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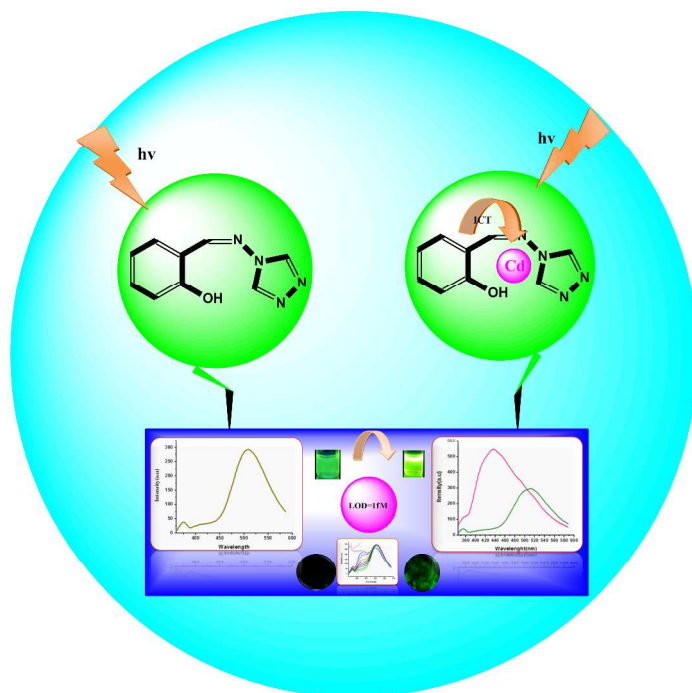
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## Graphical image



1 **A Simple Highly Sensitive and Selective TURN-ON Fluorescent Chemosensor for the**  
2 **Detection of Cadmium Ions in Physiological Condition**

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## 28 Abstract

29 A very simple fluorophore was synthesized and applied as chemosensor for Cd<sup>2+</sup> ions detection.  
30 Structural characterization was done by <sup>1</sup>HNMR <sup>13</sup>C and ESI-Mass techniques. All the  
31 fluorimetric titrations were carried out by using spectrofluorimetry. The fluorophore has showed  
32 a very good with LOD of 1 fM towards Cd<sup>2+</sup> ions. The linear range of detection is 5 fM<sup>-1</sup> to  
33 1mM. Moreover, it has successfully discriminate Zn<sup>2+</sup> ions and all interfering metal ions in  
34 physiological pH (7.5). The quantum yield of the fluorophore increased from 0.4 to 0.78 after the  
35 binding of Cd<sup>2+</sup> ions. The formation of 1:1 complex is confirmed by Job's plot, <sup>1</sup>H NMR and  
36 ESI-Mass which have strongly manifests the formation of 1:1 complex. The binding constant is  
37 also calculated for the probe-Cd<sup>2+</sup> ions in lower (femto) concentrations as 2.2 x10<sup>3</sup> M<sup>-1</sup>. The  
38 chemosensor has also exhibited very good results in He-La Cells imaging under physiological  
39 pH. So the chemosensor that has developed can be recommended for the practical biological  
40 applications especially in oncology.

41 **Keywords:** Chemosensors, Internal Charge Transfer, Discrimination, Turn ON, Cell imaging

42

## 43 Introduction

44 Design of a metal ion sensor has been allured through many researchers related to various  
45 applications such as clinical toxicology, environmental bioinorganic chemistry, bioremediation,  
46 and waste management.<sup>1-3</sup> There were so many sensors have been developed for different metal  
47 ions since from the last 10 decades. The need for the development of metal sensor has been  
48 increasing till now for their peculiar sensitivity in order to applications concern. Among various  
49 metal ions Cd<sup>2+</sup> ions, is highly noticed due to its high toxicity and carcinogenicity. Cadmium,  
50 whose half-life in humans is estimated to be between 15 and 20 years is listed by the U.S.  
51 Environmental Protection Agency as one of 126 priority pollutants. Cadmium was listed a  
52 number 7 on ATSDR's "CERCLA Priority List of Hazardous Substances". In addition, it can  
53 accumulate in the human body for >10 years.<sup>4</sup> The extensive use of cadmium metal in industrial  
54 and agriculture fields lead to water and soil contamination. Cadmium ions contamination is  
55 mainly derived from phosphate fertilizers, metal alloys, Ni-Cd batteries, paint pigments, ceramic  
56 enamels and natural factors like erosion, abrasion and volcanic eruption.<sup>5</sup> The intake of cadmium

ions by the cells has showed an adverse effect in cellular functions, molecular mechanism and causes so many acute health diseases like lung cancer, kidney cancer, renal cancer and prostate cancers. Smoking and inhalation of cadmium-containing dust represent additional sources of cadmium uptake in humans. Cadmium ions accumulation is involved in neurological, reproductive, cardiovascular, and developmental disorders.<sup>6</sup> Even the fertility of the soil is affected primarily with cadmium ions contamination which leads to the poor growth of crops. In order to quantification of cadmium ions, numerous high cost and sophisticated methods were available such as inductively coupled atomic plasma mass spectroscopy (ICP-MS),<sup>7</sup> inductively coupled plasma emission spectroscopy (ICP-AES),<sup>8</sup> atomic absorption spectroscopy (AAS) and anode stripping voltammetry.<sup>9</sup> Amid these, the fluorescence spectral analysis has been used as a powerful analytical tool owing to its operational simplicity, cost effectiveness, high sensitivity and selectivity.<sup>10</sup> Many fluorescent sensors has been successfully developed for cadmium ions based on the mechanism like PET, ICT, CHEF, FRET etc.<sup>11-12</sup> But only a few of them are applicable to live cell imaging.<sup>13</sup> It is due to poor water solubility of probe, UV-excitation and pH-dependent fluorescence in physiological environments. However the need for developing the cadmium ion sensor in aqueous and biological media has been gradually increasing for its potential applications in living cell imaging. Besides, there is a challenge persisting till now in discriminating ability of the probe towards  $Zn^{2+}$  ions in selectivity point of view concern since these two metal ions are having similar physical and chemical properties. Hence, most of the developed sensors showed a poor selectivity between  $Cd^{2+}$  and  $Zn^{2+}$  ions.<sup>14</sup> In some cases, the fluorescence enhancement was achieved for probe after the addition of various nanomaterials like silver nanoparticles, quantum dots and graphene sheets.<sup>15</sup> But these type of nanoparticles based cadmium ions sensors are belonging to the hard matter.

Recently few chemosensors have been developed for distinguish the  $Cd^{2+}$  ions from  $Zn^{2+}$  ions with different mechanisms.<sup>16-18</sup> Amidst these, only a very few reports had showed a good discrimination for  $Cd^{2+}$  ions from  $Zn^{2+}$  ions.<sup>19-20</sup> Lu and co-workers had reported a selective ratiometric sensor for  $Cd^{2+}$  ions which were detected in neutral aqueous medium based on two different ICT mechanism.<sup>21</sup> By utilizing the unique role of 2-picoyl group, they demonstrated a new discrimination strategy for  $Cd^{2+}$  ions from  $Zn^{2+}$  ions with the LOD of  $0.1\mu M$ . For the first time Peng and co-workers had developed  $Cd^{2+}$  ions sensor in water/acetone mixture which had showed a good discrimination of  $Cd^{2+}$  ions from  $Zn^{2+}$  ions and enhanced fluorescence intensity

88 with shift in emission using BODIPY (4,4-difluoro-1,3,4,5,7-quantmethyl-4- bora-3a,4a-diaza-*s*-  
89 indacene) as fluorophore.<sup>22</sup> Table.1 shows the different types Cd<sup>2+</sup> ion sensors reported so far  
90 with their LOD. Keeping these points in mind, we have synthesized a very simple Schiff base  
91 ligand [(*Z*)-2-(4H-1, 2, 4-triazol-4-yl) imino methylphenol] as a probe and utilized successfully  
92 for the sensing of Cd<sup>2+</sup> ions over Zn<sup>2+</sup> ions. The developed fluoro sensor has showed a clear  
93 discrimination for Cd<sup>2+</sup> ions towards Zn<sup>2+</sup> ions when compare to the other sensors mentioned  
94 above. Besides, the sensor has showed a very good linear range of detection from 5fM to 1mM  
95 with an excellent LOD of 1fM.

