

# RSC Advances



This is an *Accepted Manuscript*, which has been through the Royal Society of Chemistry peer review process and has been accepted for publication.

*Accepted Manuscripts* are published online shortly after acceptance, before technical editing, formatting and proof reading. Using this free service, authors can make their results available to the community, in citable form, before we publish the edited article. This *Accepted Manuscript* will be replaced by the edited, formatted and paginated article as soon as this is available.

You can find more information about *Accepted Manuscripts* in the [Information for Authors](#).

Please note that technical editing may introduce minor changes to the text and/or graphics, which may alter content. The journal's standard [Terms & Conditions](#) and the [Ethical guidelines](#) still apply. In no event shall the Royal Society of Chemistry be held responsible for any errors or omissions in this *Accepted Manuscript* or any consequences arising from the use of any information it contains.

## A Chemosensor for Al<sup>3+</sup> Ion in Aqueous Ethanol Medium: Photophysical and Live Cell Imaging Studies

Nabanita Chatterjee,<sup>a</sup> Shubhra Bikash Maity,<sup>a</sup> Asmita Samadder,<sup>b,c</sup> Puspall Mukherjee,<sup>a</sup> Anisur Rahman Khuda-Bukhsh,<sup>b</sup> Parimal K. Bharadwaj\*<sup>a</sup>

<sup>a</sup> Department of Chemistry, Indian Institute of Technology Kanpur, Kanpur 208016, India

<sup>b</sup> Cytogenetics and Molecular Biology Laboratory, Department of Zoology, University of Kalyani, Kalyani 741235, India

<sup>c</sup> Department of Zoology, Dum Dum Motijheel College, West Bengal State University, Kolkata 700074, India

\* Email: pkb@iitk.ac.in

A novel fluorescent chemosensor **L** has been designed and synthesized based on rhodamine-B moiety. The dye **L** exhibits selective fluorescence enhancement towards the Al<sup>3+</sup> ion over other biologically relevant metal ions in aqueous ethanolic medium (EtOH–H<sub>2</sub>O, 2:3, v/v). The structure of the probe **L** has been established by <sup>1</sup>H- and <sup>13</sup>C-NMR spectroscopy, single crystal X-ray diffraction, ESI-mass spectrometry and elemental analysis. The cleavage of the spirolactam bond of the rhodamine moiety induced by the Al<sup>3+</sup> ion generates the delocalized xanthenes fluorophore that is responsible for the emission enhancement of the probe **L**. Recognition behavior of the receptor **L** has been investigated experimentally with supports from theoretical DFT studies. Furthermore, the efficacy of **L** in cell imaging studies is also probed by confocal microscopy.

### Introduction

Design and synthesis of fluorescent probes for environmentally hazardous metal ions has achieved considerable current interest in terms of using these probes for chemosensing, live cell imaging, ion transport, metalloenzyme mimics, catalysis and nuclear waste treatment.<sup>1–3</sup>

Extensive use of aluminum in food additives, pharmaceuticals, electrical equipments, cooking utensils, computer parts etc.<sup>4</sup> leads to environmental problems. In human, it exists<sup>5</sup> in its ionic form  $\text{Al}^{3+}$  with long residence time before it excretes through urine. Accumulation of this ion can be potentially hazardous affecting neurological order that may lead to Parkinson's disease, Alzheimer's disease etc.<sup>6-7</sup> It also hampers the growth of plants and biological functions of metalloenzymes.<sup>8</sup> Therefore, spatial as well as temporal distribution of  $\text{Al}^{3+}$  ions even at low concentration is highly desirable.

Our interest in developing chemosensors for the  $\text{Al}^{3+}$  ion led us to synthesize the dye **L** to utilize the excellent photophysical properties of rhodamine B as a turn-on sensor.<sup>9-11</sup> Several rhodamine derivatives have been reported as sensors for the  $\text{Al}^{3+}$  ions.<sup>12-17</sup> Most of them are functional only in organic solvents due to strong hydration of  $\text{Al}^{3+}$  in water thus making them difficult for practical applications.<sup>18</sup> To increase aqueous solubility of the chemosensor, 2-methoxy-4-nitroaniline group, has been appended as the receptor to the rhodamine B moiety (Scheme 1) as it can increase the hydrophilic interaction by enhancing the polar character in the molecule.<sup>19</sup>

The dye is well-characterized by  $^1\text{H}$ - and  $^{13}\text{C}$ -NMR spectroscopy, single crystal X-ray diffraction, ESI-mass spectrometry and elemental analyses. It acts as a fluorescence turn-on chemosensor for the  $\text{Al}^{3+}$  ion in aqueous ethanol medium.

## Experimental section

**Materials and methods.** Reagent grade rhodamine B and all metal perchlorate and nitrate salts were from Sigma Aldrich (USA). 2-Methoxy-4-nitroaniline was obtained from Alfa Aesar (USA). Other chemicals such as triethylamine,  $\text{POCl}_3$  and EDTA were obtained from S. D. Fine Chemicals (India). These chemicals were used as received without further purification. All

the solvents were from S. D. Fine Chemicals and were dried following standard protocols prior to use. Chromatographic separation was done by column chromatography using basic  $\text{Al}_2\text{O}_3$  from S. D. Fine Chemicals (India).

***Caution! Perchlorate salts are potentially explosive especially if they are mixed with organic solvents and should be handled with utmost care.***

### **Analysis and measurements**

Both  $^1\text{H}$  NMR (500 MHz) and  $^{13}\text{C}$  NMR (125 MHz) spectra of the compounds were recorded on a JEOL JNM-LA500 FT spectrometer in  $\text{CDCl}_3$  with tetramethylsilane as the internal standard. The ESI-Mass data were obtained in acetonitrile using a WATERS-Q-TOF Premier Mass Spectrometer. UV-Vis absorption spectra were recorded on a Shimadzu 2450 UV-vis spectrophotometer in aqueous ethanol medium at 298 K. Steady-state fluorescence spectra were obtained using a Perkin-Elmer LS 50B Luminescence spectrometer at 298 K with excitation and emission band-pass of 10 nm. The excitation wavelength was 520 nm and the spectra were recorded in the range 540-700 nm. Fluorescence quantum yields were determined by comparing the corrected spectra with that of pure rhodamine B in ethanol (ESI†).

