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Enhanced reductive transformation of 2,4-dinitroanisole in the anaerobic system: the key role of zero valent iron

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1

2 **Abstract**

3 Accelerated reduction of a typical multi-substituted nitroaromatic compounds
4 (NACs), i.e., 2,4-dinitroanisole (DNAN), was achieved in an anaerobic system
5 coupled with zero valent iron (ZVI), with the underlying role of ZVI in this process
6 elucidated. Both removal of DNAN and formation of its final reductive product
7 2,4-diaminoanisole (DAAN) were notably improved in the ZVI coupled biosystem. In
8 the ZVI coupled biosystem and biotic control system, complete removal of DNAN
9 could be achieved within 4 h and 20 h, respectively. However, only 28.71 ± 5.06 % of
10 DNAN could be removed in the ZVI control system after 20 h. Correspondingly, the
11 formation efficiencies of DAAN in ZVI coupled biosystem, biotic control system and
12 ZVI control system were 99.66 ± 0.70 %, 16.99 ± 1.73 % and 0.00 ± 0.00 %,
13 respectively. The increased DNAN removal and DAAN formation in the ZVI coupled
14 biosystem was linked to the high accumulation of formate, low oxidation-reduction
15 potential (ORP) and great pH self-buffering capability, which was provided by the
16 addition of ZVI. Compared with the biotic control system, the production of CH₄ was
17 significantly accelerated in the ZVI coupled biosystem, indicating that a favorable
18 environmental for methanogens was created at the presence of ZVI. Specially, the
19 ZVI coupled biosystem displayed a more stable performance in terms of DNAN
20 reduction with the coexistence of the competitive electron acceptors, such as nitrate
21 and sulfate. Therefore, the ZVI coupled biosystem could be a promising alternative to
22 the conventional anaerobic reduction process for the removal of multi-substituted
23 NACs from wastewater.

24

25

26 **Keywords:** Anaerobic reduction; Zero valent iron; 2,4-Dinitroanisole; Electron
27 acceptor

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2 **1. Introduction**

3 2,4-Dinitroanisole (DNAN) is an important ingredient in the production of dyes and
4 insecticides.¹ Recently, DNAN is being considered as a replacement for the sensitive
5 explosives such as 2,4,6-trinitrotoluene (TNT), because of its insensitive properties.²
6 Compared to the TNT, the detonation temperature of DNAN is higher, which is
7 beneficial for manufacture, transport and store process.³ However, considering its
8 potential environmental risk, high toxicity, poor biodegradability and wide usage,
9 improper disposal of DNAN containing waste can lead to tremendous environmental
10 pollution, including water contamination and soil problem. Due to the pronounced
11 electron-withdrawing character of the nitro groups on the benzene ring, nitroaromatic
12 compounds (NACs) harbors a highly electron deficient π -electron system, resulting
13 into the difficulty in chemical oxidation or biological oxidation.⁴ Moreover, with the
14 increase of the nitro group number, mineralization of multi-substituted NACs such as
15 DNAN through oxidative pathways becomes more resistant.⁵ Thus, there are
16 significant needs of appropriate methods for the remediation of the sites contaminated
17 by DNAN.

18 Under anaerobic or anoxic conditions, NACs succumb to electrophilic attack and
19 can be transformed to their corresponding aromatic amines but without cleaving the
20 aromatic ring. Generally, the produced aromatic amines are less toxicity and easier to
21 mineralize than their parent compounds.⁶ Nevertheless, due to the highly recalcitrant
22 and toxicological nature of NACs, the anaerobic reduction is usually limited by low
23 degradation rate and poor stability. Therefore, it is important to improve anaerobic
24 reduction performance to achieve more effective reduction of NACs such as DNAN.

25 Zero valent iron (ZVI) is currently attracting wide interest in the treatment of
26 wastewater and groundwater due to its inexpensive, reliable and moderately strong
27 reduction properties. Some refractory contaminants at oxidative state, such as NACs,
28 azo dyes and halogenated organic compounds, could be effectively reduced in the ZVI
29 process.⁷⁻⁹ For the treatment of the wastewater containing these refractory
30 contaminants, ZVI reduction process is often used prior to the biological process for

1 reducing toxicity and improving biodegradability.^{4,10} Ahn et al.¹¹ reported that the iron
2 pretreatment not only removed energetic compounds but also eliminated the toxic
3 effect on perchlorate reducing bacteria. Oh et al.¹² also showed that the ZVI
4 pretreatment transformed recalcitrant hexahydro-1,3,5-trinitro-1,3,5-triazine (RDX) to
5 ring-opening products, i.e., formaldehyde, which are more amenable to mineralization
6 by aerobic bacteria. Therefore, the combined ZVI and biological process offers bright
7 prospects for the treatment of highly recalcitrant industrial wastewater.¹³ In general,
8 ZVI process and biological process was often operated in sequence, however,
9 coupling of ZVI into the biological process could facilitate the degradation of
10 pollutants in a single reactor, which might take full advantage of both ZVI and
11 biological process.¹⁴ Considering its reductive property, ZVI is expected to be helpful
12 for creating an enhanced anaerobic environment which may be beneficial to improve
13 the performance of an anaerobic reactor in wastewater treatment.¹⁵ Meanwhile, ZVI
14 corrosion products, especially alkaline byproducts $\text{Fe}(\text{OH})_2$ or $\text{Fe}(\text{OH})_3$, can not only
15 act as the acid buffers, but also provide another alternative for the contaminant
16 removal through flocculation, adsorption and precipitation.¹⁶

17 Given these, attentions have been increasingly paid to the combined use of ZVI and
18 microbe for enhanced degradation of recalcitrant contaminants from wastewater.^{8,17}
19 Liu et al.¹⁸ reported that both azo dye decolorization and COD removal were
20 remarkably improved in an acidogenic reactor packed with ZVI. At the presence of
21 ZVI, the abundance of methanogens was significantly increased and microbial strains
22 responsible for azo dye decolorization were enriched in the anaerobic reactor.¹⁵ Of
23 even greater importance, the release of H_2 during ZVI corrosion became an alternative
24 electron donor for hydrogen-consuming microorganisms, such as methanogenic and
25 denitrifying bacteria, as well as some reduction related species.¹⁹ Even though these
26 physicochemical and microbial interactions are highly important for the overall
27 performance of the coupled system, systematic investigation on the ZVI coupled
28 anaerobic reduction system is still limited. In addition, coupling of ZVI into an
29 anaerobic biological system for the treatment of multi-substituted NACs containing
30 wastewater has been rarely investigated, and the underlying role of ZVI in the coupled

1 system treating NACs containing wastewater is not fully understood.

2 Therefore, in this study, coupling of ZVI into the anaerobic system was established
3 with the goal of accelerating the DNAN removal from wastewater. Specially, the key
4 role of ZVI in the coupled system was investigated in terms of the intermediate
5 products, ORP, pH and biogas analysis. The performance of the ZVI coupled
6 biosystem at the presence of competitive electron acceptors, such as nitrate and
7 sulfate, was also evaluated.