96

## 97 **Experimental**

### 98 **Materials and general methods**

99 Analytical grade solvents and double distilled water were used in all steps. The acetate salts of  
100 different metal ions were used. <sup>1</sup>H-NMR and <sup>13</sup>C-NMR were measured on a BrukerAV-400  
101 spectrometer with chemical shifts reported in ppm (in (CD<sub>3</sub>)<sub>2</sub>S=O, or DMSO; TMS as internal  
102 standard). Mass spectra were measured on a Thermofleet LC-MS spectrometer. All pH  
103 measurements were made with a Eutek pH-Tutor. All the UV studies were done with JASCO  
104 550 and Fluorescence spectra titrations were carried out with CARY Eclipse Fluorescence  
105 spectrophotometer. Fluorescence microscope (Model: LEICA DMLS) with an objective lens of  
106 20 magnification was used for the cell imaging studies.

### 107 **Synthetic route for the fluorophore**

108 A solution of I (1.0 mL, 0.94 mmol, 1 equiv.) dissolved in absolute ethanol (10 mL) was  
109 added to a solution of II (1 equiv.) in ethanol (5 mL). The resulting slightly yellow solution was  
110 irradiated in microwave oven for 10 mins and the resulting pale-yellow solution was allowed to  
111 cool to room temperature to give a crystalline white powder III (Scheme 1). The product was  
112 filtered, washed with ethanol and diethyl ether and dried over MgSO<sub>4</sub> vacuum (2.5 g, 95%). The  
113 method of preparation was simple and fast as the totally 10mins was required for this synthesis,  
114 when compare to other previous reports.<sup>27, 28</sup> <sup>1</sup>H NMR (400 MHz, [D6] DMSO, 298 K): δ =  
115 10.47 (s, 1 H), 9.18 (D, 2H), 7.80 (m, 1 H), 7.44 (m, 1 H), 7.02 (m, 2 H) ppm.(Fig. S1) <sup>13</sup>C NMR  
116 (400 MHz, [D6] DMSO, 298 K): δ = 158.57 (C9), 155.31 (C7), 139.43 (C3 and C5), 134.36  
117 (C11), 128.13 (C13), 120.11 (C10), 118.65 (C8), 117.17 (C12) ppm.(Fig. S2) C<sub>9</sub>H<sub>8</sub>N<sub>4</sub>O  
118 (calculated mass 188.19): Observed Mass [M+H]<sup>+</sup>=187.09 (Fig. S3)

119

## 120 **Result and Discussion**

### 121 **Optimization of pH for the Probe**

122 In general the protons in probe/fluorophore determine the fluorescence response of the probe and  
123 the detection of metal ions, so it is necessary to investigate the effect of pH. Fig. 1 shows the  
124 fluorescence response of probe as a function of pH. Initially the probe showed an emission peak  
125 at 511 nm (Fig. 2b) which is mainly owing to the ICT mechanism. The donating nature of  
126 hydroxyl and imine functional groups are took part into this mechanism. From the optimization  
127 graph, it was clearly seen that under acidic conditions the functional groups in the fluorophore  
128 gets protonated. So the initial ICT mechanism is arrested. When the pH is increased from 1.0 to  
129 7.0 there is a gradual increment in fluorescent intensity. This is mainly due to the gradual  
130 relaxation of arrested ICT mechanism. At pH 7.0, a maximum emission was observed due to the  
131 free hydroxyl and imine functional groups under neutral medium which initiates the ICT  
132 mechanism once again. Under this neutral aqueous media, in the ground state, the hydroxyl  
133 group promotes the electron transfer. When the pH is increased from 7.0 to 13.5, a decrease in  
134 fluorescence signal (excited at 332 nm) could be observed for the probe suggesting that  
135 deprotonation of hydroxyl group takes place. Meanwhile, the imine group gets protonated by  
136 absorbing this deprotonated hydrogen from the hydroxyl group. As a result, a gradual decrease in  
137 emission intensity is observed which mainly suggests the inhibition of ICT mechanism. No  
138 drastic change in wavelength shift is observed while varying the pH from 1 to 13.5. Under acidic  
139 condition (pH 1), the probe shows an emission peak at 501nm while the pH is adjusted from 2 to  
140 6, the emission peak is slightly red shifted from 501 nm to 508 nm. Under neutral medium (pH  
141 7), the probe gives its original fluorescence peak at 511 nm. When the pH is further varied from  
142 8 to 13.5, the emission peak at 511 nm is blue shifted from 511 nm to 490 nm. (Fig. 1)

143

### 144 **Absorption and Emission studies for the probe – Cd<sup>2+</sup> ions**

145 In absorption spectra, initially two peaks are observed for the probe at 273 nm and 332 nm which  
146 corresponds to the  $\pi$ - $\pi^*$  and  $n$ - $\pi^*$  transitions respectively (Fig.2a). Upon the addition of  
147 Cd<sup>2+</sup> ions, the intensities of absorption peaks at 273nm and 332nm was increased gradually (Fig.  
148 3 a) and they didn't show any remarkable shift. In emission spectra only one peak is observed at  
149 511 nm with the excitation wavelength of 332 nM (Fig.2b). Upon the addition of Cd<sup>2+</sup> ions, a



150 new peak is observed at 433 nm with remarkable enhancement of fluorescence intensity (TURN-  
151 ON) along with a notable blue shift from the peak at 511 nm. Because, the electron donating  
152 ability of the probe is inhibited when  $\text{Cd}^{2+}$  ions bind with it and the corresponding ICT  
153 mechanism is inhibited once again, resulting the blue shift in emission spectra (Fig. 4). In case of  
154 fluorescent signaling, molecular switching and metal ions sensing, the Internal Charge Transfer  
155 mechanism has been widely exploited owing to its extraordinary spectral shift followed by  
156 quantitative detection. Generally, if a fluorophore contains an electron donating group like  
157 amino, it undergoes ICT from the donor to the fluorophore upon light excitation, which provides  
158 a red-shifted emission. When coordinated with a metal ion, the amino group loses its donating  
159 ability. Consequently, the ICT is inhibited and the emission shows blue shift. So that quantum  
160 yields always change in the processes. In this chemosensor, triazole part could act as a receptor  
161 and the hydroxyl group of the salicyaldimine as ICT donor. An imine group between the receptor  
162 and the donor part can induce longer wavelengths in absorption and fluorescence spectra. Hence,  
163 the possibility of FRET is ruled out as there is no donor and acceptor separately located in our  
164 probe.<sup>29</sup> So the ICT mechanism is followed here. The fluorescence intensity of the new peak at  
165 436 nm is significantly enhanced and the quantum yield is increased from 0.4 to 0.8 with a  
166 remarkable blue shift in wavelength. When the concentration of  $\text{Cd}^{2+}$  ions is gradually increased,  
167 the intensity of the emission peak at 436 nm is also increased with a remarkable blue shift from  
168 the peak at 511 nm (Fig.3b). The linear range of the developed sensor is varied from 5fM to  
169 1mM Fig 3b and inset in Fig. 3b show the selective overlay and the corresponding linearity plot  
170 with varying the  $\text{Cd}^{2+}$  ions concentration. For the very first time our simple probe is showed an  
171 excellent LOD of 1fM. Even for this femto level range a pronounced increased in intensity was  
172 observed (Fig. 5)

173

#### 174 **Calculation of quantum yields ( $\Phi$ ) by Emission Spectra**

175 Fluorescence quantum yields ( $\Phi$ ) were estimated from the area under the corresponding  
176 fluorescence curves recorded for probe as well as probe- $\text{Cd}^{2+}$  ions. The absorbance value of  
177 probe was chosen from the absorption spectra (Fig. 2a).The concentration of probe is 5  $\mu\text{M}$  and  
178 the integrated area is calculated from the emission spectra of probe and probe- $\text{Cd}^{2+}$  ions (Fig. 2b  
179 &3) with excitation wavelength of 332 nm in phosphate buffer pH=7.0 . For the standard



180 Rhodamine 6G, all the values were taken from the previous report. The quantum yield  
 181 calculation was done based on the following equation.<sup>30, 31</sup>

$$\phi_u = \frac{\phi_s I_u A_s \lambda_{exs} \eta_u}{I_s A_u \lambda_{exu} \eta_s}$$

182  
 183  
 184  
 185 Where,  $\phi$  is the quantum yield;  $I$  is integrated area under the corrected emission spectra;  $A$  is  
 186 absorbance at the excitation wavelength;  $\lambda_{ex}$  is the excitation wavelength;  $\eta$  is the refractive  
 187 index of the solution; the subscripts  $u$  and  $s$  refer to the unknown and the standard, respectively.  
 188 For this study, Rhodamine 6G in Methanol is taken as a standard, which has the quantum yield  
 189 of 0.91.