Single-crystal X-ray data of compound **L** was collected at 100 K on a Bruker SMART APEX CCD diffractometer using graphite monochromated  $\text{MoK}_\alpha$  radiation ( $\lambda = 0.71073 \text{ \AA}$ ). The data reduction, structure solution and refinements were carried out using the SHELXL-97 package. The details are also provided in the supplementary information†.

### **Jobs plot and Benesi-Hildebrand plot**

Job's plot experiment was conducted, keeping the total concentration of **L** and  $\text{L-Al}^{3+}$  complex at  $1 \times 10^{-4} \text{ mol L}^{-1}$  and the mole fraction of  $\text{Al}^{3+}$  varied from 0 to 1. The sensor **L** (2.96

mg) was dissolved in 5 mL ethanol to make a solution of  $10^{-3}$  M order and diluted to 25 mL  $1.0 \times 10^{-4}$  M order by addition of ethanol and water (EtOH:H<sub>2</sub>O = 2:3, v/v). 2.7, 2.4, 2.1, 1.8, 1.5, 1.2, 0.9, 0.6 and 0.3 mL stock solution of L were taken and kept them into vials. 25 mL  $1.0 \times 10^{-4}$  M stock solution of Al(NO<sub>3</sub>)<sub>3</sub>.9H<sub>2</sub>O was also prepared in similar way in ethanol. 0.3, 0.6, 0.9, 1.2, 1.5, 1.8, 2.1, 2.4 and 2.7 mL Al<sup>3+</sup> stock solution were to the vials containing solution of sensor in such a way that total volume of each vial were 3 mL. After shaking the vials UV-vis absorbance were performed at room temperature.

Similarly for Benesi-Hildebrand experiment a series of stock solution of Al(NO<sub>3</sub>)<sub>3</sub>.9H<sub>2</sub>O were prepared from  $1 \times 10^{-6}$  to  $1 \times 10^{-3}$  M range in different vials and added separately to  $1 \times 10^{-5}$  M solution of L. After mixing the vials fluorescence spectra were recorded at room temperature.

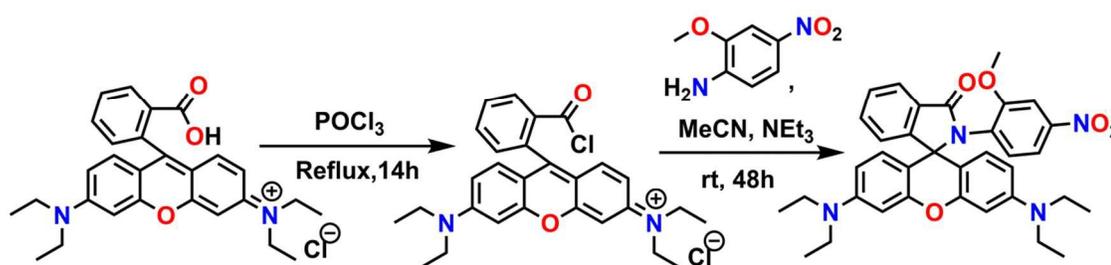
### Cell culture and imaging

MTT [3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyltetrazolium bromide] and Dimethyl sulphoxide (DMSO) of analytical grade were procured from Sigma-Aldrich Inc. (St-Louis, MO, USA); Dulbecco's modified Eagle's medium (DMEM), fetal bovine serum, and antibiotics were procured from Gibco BRL (Grand Island, NY, USA). All organic solvents used were of HPLC grade. Briefly, the rat skeletal muscle cells of L6 cell line obtained from National Centre for Cell Science, Pune, India, were grown in Dulbecco's modified Eagle's Medium (DMEM) supplemented with 10% foetal bovine serum and 1% antibiotic in a 5% carbon dioxide incubator maintained at 37°C and plated in different petri-plates.<sup>20</sup> All the control and experimental sets of cells were grown till they attained 70–80% confluence, and then the different treatment procedures commenced. The concentration of the sensor optimized for the study was estimated at  $7 \times 10^{-5}$  M (20 mL of total volume of stock solution); it was incubated for 15 min and the

concentration of that of  $\text{Al}(\text{NO}_3)_3$  salt solution standardized at  $4 \times 10^{-3} \text{M}$  (5 mL of total volume of stock solution) was also incubated for 15 min. Cell images were taken under an Andor Spinning Disk Confocal Microscope. The images were then evaluated against untreated control set of cells. The fluorescence intensity profile was derived using Analysis Software, NIS Elements AR Version 4.00.

### Synthesis of the chemosensor L

Synthesis of the chemosensor was easily achieved in two steps as illustrated in Scheme 1.



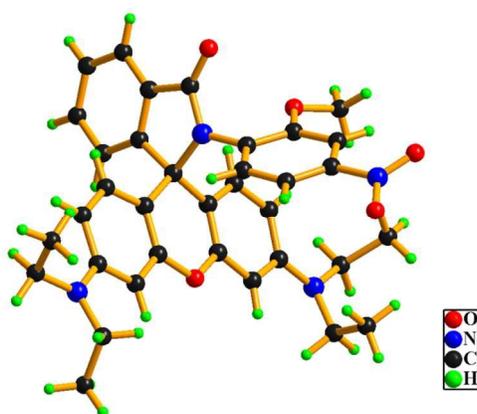
**Scheme 1** Synthetic route of the chemosensor L

To rhodamine B (2 g, 4.2 mmol) taken in a three-necked round bottom flask, 50 mL of freshly distilled phosphorous oxychloride was added and the mixture was heated reflux for 24 h under a dinitrogen blanket. Excess  $\text{POCl}_3$  was then distilled off and the acid chloride formed was dried under reduced pressure and used for the next step without further purification. The crude acid chloride was dissolved in dry acetonitrile (70 mL) and treated with 2-methoxy-4-nitroaniline (1.2 equivalents) in presence of triethylamine (2 mL) taken in the same solvent (50 mL) over a period of 20 min. After the addition was complete, the reaction mixture was allowed to stir at room temperature for 48 h and then all the solvent was removed under reduced pressure. The crude reaction mixture was dissolved in dichloromethane ( $3 \times 50 \text{ mL}$ ) and washed several times with water. The organic layer after drying over anhydrous  $\text{Na}_2\text{SO}_4$ , was evaporated off

