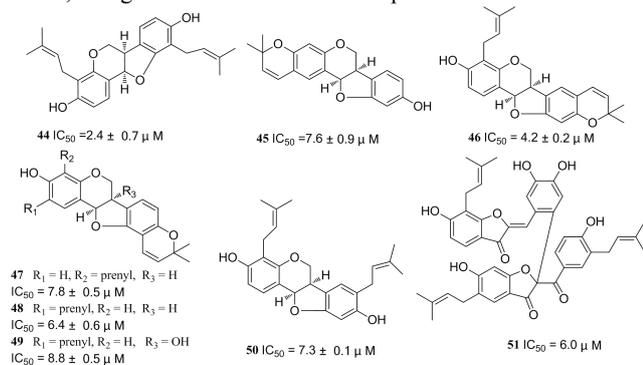


Fig. 4 Structures of compounds 44–51

5 Five isoflavonoids isolated from the EtOAc extract of roots of *Erythrina addisoniae* (Leguminosae) were found to be significant PTP1B inhibitors.⁵⁷ Among them, erythraddison III (52, Fig. 5) with a conjugated hydroxy group at C-5, a prenyl group at C-3' and a substituted 4'-methyl ether, showed potent activity against PTP1B ($IC_{50} = 4.6 \pm 0.1 \mu M$). The IC_{50} value of erysubin F (53, Fig. 5) for inhibition of PTP1B was $7.8 \pm 0.5 \mu M$.

2'-Methoxykurarinone (54, Fig. 5), which was isolated from the roots of *Sophora flavescens*, exhibited potent activity ($IC_{50} = 5.26 \pm 0.24 \mu M$) as a novel noncompetitive PTP1B inhibitor. Moreover, compound 54 exhibited cellular activity in the insulin signalling pathway by increasing the insulin-stimulated Akt phosphorylation level in human hepatocellular liver carcinoma HepG2 cells.⁵⁸

Luteolin (55, Fig. 5), an important bioflavonoid, is abundantly present in various fruits and vegetables and exhibits potent inhibitory activity against PTP1B, with an IC_{50} value of $6.70 \pm 0.03 \mu M$ (the positive control ursolic acid has an IC_{50} value of $8.20 \pm 0.55 \mu M$). However, its two C-glycosylated derivatives, orientin and isoorientin, were inactive at the same concentration, which indicates that C-glycosylation at different positions on luteolin may strongly affect the PTP1B inhibitory activity of luteolin and its C-glycosylated derivatives.⁵⁹

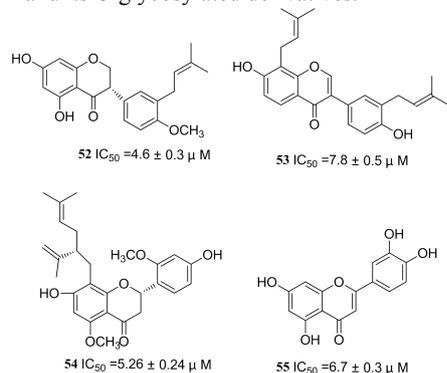


Fig. 5 Structures of compounds 52–55

2.3 Phenolic Compounds

Hoang *et al.* reported that the compounds albafuluran A (56, Fig. 6), mulberrofuluran W (57, Fig. 8), mulberrofuluran D (58, Fig. 6), kuwanon J (59, Fig. 6) and kuwanon R (60, Fig. 6) isolated from the chloroform-soluble fraction of *Morus bombycis* showed 35 remarkable inhibitory activity against PTP1B with IC_{50} values of

$9.2 \pm 0.7 \mu M$, $2.7 \pm 0.3 \mu M$, $4.3 \pm 0.5 \mu M$, $2.7 \pm 0.6 \mu M$ and $8.2 \pm 0.9 \mu M$, respectively.⁶⁰ Albafuluran B (61, Fig. 6), isolated from the root bark of *Morus alba* var. *tatarica*, exhibited strong inhibitory effects against PTP1B with an IC_{50} value of $8.9 \pm 1.1 \mu M$.⁶¹ From these results, it can be concluded that (I) increased lipophilicity and decreased charge of aliphatic side chains leads to stronger activity and (II) increased numbers of OH groups in chalcone-derived Diels–Alder-type compounds could improve potential inhibitory effects against PTP1B.

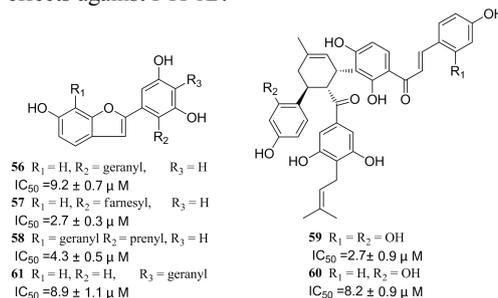


Fig. 6 Structures of compounds 56–61

Cinnamic acid analogues (esters, amides and glycosides) show various activities, including antiviral, anti-atherogenic, antitumor, antituberculosis, antioxidant, and antibacterial properties, and have attracted much attention in biology and medicine.⁶² Caffeic acid (62, Fig. 7) was proven to have a potent inhibitory effect on the activity of PTP1B, with an IC_{50} value of $3.06 \mu M$. Islam *et al.* reported that quinic acid derivatives, including 3, 4-dicaffeoylquinic acid (63, Fig. 7), 3, 5-dicaffeoylquinic acid (64, Fig. 7), 3, 5-dicaffeoylquinic acid methyl ester (65, Fig. 7), and 4, 5-dicaffeoylquinic acid (66, Fig. 7) show potent inhibitory activity against PTP1B, with IC_{50} values of $2.60 \pm 0.24 \mu M$, $2.02 \pm 0.46 \mu M$, $2.99 \pm 0.42 \mu M$ and $3.21 \pm 0.23 \mu M$, respectively.⁶³

The roots of *Paeonia lactiflora* Pall. or *Paeonia suffruticosa* Andrews (Ranunculaceae) are used in TCM as “*Paeoniae radix rubra*” (as deduced from the reddish colour of the dried root bark) or “*Cortex Moutan*”, which can be used to treat symptoms related to metabolic syndrome and T2DM. Baumgartner *et al.* found that a crude methanol extract of *P. lactiflora* roots could reduce the residual activity of human recombinant PTP1B. Subsequently 1, 2, 3, 4, 6-penta-*O*-galloyl-*D*-glucopyranose (67, Fig. 7) was isolated from the roots of *P. lactiflora*, and it showed potent activity against PTP1B with an IC_{50} value of $4.8 \mu M$; moreover, it could act as an insulin sensitizer at a concentration of $10 \mu M$ in 70 human hepatoma cells (HCC-1.2).⁶⁴