190

### 191 **Determination of Binding Constant Probe-Cd<sup>2+</sup> ions Complex**

192 The binding constant  $K_s$  of the probe- Cd<sup>2+</sup> ions are determined in both lower and higher  
 193 concentration ranges (Fig. 5a and 5b). Benesi-Hildebrand modified equation was used for the  
 194 calculation of binding constant, where the binding constant was calculated from the ratio of  
 195 intercept to slope.

196

$$K_s = \text{Intercept} / \text{Slope}$$

197 And it is found to be  $2.2 \times 10^3 \text{ M}^{-1}$  for femto molar range. From this data, it is clearly revealed  
 198 that the developed sensor exhibited a very good binding even in lower concentration of Cd<sup>2+</sup> ions.  
 199 The stoichiometry of the probe- Cd<sup>2+</sup> ion is confirmed through a continuous variation method  
 200 i.e., the Job's Plot.<sup>32</sup> From the figure 6, it is clearly revealed the formation of 1:1 stoichiometry  
 201 of with a 0.5 molar ratio of probe- Cd<sup>2+</sup> complex. In addition to that it is also confirmed via the  
 202 mass spectrometry studies (Fig. S4) and <sup>1</sup>H NMR titrations (Fig. S5).

203 The reversibility experiment was also done by EDTA titration.<sup>33</sup> The initial fluorescence  
 204 intensity at 436 nm of probe-Cd<sup>2+</sup> was gradually diminished while vary the EDTA from zero to  
 205 one equivalent. (Inset Fig. S6). While adding the one equivalent of EDTA to the solution  
 206 containing probe-Cd<sup>2+</sup>, the initial fluorescence intensity of the probe at 511 nm is recovered (Fig.  
 207 S6). These results suggested that the developed sensor could be reusable for real time analysis.

208 [Detailed experimental condition was given in supporting information.] (Scheme 2)

209

210

211

### 212 **<sup>1</sup>H NMR titrations of probe-Cd<sup>2+</sup> ions complex formation**

213 In further, the binding of Cd<sup>2+</sup> ions with the probe was confirmed by NMR titrations. Upon the  
214 addition of 1 equivalent of Cd<sup>2+</sup> ions to the 1 equivalent of probe, the peak at 10.5 ppm (imine  
215 peak) and peak at 3.4ppm (OH) is completely quenched. This study clearly reveals that after the  
216 binding of Cd<sup>2+</sup> ions, the deprotonation takes place. And there is a slight downfield shift  
217 experienced by the remaining characteristic peaks which is mainly due to the deshielding effect  
218 of the Cd<sup>2+</sup> ions through the direct N and O-metal interactions (Fig. S5).

219

### 220 **Interference studies of other metal ions**

221 The specific selectivity of the probe was also determined by recording the florescent spectra in  
222 the presence of series of other metal cations. The interference of other heavy metal ions has  
223 studied and observed at the emission peak of Cd<sup>2+</sup> ion –probe at 436 nm. Except Cd<sup>2+</sup> ions, other  
224 ions like Au<sup>3+</sup>, Zn<sup>2+</sup>, Pb<sup>2+</sup>, Hg<sup>2+</sup>, Cr<sup>3+</sup>, Fe<sup>2+</sup>, Cu<sup>2+</sup>, Ca<sup>2+</sup>, Ba<sup>2+</sup>, Mg<sup>2+</sup>, Co<sup>2+</sup>, Ni<sup>2+</sup>, Ag<sup>+</sup>, Na<sup>+</sup> and K<sup>+</sup>  
225 ions do not show any fluorescence effect at 436 nm. Only the peak intensity at 511 nm slightly  
226 decreases for some metal ions without the formation of new peak (Fig. 4). This is due to the  
227 formation on non fluorescent complexes with other metal ions.<sup>10,34</sup> In order to further support,  
228 the color changes were observed both visual eye as well as under UV lamp, while the interferents  
229 is added to the probe Cd<sup>2+</sup> ion alone showed an increase in fluorescence when compare to all  
230 other interfering metal ions in both visualization.(Fig. S7). Moreover for the Zn<sup>2+</sup> ions, the  
231 synthesized probe has showed an excellent discrimination over Cd<sup>2+</sup> ions when compare to the  
232 other Cd<sup>2+</sup> ions sensors reported. Under optimized conditions, the developed chemosensor has  
233 exhibited a significant enhancement of fluorescence intensity in presence of Cd<sup>2+</sup> ions. The  
234 competitive fluorimetric titrations were also conducted and the results showed a very good  
235 selectivity towards Cd<sup>2+</sup> ions rather than other metal ions. Two types of titrations were done  
236 here. Initially 1 equivalent of Cd<sup>2+</sup> ions (10 μL) was added into the solution of probe (5μM) in  
237 the presence of one equivalent of other metal ions. Simultaneously, the same competitive  
238 titrations were also carried out with two equivalents of other metal ions (20 μL) (Fig. 7a & 7b).  
239 In both the cases, Hg<sup>2+</sup> ions induce minimal quenching of fluorescence, whereas Li<sup>+</sup> Ca<sup>2+</sup> and  
240 Na<sup>+</sup> ions induce a slight enhancement of the fluorescence.

241

### 242 **Competitive Spectral Studies in lower concentration (fM) of Cd<sup>2+</sup> ions**

243 Additionally, the interference of other metal ions were also cross checked for very low  
244 concentration of  $\text{Cd}^{2+}$  ions ( $f\text{M}$ ). For this, 1 ml of  $\text{Cd}^{2+}$  ions (in Femto molar concentration) was  
245 added to the 1 ml of probe in the presence of 1 equivalent (10  $\mu\text{L}$ ) and two equivalents (20  $\mu\text{L}$ )  
246 of other metal ions (Fig. 8a & 8b). In the first test, a  $\text{Co}^{2+}$  ion has showed a slight increase in  
247 fluorescence. Whereas in presence of 2 equivalents of other metal ions, there is a slight  
248 increment of fluorescence intensity observed. Hence from these two competitive experiments, it  
249 is clearly showed that our probe showed a very good selectivity towards  $\text{Cd}^{2+}$  ions in a very low  
250 concentration ( $f\text{M}$ ) in presence of other metals, even in higher concentrations (Fig. 9).