completely to afford a dark pink solid. Column chromatography on basic  $\text{Al}_2\text{O}_3$  using DCM/Hexane (70: 30, v/v) as the eluent provided the desired product as a pale yellow crystalline solid in ~ 58% yield. M.p. ~ 210 °C (uncorrected);  $^1\text{H}$  NMR (500 MHz,  $\text{CDCl}_3$ , 25 °C,  $\text{Si}(\text{CH}_3)_4$ )  $\delta$ : 1.13 (t, 12 H,  $J = 7.45$  Hz), 3.29 (q, 8 H,  $J = 6.85$  Hz), 3.58 (s, 3 H), 6.19 (d, 2 H,  $J = 2.3$  Hz), 6.29 (dd, 2 H,  $J = 2.3, 8.75$  Hz), 6.38 (d, 1H,  $J = 8.05$  Hz), 6.65 (d, 2 H,  $J = 8.55$  Hz), 7.21(s, 1 H), 7.53-7.55 (m, 4 H), 8.00-8.02 (m, 1H) (Fig. S1 $\dagger$ );  $^{13}\text{C}$  NMR (125 MHz,  $\text{CDCl}_3$ , 25 °C,  $\text{Si}(\text{CH}_3)_4$ )  $\delta$ : 12.66, 44.44, 56.07, 68.38, 97.48, 105.76, 106.85, 107.68, 115.51, 123.59, 124.39, 128.47, 129.59, 129.72, 131.46, 131.92, 133.02, 147.99, 148.94, 152.89, 153.54, 156.56, 166.69 (Fig. S2 $\dagger$ ). ESI MS:  $m/z$  (%): 593.2761 (100)  $[\text{M} + \text{H}]^+$  (Fig. S3 $\dagger$ ); Anal. Calcd. for  $\text{C}_{35}\text{H}_{36}\text{N}_4\text{O}_5$ : C, 70.93; H, 6.12; N, 9.45%. Found: C, 71.07; H, 6.28; N, 9.32%.

## Results and discussion

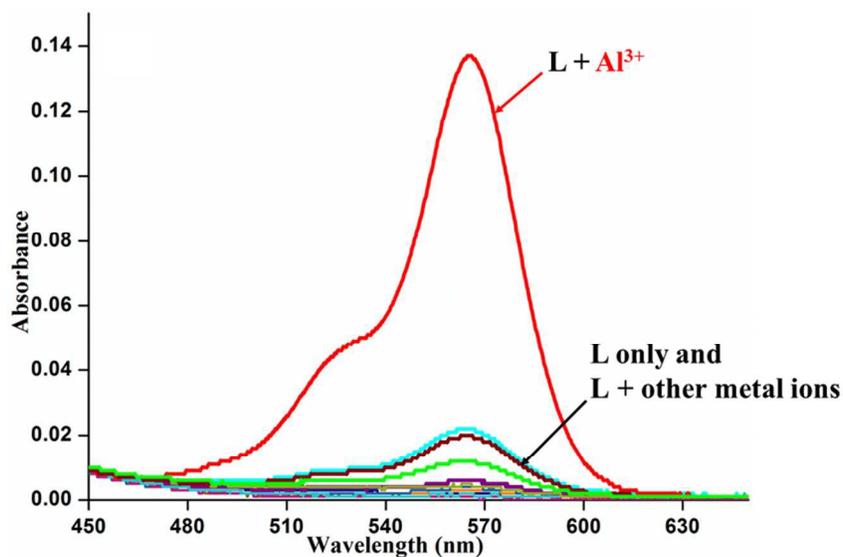
The chemosensor **L** can be synthesized from readily available chemicals in two steps. The rectangular parallelepiped pale yellow single crystals, suitable for X-ray crystallography were obtained by slow evaporation of the solution of the compound in ethyl acetate at room temperature. A perspective view of the molecule is shown in Fig. 1 while the crystallographic data are collected in Table S1 in the supporting information (ESI $\dagger$ ).



**Fig. 1** X-ray single crystal structure of **L**

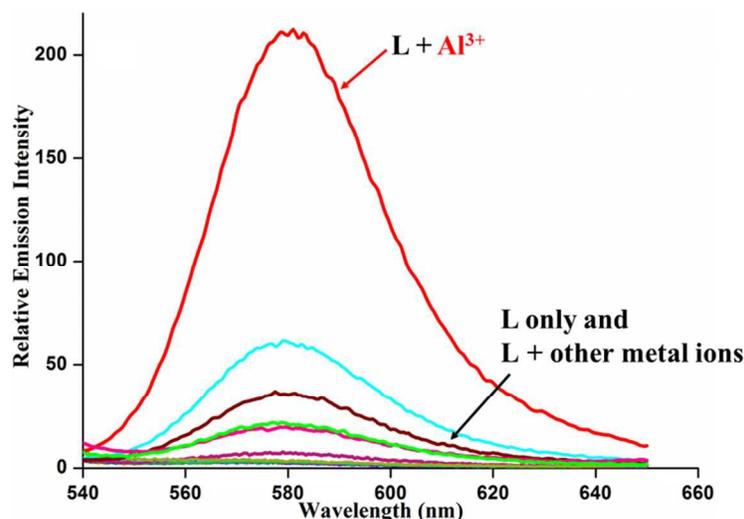
## Photophysical properties

The probe is readily soluble in common organic solvents and also has sufficient solubility in aqueous-organic medium. All spectroscopic measurements were carried out in aqueous ethanol (EtOH–H<sub>2</sub>O, 2:3 v/v) medium. Perchlorate salts of metal ions that include Na<sup>+</sup>, K<sup>+</sup>, Mg<sup>2+</sup>, Ca<sup>2+</sup>, Mn<sup>2+</sup>, Fe<sup>2+</sup>, Fe<sup>3+</sup>, Co<sup>2+</sup>, Ni<sup>2+</sup>, Cu<sup>2+</sup>, Zn<sup>2+</sup>, Cd<sup>2+</sup>, Ag<sup>+</sup>, Pb<sup>2+</sup> and Hg<sup>2+</sup> were used in the photophysical studies. For Al<sup>3+</sup> and Cr<sup>3+</sup> ions nitrate salts were used. The metal free dye **L** exhibits negligible absorption and fluorescence above 500 nm implies the retention of the spirolactam bond in this region. However, addition of 10 equivalents of the Al<sup>3+</sup> ion to a solution of **L** makes the solution from colorless to pink in about 20 min that can be detected with a naked eye. In the visible region, a strong band appears at 565 nm with a shoulder around 525 nm (Fig. 2) indicating breaking of the spirolactam bond in presence of the Al<sup>3+</sup> ion. No such significant change is observed in presence of any of the aforementioned metals ions.