8

9 **2. Materials and Methods**

10 *2.1 Chemicals*

11 DNAN was a gift from Hubei Dongfang Chemical Co. Ltd in Hubei province,
12 China. 2-Nitro-4-aminoanisole (2-N-4-AAN) and 2-amino-4-nitroanisole
13 (2-A-4-NAN) were purchased from Bepfarm Co. Ltd (Shanghai, China). DAAN was
14 purchased from Sun Chemical Technology Co. Ltd (Shanghai, China). ZVI powder
15 with analytical purity was purchased from Sinopharm Chemical Reagent Co. Ltd
16 (Shanghai, China) and was used without pretreatment.

17

18 *2.2 Synthetic wastewater and sludge cultivation*

19 The composition of the synthetic wastewater used in this study was as follows:
20 DNAN (50 mg/L), methanol (0.84 mL/L), KH_2PO_4 (25 mg/L), NH_4Cl (100 mg/L),
21 $\text{MgSO}_4 \cdot 7\text{H}_2\text{O}$ (200 mg/L), CaCl_2 (30 mg/L), and trace element solution (SL-4, 10
22 mL/L). In order to give sufficient electron donor for DNAN reduction, methanol was
23 added excessively. The composition of SL-4 was as described previously by Shen et
24 al.²⁰

25 Anaerobic sludge taken from an anaerobic baffled reactor treating real NACs
26 containing wastewater was used as the seed sludge. Before inoculation, the seed
27 sludge was acclimated for about three months using the synthetic wastewater as the
28 influent. Once stable reduction performance of the acclimation system was achieved,
29 the acclimatized sludge could be used as the inoculum.

30

1 *2.3 Experimental procedure*

2 In this study, DNAN reduction was performed in batch mode, which was carried
3 out in a series of 100 mL serum bottles. 80 mL synthetic wastewater containing 50
4 mg/L DNAN was added into each serum bottle. To remove any residual dissolved
5 oxygen, the synthetic wastewater was purged with nitrogen for at least 20 min,
6 followed by the addition of the 0.1 g ZVI and 20 mL seed sludge prepurged with
7 nitrogen. The initial DNAN and MLSS concentrations in each serum bottle were
8 calculated to be 40 mg/L and 13 g/L, respectively. Then, the serum bottles were
9 sealed with polytetrafluoroethylene/silica plugs and aluminum crimp seals. All serum
10 bottles were incubated on a rotary shaker at 200 rpm and 30 °C. At every
11 predetermined sampling time, one serum bottle was sacrificed, and 10 mL of the
12 solution was filtered through a 0.22 µm membrane for analysis. The control systems,
13 i.e., the ZVI control system with the addition of 0.1 g ZVI but without sludge, and the
14 biotic control system with the addition of 20 mL anaerobic sludge but without ZVI,
15 were operated according to the same experimental procedures as the coupled system.

16 To evaluate the competitive effect of other electron acceptors on the microbial
17 transformation of DNAN, two common competing electron acceptors, i.e., nitrate and
18 sulfate, were added respectively to the batch anaerobic reactors at the concentration of
19 500 mg/L. DNAN reduction performance at the presence of nitrate and sulfate was
20 evaluated.

21 All experimental runs were performed in triplicate and the results were reported as
22 an average of the three independent determinations.

23

24 *2.4 Analytical methods*

25 DNAN and its intermediate products were identified and quantified by high
26 performance liquid chromatography (HPLC) (Waters 2996, Waters Incorporation,
27 USA). The HPLC analysis was conducted at room temperature using a RP18 column
28 (5 mm, 4.6×250 mm) and a UV-vis detector. The mobile phase was a mixture of 45%
29 methanol and 55% water pumped at a flow rate of 1.00 mL min⁻¹. The analysis was
30 performed at 254 nm with a column temperature of 35°C. The analysis of formate ion

1 was performed on an ion chromatograph (ICS-2100, DIONEX, USA) using an Ion
2 Pac[®] As11-HC (4×250 mm) column and a suppressed conductivity detector. The pH
3 and oxidative-reductive potential (ORP) were measured by a pH meter (FE20K,
4 Mettler-Toledo instruments, CH) with a redox electrode. At given time intervals, the
5 volume of biogas produced was measured using a syringe after the gas pressure in the
6 headspace was brought to atmospheric pressure. The composition of biogas was
7 analyzed by gas chromatography (Agilent 6820, Agilent Technologies, USA)
8 equipped with a thermal conductivity detector (TCD) using molecular sieve 5A-60/80
9 mesh column (ANPEL Laboratory Technologies Inc., Shanghai, China) as a
10 separation column. N₂ was the carrier gas, and the operating temperature of the
11 injection port, oven, and detector was 150 °C, 60 °C and 200 °C, respectively. Iron
12 concentration in the reactors was determined by Inductively Coupled Plasma
13 (Optima7000, PerkinElmer instruments, USA). Scanning electron microscopy
14 coupled with energy dispersive spectroscopy (SEM-EDS) (Quanta 250FEG, FEI, USA)
15 was applied to characterize the morphology and chemical composition of anaerobic
16 sludge after ZVI treatment.

17

18 2.5 Data analysis

19 DNAN reduction rates in the reduction systems were described by the pseudo
20 zero-order kinetic model (Eq. (1)) and pseudo first-order kinetic model (Eq. (2)),
21 respectively.

$$22 \quad C_0 - C_t = k_0 t \quad (1)$$

$$23 \quad \ln(C_0/C_t) = k_1 t \quad (2)$$

24 where C_0 is the initial DNAN concentration (mg/L), C_t is the DNAN concentration
25 (mg/L) at reaction time t (h), k_0 is the pseudo zero-order rate constant (mg/h), and k_1 is
26 the pseudo first-order rate constant (h⁻¹).

27

28 3. Results and discussion

29 3.1 Performance of DNAN reduction in the ZVI coupled biosystem

30 To verify whether the anaerobic reduction of DNAN could be enhanced by ZVI,

1 removal of DNAN and formation of its reduction intermediates in ZVI coupled
2 biosystem, ZVI control system and biotic control system, were compared. As shown
3 in Fig. 1a, only 4 h was required for complete DNAN reduction in the ZVI coupled
4 biosystem, while as long as 20 h was required for complete DNAN reduction in the
5 biotic control system. The difference in terms of DNAN removal was significant,
6 probably due to the key role of ZVI in DNAN reduction. However, only $28.71 \pm$
7 5.06 % of the total DNAN could be removed in the ZVI control system after 20 h,
8 indicating that the ZVI alone could not serve as an efficient and sufficient electron
9 donor for abiotic reduction of DNAN, especially at the neutral pH condition adopted
10 in this study. Therefore, it could be inferred that there existed some synergistic effects
11 between ZVI and anaerobic sludge for DNAN reduction.