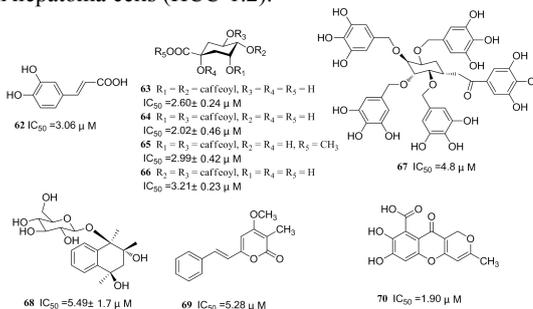


Fig. 7 Structures of compounds 62–70

Cyclonoside A (68, Fig. 7), a naphthoquinone derivative with an IC_{50} value of $5.49 \pm 1.7 \mu M$, was found in the ethanol extract of

leaves of *Cyclocaryapaliurus* (Batal.) Ijinskajavia activity-guided bioassay.⁶⁵ Lee *et al.* investigated the chemical constituents of crude extracts obtained from cultures of the marine-derived fungus *Penicillium* sp. JF-55 cultures afforded two PTP1B inhibitors named penstyrylpyrone (**69**, Fig. 7) and anhydrofulvic acid (**70**, Fig. 7), which had IC₅₀ values of 5.28 μM and 1.90 μM. These compounds inhibited PTP1B activity in a competitive manner; their linear tricyclic system and the positions of the carbonyl groups were the important structural features for their binding to the active site of PTP1B.⁶⁶

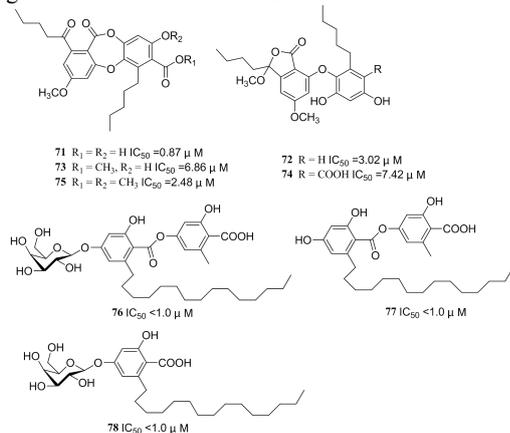
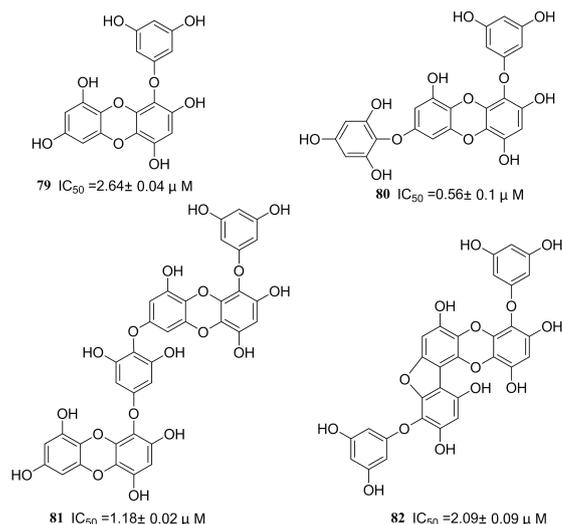


Fig. 8 Structures of compounds **71-78**

Seo *et al.* studied compounds that are metabolites belonging to the depsidone class and that behave as PTP1B inhibitors. Among them, lobaric acid (**71**, Fig. 8) and two pseudodepsidone-type metabolites (**72** and **73**, Fig. 8) exhibited potent PTP1B inhibitory activity in a dose-dependent manner, with IC₅₀ values of 0.87 μM, 6.86 μM, and 2.48 μM, respectively. Derivatives **74** and **75** (Fig. 8) derived from compound **71** by chemical modification also showed potent inhibitory activity with IC₅₀ values of 3.02 and 7.42 μM. These results proved that the carboxylic acid and hydroxyl group play roles in the inhibition mechanism because they provide hydrogen-bonding sites that are relevant to the interaction with PTP1B.⁶⁷ Four active compounds were isolated from the EtOAc extract of culture broth of the marine-derived fungus *Cosmospora* sp. SF-5060. Of these, three compounds (**76-78**, Fig. 8) had IC₅₀ values under 1.00 μM. Aquastatin A (**76**, Fig. 8) was identified as a competitive and selective inhibitor of PTP1B over other protein tyrosine phosphatases such as TCPTP, SHP-2, LAR, and CD45, and the dihydroxy pentadecyl benzoic acid was a key pharmacophore.⁶⁸

Moon *et al.* investigated the MeOH extract and solvent-soluble fractions of two brown algae, *E. stolonifera* and *E. bicyclis*. 4-Phlorotannin [eckol (**79**, Fig. 9), 7-phloroeckol (**80**, Fig. 9), dieckol (**81**, Fig. 9), and phlorofurofucoeckol-A (**82**, Fig. 9)] showed potent and noncompetitive inhibitory activity against PTP1B with IC₅₀ values of 2.64 ± 0.04 μM, 0.56 ± 0.10 μM, 1.18 ± 0.02 μM, and 2.09 ± 0.09 μM, respectively.⁶⁹



10

Fig. 9 Structures of compounds **79-82**

2.4 Bromophenols

Bromophenols with one or several benzene rings substituted with varying degree of bromine and hydroxyl groups exist in marine algae, and they exhibit a wide spectrum of beneficial biological activities (including antioxidant, antibacterial, anticancer, and anti-diabetic activity) based on recent studies of marine bromophenols.⁷⁰

In searching for PTP1B inhibitors from marine algae, the ethanol-soluble extract of *Rhodomela confervoides* exhibited significant inhibitory activity against PTP1B *in vitro*. Using a variety of chromatographic techniques in bioassay-guided separation of ethanol extract resulted in a series of bromophenol derivatives, including 3, 4-dibromo - 5 - (2 - bromo - 3, 4 - dihydroxy - 6 - (isobutoxymethyl) benzyl) benzene - 1, 2 - diol (**83**, Fig. 10), 5, 5' - ((3 - bromo - 4, 5 - dihydroxy - 1, 2 - phenylene) bis (methylene)) bis (3, 4 - dibromobenzene - 1, 2 - diol) (**84**, Fig. 10), 5, 5' - (oxybis (methylene)) bis (3, 4 - dibromobenzene - 1, 2 - diol) (**85**, Fig. 10), 3, 4 - dibromo - 5 - (2 - bromo - 6 - (ethoxymethyl) - 3, 4 - dihydroxybenzyl) benzene - 1, 2 - diol (**86**, Fig. 10), 3, 4 - dibromo - 5 - (methoxymethyl) benzene - 1, 2 - diol (**87**, Fig. 10), 3 - (2, 3 - dibromo - 4, 5 - dihydroxyphenyl) - 2 - methylpropanal (**88**, Fig. 10) and 7 - bromo - 1 - (2, 3 - dibromo - 4, 5 - dihydroxyphenyl) - 2, 3 - dihydro - 1H - indene - 5, 6 - diol (**89**, Fig. 10) with potent inhibitory activity with IC₅₀ values of 2.4 μM, 1.7 μM, 1.5 μM, 0.84 μM, 3.4 μM, 4.5 μM, and 2.8 μM, respectively.^{71, 72}