251

### 252 **Naked Eye Discrimination of $\text{Cd}^{2+}$ from $\text{Zn}^{2+}$ ions**

253 As we pointed out early, the discrimination of  $\text{Cd}^{2+}$  ions from  $\text{Zn}^{2+}$  ions is very important on  
254 cadmium ion based chemosensors. There are many similarities between  $\text{Cd}^{2+}$  and  $\text{Zn}^{2+}$  ions.  $\text{Cd}^{2+}$   
255 ion is commonly found in Zn ores, which are the principal commercial sources of  $\text{Cd}^{2+}$ . Both  
256 metals are classified, commonly with mercury (Hg), in group II B of post-transition elements of  
257 the periodic table. Besides,  $\text{Cd}^{2+}$  ion are intermediate in size between  $\text{Zn}^{2+}$  and  $\text{Hg}^{2+}$  ion.  $\text{Cd}^{2+}$  and  
258  $\text{Zn}^{2+}$  ions have a similar electron configuration on their outer shells. Both metals have filled inner  
259 electron shells (d shells) with the outermost shell having two s electrons.  $\text{Zn}^{2+}$  ion, unlike  $\text{Cd}^{2+}$   
260 ion is relatively stable in its divalent state and does not undergo redox changes.  $\text{Cd}^{2+}$  ion is  
261 generally classified as a ‘soft’ metal that is more likely to form covalent linkages with electron-  
262 donating ligands, whereas  $\text{Zn}^{2+}$  ion is classified as ‘intermediate in softness.’<sup>35</sup> In this developed  
263 chemosensor, the binding site i.e. imine nitrogen and hydroxyl group both are coming under soft  
264 base categories. So it has selectively binds  $\text{Cd}^{2+}$  ion (soft acid) rather than  $\text{Zn}^{2+}$  ion (intermediate  
265 base). This phenomenon was also confirmed by the selectivity fluorimetric titrations itself.  
266 Moreover, it is clearly distinguished by both naked eye and UV lamp analysis. Initially the probe  
267 was colorless under normal vision but it showed an intense green color after the binding of  $\text{Cd}^{2+}$   
268 ions with the probe. The same trend was also observed when the probe is subjected to UV lamp.  
269 The slight green fluorescence color of the probe is increased immensely (very bright green  
270 fluorescence) after the addition of  $\text{Cd}^{2+}$  ions (Fig.10a & 10b). In both the visualization,  $\text{Zn}^{2+}$  ions  
271 have showed only a negligible change. So our synthesized fluorophore is successfully  
272 discriminates the  $\text{Cd}^{2+}$  ions from the  $\text{Zn}^{2+}$  ions in both naked eye and under UV Lamp.

273

### 274 **Imaging studies at Living cells**

275 To further highlight the practical application of the probe, we carried out living cells imaging  
276 studies by using the confocal fluorescence microscopy. Incubation of He-La cells with probe (5  
277  $\mu\text{M}$ ) for 0.5 h at 37 °C was followed by the addition of  $\text{Cd}^{2+}$  ions (5  $\mu\text{M}$ ) and then was incubated  
278 for another 0.5 h. Initially the cell exhibits a moderate weak fluorescence and then the great  
279 enhancement of fluorescence is observed (Fig. 11). These results are suggested that the ability of  
280 the probe to penetrate the cell membrane and can be used for tracking the of  $\text{Cd}^{2+}$  ions in living  
281 cells and in vivo potentially.

282

### 283 **Conclusion**

284 In conclusion, we have designed a new simple fluorescent sensor platform based on the simple  
285 salicyaldimine based fluorophore. It shows high sensitivity in femto level and selectivity toward  
286  $\text{Cd}^{2+}$  ion in physiological pH. The fluorescence intensity was significantly enhanced and the  
287 quantum yield was increased from 0.4 to 0.8. Moreover, its fluorescence intensity is enhanced in  
288 a linear fashion with lower concentration (fM) of  $\text{Cd}^{2+}$  ion and thus can be potentially used for  
289 quantification of  $\text{Cd}^{2+}$  ion even it will present in femto molar concentration. The living cell  
290 imaging studies under physiological pH further demonstrate its value in the practical applications  
291 of biological systems.

292

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296

### 297 **Abbreviations**

298 PET- Photoinduced Electron Transfer, ICT- Internal Charge Transfer, FRET- Fourier Energy  
299 Transfer, CHEF- Chealtion Enhanced Fluorescence.

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304 **References**

- 305 (1) (a) G. K. Walkup and B. Imperiali, *J. Am. Chem. Soc.*, 1996, **118**, 3053. (b) Liu, J.; Lu, Y. *J.*  
306 *Am. Chem. Soc.*, 2004, **126**, 12298.
- 307 (2) (a) S. Deo and H. A. Godwin, *J. Am. Chem. Soc.*, 2000, **122**, 174. (b) J. Li, Y. Lu, *J. Am.*  
308 *Chem. Soc.*, 2000, **122**, 10466.
- 309 (3) (a) A. Miyawaki, J. Llopis, R. Helm, J. M. McCaffery, J. A. Adams, M. Ikura, R. Y. Tsien,  
310 *Nature.*, 1997, **388**, 882. (b) P. Chen, C. He, *J. Am. Chem. Soc.*, 2004, **126**, 728.
- 311 (4) (a) H. Wang, L. Xue, H. Jiang, *Org. Lett.*, 2011, **13**, 3844. (b) P. N. Basa, A. Bhowmick,  
312 M. M. Schulz, A. G. Sykes, *J. Org. Chem.*, 2011, **76**, 7866. (c) A. E. Palmer, *ACS*  
313 *Chem. Biol.*, 2009, **3**, 157. (d) J. H. Lee, A. R. Jeong, I. S. Shin, H. J. Kim, J. I. Hong,  
314 *Org. Lett.*, 2010, **12**, 764. (e) M. Zheng, Z. Xie, D. Qu, D. Li, P. Du, X. Jing, Z. Sun, *ACS*  
315 *Appl. Mater. Interfaces.*, 2013, **5**, 13242. (f) R. R. Lauwerys, A. M. Bernard and H. A.  
316 Reels, *Clin. Chem.*, 1994, **40**, 1391. (g) T. Jin, J. Lu, M. Nordberg, *Neurotoxicology.*,  
317 1998, **19**, 529. (h) Agency for Toxic Substances and Disease Registry, 4770 Buford Hwy  
318 NE, Atlanta, GA 30341. <http://www.atsdr.cdc.gov/cercla/07list.html>.
- 319 (5) (a) R. E. Clement, P. W. Yang, C. Koester, *J. Anal. Chem.*, 1999, **71**, 257.  
320 (b) L. H. Thaller, A. H. Zimmerman, *J. Power Sources.*, 1996, **63**, 53.  
321 (c) R. E. Hamon, M. J. McLaughlin, R. Naidu, R. Correll, *Environ. Sci. Technol.*, 1998, **32**,  
322 3699.
- 323 (d) S. E. Bailey, T. J. Olin, R. M. Bricka, D. D. Adrian, *Water Res.*, 1999, **33**, 2469.
- 324 (6) (a) M. P. Waalkes, *J. Inorg. Biochem.*, 2000, **79**, 241.  
325 (b) M. P. Waalkes, T. P. Coogan, R. A. Barter, *Crit. Rev. Toxicol.*, 1992, **22**, 175.
- 326 (c) W. de Vries, P. F. Römken and G. Schütze, *Rev. Environ. Contam. Toxicol.*, 2007, **191**,  
327 91. (d) H. N. Kim, W. X. Ren, J. S. Kim and J. Yoon, *Chem. Soc. Rev.*, 2012, 3210 and  
328 references therein.
- 329 (7) (a) C. Cloquet, J. Corrigan, G. Libourel, T. Sterckeman, E. Perdrix, *Environ. Sci. Technol.*,  
330 2006, **40**, 2525. (b) S. P. Dolan, D. A. Nortrup, P. M. Bolger, S. G. Capar, S. G. *J. Agric.*  
331 *Food Chem.*, 2003, **51**, 1307.
- 332 (8) (a) N. H. Bings, A. Bogaerts, J. A. C. Broekaert, *Anal. Chem.*, 2002, **74**, 2671. (b) S. M. Pyle,  
333 J. M. Nocerino, S. N. Deming, J. A. Palasota, J. M. Palasota, E. L. Miller, D. C. Hillman,