12 Under anaerobic condition, DNAN could be reductively transformed into DAAN
13 with 2-amino-4-nitroanisole (2-A-4-NAN) and 2-nitro-4-aminoanisole (2-N-4-AAN)
14 as the intermediates,² which was confirmed by HPLC analysis (Fig. S1). In the ZVI
15 coupled biosystem and biotic control system, the maximum accumulation
16 concentrations of 2-A-4-NAN and 2-N-4-AAN were 9.65 ± 1.09 mg/L and $2.40 \pm$
17 0.15 mg/L, 23.11 ± 0.77 mg/L and 3.88 ± 0.22 mg/L, respectively, indicating the
18 relatively low accumulation of reduction intermediates in the ZVI coupled biosystem
19 (Fig. 1b and 1c). However, only 3.65 ± 0.62 mg/L 2-A-4-NAN was detected in the
20 ZVI control system (Fig. 1b). Moreover, the maximum concentration of final product
21 DAAN in the ZVI coupled biosystem was as high as 27.79 ± 0.69 mg/L, which was
22 much higher than 4.74 ± 0.48 mg/L in the biotic control system and 0.00 ± 0.00 mg/L
23 in the ZVI control system (Fig. 1d). Correspondingly, the formation efficiencies of
24 DAAN in ZVI coupled biosystem, biotic control system and ZVI control system were
25 99.66 ± 0.70 %, 16.99 ± 1.73 % and 0.00 ± 0.00 %, respectively. It could be seen that
26 DNAN was only partially reduced in the biotic control system, with more 2-A-4-NAN
27 accumulated but less DAAN produced. These results further indicated that coupling
28 of ZVI into the anaerobic system could accelerate the degradation of DNAN,
29 particularly the formation of its final reductive product DAAN. In addition,
30 accounting for these intermediate species and end products gave good mass balance

1 (greater than 85%) for the three individual batch systems, suggesting that other
2 reaction products were negligible.

3

4 *3.2 Reduction pathway of DNAN in the ZVI coupled biosystem*

5 Based on the evolution of intermediate products, it is interesting to note that the
6 reduction of -NO₂ on DNAN was preferential at *ortho* position in the ZVI coupled
7 biosystem, resulting in the formation of 2-A-4-NAN, which could be subsequently
8 reduced to DAAN. This phenomenon was in accordance with the result in the
9 previous study. Olivares et al.²¹ also found that the reduction of the nitro groups on
10 DNAN under anaerobic condition followed the order of *ortho*-position >
11 *para*-position. According to the density function theory computations analysis, the
12 charge densities of the N atoms at *para*-NO₂ and *ortho*-NO₂ positions of DNAN were
13 0.211 and 0.215, following the order of *ortho*-position > *para*-position. Electron
14 attacks, which were nucleophilic, would occur preferentially at the N atom with more
15 positive charge density. Therefore, the reduction of DNAN was selectively favored at
16 the *ortho*-NO₂ group.

17

18 *3.3 Kinetics of DNAN reduction in the coupled ZVI anaerobic system*

19 Pseudo first-order and pseudo zero-order kinetic models were employed to
20 elucidate the DNAN reductive transformation process in the three individual systems
21 (i.e., ZVI coupled biosystem, biotic control system and ZVI control system). The rate
22 constant and regression coefficient (R^2) relevant to the zero and first kinetic models
23 were shown in Table 1. It was noteworthy that the removal of the DNAN in either
24 ZVI coupled biosystem or biotic control system could be appropriately simulated by
25 the pseudo first-order kinetic model, while the removal of DNAN in ZVI control
26 system followed the pseudo zero-order kinetic model. This result suggested that there
27 existed different mechanisms between biotic system and abiotic system for DNAN
28 removal.

29 For the solid-liquid heterogeneous reaction system, such as the ZVI reduction
30 system or the ZVI coupled biosystem, if the adsorption of contaminants onto the solid

1 surface played a minor role in the reductive process, the contaminant removal often
2 followed the pseudo zero-order kinetic model, otherwise pseudo first-order kinetic
3 model was more appropriate for the removal kinetics.^{22,23} Since no removal of DNAN
4 was observed at the initial stage in the ZVI control system (Fig. 1a), the removal of
5 DNAN in ZVI control system could be attributed to the reduction by ZVI rather than
6 adsorption by ZVI. As a result, the pseudo zero-order kinetics model could be applied
7 to the DNAN removal process in the ZVI control system.^{23,24} However, a sharp
8 decrease of DNAN concentration was observed in either ZVI coupled biosystem or
9 biotic control system within the first hour (Fig. 1a), probably due to the strong
10 adsorption of DNAN by the sludge inoculated in these two systems, which was
11 confirmed by the good match between DNAN removal and first-order kinetic in either
12 ZVI coupled biosystem or biotic control system.²⁵

13 As was indicated in Table 1, the pseudo first-order rate constant for DNAN removal
14 in the ZVI coupled biosystem was as high as 1.263 h^{-1} , which was much higher than
15 0.217 h^{-1} in the biotic control system. This result strongly confirmed that the removal
16 of DNAN in anaerobic system could be largely promoted by the addition of ZVI,
17 probably due to the synergistic interaction between anaerobic microbes and ZVI.
18 However, the surface adsorption by ZVI was negligible, since the ZVI used in this
19 study had few surface sites amenable for DNAN adsorption, as was indicated by the
20 slight removal of DNAN at the early stage in the ZVI control system.

21

22 *3.4 The key role of ZVI in the coupled system*

23 As indicated in previous study, pH is one of the most important parameters
24 affecting the community structure and activity of anaerobic microorganisms.¹⁴ After
25 the 20h reaction, the pH in the biotic control system and ZVI coupled biosystem
26 shifted from 7.12 ± 0.01 to 6.35 ± 0.04 and 6.67 ± 0.02 (Table 2), respectively,
27 suggesting that the ZVI coupled biosystem seemed to have a greater pH self-buffering
28 capability than the biotic control system. This phenomenon could be linked to the ZVI
29 corrosion in the anaerobic system, which consumed the acidity produced from the
30 anaerobic acidification process.

1 Furthermore, ZVI corrosion process could create a more stable and favorable
2 anaerobic environment for microorganisms by lowering the ORP.¹⁵ As shown in Fig.
3 2, the ORP in the ZVI coupled biosystem approximately ranged from -128.5 ± 12.0
4 mV to -265.0 ± 19.8 mV, while it ranged from -122.5 ± 3.5 mV to -215.5 ± 20.5 mV
5 in the biotic control system. Lower ORP value means a better reductive environment,
6 which could exert a positive effect on the reduction of nitro group.^{26,27} Additionally, a
7 sharp decrease of ORP was observed in ZVI coupled biosystems within the first 4 h,
8 implying that there was a substantial depletion of the oxidative compounds in aqueous
9 solution. Such a phenomenon was well in agreement with the DNAN reduction in the
10 ZVI coupled biosystem, confirming the effective reduction of DNAN in the ZVI
11 coupled system.

12 To further clarify the effect of ZVI on methanol metabolism, the production of the
13 methanol metabolism product, i.e., formate, was investigated. As shown in Fig. 3, the
14 production of formate in ZVI coupled biosystem was significantly higher than that in
15 the biotic control system. A previous work showed that both acidogenesis and activity
16 of fermentative bacteria could be effectively improved by lowering ORP, which was
17 provided by the addition of ZVI.²⁸ On the other hand, the ferrous ions from ZVI
18 corrosion could stimulate the synthesis of key enzymes in the hydrolysis-acidification
19 process, resulting in the accumulation of volatile fatty acids.²⁹ Considering that
20 formate was an effective electron donor for the reduction process,³⁰ the increased
21 production of formate in the ZVI coupled biosystem could be beneficial for the
22 efficient reduction of DNAN.