Two brominated metabolites, 3', 5', 6', 6-tetrabromo-2, 4-dimethyldiphenyl ether (**90**, Fig. 10) and 2', 5', 6', 5, 6-pentabromo-3', 4', 3, 4-tetramethoxybenzo-phenone (**91**, Fig. 10), were isolated from the red alga *Laurencia similis*, and they showed potent inhibitory activity against PTP1B protein, with IC₅₀ values of 2.97 and 2.66 μM, respectively.⁷³ Liu *et al.* investigated chemical constituents of the marine red alga *Symphycloa dialatiuscula* collected from the Weihai coastline of Shandong Province of China. Three bromophenols (**92**, **93**, Fig. 10, and **130**) were found that showed strong activity against PTP1B, with IC₅₀ values of 3.9, 4.3, and 3.5 μM, respectively.⁷⁴

Compounds **94** and **95** (Fig. 10) markedly inhibit PTP1B activity (IC₅₀ = 0.85 and 1.7 μM) and were isolated from the ethanol extract of the Indonesian marine sponge *Lamellodysidea*

herbacea. Four ester derivatives [acetyl (**96**), butyryl (**97**), hexanoyl (**98**), and benzoyl (**99**), Fig. 10] were prepared from compound **94**, which revealed comparable to stronger inhibitory activity against PTP1B than that of compound **94**, with IC_{50} values of 0.62, 0.68, 0.69 and 0.97 μ M, respectively.⁷⁵

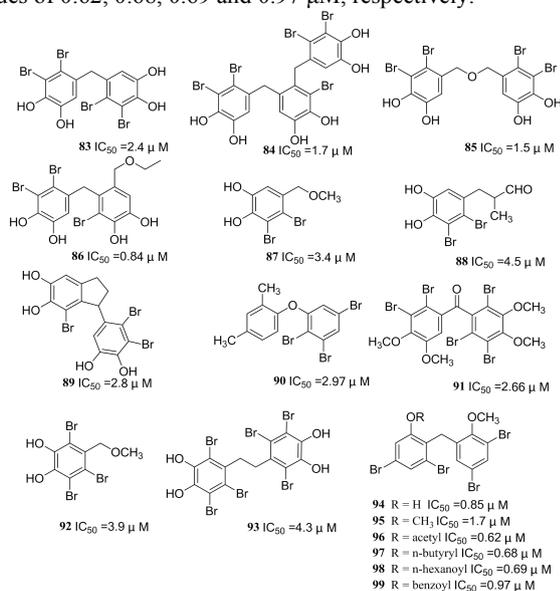


Fig. 10 Structures of compounds **83-99**

2.5 Alkaloids

Berberine (**100**, Fig. 11) is an interesting natural isoquinoline alkaloid that is widely used in traditional medicine for its potent activities. Berberine was found to potently competitively inhibit recombinant h-PTP1B *in vitro* (IC_{50} = 156.9 nM, K_i value = 91.3 nM).⁷⁶ Chen *et al.* demonstrated that berberine has insulin-mimicking effects on both adipocytes and myocytes, which may occur through the inhibition of PTP1B activity and significantly lower the blood glucose of diet-induced-obese (DIO) mice and db/db mice, but they do not increase insulin release and synthesis *in vivo*. These results indicate that berberine is an insulin signalling activator and could be used to treat T2DM.⁷⁷

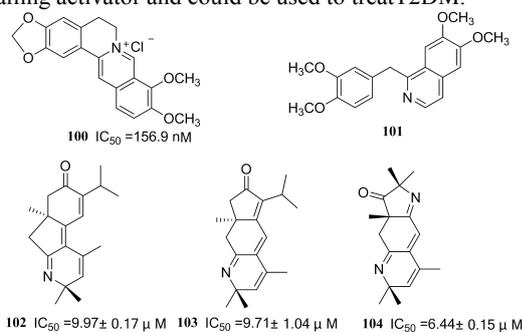


Fig. 11 Structures of compounds **100-104**

Papaverine (**101**, Fig. 11), a prominent member of the isoquinoline alkaloids, was isolated from opium poppy (*Papaver somniferum* L.), and it showed potent inhibitory effects against recombinant h-PTP1B *in vitro* and significantly decreased the fasting blood glucose level of Balb/c mice after a single intraperitoneal injection. A docking investigation indicated that both ionized and non-ionized forms of papaverine fit in the binding pocket of h-PTP1B in a relatively similar manner and interact with nearly the same set of amino acids that extend from

ARG47 to GLY220.⁷⁸

Three norditerpenoid alkaloids, nigelladines A–C, and one pyrroloquinoline alkaloid, nigellaquinomine, possessing new skeletons were isolated from the seeds of *Nigella glandulifera* Freyn. Of them, nigelladines A, B and nigellaquinomine (**102-104**, Fig. 11) exhibit potent PTP1B inhibitory activity (IC_{50} = 9.97 ± 0.17, 9.71 ± 1.04, and 6.44 ± 0.15 μ M) but are devoid of cytotoxicity against the A431 cell line at 100 μ M.⁷⁹

2.6 Steroids

A new pregnane steroid, (Z)-aglawone [**105**, Fig. 12], was isolated from the stem bark of *Toona ciliata* var. *pubescens* and exhibits potent inhibition of PTP1B with an IC_{50} value of 3.54 μ M; its inhibition mode is competitive toward p-nitrophenyl phosphate (pNPP) based on kinetic analysis results.⁸⁰ *Antrodiella albocinnamomea* is a basidiomycetous fungus that is widely distributed in temperate to subtropical areas of China. Two novel degraded steroids (**106**, **107**, Fig. 12) that possess significant inhibitory activity against PTP1B (IC_{50} = 1.1 μ g/mL) were isolated from cultures of *Antrodiella albocinnamomea*.⁸¹

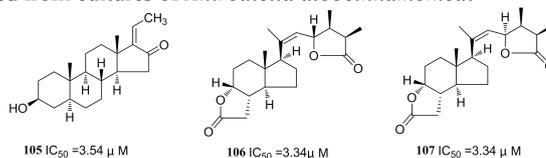


Fig. 12 Structures of compounds **105-107**

3 Semi-synthetic PTP1B inhibitors

Total synthesis and modification of active natural products are better ways to develop lead drugs. In this section, five types of derivatives and their SARs are discussed.