- 334 C. A. Kuharic, W. H. Cole, P. M. Fitzpatrick, M. A. Watson, K. D. Nichols, K. D.  
335 *Environ. Sci. Technol.*, 1996, **30**, 204.
- 336 (9) (a) Y. Wang, Y. H. Wang, Z. L. Fang, *Anal. Chem.*, 2005, **77**, 5396. (b) Q. Y. Ye, Y. Li, Y.  
337 Jiang, X. P. Yan, *J. Agric. Food Chem.*, 2003, **51**, 2111.
- 338 (10) (a) K. Komatsu, Y. Urano, H. Kojima, T. Nagano, *J. Am. Chem. Soc.* 2007, **129**, 13447. (b) E.  
339 M. Nolan, S. J. Lippard, *J. Am. Chem. Soc.*, 2007, **129**, 5910. (c) S. Yoon, A. E. Albers,  
340 A. P. Wong, C. J. Chang, *J. Am. Chem. Soc.*, 2005, **127**, 16030. (d) J. Y. Kwon, Y. J.  
341 Jang, Y. J. Lee, K. M. Kim, M. S. Seo, W. Nam, J. Yoon, *J. Am. Chem. Soc.*, 2005, **127**,  
342 10107. (e) J. Wang, X. Qian, J. Cui, *J. Org. Chem.*, 2006, **71**, 4308. (f) X. Guo, X. Qian,  
343 L. Jia, *J. Am. Chem. Soc.*, 2004, **126**, 2272. (g) C. Lu, Z. Xu, J. Cui, R. Zhang, X. Qian,  
344 *J. Org. Chem.*, 2007, **72**, 3554. (h) M. Royzen, Z. Dai, J. W. Canary, *J. Am. Chem. Soc.*,  
345 2005, **127**, 1612. (i) W. Jiang, Q. Fu, H. Fan, J. Ho, W. Wang, *Angew. Chem., Int. Ed.*,  
346 2007, **46**, 8445. (j) C. W. Rogers, M. O. Wolf, *Angew. Chem., Int. Ed.*, 2002, **41**, 1898.  
347 (k) A. Coskun, E. U. Akkaya, *J. Am. Chem. Soc.*, 2006, **128**, 14474. (l) J. F. Callan, A. P.  
348 de Silva, D. C. Magri, *D. C. Tetrahedron.*, 2005, **61**, 8551.
- 349 (11) C. Lu, Z. Xu, J. Cui, R. Zhang, X. Qian, *J. Org. Chem.*, 2007, **72**, 3554.
- 350 (12) J. Jia, Q. C. Xu, R. C. Li, X. Tang, Y. F. He, M. Y. Zhang, Y. Zhang, G. W. Xing, G.  
351 *Org. Biomol. Chem.*, 2012, **10**, 6279.
- 352 (13) W. Wang, Q. Wen, Y. Zhang, X. Fei, Y. Li, Q. Yang, X. Xu, *Dalton Trans.*, 2013, **42**,  
353 1827.
- 354 (14) (a) F. A. Cotton and G. Wilkinson, *Advances in Inorganic Chemistry.*, 5<sup>th</sup> ed.; Wiley: New  
355 York., 1988, 957. (b) D. Dakternieks, *Coord. Chem. Rev.*, 1990, **98**, 279.  
356 (c) M. Li, H. Y. Lu, R. L. Liu, J. D. Chen, C. F. Chen, *J. Org. Chem.*, 2012, **77**, 3670.
- 357 (15) A. Bencini, F. Caddeo, C. Caltagirone, A. Garau, M. B. Hurstouse, F. Isaia, S. Lampis, V.  
358 Lippolis, F. Lopez, V. Meli, M. Monduzzi, M. C. Mostallino, S. Murgia, S. Puccioni, J.  
359 Schmidt, P. P. Seccif, Y. Talmong, *Org. Biomol. Chem.*, 2013, **11**, 7751.
- 360 (16) L. Xue, C. Liu, H. Jiang, *Org. Lett.*, 2009, **11**, 1656.
- 361 (17) R. T. Bronson, D. J. Michaelis, R. D. Lamb, G. A. Hussein, P. B. Farnsworth, M. R. Linford, M.  
362 R. M. Izatt, J. S. Bradshaw, P. B. Savage, *Org. Lett.*, 2005, **7**, 1106.
- 363 (18) S. Sarkar, R. Shunmugam, *Appl. Mater. Interfaces.*, 2013, **5**, 7379.
- 364 (19) L. Xue, C. Liu, H. Jiang, *H. Org. Lett.*, 2009, **11**, 1655.

- 365 (20) L. K. Zhang, Q. X. Tong, L. J. Shi, *Dalton Trans.*, 2013, **42**, 8567.
- 366 (21) C. Lu, Z. Xu, J. Cui, R. Zhang, X. Qian, *J. Org. Chem.*, 2007, **72**, 3554.
- 367 (22) X. Peng, J. Du, J. Fan, J. Wang, Y. Wu, J. Zhao, S. Sun, T. J. Xu, *J. Am. Chem. Soc.*,  
368 2007, **129**, 1500.
- 369 (23). T. Cheng, Y. Xu, S. Zhang, W. Zhu, X. Qian, L. Duan, *J. Am. Chem. Soc.*, 2008, **130**,  
370 16160.
- 371 (24). X. Liu, N. Zhang, J. Zhou, T. Chang, C. Fangab, D. Shangguan, *Analyst.*, 2013, **138**, 901.
- 372 (25). P. G. Sutariya, A. Pandya, N. R. Modi, S. K. Menon, *Analyst.*, 2013, **138**, 2244.
- 373 (26). M. Taki, M. Desaki, A. Ojida, S. Iyoshi, T. Hirayama, I. Hamachi, Y. Yamamoto, *J. Am.*  
374 *Chem. Soc.*, 2008, **130**, 12564.
- 375 (27). F. Robert, A. D. Naik, F. Hidara, B. Tinant, R. Robiette, J. Wouters, Y. Garcia, Y. *Eur. J.*  
376 *Org. Chem.*, 2010, **10**, 621.
- 377 (28). R. Herchel, L. Pavelek, Z. Tr'avn'ı'cek, *Dalton Trans.*, 2011, **40**, 11896.
- 378 (29). C. Kaewtong, J. Noiseephum, Y. Uppa, N. Morakot, N. Morakot, B. Wannu, T.  
379 Tuntulani, B. Pulpoka, *New J. Chem.*, 2010, **34**, 1104.
- 380 (30). D. Magde, R. Wong, P. G. Seybold, *Photochemistry and Photobiology.*, 2002, **75**(4), 327.
- 381 (31). (a) C. Tanyu, X. Yufang, Z. Shenyi, Z. Weiping, Q. Xuhong, D. Liping, *J. Am. Chem.*  
382 *Soc.*, 2008, **130**, 16160. (b) J. R. Lakowicz, *Principles of Fluorescence Spectroscopy*,  
383 second edition, Kluwer Academic/Plenum, 1999, 725 pp. (c) M. Fischer and J. Georges,  
384 *Chemical Physical Letters.*, 1996, **260**, 115.
- 385
- 386 (32). (a) J. Hatai, S. Pal, S. Bandyopadhyay, *Tetrahedron Lett.*, 2012, **53**, 4357. (b) C. Alexander,  
387 F. M Mary, A. De Bank. Paul, C. Lorenzo, *Org. Biomol. Chem.*, 2012, **10**, 8753. (c) G. S.  
388 Pinkesh, R. M. Nishith, P. Alok, K. J. Bhoomika, V. J. Kuldeep, K. M. Shobhana, *Analyst.*, 2012,  
389 **137**, 5491. (d) W. Wei, W. Qian, Z. Yue, F. Xiaoliang, L. Yaoxian, Y. Qingbiao Yang, X.  
390 Xiaoyi Xu, *Dalton Trans.*, 2013, **42**, 1827.
- 391 (33). S. Muthaiah Shellaiah, W. Yen-Hsing, L. Hong-Cheu, *Analyst*, 2013, **138**, 2931.
- 392 (34). (a) M. Hiroshi, S. Hiroshi, S. Tokashi, H. Kei, Y. Seikichi, M. Noboru, *J. Phys. Chem.* 1984,**88**,  
393 5868. (b) S. T. Samuel, J. K. Su, T. K. Eric, *J. Am. Chem. Soc.*, 2011, **133**, 2664. (c) Y. P.



394 Sun, H. Y. Jung Hee, S. H. Chang, S. Rachid, S. K. Jong, E. M. Susan, V. Jacques, *J. Org. Chem.*  
395 2008, **73**, 8212.