**Fig. 2** Absorption spectra of **L** in EtOH–H<sub>2</sub>O (2:3, v/v) medium in the presence of 10 equivalents of metal ions. [**L**] = 1 × 10<sup>−4</sup> M.

Upon excitation at 520 nm, the metal-free chemosensor **L** ( $\Phi = 0.0065$ ) exhibit only a very weak fluorescence. In sharp contrast, addition of  $\text{Al}^{3+}$  ion to the solution of **L** leads to a strong emission ( $\sim 50$  fold enhancement) giving a band centering around 580 nm ( $\Phi = 0.328$ ).

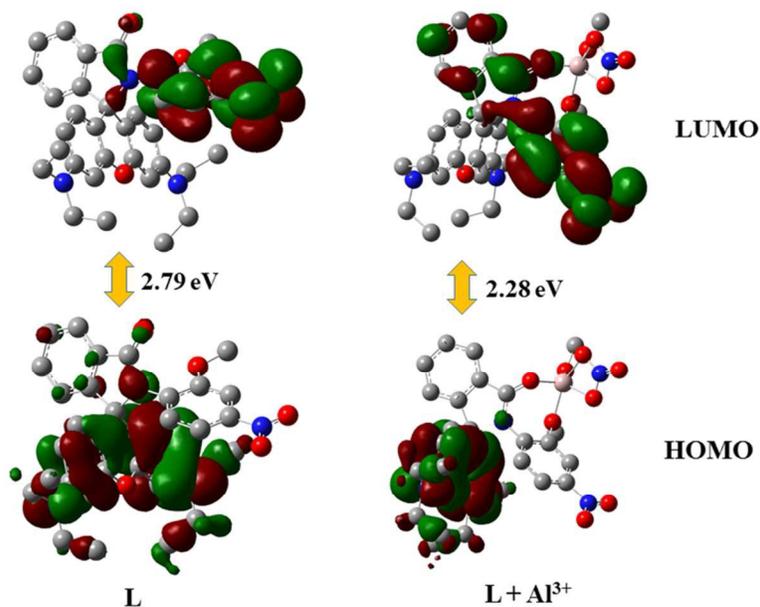


**Fig. 3** Relative emission spectra of **L** in EtOH–H<sub>2</sub>O (2:3, v/v) in presence of 10 equivalents of metal ions. [**L**] = 10  $\mu\text{M}$ . Slit width = 10/10 nm.  $\lambda_{\text{ex}}$  = 520 nm.

Other metal ions listed above do not elicit any noticeable emission when added to the solutions of **L**. These results are consistent with the absorption spectral results. To confirm the spirocyclic ring opening, the  $^{13}\text{C}$  NMR spectra of the complexes were also recorded. The absence of the characteristic peak of the quaternary carbon in the region, 67.0–69.0 ppm<sup>21–22</sup> in the metal complex (Fig. S4†) reveals the rupture of the spirolactam bond to the ring-open form. In the IR spectra of the metal free sensor **L** the amide carbonyl frequency appears at 1709  $\text{cm}^{-1}$ . The downward shift around 1589  $\text{cm}^{-1}$  (Fig. S5†) of the amide carbonyl stretching in the complexes is consistent with fact that the carbonyl oxygen is involved in the coordination. In addition, a peak in the region 1370–1384  $\text{cm}^{-1}$  suggests the presence of coordinated nitrate ion ( $\text{NO}_3^-$ ) in the complex.

## Theoretical study

For better understanding about the nature of binding of the ligands, theoretical calculations were performed using Gaussian-09 software. For the comparative study, various molecular interactions of the ligand **L** and **L-Al<sup>3+</sup>** complex has been studied using density functional theory (DFT) with the B3LYP/6-31G (d,p+) functional model and basis set. Energy optimization calculations have revealed that **L-Al<sup>3+</sup>** complex is energetically much more stable compared to the ligand **L**. Here the energy gap between the HOMO and LUMO of the complex (2.28 eV) was less compared to that of the corresponding sensor **L** (2.79 eV) (Fig. 4).



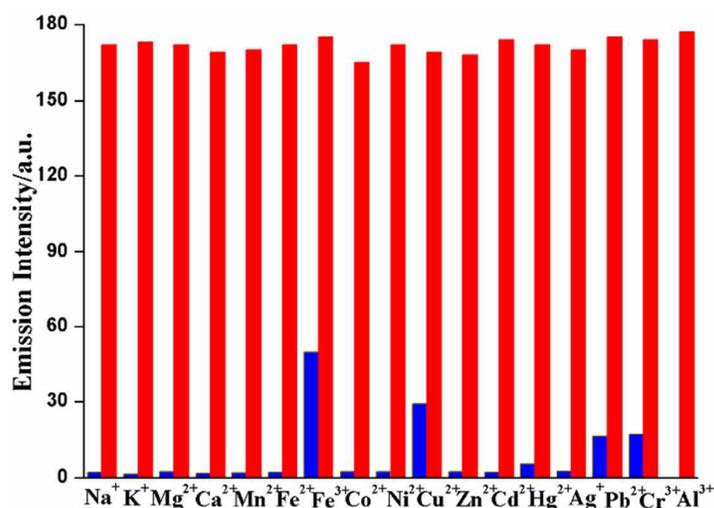
**Fig. 4** Optimized structures of HOMO and LUMO s of the ligand **L** and **L-Al<sup>3+</sup>** complex.

From the spatial electronic distribution in the frontiers orbitals of the ligand **L** it can be easily pointed out that in the free ligand the electron density mainly resides on the xanthene moiety and some electron clouds on C=O moiety. In the complexes, spirolactam bonds were opened when the **Al<sup>3+</sup>** ions bind to the ligand. These structural change introduced by the attachment of the

metals ions lead to significant charge transfer from the xanthene moiety towards the metal binding moiety that could be well observed from the frontiers orbitals of the complex. Here the electron charge cloud was pulled towards the electron withdrawing nitro group in the LUMO. So here the extent of these transfer of electron cloud have played crucial role to this tuning of selectivity towards the tripositive ions. The Optimized structures of the sensor **L** and **L-Al<sup>3+</sup>** complexes are summarized in Fig. S9†.