23 Generally, methanol as well as the ZVI in anaerobic system may serve as
24 precursors for the formation of an intermediate H_2 pool, which could be utilized as the
25 electron donor for the reduction process.^{19,31} However, no hydrogen was produced in
26 either biotic control system or ZVI coupled biosystem (Fig. 4). This might be
27 attributed to the slow corrosion rate of ZVI and high consumption rate of methanol in
28 the anaerobic system. Under anaerobic conditions, the accumulated VFAs could be
29 further bioconverted to methane. It was observed that the cumulative CH_4 in the ZVI
30 coupled biosystem was about 0.638 ± 0.017 mmol/mmol methanol, while, it was only

1 0.017 ± 0.002 mmol/mmol methanol in the biotic control system, indicating that the
2 methanogenic activity of the sludge could be effectively improved by addition of ZVI.
3 Previous studies have shown that an appropriate amount of ferrous ions released from
4 ZVI corrosion could be involved in energy metabolism as a cytochrome and
5 ferredoxin in methylotrophic methanogens.^{32,33} Meanwhile, the CO₂ produced in ZVI
6 coupled biosystem could be further converted to methane through methanogenesis
7 using ZVI as the direct electron donor, which was necessarily beneficial for the
8 increase of methane production.³⁴ Furthermore, the rapid reduction of DNAN in the
9 ZVI coupled biosystem alleviated the inhibitive effect of DNAN on methanogens,
10 since DNAN was much more toxic to methanogenic microorganisms than its
11 reduction products.^{6,35}

12 Additionally, the electronegative anaerobic bacteria could be easily attached on the
13 surfaces of ZVI due to the static function in a mixed anaerobic culture, and a stable
14 ZVI-microbial zoogloea could be gradually formed, which was beneficial for NACs
15 reduction.³⁶ The SEM-EDS analysis confirmed the presence of Fe element on the
16 outer layer and inner parts of anaerobic granules, indicating that ZVI could be a ideal
17 site for the formation of ZVI-microbial zoogloea complex (Fig. S2). Moreover, under
18 anaerobic condition, the ZVI surface area might be increased by etching and pitting
19 through corrosion, which was further beneficial for mass transfer and reductive
20 transformation of pollutants on it.³⁶

21

22 *3.5 DNAN reduction at the presence of competitive electron acceptors*

23 Previous studies suggested that the competitive electron acceptors in wastewater,
24 such as nitrate and sulfate, have profound impact on the anaerobic reduction of
25 pollutants, such as nitrobenzene, pentachloroaniline and 4-chloronitrobenzene, et
26 al.^{26,31,37} Therefore, it was essential to investigate the DNAN reduction at the presence
27 of these competitive electron acceptors.

28

29 *3.5.1 Effect of nitrate on DNAN reduction*

30 As shown in Fig. 5, sharp decrease in terms of DNAN removal was observed when

1 500 mg/L nitrate was introduced to the biotic control system. Only 55.79 ± 1.54 % of
2 the total DNAN could be removed after 20 h at the presence of 500 mg/L nitrate,
3 while complete DNAN removal could be observed after incubation time of 20 h at the
4 absence of nitrate. Correspondingly, the constant rate (k_r) of DNAN reduction
5 decreased from 0.217 h^{-1} to 0.024 h^{-1} due to the introduction of nitrate in the biotic
6 control system. However, after 500 mg/L nitrate was introduced, the incubation time
7 required for the complete DNAN removal in the ZVI coupled biosystem slightly
8 increased from 4 h to 8 h, indicating the low competitiveness of nitrate for the
9 electron donor at the presence of ZVI.

10 In the biotic control system, the inhibitory effect of nitrate on DNAN removal
11 could be expected considering the much higher oxidation potential of nitrate
12 compared to DNAN. The standard electrode potentials for the reduction of NO_3^- to
13 NO_2^- at neutral pH were reported to be 0.43 V vs standard hydrogen electrode (SHE),
14 while the one-electron reduction of DNAN was as low as -0.40 V.³⁸⁻⁴⁰ As a result,
15 nitrate reduction had an advantage over DNAN reduction in the competition for the
16 limited electron donor. The ORP increase in the anaerobic system after the addition of
17 nitrate was another reason for the decreased DNAN removal (Fig. S3a). However, at
18 the presence of ZVI, the situation was different. The standard electrode potential of
19 $\text{NO}_3^-/\text{NO}_2^-$, i.e., 0.43 V, was higher than that of Fe^{2+}/Fe , i.e., -0.44 V. Therefore, the
20 competitive electron acceptor, i.e., nitrate, could be theoretically reduced by ZVI in
21 ZVI coupled biosystem.⁴¹ As was reported in previous study, at the presence of nitrate
22 or nitrite, corrosion of iron might be alleviated, especially under neutral or alkaline
23 condition.^{42,43} However, in this ZVI coupled biosystem, slightly acidic condition was
24 well maintained, probably due to the acidification reaction in this anaerobic system.
25 Therefore, corrosion of ZVI would make an important contribution for both nitrate
26 reduction and DNAN reduction. In addition, ZVI surface area could be increased by
27 etching and pitting through anaerobic corrosion, which was further beneficial for mass
28 transfer and reductive reduction on it.³⁶ More importantly, at near-neutral pH
29 condition, nitrate as a less strong oxidant could oxidize ZVI to form the magnetite,⁴⁴
30 overcoming the obstacle from the electron transfer barrier over the corrosion coating,

1 which might be beneficial for NACs reduction.⁴⁵ Therefore, the ZVI coupled
2 biosystem showed excellent performance in terms of DNAN reduction at the presence
3 of nitrate.

4

5 3.5.2 Effect of sulfate on DNAN reduction

6 It was interesting to observe that DNAN removal was significantly accelerated in
7 the biotic control system at the presence of 500 mg/L sulfate (Fig. 6), with the pseudo
8 first-order rate constant (k_1) for DNAN removal increased obviously from 0.217 h⁻¹ to
9 0.432 h⁻¹. In addition, slight increase in terms of DNAN removal was also observed in
10 the ZVI coupled biosystem after the introduction of 500 mg/L sulfate. These results
11 indicated that the presence of sulfate had no adverse effect on DNAN removal. On the
12 contrary, DNAN removal could be enhanced to some extent at the presence of sulfate,
13 which was rather different from the results at the presence of nitrate.