3.1 Maslinic Acid Derivatives

Maslinic acid (MA, **108**, Fig. 13), a natural pentacyclic triterpene acid present in many plant species, has various pharmacological activities including anti-tumour, antioxidant, anti-HIV, antimicrobial and anti-diabetic activities. Qiu *et al.* found that MA exhibits potent inhibitory activity against PTP1B (IC_{50} = 5.93 μ M) and selected it as a promising lead compound to synthesize a series of MA derivatives modified with various fused heterocyclic rings at positions C-2 and C-3. The derivatives exhibited a dramatic increase in inhibitory potency (IC_{50} values all under 10 μ M) along with better selectivity. The two most potent PTP1B inhibitors **109** (IC_{50} = 0.61 μ M, Fig. 13) and **110** (IC_{50} = 0.64 μ M, Fig. 13) showed approximately 10-fold more potency than did the lead compound MA, particularly compound **110**, which possesses the best selectivity (6.9-fold) for PTP1B over TCPTP.⁸²

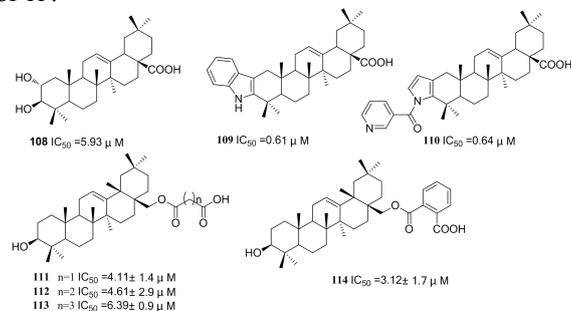


Fig. 13 Structures of compounds **108-114**

3.2 Oleanolic Acid Derivatives

A series of OA derivatives were obtained by Qian *et al.*, who evaluated their anti-PTP1B activity. Of them, four derivatives (**111** - **114**, Fig. 13) showed potent activity, with IC_{50} values of $4.11 \pm 1.4 \mu M$, $4.61 \pm 2.9 \mu M$, $6.39 \pm 0.9 \mu M$, and $3.12 \pm 1.7 \mu M$, respectively. The binding models of compound **114** from molecular docking simulations revealed that (I) the carboxylic group of the triterpene moiety and the hydroxy group of Tyr-46 plus the amino group of Lys-120 form hydrogen bonds and (II) a network of hydrophobic interactions between **114** and Arg-24, Tyr-46, Asp-48, Val-49, Phe-182, Ala-217, Ile-219, and Gln-262 is important for the activity. SAR analysis of these derivatives demonstrated that (I) the integrity of the A ring and 12-ene moieties is important for inhibitory activity; (II) the hydrophilic and acidic groups are an important feature in the anti-PTP1B activity of these derivatives; and (III) the distance between the oleanene and acid moieties is crucial for PTP1B inhibition.⁸³

Twenty-four sugar-substituted OA derivatives were synthesized by Liu and co-workers. Of them, compounds **115-117** (Fig. 14) exhibited the most potent inhibitory activity against PTP1B with IC_{50} values of 1.91, 0.56 and $9.21 \mu M$, respectively.⁸⁴ In 2014, sixteen novel OA derivatives were synthesized, and their inhibitory activities against PTP1B were evaluated in a continuous study. Compounds **118-122** (Fig. 14) exhibited remarkably potent inhibitory activities with IC_{50} values of 6.53, 5.67, 1.03, 0.78 and $3.12 \mu M$, respectively.⁸⁵ From the above results, it is suggested that (I) introducing an acidic chain and sugar-substituted moiety at C-28 could enhance activity against PTP1B; (II) modifying C-3 with a sugar moiety could significantly improve activity against PTP1B and the insulin-sensitizing response; (III) attaching a sugar-substituted moiety to C-3 and C-28 could greatly affect selectivity over TCPTP; and (IV) the lipophilicity of OA derivatives may correlate with their evaluated biological potency.

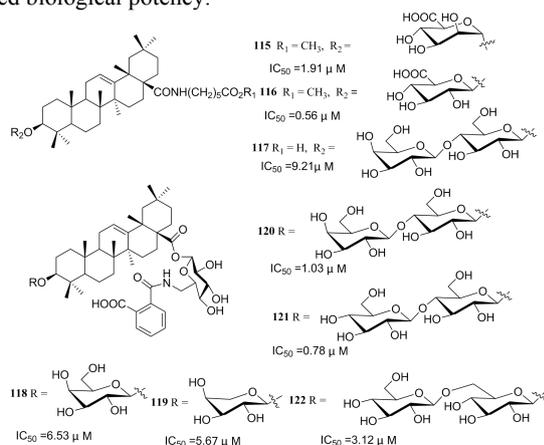


Fig. 14 Structures of compounds **115-122**

3.3 TriterpeneSaponin Derivatives

Xu *et al.* modified the natural triterpenesaponin **123** (Fig. 15) at 3-position, which yielded fourteen potent inhibitory derivatives whose IC_{50} values were under $5 \mu M$. 3-O-para-carboxylphenyl substituted derivative **124** (Fig. 15) was proven to show the best inhibition activity ($IC_{50} = 0.27 \mu M$), whereas 3-O-meta-carboxylphenyl **125** (Fig. 15) exhibited the best selectivity between PTP1B and TCPTP. The SARs of these derivatives were summarized as (I) the hydroxyl group might not influence anti-

PTP1B activity; (II) the activity gradually increased along with increasing length of the introduced fatty chain until the carbon number of the chain reached six; (III) introduction of an aroyl with electron-withdrawing groups was more beneficial for the inhibitory activity than for that of moieties with electron-donating groups; and (IV) introduction of an aryl with a carboxyl group could largely enhance the activity.⁸⁶

3.4 Lithocholic Acid Derivatives

The steroid acid lithocholic acid (LCA, **126**, Fig. 15) with PTP1B inhibitory activity of $12.7 \mu M$ was used as a lead compound to develop potent PTP1B inhibitors bearing two methyls at position C-4 and a fused heterocyclic ring A. Most of them had improved inhibitory activity against PTP1B, and some of them had better selectivity. The most potent compound **127** (Fig. 15) showed excellent activity with an IC_{50} value of $1.62 \pm 0.08 \mu M$, which was about eight-fold greater than LCA, and 14-fold selectivity over the homogenous enzyme TCPTP, with no obvious inhibition against SHP-1, SHP-2 or LAR. It was found that the SARs of these compounds were the following: (I) the charge effect of the substitutions on the fused heterocycle play a key role in the activity; (II) the size of the fused cycles has no obvious effect on the activity; and (III) the 24-carboxyl is important for the activity.⁸⁷