396 (35). K. B. Jacobson, J. E. Turner, J.E., 1980. *The interaction of cadmium and certain other*  
397 *metal ions with proteins and nucleic acids.* Toxicology **16**, 1.

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399 **Schemes**

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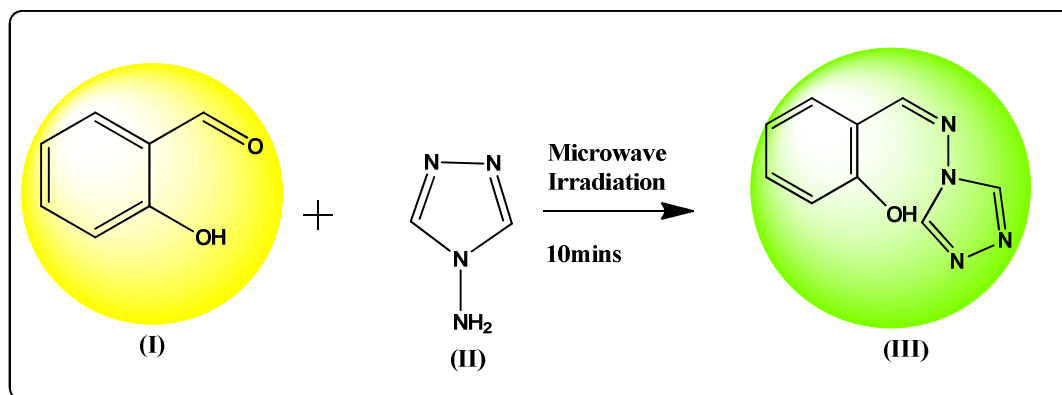
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**Scheme 1** Synthesis of the probe [(Z)-2-(4H-1, 2, 4-triazol-4-yl) imino methylphenol]

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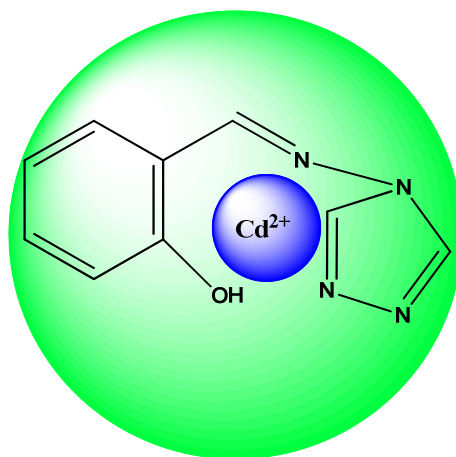
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**Scheme 2** Development of the chemosensor probe [(Z)-2-(4H-1, 2, 4-triazol-4-yl) imino methylphenol] – Cadmium ions (II)

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425 **Table**

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Sl.No	Type of the Chemosensor	Mechanism	Selectivity	Sensitivity	Quantum yield	Reference
1.	OFF - ON	CHEF	Cd <sup>2+</sup>	53nM	---	14
2.	TURN ON	PET	Cd <sup>2+</sup>	0.25 + 0.03 pM	0.2	15
3.	TURN ON	PET	Cd <sup>2+</sup>	1.6nM	0.3	17
4.	TURN ON	ICT	Cd <sup>2+</sup>	0.1 μM	0.6	21
5.	TURN ON	PET	Cd <sup>2+</sup>	0.6μM	0.3	23
6.	TURN ON	PET	Cd <sup>2+</sup>	48 nM	---	24
7.	ON-OFF-ON	PET	Cd <sup>2+</sup> Sr <sup>2+</sup>	0.94pM	---	25
8.	TURN ON	ICT	Cd <sup>2+</sup>	100pM	0.7	26
9.	TURN ON	ICT	Cd <sup>2+</sup>	1fM	0.8	In this work

428 **Table 1** Various types of Cd<sup>2+</sup> ions sensors and their lower limit of detection.

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450 **Figures**

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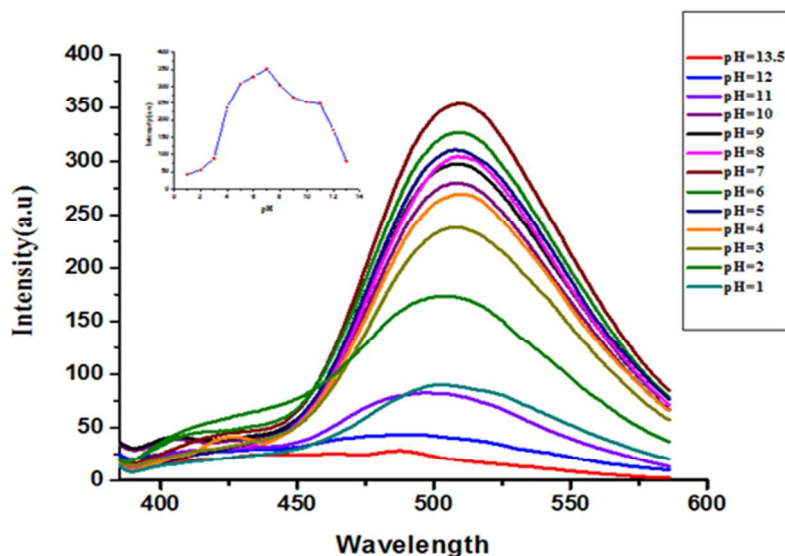
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462 **Fig. 1** Effect of pH on fluorescence Intensity of sensor in buffer (pH=7.0). The concentration of  
463 probe is 5  $\mu$ M, excitation wavelength was 332nm. Emission spectra of probe at pH from 1 to 13.5  
464 [Inset: Fluorescence Intensity at 511 nm versus pH. Excitation and emission slit widths are 5 nm  
465 and 2.5 nm respectively].

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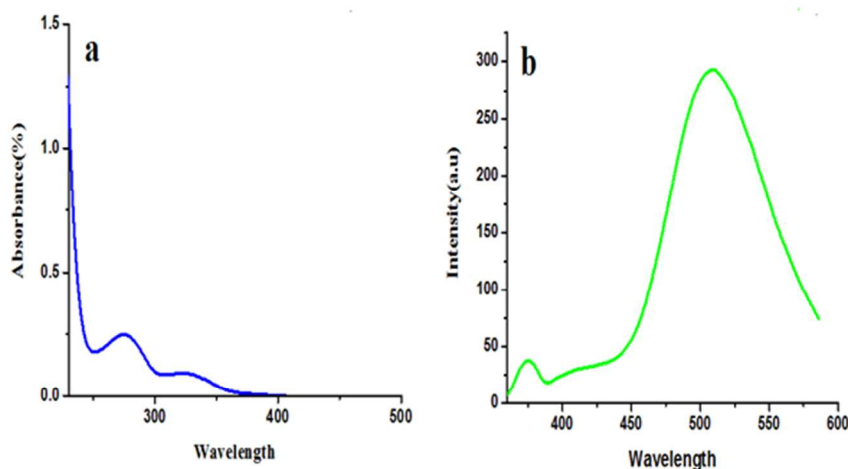
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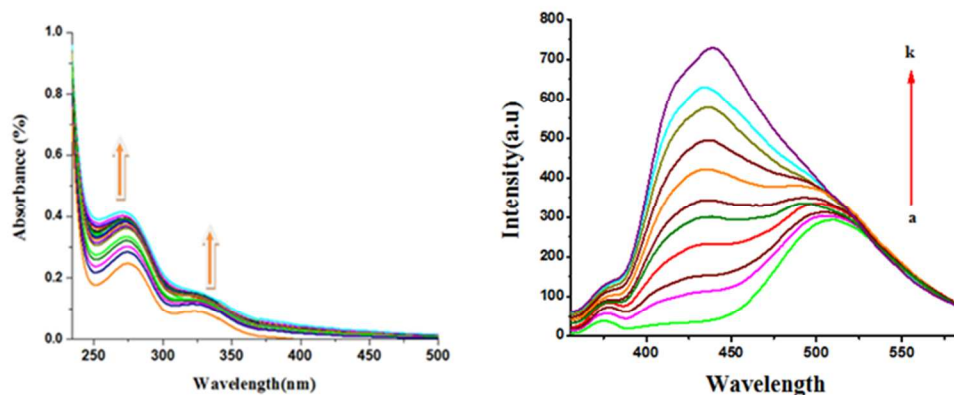
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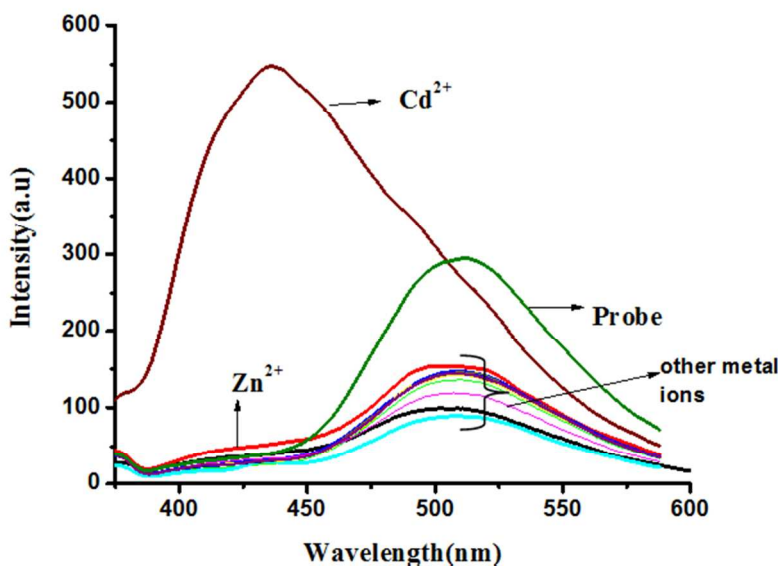