14 Since the inhibitory effect of sulfate on NACs degradation had been well
15 recognized,²⁶ the phenomenon observed in this study was rather interesting. The
16 reason could be ascribed to the generation of some reducing agents, e.g., sulphide,
17 which could be used as the electron donor for DNAN. Similar result was also reported
18 by van der Zee et al.,⁴⁶ where the reduction of azo dye could be significantly
19 enhanced at the presence of sulfate, especially under the anoxic condition. The
20 standard electrode potential of SO₄²⁻/HSO₃⁻ was -0.52 V vs SHE at neutral pH,³⁸
21 which was lower than the one-electron standard reduction potential of DNAN.
22 Compared with sulfate, DNAN was more subjected to reduction in this study, as
23 DNAN showed higher competitiveness for the electron. Besides, another compelling
24 evidence hypothesized by Ismail and Pavlostathis,³¹ was that the growth of sulfate
25 reducers was relatively slow when methanol was used as the energy source. Therefore,
26 the electron donors used for the reduction of sulfate could be limited, which was
27 further beneficial for DNAN reduction. Meanwhile, the poisonous effect of sulphide
28 on microorganisms might be ignored under near-neutral pH condition.⁴⁷ More
29 importantly, the iron oxides and hydroxides on the surface of ZVI could be eliminated
30 by sulphide, with the formation of mackinawite or pyrite.^{48,49} Compared with the iron

1 oxides and hydroxides, the mackinawite or pyrite was a better promotor of electron
2 transfer to organic pollutants.⁵⁰ Unfortunately, the quantification of sulphide and
3 elemental sulfur during DNAN reduction has been unsuccessful in this study. This
4 might be due to the low concentrations of the sulphide and elemental sulfur in the
5 anaerobic system, which needs further investigation.

6 Additionally, the ORP in either biotic control system or ZVI coupled biosystem was
7 significantly decreased at the presence of sulfate, indicating that a more reductive
8 condition was created for DNAN reduction (Fig. S3b). In terms of the fore-mentioned
9 discussion, it could be concluded that the DNAN removal was accelerated by addition
10 of sulfate in either ZVI coupled biosystem or biotic control system.

11

12 *3.6 Implication of this work*

13 Compared to the biotic control system and ZVI control system, both DNAN
14 reduction and DAAN formation were significantly improved in the ZVI coupled
15 biosystem. The efficient reduction of DNAN to DAAN in the ZVI coupled biosystem
16 would result in a significant improvement of biodegradability and reduction of
17 toxicity.⁵¹ More importantly, favorable environment for some specific microbial
18 species, such as methanogens, could be created by offsetting the possible pH decline
19 and lowering the ORP.

20 What's more, the Fe^{2+} in the effluent of the ZVI coupled biosystem was generally
21 below 1 mg/L, suggesting the slow rate of ZVI dissolution. The low consumption of
22 iron leads to easy maintenance and low operating cost. In addition, the low
23 concentration of ferrous iron was beneficial for the growth of microorganisms.^{32,33}
24 Moreover, under anaerobic condition, ZVI could be protected from oxygen, with the
25 reduced formation of iron oxides on the surface.¹⁵ Thus the frequent replacement and
26 regeneration of ZVI was not required in the ZVI coupled biosystem. What's more
27 important, under anaerobic condition, ZVI surface area could be increased by etching
28 and pitting through corrosion, which was further beneficial for mass transfer and
29 reductive reduction on it.³⁶

30 Recently, for the removal of various contaminants, the application of ZVI powder,

1 especially the nano zero valent iron (NZVI), has received increasing attention due to
2 their high surface area and high reactivity. However, coupling of ZVI powder into an
3 anaerobic system for the treatment of raw industrial wastewater was limited by far,
4 due to the inherent weakness of ZVI powder and NZVI, such as poor stability and
5 easy aggregation. To address these issues, iron shavings may be a better choice,
6 compared with ZVI powder and NZVI. The primary reason for this selection was the
7 abundant local supply, relatively low cost and fairly large surface area. Ma and his
8 co-workers have undertaken a major research and development project to investigate
9 the technical and economic feasibility of iron shavings for the enhance treatment of
10 industrial process wastes, with success achieved.¹⁰ Nowadays, coupling of iron
11 shaving into the upflow anaerobic sludge blanket (UASB) has been developed in our
12 laboratory for the treatment of high strength wastewater containing NACs. The
13 interaction between iron shaving and microorganisms, as well as the dynamic change
14 of iron surface and microbial population after long-term operation, will be
15 investigated in our future study.

16

17 **4. Conclusions**

18 Compared to the biotic control system and the ZVI control system, both DNAN
19 reduction and DAAN formation were significantly improved in the ZVI coupled
20 biosystem. The high performance of the ZVI coupled biosystem could be attributed to
21 the high accumulation of formate, low ORP and great pH self-buffering capability at
22 the presence of ZVI. Compared with the biotic control system, the survival
23 environment for methanogens was effectively improved in ZVI coupled biosystem. In
24 addition, the ZVI coupled biosystem showed high efficiency in terms of DNAN
25 removal with the coexistence of competitive electron acceptors, such as nitrate and
26 sulfate. The ZVI coupled biosystem could be a promising alternative to the
27 conventional anaerobic reduction process for the removal of recalcitrant contaminants
28 from wastewater, especially for the treatment of wastewater containing
29 multi-substituted NACs.

30

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7

8 References

- 9 1 V.M. Boddu, K. Abburi, A.J. Fredricksen, S.W. Maloney, R. Damavarapu, *Environ.*
10 *Technol.*, 2009, **30**, 173-181.
- 11 2 W.E. Platten, D. Bailey, M.T. Suidan, S.W. Maloney, *Chemosphere*, 2010, **81**,
12 1131-1136.
- 13 3 N.N. Perreault, D. Manno, A. Halasz, S. Thiboutot, G. Ampleman, J. Hawari,
14 *Biodegradation*, 2012, **23**, 287-295.
- 15 4 J.Y. Shen, C.J. Ou, Z.Y. Zhou, J. Chen, K. X. Fang, X.Y. Sun, J.S. Li, L. Zhou, L.J.
16 Wang, *J. Hazard. Mater.*, 2013, **260**, 993-1000.
- 17 5 S. Susarla, Y. Yonezawa, S. Masunaga, *Water Res.*, 1998, **32**, 639-648.
- 18 6 B.A. Donlon, E. Razo-Flores, J.A. Field, G. Lettinga, *Appl. Environ. Microbiol.*,
19 1995, **61**, 3889-3893.
- 20 7 J. Singh, S. Comfort, P. Shea, *J. Environ. Qual.*, 1998, **27**, 1240-1245.
- 21 8 B.T. Oh, C.L. Just, P.J.J. Alvarez, *Environ. Sci. Technol.*, 2001, **35**, 4341-4346.
- 22 9 C.B. Wang, W. X. Zhang, *Environ. Sci. Technol.*, 1997, **31**, 2154-2156.
- 23 10 L.M. Ma, W.X. Zhang, *Environ. Sci. Technol.*, 2008, **42**, 5384-5389.
- 24 11 S.C. Ahn, D. K. Cha, B. J. Kim, S.-Y. Oh, *J. Hazard. Mater.*, 2011, **192**, 909-914.
- 25 12 S.Y. Oh, P.C. Chiu, B.J. Kim, D.K. Cha, *Water Res.*, 2005, **39**, 5027-5032.
- 26 13 J. Shen, Z. Zhou, C. Ou, X. Sun, J. Li, W. Han, L. Zhou, L. Wang, *J. Environ. Sci.*,
27 *2012*, **24**, 1900-1907.
- 28 14 W.W. Li, Y. Zhang, J.B. Zhao, Y.L. Yang, R. J. Zeng, H.Q. Liu, Y.J. Feng,
29 *Bioresour. Technol.*, 2013, **149**, 38-43.
- 30 15 Y.B. Zhang, Y.W. Liu, Y.W. Jing, Z.Q. Zhao, X. Quan, *J. Environ. Sci.*, 2012, **24**,