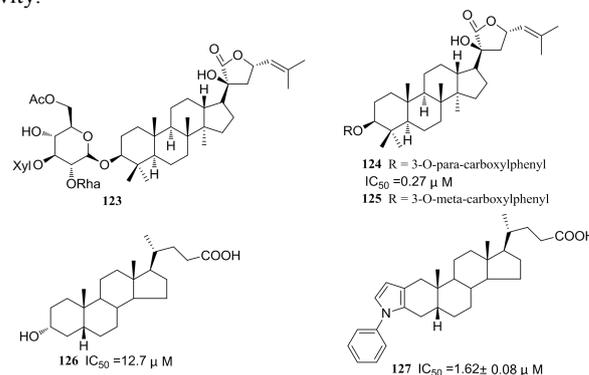


Fig. 15 Structures of compounds **123-127**

The preferred coordination modes of **127** with PTP1B and TCPTP are as follows: the carboxylic acid group of **127** is bound into the active site; the carbonyl of the $-COOH$ may interact with Arg221 via a salt bridge; the hydroxyl of the $-COOH$ shows an H-bond interaction with Ser216; and the phenyl group of **127** binds in the second phosphotyrosine (pTyr) binding site of PTP1B by ion-p interaction with the guanidine group of Arg24.

3.5 Bromophenol Derivatives

Based on the promising finding that bromophenols possess potent inhibitory activity against PTP1B and to find a potential drug to treat T2DM, a series of bromophenol derivatives were synthesized by using active bromophenols as lead compounds, and their SARs are discussed here.⁸⁸⁻⁹³ The preliminary SARs indicated that (I) the tricyclic scaffold promotes the activity; (II) multiple bromine atoms (four to five) incorporated into the tricyclic scaffold are favourable for the activity; and (III) a alkoxy methylene group attached to the phenyl ring and the diaryl-methane scaffold are favourable for inhibitory activity against PTP1B based on the activity of these analogues. Among the potent active derivatives, compounds **128-130** (Fig. 16) exhibited remarkable inhibitory activity against PTP1B, with IC_{50} values of $0.89 \mu M$, $1.50 \mu M$ and $0.68 \mu M$, respectively, which are

all better than that of compound **83**. Moreover, compounds **129** and **130** showed high selectivity against other PTPs (TCPTP, leukocyte antigen-related tyrosine phosphatase (LAR), src homology 2-containing protein tyrosine phosphatase-1 (SHP-1 and SHP-2)) and could decrease levels of glucose and HbA1c in adb/db mouse model.

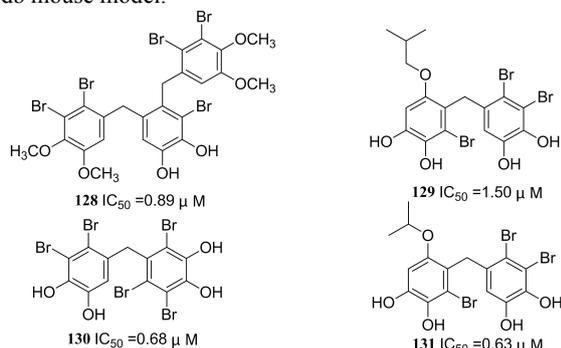


Fig. 16 Structures of compounds **128-131**

We were inspired by the optimization of compound **86**, which provided a novel bromophenol analogue, 3, 4-dibromo-5-(2-bromo-3, 4-dihydroxy-6-(isopropoxymethyl)benzyl)benzene-1, 2-diol (HPN, **131**, Fig. 16), and showed inhibitory activity against PTP1B with IC_{50} 0.63 μM and high selectivity against other PTPs (TCPTP, LAR, SHP-1) and SHP-2). Anti-hyperglycaemic activity assessment demonstrated that HPN significantly decreased the concentration of plasma glucose, lowered serum triglycerides and total cholesterol in a dose-dependent manner, and decreased the levels of HbA1c and insulin compared with the control group. In addition, HPN could decrease PTP1B levels in pancreatic tissue based on western blotting results, and it had anti-hyperglycaemic activity similar to that of rosiglitazone in an intraperitoneal glucose tolerance test in Sprague–Dawley rats.⁹⁴ Assessment of the other pharmacological properties of HPN is in progress, and they will be reported in the future.

25 4 Conclusion and perspective

Over the six years from 2009 to 2014, a variety of potent PTP1B inhibitors were isolated from natural resources based on bioactivity-guided investigations or were prepared by synthesis/semi-synthesis. In this mini-review, the development of diverse small molecule inhibitors with IC_{50} values under 10 μM has been summarized with an emphasis on their structural features, their relevant biological activities, and their SARs.

Diabetes mellitus, including T2DM, is the main threat to human health. Although there are some therapies that can treat this disease, they are unsatisfactory because they do not mimic insulin signalling. PTP1B has been proven to be a promising target for treating T2DM. PTP1B inhibitors were developed very quickly, and they exhibit potential ability to manage diabetes. Nevertheless, it should be noted that these PTP1B inhibitors still retain significant problems due to their poor selectivity over closely related PTPs, such as TCPTP, SHP-1, SHP-2 and PTPRR, which is the key point to overcoming adverse side effects. The secondary binding pockets and the peripheral binding sites around the conserved active site should be used to develop novel PTP1B inhibitors with minimal side effects.

Natural products with various skeletons and diverse bioactivities offer opportunities for the development of new drugs and lead

compounds as potential and novel PTP1B agents. Among these, traditional medicines have been important sources because they have been used for a long time and have already shown therapeutic effects, which would enhance the probability of finding a PTP1B inhibitory drug. In addition, marine organisms are one of the richest sources of natural products with various skeletons and diverse bioactivities and offer more opportunities to develop new drugs and lead compounds with potent activity against PTP1B *in vitro* and *in vivo*. High-throughput screening, which is an effective tool for discovering potent PTP1B inhibitors, combined with the use of natural products, would enhance the speed of development of potent PTP1B inhibitors.

Organic synthesis, especially involving chemical modification, could rapidly provide plenty of candidates from initial substrates (generally active natural or synthesized compounds). The strategies of synthesis and molecular design of PTP1B inhibitors based on rational drug design using computer-assisted docking could rationally increase their bioactivity, decrease their side effects, and improve their physicochemical properties under the guidance of SARs.