475 **Fig. 2** (a) Absorption spectra and (b) Emission spectra [with excitation wavelength of 332 nm in  
476 phosphate buffer pH=7.0) concentration of probe is 5  $\mu$ M].

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484 **Fig. 3 (a) Absorption spectra of the probe in presence of  $\text{Cd}^{2+}$  ions from 0-20 $\mu\text{M}$  and (b) Emission**  
485 **spectra with excitation wavelength of 332 nm in phosphate buffer (pH=7.0) in the presence of**  
486 **increasing the concentration of  $\text{Cd}^{2+}$  ions from ( $5 \times 10^{-15} \text{M}$  to  $1 \times 10^{-3} \text{M}$ ). (a) probe; (b)  $5 \times 10^{-15}$  (c)  $1 \times 10^{-$**   
487  **$14$ ; (d)  $1 \times 10^{-12}$ : (e)  $1 \times 10^{-9}$ ; (f)  $5 \times 10^{-9}$ ; (g)  $1 \times 10^{-8}$ ; (h)  $1 \times 10^{-6}$ : (i)  $5 \times 10^{-6}$ ; (j)  $1 \times 10^{-5}$ ; (k)  $1 \times 10^{-3}$ .The**  
488 **concentration of the probe is 5  $\mu\text{M}$ . [Excitation and emission slit widths are 5 nm and 2.5 nm**  
489 **respectively Inset: corresponding linearity plot for the selective concentrations].**



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499 **Fig. 4 Selectivity of probe in phosphate buffer (pH = 7.0) in the presence of different metal ions.**  
500 **Excitation wavelength is 332 nm In this emission plot,  $\text{Zn}^{2+}$  ions and other metal ions are quenches**  
501 **the fluorescence intensity of the probe with little change in the emission maximum but**  
502 **enhancement with enormous blue shift is only observed in the presence of  $\text{Cd}^{2+}$  ions. [The total**  
503 **concentration of sensor is 5  $\mu\text{M}$ . Excitation and emission slit widths are 5 and 2.5 nm respectively].**

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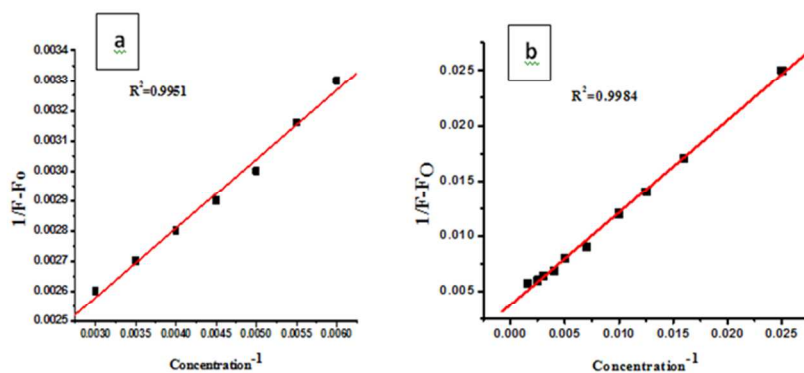
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512 **Fig. 5 (a) Linear response of Sensor as a function of higher concentration of  $\text{Cd}^{2+}$  ions in buffer**  
 513 **solution (pH=7.0). (b) Linear response of Sensor as a function of lower concentration of  $\text{Cd}^{2+}$  ions**  
 514 **(fM) in buffer solution (pH=7.0).**

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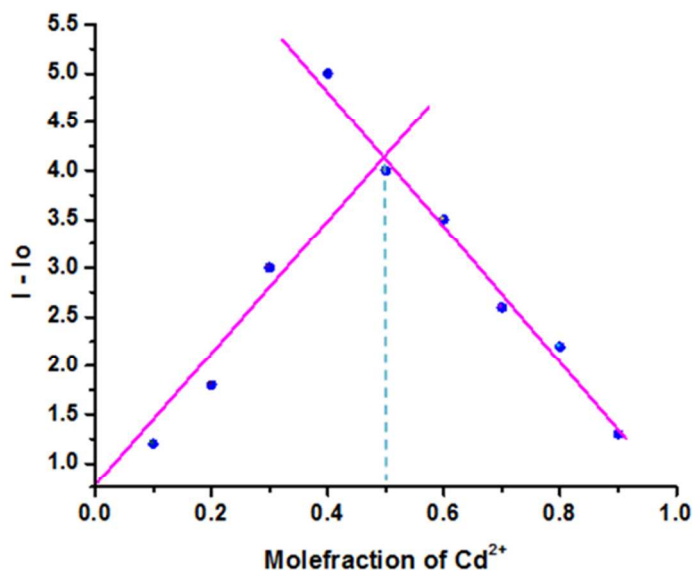
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525 **Fig. 6 Job's Plot of probe (1.0 equiv) in presence of  $\text{Cd}^{2+}$  ions (1.0 equiv). [Confirms the formation of**  
 526 **1:1 complex of Probe- $\text{Cd}^{2+}$  ion]**

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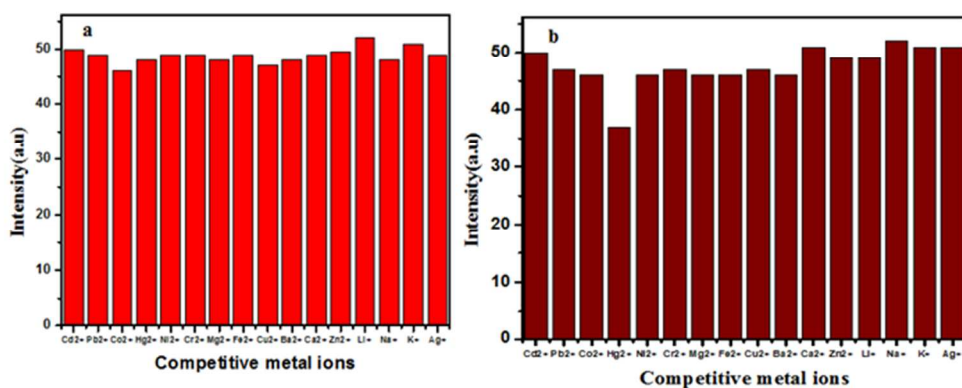
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539 **Fig. 7 (a) The fluorescence intensity of sensor at 511 nm with 1 equiv (10  $\mu$ L) of other metal ions**  
 540 **followed by 1 equiv Cd<sup>2+</sup> ions (10  $\mu$ L) (b) Fluorescence intensity of sensor at 511 nm with 2 equiv (20**  
 541  **$\mu$ L) of other metal ions followed by 1 equiv Cd<sup>2+</sup> ions (10  $\mu$ L). [Concentration of probe is 5  $\mu$ M,**  
 542 **Phosphate buffer, pH 7.5 and Slit widths were 5 nm and 2.5nm respectively. Excitation wavelength**  
 543 **is 332 nm, concentration of Cd<sup>2+</sup> ions and other metal ions are 1X10<sup>-3</sup> M)**