- 1 720-727.
- 2 16 X. Xiao, G.P. Sheng, Y. Mu, H.Q. Yu, *Water Res.*, 2013, **47**, 6007-6013.
- 3 17 W. Yin, J. Wu, P. Li, G. Lin, X. Wang, B. Zhu, B. Yang, *Chem. Eng. J.*, 2012, **210**,
- 4 309-315.
- 5 18 Y. Liu, Y. Zhang, Z. Zhao, Y. Li, X. Quan, S. Chen, *Bioresour. Technol.*, 2012, **121**,
- 6 148-153.
- 7 19 X.Y. Yu, C. Amrhein, M. A. Deshusses, M. R. Matsumoto, *Environ. Sci. Technol.*,
- 8 2006, **40**, 1328-1334.
- 9 20 J. Shen, R. He, H. Yu, L. Wang, J. Zhang, X. Sun, J. Li, W. Han, L. Xu, *Bioresour.*
- 10 *Technol.*, 2009, **100**, 1922-1930.
- 11 21 C. Olivares, J. Liang, L. Abrell, R. Sierra-Alvarez, J.A. Field, *Biotechnol. Bioeng.*,
- 12 2013, **110**, 1595-1604.
- 13 22 J. Fan, Y. Guo, J. Wang, M. Fan, *J. Hazard. Mater.*, 2009, **166**, 904-910.
- 14 23 M. Hou, F. Li, X. Liu, X. Wang, H. Wan, *J. Hazard. Mater.*, 2007, **145**, 305-314.
- 15 24 L. Liang, W. Sun, X. Guan, Y. Huang, W. Choi, H. Bao, L. Li, Z. Jiang, *Water Res.*,
- 16 2014, **49**, 371-380.
- 17 25 L. Zhu, H. Lin, J. Qi, X. Xu, *Environ. Sci. Pollut. R.*, 2013, **20**, 6119-6127.
- 18 26 J. Huang, Y. Wen, N. Ding, Y. Xu, Q. Zhou, *Water Res.*, 2012, **46**, 4361-4370.
- 19 27 V. Murali, S.-A. Ong, L.-N. Ho, Y.S. Wong, *Bioresour. Technol.*, 2013, **143**,
- 20 104-111.
- 21 28 Y. Liu, Y. Zhang, X. Quan, Y. Li, Z. Zhao, X. Meng, S. Chen, *Chem. Eng. J.*, 2012,
- 22 **192**, 179-185.
- 23 29 X. Meng, Y. Zhang, Q. Li and X. Quan, *Biochem. Eng. J.*, 2013, **73**, 80-85.
- 24 30 X. Quan, X. Zhang, H. Xu, *Water Res.*, 2015, **78**, 74-83.
- 25 31 Z.Z. Ismail, S. G. Pavlostathis, *Biodegradation*, 2010, **21**, 43-57.
- 26 32 C.F. Shen, N. Kosaric, R. Blaszczyk, *Water Res.*, 1993, **27**, 25-33.
- 27 33 G. Zhen, X. Lu, Y.Y. Li, Y. Liu, Y. Zhao, *Chem. Eng. J.*, 2015, **263**, 461-470.
- 28 34 S. Karri, R. Sierra-Alvarez, J.A. Field, *Biotechnol. Bioeng.*, 2005, **92**, 810-819.
- 29 35 D.O. Tas, S.G. Pavlostathis, *Environ. Sci. Technol.*, 2005, **39**, 8264-8272.
- 30 36 W. Zhang, L. Chen, H. Chen, S.Q. Xia, *J. Hazard. Mater.*, 2007, **143**, 57-64.

- 1 37 J.F. Devlin, K.O. Allin, *Environ. Sci. Technol.*, 2005, **39**, 1868-1874.
- 2 38 R.K. Thauer, K. Jungermann, K. Decker, *Bacteriol. Rev.*, 1977, **41**, 100-180.
- 3 39 P. Clauwaert, K. Rabaey, P. Aeltermann, L. De Schamphelaire, T.H. Ham, P. Boeckx,
4 N. Boon, W. Verstraete, *Environ. Sci. Technol.*, 2007, **41**, 3354-3360.
- 5 40 M.Uchimiya, L. Gorb, O. Isayev, M.M. Qasim, J. Leszczynski, *Environ. Pollut.*,
6 2010, **158**, 3048-3053.
- 7 41 Y.H. Hwang, D.G. Kim, H.S. Shin, *J. Hazard. Mater.*, 2011, **185**, 1513-1521.
- 8 42 M.J. Alowitz, M. M. Scherer, *Environ. Sci. Technol.*, 2002, **36**, 299-306.
- 9 43 K. Ritter, M.S. Odziemkowski, R. Simpraga, R.W. Gillham, D.E. Irish, *J. Contam.*
10 *Hydrol.*, 2003, **65**, 121-136.
- 11 44 T. Suzuki, M. Moribe, Y. Oyama, M. Niinae, *Chem. Eng. J.*, 2012, **183**, 271-277.
- 12 45 Y.H. Huang, T.C. Zhang, *Water Res.*, 2006, **40**, 3075-3082.
- 13 46 F.P. van der Zee, I.A.E. Bisschops, V. Blanchard, R.H.M. Bouwman, G. Lettinga,
14 J.A. Field, *Water Res.*, 2003, **37**, 3098-3109.
- 15 47 M.A. Reis, J.S. Almeida, P.C. Lemos, M.J. Carrondo, *Biotechnol. Bioeng.*, 1992,
16 **40**, 593-600.
- 17 48 A.J. Pyzik, S. E. Sommer, *Geochim. Cosmochim. Ac.*, 1981, **45**, 687-698.
- 18 49 L.A. Hoferkamp, E. J. Weber, *Environ. Sci. Technol.*, 2006, **40**, 2206-2212.
- 19 50 M. Elsner, R.P. Schwarzenbach, S.B. Haderlein, *Environ. Sci. Technol.*, 2004,
20 **38**, 799-807.
- 21 51 J. Hawari, F. Monteil-Rivera, N.N. Perreault, A. Halasz, L. Paquet, Z.
22 Radovic-Hrapovic, S. Deschamps, S. Thiboutot, G. Ampleman, *Chemosphere*,
23 2015, **119**, 16-23.

24

1 **Figure captions**

2

3 **Figure 1** Concentration evolution of DNAN and its corresponding reduction
4 intermediates as a function of reduction time (—■— ZVI coupled biosystem,
5 —●— biotic control system, —▲— ZVI control system).

6

7 **Figure 2** Evolution of the ORP during reduction of DNAN as a function of time.

8

9 **Figure 3** Concentration evolution of formate in the ZVI coupled biosystem and biotic
10 control system.