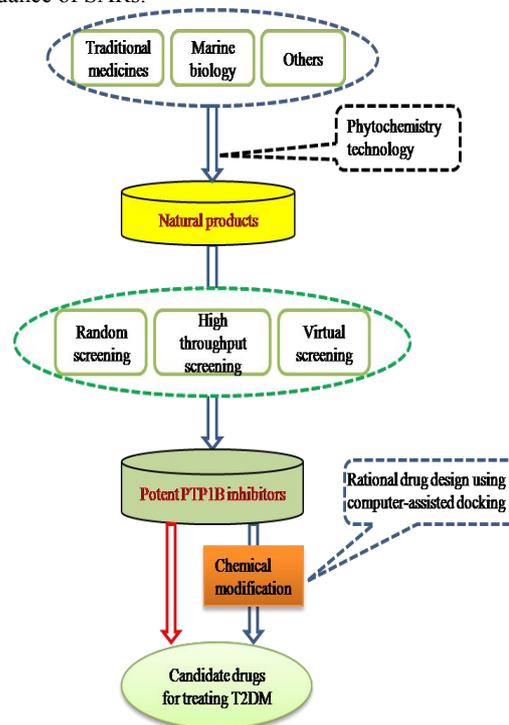


Fig. 17 The strategies for identifying potent and selective PTP1B inhibitors from natural products

The strategies for identifying potent and selective PTP1B inhibitors from natural products are summarized in Fig. 17. It is likely that more potent and selective PTP1B inhibitors with excellent pharmacological properties will be derived from natural products or from the structural optimization of natural products, which might result in novel drugs for treating T2DM by inhibiting PTP1B activity in the future.

Acknowledgements

The work was supported by the National Natural Science Foundation of China (No. 41276167, 41206066 and 41306157), China Postdoctoral Science Foundation (No. 2014M551971 and 2014M551972), Innovation Project Special Fund for Post Doctors

of Shandong Province (No. 201302017 and 201401019), the National science and technology support project (2013BAB01B02-2) and the Natural Science Foundation of Jiangsu Province (Grant No. BK 2012223).