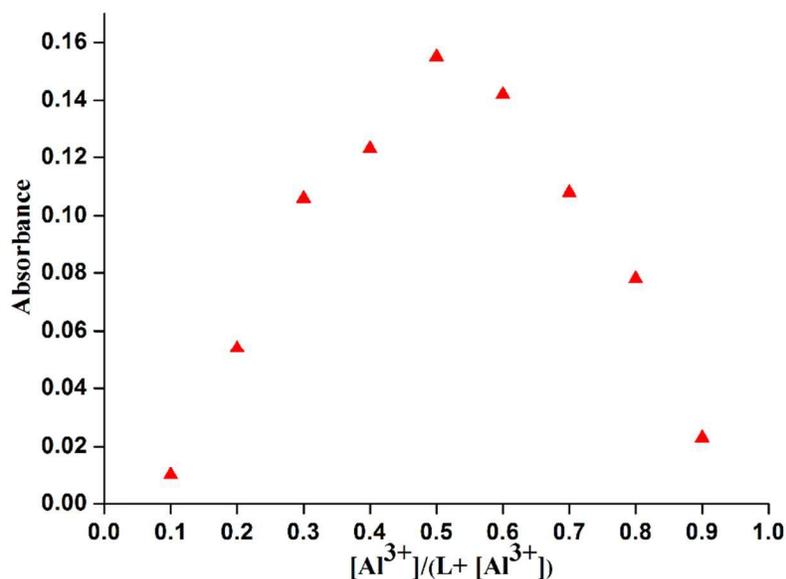
### Selectivity Experiment:

In order to check the selectivity of **L** towards **Al<sup>3+</sup>** ions over the other tested metal ions competitive studies were carried out (Fig. 5). The emission responses of the sensor **L** towards **Al<sup>3+</sup>** ion in the presence of excess of background metal ions are also recorded. In presence of other competing metal ions, no significant change in the emission intensity has been noticed in the sensing of **Al<sup>3+</sup>** ion confirming its selectivity towards the **Al<sup>3+</sup>** ion.



**Fig. 5** Selectivity of the dye **L** for **Al<sup>3+</sup>** ion in EtOH–H<sub>2</sub>O (2:3, v/v) mixture. Blue bars indicate the fluorometric response of **L** with 50 equivalents of the metal ion of interest; red bars represent the final integrated fluorescence response after the addition of 10 equivalents of **Al<sup>3+</sup>** ion to each solution containing other metals over the initial integrated emission. Dye concentration = 10  $\mu$ M;  $\lambda_{\text{exc}}$  = 520 nm.

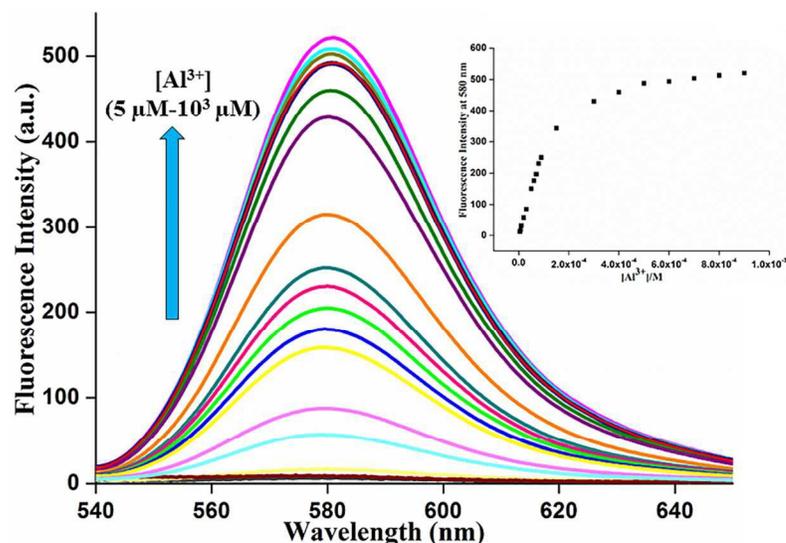
The binding stoichiometry was determined by the Job's plot experiments based on absorption keeping the total concentration of **L** and  $\text{Al}^{3+}$  ion at  $1 \times 10^{-4}$  M (Fig. 6).<sup>23</sup> The experiments support 1:1 complexation between **L** and  $\text{Al}^{3+}$ . A peak at 743.3220 due to  $[\text{L} + \text{Al}^{3+} + 2\text{NO}_3^-]^+$  in the ESI-mass spectrum of the  $\text{Al}^{3+}$  complex further confirms this 1:1 binding stoichiometry (Fig. S6†).



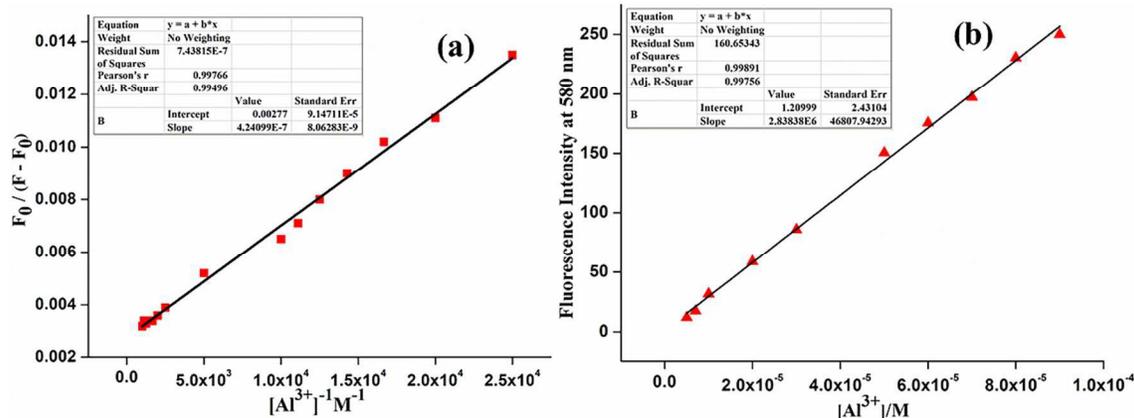
**Fig. 6** Job's plot of absorbance of **L** and  $\text{Al}^{3+}$  in EtOH–H<sub>2</sub>O (2:3, v/v) medium. Total concentration =  $1 \times 10^{-4}$ M.