11

12 **Figure 4** Dynamics of biogas production in the ZVI coupled biosystem (a) and biotic
13 control system (b).

14

15 **Figure 5** Effects of nitrate on DNAN reduction as a function of time (—■— ZVI
16 coupled biosystem, —●— ZVI coupled biosystem with nitrate, —▼— biotic
17 control system, —▲— biotic control system with nitrate).

18

19 **Figure 6** Effects of sulfate on DNAN reduction as a function of time (—■— ZVI
20 coupled biosystem, —●— ZVI coupled biosystem with sulfate, —▼— biotic
21 control system, —▲— biotic control system with sulfate).

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Table 1 Constants of pseudo zero-order and pseudo first-order models for the reduction of the DNAN in three individual systems

Experiment condition	Pseudo zero-order kinetics		Pseudo first-order kinetics	
	k_0 (mg h ⁻¹)	R^2	k_1 (h ⁻¹)	R^2
ZVI coupled biosystem	9.382	0.943	1.263	0.951
Biotic control system	1.910	0.867	0.217	0.993
ZVI control system	0.625	0.990	0.018	0.988

Table 2 Change of pH and Fe²⁺ concentrations in different systems

Parameter	Influent of wastewater	Effluent of biotic control system	Effluent of ZVI control system	Effluent of ZVI coupled biosystem
pH	7.12 ± 0.01	6.35 ± 0.04	7.41 ± 0.01	6.67 ± 0.02
Fe ²⁺ (mg/L)	n.d.	n.d.	0.06 ± 0.01	0.34 ± 0.02

n.d. Means not detectable.