5 Abbreviations

PTP1B = protein tyrosine phosphatase 1B

T2DM = type 2 diabetes mellitus

SARs = structure-activity relationships

IR = insulin receptor

10 WHO = World Health Organization

PTPs = protein tyrosine phosphatases

TCPTP = T-cell protein tyrosine phosphatase

TCM = traditional Chinese medicine

DIO = diet-induced-obese

15 pNPP = p-nitrophenyl phosphate

OA = oleanolic acid

LCA = lithocholic acid

TCPTP = T cell protein tyrosine phosphatase

LAR = leukocyte antigen-related tyrosine phosphatase

20 SHP-1=srchomology 2-containing protein tyrosine phosphatase-1

SHP-2=srchomology 2-containing protein tyrosine phosphatase-2

PTPRR = tyrosine phosphatase receptor type R

References

- G. Roglic, N. Unwin, P. H. Bennett, C. Mathers, J. Tuomilehto, S. Nag, V. Connolly, and H. King, *Diabetes Care*, 2005, **28**, 2130.
- J. Schwarz, S. R. Bornstein, and J. Schulze, *J. Public Health*, 2005, **13**, 303.
- A.A. Tahrani, C.J. Bailey, S. Del Prato, and A. H. Barnett, *Lancet*, 2011, **378**, 182.
- F.Y. Yao, and R.G. Mackenzie, *Pharmaceuticals*, 2010, **3**, 3494.
- J.P. Despres, and I. Lemieux, *Nature*, 2006, **444**, 881.
- K.C. Verbeeten, C.E. Elks, D. Daneman, and K.K. Ong, *Diabet. Med.*, 2011, **28**, 10.
- B.P. Kennedy, *Biomed. Pharmacother.* 1999, **53**, 466.
- H. Sakurai, *Chem. Rec.* 2002, **2**, 237.
- D.W. Haslam, W. Philip, and T. James, *Lancet*, 2005, **366**, 1197.
- M.L. Mohler, Y. He, Z. Wu, D. J.H wang, and D. D. Miller, *Med. Res. Rev.* 2009, **29**, 125.
- D. E. Moller, *Nature* 2001, **414**, 821.
- T. Tiganis, *FEBS J.* 2013, **280**, 445.
- A.J. Nichols, R. D. Mashal, and B. Balkan, *Drug Develop. Res.* 2006, **67**, 559.
- S. Koren, and Fantus, I. G. *Best Pract. Res. Clin. Endocrinol. Metab.* 2007, **21**, 621.
- J. M. Zabolotny, F. G. Haj, Y. B. Kim, B.G. Neel, and B.B. Kahn, *J. Biol. Chem.* 2004, **279**, 24844.
- A.J. Stull, Z. Q. Wang, X. H. Zhang, J. C. Russell, M. Hulver, and W.T. Cefalu, *Diabetes*, 2012, **61**, 1415.
- M. Elchebly, P. Payette, E. Michaliszyn, W. Cromlish, S. Collins, A.L. Loy, D. Normandin, A. Cheng, J. Himms-Hagen, and C.C. Chan, *Science*, 1999, **283**, 1544.
- B. A. Zinker, C. M. Rondinone, J. M. Trevillyan, R. J. Gum, J.E. Clampit, J.F. Waring, N. Xie, D. Wilcox, P. Jacobson, L. Frost, P.E. Kroeger, R.M. Reilly, S. Koterski, T.J. Oppenorth, R.G. Ulrich, S. Crosby, M. Butler, S.F. Murray, R.A. McKay, S. Bhanot, B.P. Monia, and M.R. Jirousek, *Proc. Natl. Acad. Sci. USA.* 2002, **99**, 11357.
- A.P. Combs, *J. Med. Chem.* 2010, **53**, 2333.
- R.J. He, Zeng, He, L.-F. Zhang, Y.T. Jones, Z.-Y. R.M. Ed.; the Royal Society of Chemistry, 2012; Vol. 6, pp. 142.
- G.M. Cragg, P.G. Grothaus, and D.J. Newman, *Chem. Rev.* 2009, **109**, 3012.
- C.S. Jiang, L.F. Liang, and Y.W. Guo, *Acta Pharmacol. Sin.* 2012, **33**, 1217.
- S. Thareja, S. Aggarwal, T.R. Bhardwaj, and M. Kumar, *Med. Res. Rev.* 2012, **32**, 459.
- E. Panzhinskiy, J. Ren, and S. Nair, *Curr. Med. Chem.* 2013, **20**, 2609.
- M. N. Rao, A. E. Shinnar, L. A. Noecker, T.L. Chao, B. Feibush, B. Snyder, I. Sharkansky, A. Sarkahian, X. Zhang, S.R. Jones, W.A. Kinney, and M. Zasloff, *J. Nat. Prod.* 2000, **63**, 631.
- S. Shrestha, B. R. Bhattarai, H. Cho, J.K. Choi, and H. Cho, *Bioorg. Med. Chem. Lett.* 2007, **17**, 2728.
- K.A. Lantz, S.G. Hart, S.L. Planey, M.F. Roitman, I. A. Ruiz-White, H. R. Wolfe, and M. P. McLane, *Obesity*, 2010, **18**, 1516.
- M.P. McLane, I. Ruiz-White, H.R. Wolfe, U.S. Patent 61/129,697, July 14, 2008.
- R. Paduch, M. Kandefer-Szerszen, M. Trytek, and J. Fiedurek, *Arch. Immunol. Ther. Exp.* 2007, **55**, 315.
- C. Sanchez-Quesada, A. Lopez-Biedma, F. Warleta, M. Campos, G. Beltran, and J.J. Gaforio, *J. Agr. Food. Chem.* 2013, **61**, 12173.
- J. J. Ramirez-Espinosa, M. Y. Rios, S. Lopez-Martinez, F. Lopez-Vallejo, J. L. Medina-Franco, P. Paoli, G. Camici, G. Navarrete-Vazquez, R. Ortiz-Andrade, and S. Estrada-Soto, *Eur. J. Med. Chem.* 2011, **46**, 2243.
- T.M. Hung, D.M. Hoang, J.C. Kim, H.S. Jang, J.S. Ahn, and B.S. Min, *J. Ethnopharmacol.* 2009, **124**, 240.
- Y.H. Choi, W. Zhou, J. Oh, S. Choe, D.W. Kim, S.H. Lee, and M. Na, *Bioorg. Med. Chem. Lett.* 2012, **22**, 6116.
- D.Q. Xue, S.C. Mao, X.Q. Yu, and Y.W. Guo, *Biochem. Syst. Ecol.* 2013, **49**, 101.
- M. Na, P.T. Thuong, I.H. Hwang, K. Bae, B.Y. Kim, H. Osada, and J.S. Ahn, *Phytother. Res.* 2010, **24**, 1716.
- Uddin, M. N.; Sharma, G.; Yang, J.L.; Choi, H.S.; Lim, S.-I.; Kang, K. W.; and Oh, W. K. *Phytochemistry* 2014, **103**, 99.
- M. Na, L. Cui, B.S. Min, K. Bae, J.K. Yoo, B.Y. Kim, W.K. Oh, and J.S. Ahn, *Bioorg. Med. Chem. Lett.* 2006, **16**, 3273.
- J.Y. Choi, M. Na, I.H. Hwang, S.H. Lee, E.Y. Bae, Y. Kim, and J.S. Ahn, *Molecules*, 2009, **14**, 266.
- W.B.; Wu, H. Zhang, S.H. Dong, L. Sheng, Y. Wu, J. Li, and J. M. Yue, *J. Asian Na. Prod. Res.*, 2014, **14**, 709.
- L.L. Fang, J. Q. Cao, L.L. Duan, Y. Tang, and Y.Q. Zhao, *J. Funct. Foods.* 2014, **9**, 264.
- W.H. Jiao, X.J. Huang, J.S. Yang, F. Yang, S.J. Piao, H. Gao, J. Li, W.C. Ye, X.S. Yao, W.S. Chen, and H.W. Lin, *Org. Lett.* 2012, **14**, 202.
- Y. Li, Y. Zhang, X. Shen, and Y.W. Guo, *Bioorg. Med. Chem. Lett.* 2009, **19**, 390.
- Y. Zhang, Y. Li, Y.W. Guo, H.L. Jiang, and X. Shen, *Acta. Pharmacol. Sin.* 2009, **30**, 333.
- H. Yamazaki, T. Nakazawa, D.A. Sumilat, O. Takahashi, K. Ukai, S. Takahashi, and M. Namikoshi, *Bioorg. Med. Chem. Lett.* 2013, **23**, 2151.
- M.C. Deng, W.W. Dong, W. Jiao, and R.H. Lu, *Helv. Chim. Acta*, 2009, **92**, 495.
- L. F. Liang, T. Kurtan, A. Mandi, L. X. Gao, W. Zhang, and Y. W. Guo, *Eur. J. Org. Chem.* 2014, **9**, 1841.
- L.F. Liang, L.X. Gao, J. Li, O. Tag lialatela-Scafati, and Y.W. Guo, *Bioorg. Med. Chem.* 2013, **21**, 5076.
- C. C. Liu, C. Lei, Y. Zhong, L. X. Gao, J. Y. Li, M. H. Yu, J. Li, and A. J. Hou, *Tetrahedron* 2014, **70**, 4317.
- H.J. Jung, H.A. Jung, S.S. Kang, J.H. Lee, Y.S. Cho, K.H. Moon, and J.S. Choi, *Arch. Pharm. Res.* 2012, **35**, 1771.
- H.A. Jung, Y.S. Cho, S.H. Oh, S. Lee, B.-S. Min, K.H. Moon, and J.S. Choi, *Arch. Pharm. Res.* 2013, **36**, 957.
- H.A. Fu, Y.M. Luo, C.J. Li, J.Z. Yang, and D.M. Zhang, *Org. Lett.* 2010, **12**, 656.
- S. J. Piao, W.H. Jiao, F. Yang, Y. H. Yi, Y. T. Di, B.N. Han, and H.W. Lin, *Mar. Drugs* 2014, **12**, 4096.
- H.A. Jung, Md. N. Islam, C.M. Lee, H.O. Jeong, H.Y. Chung, H.C. Woo, and J.S. Choi, *Fish Sci.* 2012, **78**, 1321.
- L. H. Yao, Y. M. Jiang, J. Shi, F.A.T. S-Barberan, N. Datta, R. Singanusong, and S.S. Chen, *Plant Food Hum. Nutr.* 2004, **59**, 113.
- T.T. Dao, P.H. Nguyen, P.T. Thuong, K.W. Kang, M. Na, D.T. Ndinteh, J.T. Mbafor, W.K. Oh, *Phytochemistry* 2009, **70**, 2053.
- W. Li, S.P. Li, K. Higai, T. Sasaki, Y. Asada, S. Ohshima, and K. Koike, *Bioorg. Med. Chem. Lett.* 2013, **23**, 5836.
- P.H. Nguyen, G. Sharma, T.T. Dao, M.N. Uddin, K.W. Kang, D.T.