The association constant for the complex and the detection limit were determined from emission titration data (Fig. 7).<sup>24</sup> Following the Benesi-Hildebrand plot<sup>25</sup> the association constant of **L** with  $\text{Al}^{3+}$  ion is found to be  $6.53 \times 10^3 \text{ M}^{-1}$  ( $R^2 = 0.99496$ ) (Fig. 8a). This value matches well with the reported range ( $10^3$ – $10^9 \text{ M}^{-1}$ ) of the association constants for the  $\text{Al}^{3+}$  specific chemosensors reported in the literature.<sup>26–27</sup> The detection limit is calculated to be  $0.316 \mu\text{M}$  (Fig. 8b) which is smaller compared to the maximum concentration of  $7.41 \mu\text{M}$  of  $\text{Al}^{3+}$  ion in

drinking water approved by WHO.<sup>28</sup> Also the calculated value of detection limit is found to lie in comparable range ( $1.0 \times 10^{-6}$  M) for the reported  $\text{Al}^{3+}$  sensors.<sup>13,29-34</sup>



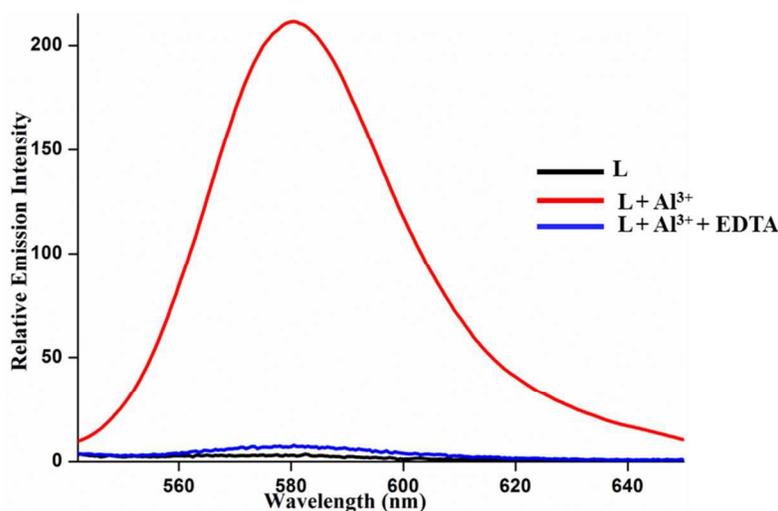
**Fig. 7** Change of fluorescence spectra of the dye **L** as a function of  $[\text{Al}^{3+}]$  ion in neutral EtOH- $\text{H}_2\text{O}$  (2:3 v/v) medium. The arrow indicates the trend in increasing  $[\text{Al}^{3+}]$ .



**Fig.8** (a) Binding constant plot of **L** for  $\text{Al}^{3+}$ . (b) Linear response curve of **L** at 580 nm depending on the  $\text{Al}^{3+}$  ion concentration for determination of lowest detection limit.

Reversibility of fluorescence is an imperative characteristic in developing chemosensors for practical aspects. The reversible nature of the sensor **L** is checked by adding an aqueous

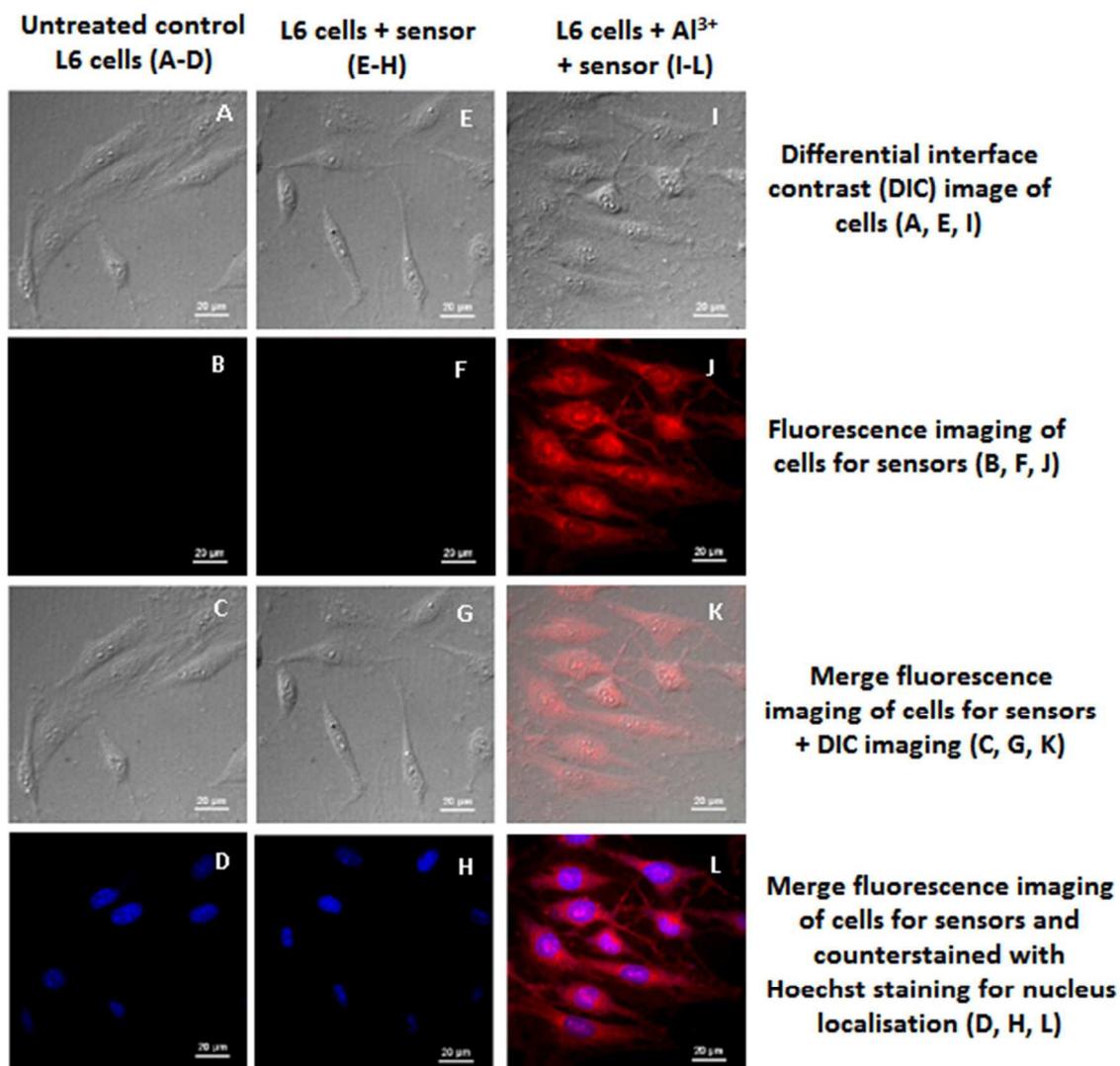
solution of  $\text{Na}_2\text{EDTA}$  to the solution of metal complex. Addition of excess EDTA salt solution results in the lowering of emission intensity to the levels of metal-free sensors (Fig. 9).