Fig. 1

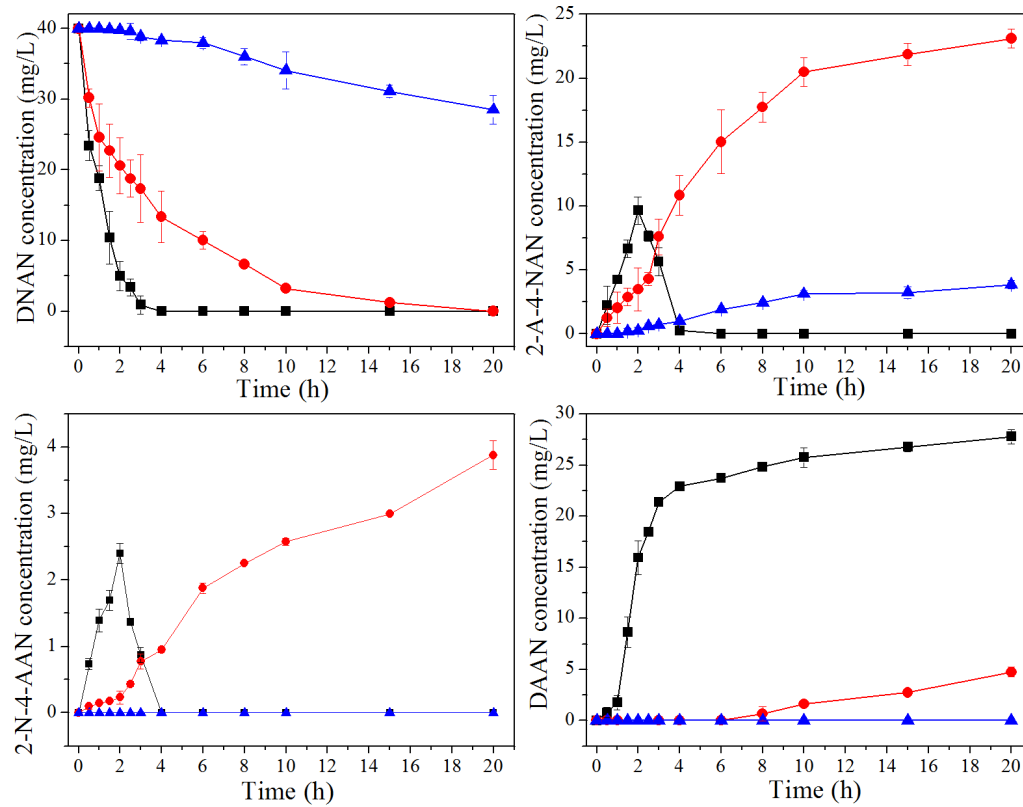


Fig. 2

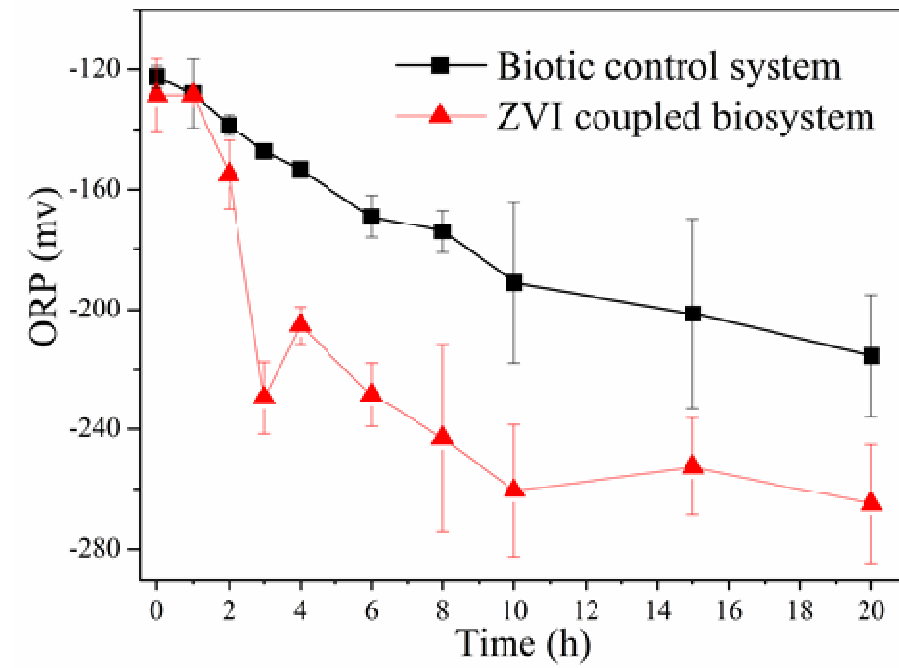


Fig. 3

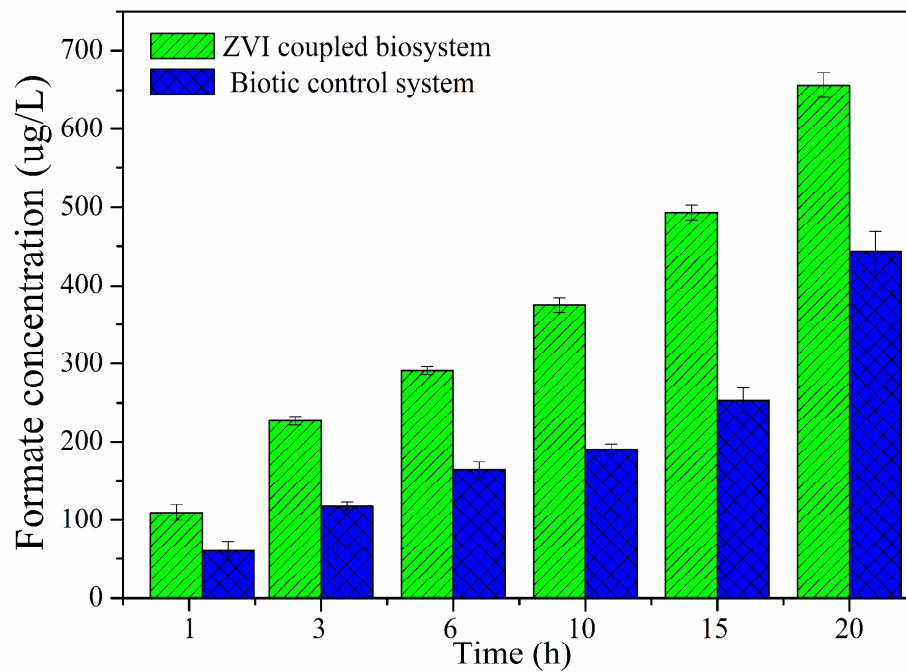


Fig. 4

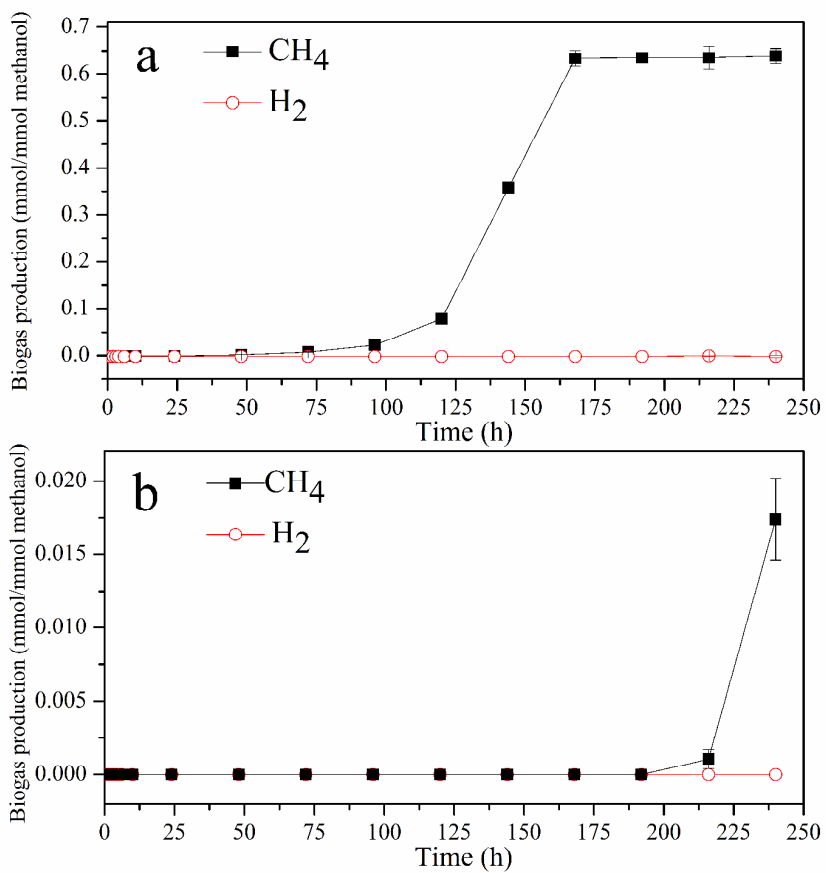


Fig. 5

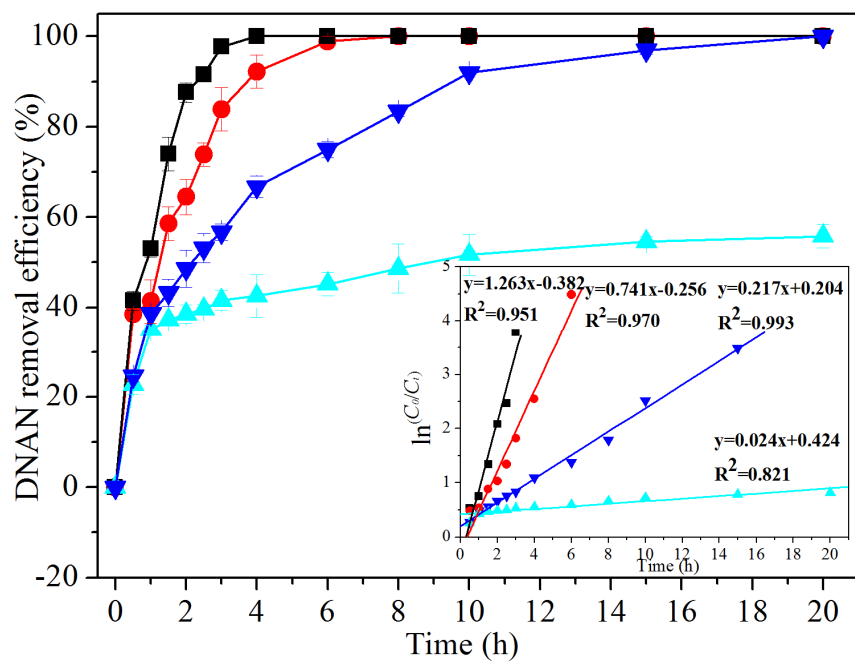
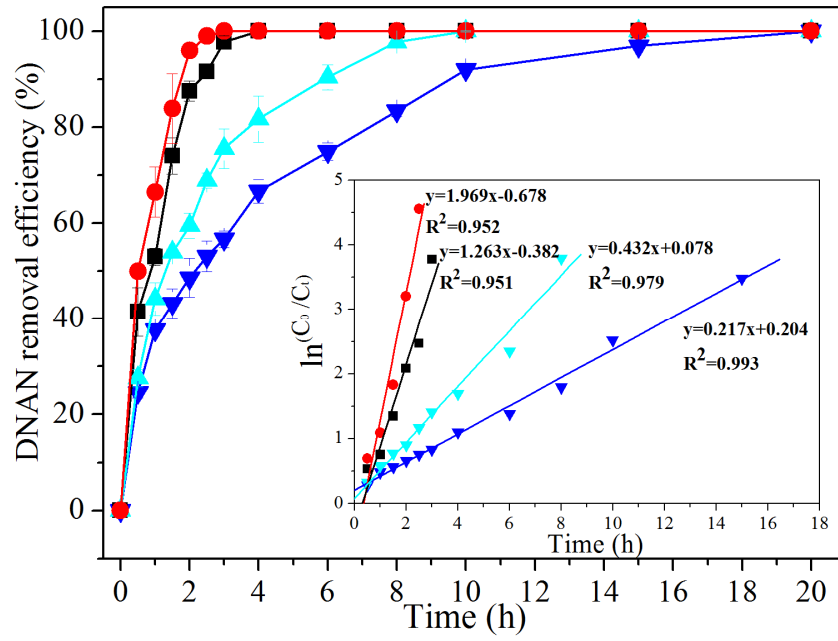


Fig. 6



Graphical abstract