- Ndinteh, J.T. Mbafor, and W.K. Oh, *Bioorg. Med. Chem.* 2012, **20**, 6459.
- 58 T. Sasaki, W. Li, H. Q. Tran, Y. H. Kim, and K. Koike, *Planta Med.* 2014, **80**, 557.
- 5 59 J. S. Choi, M. N. Islam, M.Y. Ali, Y. M. Kim, H. J. Park, H. S. Sohn, and H. A. Jung, *Arch. Pharm. Res.* 2014, **37**, 1354.
- 60 M.H. Duc, M.N. Tran, T.D. Nguyen, T.H. Do, Y.H. Kim, H.V. Luong, J.S. Ahn, and K. Bae, *Bioorg. Med. Chem. Lett.* 2009, **19**, 6759.
- 61 Y. L. Zhang, J. G. Luo, C. X. Wan, Z. B. Zhou, and L. Y. Kong, *Fitoterapia* 2014, **92**, 116.
- 10 62 L. J. Wang, C. A. Geng, Y. B. Ma, J. Luo, X. Y. Huang, H. Chen, N. J. Zhou, X. M. Zhang, and J. J. Chen, *Eur. J. Med. Chem.* 2012, **54**, 352-366.
- 63 M.N. Islam, H.A. Jung, H.S. Sohn, H.M. Kim, and J.S. Choi, *Arch. Pharm. Res.* 2013, **36**, 542.
- 15 64 R.R. Baumgartner, Steinmann, D.; Heiss, E.H.; Atanasov, A.G.; Ganzera, M.; H. Stuppner, and V.M. Dirsch, *J. Nat. Prod.* 2010, **73**, 1578.
- 65 J. Zhang, Q. Shen, J.C. Lu, J.Y. Li, W.Y. Liu, J.J. Yang, J. Li, and K. Xiao, *Food Chem.* 2010, **119**, 1491.
- 20 66 D.S. Lee, J.H. Jang, W. Ko, K.S. Kim, J.H. Sohn, M.S. Kang, J.S. Ahn, Y.C. Kim, and H. Oh, *Mar. Drugs*, 2013, **11**, 1409.
- 67 C. Seo, J.H. Sohn, J.S. Ahn, J.H. Yim, H.K. Lee, and H. Oh, *Bioorg. Med. Chem. Lett.* 2009, **19**, 2801.
- 25 68 C. Seo, J.H. Sohn, H. Oh, B.Y. Kim, and J.S. Ahn, *Bioorg. Med. Chem. Lett.* 2009, **19**, 6095.
- 69 H.E. Moon, M.N. Islam, B.R. Ahn, S.S. Chowdhury, H.S. Sohn, H.A. Jung, and J.S. Choi, *Biosci. Biotechnol. Biochem.* 2011, **75**, 1472.
- 70 M. Liu, P.E. Hansen, and X.K. Lin, *Mar. Drugs*, 2011, **9**, 1273.
- 30 71 D. Shi, F. Xu, J. He, J. Li, X. Fan, and L. Han, *Chin. Sci. Bull.* 2008, **53**, 2476.
- 72 D.Y. Shi, F. Xu, J. Li, S.J. Guo, H. Su, and L.J. Han, *Chin. J. Chin. Mater. Med.* 2008, **33**, 2238.
- 73 J.C. Qin, H. Su, Y.M. Zhang, J.M. Gao, L. Zhu, X. Wu, H.Y. and Pan, X. Li, *Bioorg. Med. Chem. Lett.* 2010, **20**, 7152.
- 35 74 X. Liu, X.M. Li, L.X. Gao, C.M. Cui, C.X. Li, J. Li, and B.G. Wang, *Chin. J. Oceanol. Limnol.* 2011, **29**, 689.
- 75 H. Yamazaki, D.A. Sumilat, S. Kanno, K. Ukai, H. Rotinsulu, D.S. Wewengkang, M. Ishikawa, R.E.P. Mangindaan, and M. Namikoshi, *J. Nat. Med.* 2013, **67**, 730.
- 40 76 Y. Bustanji, M. O. Taha, A. M. Yousef, and A. G. Al-Bakri, *J. Enzym. Inhib. Med. Chem.* 2006, **21**, 163.
- 77 C.H. Chen, Y.B. Zhang, and C. Huang, *Biochem. Biophys. Res. Commun.* 2010, **97**, 543.
- 45 78 Y. Bustanji, M. O. Taha, I. M. Al-Masri, and M. K. Mohammad, *Biol. Pharm. Bul.* 2009, **32**, 640.
- 79 Q. B. Chen, X. L. Xin, Y. Yang, S.-S. Lee, and H. A. Aisa, *J. Nat. Prod.* 2014, **77**, 807.
- 80 J.R. Wang, Q. Shen, L. Fang, S.Y. Peng, Y.M. Yang, J. Li, H.L. Liu, and Y.W. Guo, *Steroids*, 2011, **76**, 571.
- 50 81 Z. M. Chen, X. Y. Yang, Q. Y. Fan, Z. H. Li, K. Wei, H. P. Chen, T. Feng, and J. K. Liu, *Steroids* 2014, **87**, 21.
- 82 W.W. Qiu, Q. Shen, F. Yang, B. Wang, H. Zou, J.Y. Li, J. Li, and J. Tang, *Bioorg. Med. Chem. Lett.* 2009, **19**, 6618.
- 55 83 S. Qian, H.J. Li, Y. Chen, W.Y. Zhang, S.Y. Yang, and Y. Wu, *J. Nat. Prod.* 2010, **73**, 1743.
- 84 Q.C. Liu, T.T. Guo, L. Zhang, Y. Yu, P. Wang, J.F. Yang, and Y.X. Li, *Eur. J. Med. Chem.* 2013, **63**, 511.
- 85 Q. C. Liu, T. T. Guo, D. Li, F. H. Li, and W. H. Li, *Eur. J. Med. Chem.* 2014, **79**, 34.
- 60 86 J.Q. Xu, Q. Shen, J. Li, and L.H. Hu, *Bioorg. Med. Chem.* 2010, **18**, 3934.
- 87 H.B. He, L.X. Gao, Q.F. Deng, W.P. Ma, C.L. Tang, W.W. Qiu, J. Tang, J.Y. Li, J. Li, and F. Yang, *Bioorg. Med. Chem. Lett.* 2012, **22**, 7237.
- 65 88 B. Jiang, S.J. Guo, D.Y. Shi, C. Guo, and T. Wang, *Eur. J. Med. Chem.* 2013, **64**, 129.
- 89 Y.C. Cui, D.Y. Shi, and Z.Q. Hu, *Chin. J. Oceanol. Limn.* 2011, **29**, 1237.
- 70 90 J. Li, S.J. Guo, H. Su, L.J. Han, and D.Y. Shi, *Chin. Chem. Lett.* 2008, **19**, 1290.
- 91 D.Y. Shi, J. Li, B. Jiang, S.J. Guo, H. Su, and T. Wang, *Bioorg. Med. Chem. Lett.* 2012, **22**, 2827.
- 92 B. Jiang, D.Y. Shi, Y.C. Cui, and S.J. Guo, *Arch. Pharm. Chem. Life Sci.* 2012, **345**, 444.
- 75 93 S.J. Guo, J. Li, T. Li, D.Y. Shi, and L.J. Han, *Chin. J. Oceanol. Limn.* 2011, **29**, 68.
- 94 D.Y. Shi, S.J. Guo, B. Jiang, C. Guo, T. Wang, L.Y. Zhang, and J.Y. Li, *Mar. Drugs* 2013, **11**, 350.
- 30

Notes

Institute of Oceanology, Chinese Academy of Sciences, 7 Nanhai road, 266071, Qingdao, China. Fax: + 86 532 82898719; Tel: + 86 532 82898719; E-mail: shidayong@qdio.ac.cn