**Fig. 9** Emission responses of **L** in the presence of 10 equivalents  $\text{Al}^{3+}$  ion and subsequent addition of excess EDTA solution to the **L** complex of  $\text{Al}^{3+}$  ion in EtOH–H<sub>2</sub>O (2:3, v/v) medium.

### Cell imaging studies

The potentiality of a sensor to determine the presence of guest species in living cells is of immense practical importance.<sup>35</sup> To confirm the competence of the chemosensor in biological samples the intracellular confocal imaging studies have been carried out in the presence of  $\text{Al}^{3+}$  ion. No distinct intracellular fluorescence was observed for the series of cells incubated with normal untreated cells (B) or sensor (H) alone. However, significant increase in the fluorescence was gradually observed in case of sensor **L** when exogenous  $\text{Al}^{3+}$  stock solution was added into them *via* incubation with  $\text{Al}(\text{NO}_3)_3$  salt solution (N) (Fig. 10). This finding corroborated well with the statistically analyzed fluorescence intensity profile (Fig. S7†). Counterstaining of the cells was also done with Hoechst dye to confirm the permeability range of the sensors which showed a more prominent distribution in the cytoplasm than that of the nucleus (Fig. 10).



**Fig. 10.** Confocal imaging of control and treatment series of L6 cells for sensor L viewed under 40X magnification; the scale bar provided in each image measures 20 $\mu$ m in size:

A-D: Untreated control L6 cells; E-H: L6 cells + sensor; I-L: L6 cells + Al<sup>3+</sup> + sensor; Differential interface contrast (DIC) image of untreated control L6 cells (A), L6 cells + sensor (E) and L6 cells + Al<sup>3+</sup> + sensor (I); Fluorescence imaging of untreated control L6 cells (B), L6 cells + sensor (F) and L6 cells + Al<sup>3+</sup> + sensor (J); Merge fluorescence imaging + DIC imaging of untreated control L6 cells (C), L6 cells + sensor (G) and L6 cells + Al<sup>3+</sup> + sensor (K); Merge fluorescence imaging of cells for sensors and Hoechst staining for nucleus localisation of untreated control L6 cells (D), L6 cells + sensor (H) and L6 cells + Al<sup>3+</sup> + sensor (L).

### MTT assay for cell viability assessment

For the sensor **L**, the MTT assay (Mossman, 1983) has been performed to evaluate the extent of cytotoxicity it could induce in normal cells.<sup>36</sup> The cell viability sustains ~85-90% upon treatment of maximum dose of **L** (50  $\mu$ L of stock solution), which indicates that the sensor **L** does not affect the cell viability to a significant extent, an important criterion for biological application and also supports the intracellular  $Al^{3+}$  ion detection in living cells as well. No significant difference was observed between the percentages of viable cells in the sensor-treated and the solvent-treated lots, indicating thereby that the sensors were non-cytotoxic and safe for biological uses (Fig. S8<sup>†</sup>).

### Conclusions

In summary, a new rhodamine based chemosensor **L** has been synthesized readily. Metal assisted spirolactam ring opening of the dyes give noticeable change in fluorescence response. Sufficient water solubility, high selectivity, sensitivity, reversibility along with the noticeable color change made this probes suitable chemosensor for the  $Al^{3+}$  ion. The sensor **L** being non-cytotoxic in nature, and having easy cellular penetrability can be a possible candidate for some practical application as an additional tool for detection of cellular  $Al^{3+}$  ions.

### Acknowledgements

This work was supported by Department of Science and Technology, New Delhi, India (SERB/CHM/20130239). PKB thanks SERB for the funding. Financial support from the CSIR, New Delhi to NC and PM are gratefully acknowledged. SBM thanks IIT Kanpur for financial assistance. ARK-B and AS sincerely thank to UGC, New Delhi for awarding an Emeritus Fellowship. The authors are thankful to Department of Zoology, The University of Kalyani.

## Notes and references

Department of Chemistry, Indian Institute of Technology, Kanpur 208016, India.

E-mail: [pkb@iitk.ac.in](mailto:pkb@iitk.ac.in). Fax: (+91) 512-259-7637; Tel: (+91) 512-259-7034.

†Electronic supplementary information (ESI) available: [Materials and physical methods, general procedures, tables, schemes, figures, characterization data, and some spectra are provided in ESI]. CCDC No. for sensor **L** is 1421529.

1 K. P. Carter, A. M. Young and A. E. Palmer, *Chem. Rev.*, 2014, **114**, 4564–4601.

2 D.T. Quang and J.S. Kim, *Chem. Rev.*, 2010, **110**, 6280–6301.

3 J. P. Desvergne and A. W. Czarnik, *Chemosensors of Ion and Molecule Recognition. In NATO Science Series C: Mathematical and Physical Sciences*, ed. Kluwer Academic, Dordrecht, London, **1997**.

4 S. Kim, J. Y. Noh, K. Y. Kim, J. H. Kim, H. K. Kang, S. W. Nam, S. H. Kim, S. Park, C. Kim and J. Kim, *Inorg. Chem.*, 2012, **51**, 3597–3602.