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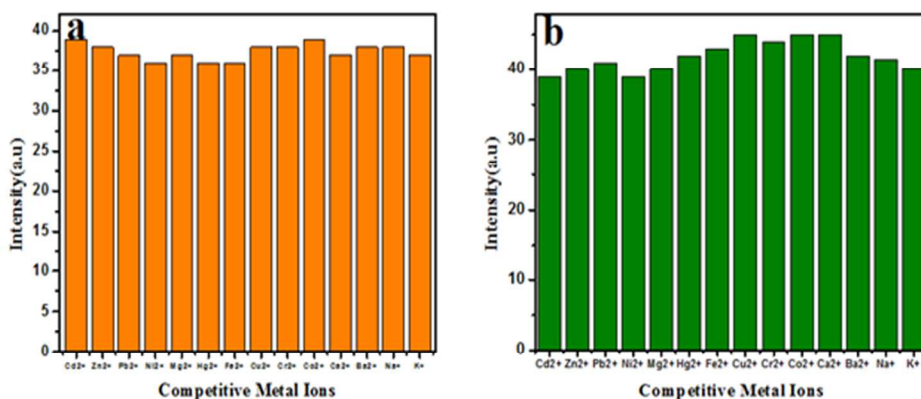
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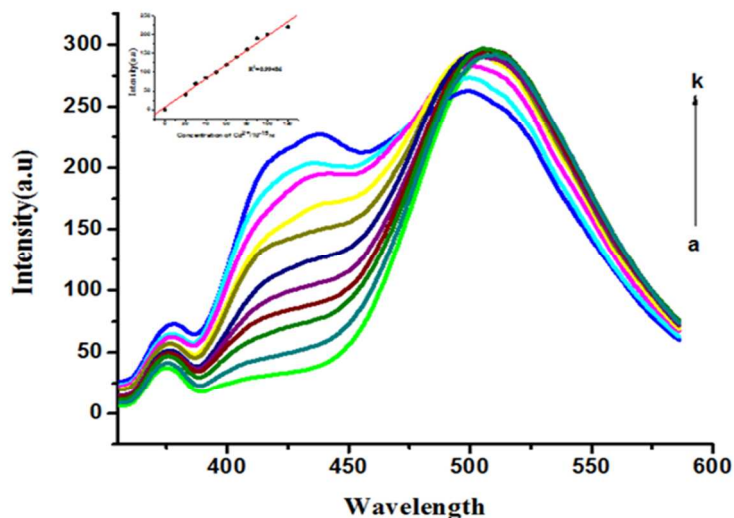
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551 **Fig. 8 (a) Fluorescence intensity of 1ml of probe with 1 ml of Cd<sup>2+</sup> ions in presence of one equivalent**  
 552 **(10  $\mu$ L) of other metal ions. (b) Fluorescence intensity of 1ml of probe with 1 ml of Cd<sup>2+</sup> ions with 2**  
 553 **equivalents (20  $\mu$ L) of other metal ions. [Excitation wavelength 332 nm in phosphate buffer solution**  
 554 **(pH=7.0), excitation and emission slit widths are 5 nm and 2.5 nm respectively, the concentration of**  
 555 **other interfering metal ions are 1x10<sup>-3</sup>M and Cd<sup>2+</sup> ions are 1x10<sup>-15</sup>M]**

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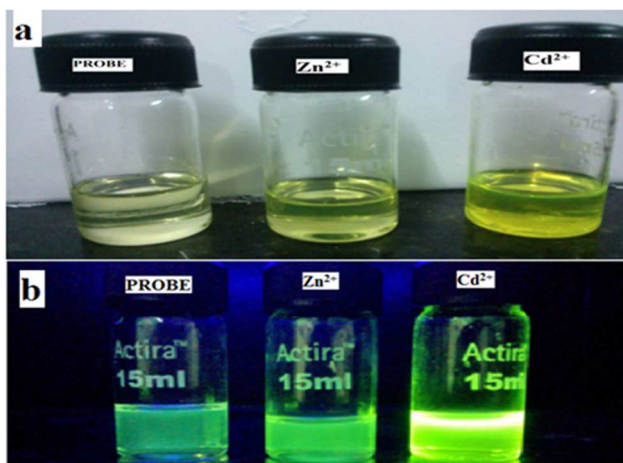
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568 **Fig. 9** Emission spectra of probe in the presence of lower concentrations of  $\text{Cd}^{2+}$  ions. [Inset:  
569 Fluorescence intensity of probe (5  $\mu\text{M}$ ) at 436 nm as a function of concentration of  $\text{Cd}^{2+}$  ions  $1 \times 10^{-15}$   
570 M to  $5 \times 10^{-12}$  M in buffer solution (pH=7.0). Excitation and emission slit widths are 5 nm and 2.5  
571 nm, respectively.] (a) probe; (b)  $1 \times 10^{-15}$  (c)  $2 \times 10^{-15}$ ; (d)  $3 \times 10^{-15}$ ; (e)  $5 \times 10^{-15}$ ; (f)  $1 \times 10^{-14}$ ; (g)  $5 \times 10^{-14}$ ;  
572 (h)  $1 \times 10^{-13}$ ; (i)  $5 \times 10^{-13}$ ; (j)  $1 \times 10^{-12}$  and (k)  $5 \times 10^{-12}$ .

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583 **Fig. 10** (a) Naked eye detection of  $\text{Cd}^{2+}$  over  $\text{Zn}^{2+}$  ions and (b) Response of the probe in presence of  
584  $\text{Cd}^{2+}$  ion under UV-Lamp.

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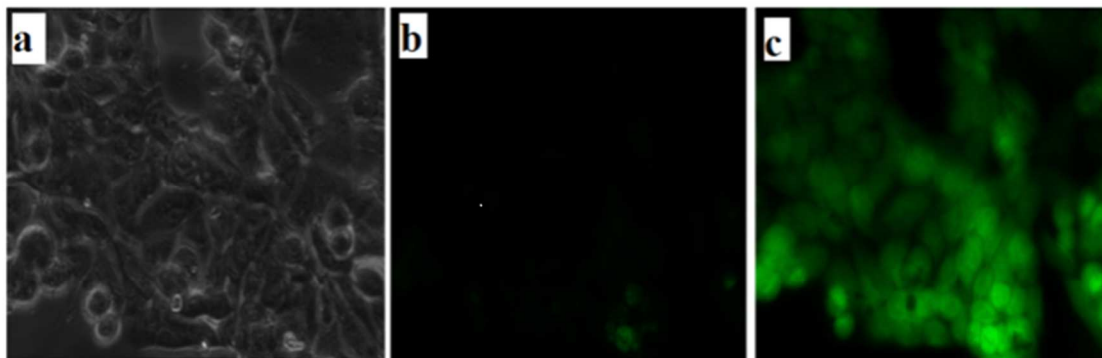
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593 **Fig.11. Fluorescent images of Cd<sup>2+</sup> ions in HeLa cells. (a) Bright-field transmission image of HeLa**  
594 **Cells incubated with probe (5 μM). (b) Fluorescence image of HeLa cells incubated with probe (5**  
595 **μM). (c) Fluorescence image of HeLa cells incubated with probe for 30 min, washed three times,**  
596 **and then further incubated with 5 μM Cd<sup>2+</sup> ions for 30 mins.**

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