5 D. Maity and T. Govindaraju, *Inorg. Chem* 2010, **49**, 7229–7231.

6 S. W. King, J. Savory and M. R. Willis, *Crit. Rev. Clin. Lab. Sci.*, 1981, **13**, 1–20.

7 I. Shcherbatykh and D. O. Carpenter, *J. Alzheimers Dis.*, 2007, **11**, 191–205.

8 E. Delhaize and P. R. Ryan, *Plant Physiol.*, 1995, **107**, 315–321.

9 X. Chen, T. Pradhan, F. Wang, J. S. Kim and J. Yoon, *Chem. Rev.*, 2012, **112**, 1910–1956.

10 J. R. Lakowicz, *Principles of Fluorescence Spectroscopy*, 3rd ed.; Springer, New York, 2006.

- 11 S. Pal, N. Chatterjee and P. K. Bharadwaj, *RSC Adv.*, 2014, **4**, 26585–26620.
- 12 S. B. Maity and P. K. Bharadwaj, *Inorg. Chem.*, 2013, **52**, 1161–1163.
- 13 I. H. Hwang, Y. W. Choi, K. B. Kim, G. J. Park, J. J. Lee, L. Nguyen, I. Noh and C. Kim, *New J. Chem.*, 2016, **40**, 171–178 and references 21– 61 therein.
- 14 B. Sen, M. Mukherjee, S. Banerjee, S. Pal and P. Chattopadhyay, *Dalton Trans.*, 2015, **44**, 8708–8717 and references therein.
- 15 K. Ghosh, A. Majumdar and T. Sarkar, *RSC Advances*, 2014, **4**, 23428–23432.
- 16 S. Sahana, S. Bose, S. K. Mukhopadhyay and P. K. Bharadwaj, *J. Lumin.*, 2016, **169**, 334–341.
- 17 Y.-W. Wang and Y. Peng, *Org. Lett.*, 2012, **14**, 3420–3423.
- 18 S. Gui, Y. Huang, F. Hu, Y. Jin, G. Zhang, L. Yan, D. Zhang and R. Zhao, *Anal. Chem.* 2015, **87**, 1470–1474 and references there in.
- 19 L. Di and E. Kerns, *Drug-Like Properties: Concepts, Structure Design and Methods from ADME to Toxicity Optimization*, Elsevier Science, London, 2nd Edition, 2015, pp 61–90.
- 20 A. Samadder, S. Das, J. Das and A.R. Khuda-Bukhsh, *Colloids Surf. B*, 2013, **109**, 10–19.
- 21 S. Saha, M. U. Chhatbar, P. Mahato, L. Praveen, A. K. Siddhanta and A. Das, *Chem. Commun.*, 2012, **48**, 1659–1661.
- 22 K. Ghosh, T. Sarkar, A. Samadder, *Org. Biomol. Chem.*, 2012, **10**, 3236–3243.
- 23 C. Yang, L. Liu, T.-W. Mu and Q.-X. Guo, *Anal. Sci.*, 2000, **16**, 537–539.

- 24 J. Bourson, J. Pouget and B. Valeur, *J. Phys. Chem.*, 1993, **97**, 4552–4557.
- 25 H. A. Benesi and J. H. Hildebrand, *J. Am. Chem. Soc.*, 1949, **71**, 2703–2707.
- 26 D. Maity and T. Govindaraju, *Chem. Commun.*, 2010, **46**, 4499–4501.
- 27 S. H. Kim, H. S. Choi, J. Kim, S. J. Lee, D. T. Quang and J. S. Kim, *Org. Lett.*, 2010, **12**, 560–563.
- 28 *Guidelines for drinking-water quality*, 2nd ed. World Health Organization; Vol. 2. Health criteria and other supporting information. Geneva, 1996.
- 29 W.-H. Ding, D. Wang, X.-J. Zheng, W.-J. Ding, J.-Q. Zheng, W.-H. Mu, W. Cao, L.-P. Jin, *Sens. Actuators B*, 2015, **209**, 359–367.
- 30 J. Kumar, M. J. Sarma, P. Phukan and D. K. Das, *Dalton Trans.*, 2015, **44**, 4576–4581.
- 31 Y. Fu, X.-J. Jiang, Y.-Y. Zhu, B.-J. Zhou, S.-Q. Zang, M.-S. Tang, H.-Y. Zhang and T. C. W. Mak, *Dalton Trans.*, 2014, **43**, 12624–12632.
- 32 M. Shellaiah, Y.H. Wu, H.C. Lin, *Analyst*, 2013, **138**, 2931–2942.
- 33 V.P. Singh, K. Tiwari, M. Mishra, N. Srivastava, S. Saha, *Sens. Actuators B*, 2013, **82**, 546–554.
- 34 Y.-Y. Guo, L.-Z. Yang, J.-X. Ru, X. Yao, J. Wu, W. Dou, W.-W. Qin, G.-L. Zhang, X.-L. Tang, W.-S. Liu, *Dyes and Pigments*, 2013, **99**, 693–698.
- 35 X. Peng, J. Du, J. Fan, J. Wang, Y. Wu, J. Zhao, S. Sun and X. Xu, *J. Am. Chem. Soc.*, 2007, **129**, 1500–1501.

36 T. Mosmann, *J. Immunol. Methods*, 1983, **65**, 55–63.

## Graphical Abstract

### A Chemosensor for Al<sup>3+</sup> Ion in Aqueous Ethanol Medium: Photophysical and Live Cell Imaging Studies

Nabanita Chatterjee,<sup>a</sup> Shubhra Bikash Maity,<sup>a</sup> Asmita Samadder,<sup>b,c</sup> Puspall Mukherjee,<sup>a</sup> Anisur Rahman Khuda-Bukhsh,<sup>b</sup> Parimal K. Bharadwaj\*<sup>a</sup>

<sup>a</sup> Department of Chemistry, Indian Institute of Technology Kanpur, Kanpur 208016, India

<sup>b</sup> Cytogenetics and Molecular Biology Laboratory, Department of Zoology, University of Kalyani, Kalyani 741235, India

<sup>c</sup> Department of Zoology, Dum Dum Motijheel College, West Bengal State University, Kolkata 700074, India

\* Email: pkb@iitk.ac.in

A new rhodamine based highly sensitive “Turn-ON” fluorescent chemosensor **L** for the selective detection of Al<sup>3+</sup> ion over other biologically competing metal ions in aqueous ethanol medium.

