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1 **An integrated system for field analysis of Cd(II) and Pb(II) via**  
2 **preconcentration using nano-TiO<sub>2</sub>/cellulose paper composite**  
3 **and subsequent detection with a portable X-ray fluorescence**  
4 **spectrometer**

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9 **Abstract** An integrative field analytical system was developed for the determination of Pb(II) and  
10 Cd(II). The system was based on the hyphenation of a preconcentration process with a portable X-ray  
11 fluorescence spectrometer. Preconcentration was accomplished with a composite consisting of TiO<sub>2</sub>  
12 and a cellulose film (TCP) which was prepared by immobilizing TiO<sub>2</sub> on cellulose filter paper. TCP is  
13 shown to be an adsorbent with high adsorption capacity, i.e., more than 254 µg per piece and 259 µg  
14 for Pb(II) and Cd(II), respectively. Under the optimum adsorption conditions, the best adsorption  
15 ratios of Pb(II) and Cd(II) were more than 95.5% and 94.4%, respectively. The preconcentration of  
16 Pb(II) and Cd(II) was not adversely affected by other metals ions and humic acid. Pb(II) and Cd(II)  
17 were then directly quantified by XRF. The calibration plots for both Pb(II) and Cd(II) were linear in  
18 the range from 1.0 to 50.0 µg L<sup>-1</sup>. The detection limits (3σ; for n = 11) for Pb(II) and Cd(II) were 0.69  
19 and 0.51 µg L<sup>-1</sup>, respectively, and the levels of quantification were 2.30 and 1.71 µg L<sup>-1</sup>, respectively.  
20 The preconcentration factor was 10<sup>3</sup>. Concentrations of Pb(II) and Cd(II) in drinking water and river  
21 waters were determined and found to be in agreement with ICP-MS assays.

22 **Keywords** TiO<sub>2</sub> immobilization; preconcentration; heavy metals; X-ray fluorescence; field analysis

## 23 1. Introduction

24 Lead and cadmium contamination in drinking water and natural water has raised public health and  
25 environmental safety concerns [1]. The drinking water standards are recommended by the World  
26 Health Organization, including 10 µg L<sup>-1</sup> for Pb and 5 µg L<sup>-1</sup> for Cd, respectively [2]. Hence, there is  
27 a great need to develop a simple, sensitive, selective and inexpensive method for field determination  
28 of Pb(II) and Cd(II) in water samples.

29 In order to determine the concentrations of Pb(II) and Cd(II) in water matrix, different  
30 instrumental methods have been developed, including atomic absorption spectrometry [3,4],

31 inductively coupled plasma mass spectrometry [5], high performance liquid chromatography [6], and  
32 anodic stripping voltammetry [7]. However, the applications of these techniques are limited on field  
33 monitoring, because they usually require expensive and large instruments, expert operators, and  
34 sophisticated sample pretreatment, which limit their applications on field monitoring. The major  
35 advantages of X-ray fluorescence (XRF) are minimal sample preparation, rapid screen of large  
36 numbers of samples, and low cost [8-10]. Moreover, field-portable XRF analyzer is small enough to  
37 be used in the field [8-10]. However, XRF exhibits several limitations on aqueous samples, such as  
38 short linear range, matrix effects, and poor sensitivity [11]. The interferences for XRF can be  
39 attributed to spectral overlaps and/or the limited resolution of the analyzer [10]. XRF has been  
40 sparingly used for quantitative analysis, primarily due to LODs that are at best in the 1-10 ppm range  
41 for Pb(II) and Cd(II) [10]. Because metal concentrations in natural waters and drinking water  
42 standards are at the level of  $\mu\text{g L}^{-1}$ , the determination of Pb(II) and Cd(II) in water samples by XRF  
43 without preconcentration and elimination of interferences is not possible. Therefore, sample  
44 pretreatment using an appropriate adsorbent is a convenient step that makes this analysis possible  
45 [12,13].

46 Nanosized  $\text{TiO}_2$  is an excellent preconcentration material because of its large surface area, high  
47 adsorption capacity and hypotoxicity [14-17]. However, nanosized  $\text{TiO}_2$  powders frequently cause  
48 secondary pollutions and an extra microfiltration is necessary for the separation and recovery of  $\text{TiO}_2$   
49 powders after sample pretreatment. Nanosized  $\text{TiO}_2$  can be immobilized on cellulose filter paper and  
50 form a  $\text{TiO}_2$  cellulose composite film (TCP), which can offer better stability, feasible continuous  
51 operations, easy separation and recycle, and significant decrease of the operation costs [18].

52 In this work, a practical procedure was developed for the field determination of trace Pb(II) and  
53 Cd(II) in water samples by XRF after a preconcentration step. The sophisticated sample pretreatment

54 (e.g., elution or digestion) was avoided and the risk of contamination and errors was reduced. TCP  
55 with different metal adsorption capacity was used for preconcentration of Pb(II) and Cd(II) and  
56 elimination of spectral overlaps on XRF and then the major obstacle of XRF was overcome.

57

## 58 **2. Experimental**

### 59 **2.1 Apparatus and reagents**

60 Both Pb(II) and Cd(II) on TCP were determined by energy dispersive XRF spectrometry using a  
61 Thermo Scientific Niton hand-held XRF analyser (XL3t 950, Niton, USA). The instrument was  
62 fitted with an X-ray tube with Ag anode target excitation source and a geometrically optimized  
63 large area drift detector, and data were transferred using Thermo Scientific Niton data transfer PC  
64 software. Inductively coupled plasma mass spectrometry (ICP-MS, 7500cx, Agilent, USA) was  
65 used as a comparative method for metal determination. The peristaltic pump (BT00-300T, Baoding  
66 Longer Peristaltic Pump Co., Ltd., China) was applied to propel water samples or standard solutions  
67 onto the circulating filtration system with TCP to enrich Pb(II) and Cd(II).

68 The standard solutions of Pb(II) and Cd(II) were prepared from stock standard solutions (GSB G  
69 62071-90 and 62040-90, China, respectively). The certified reference water samples,  
70 GBW(E)080402 and GBW(E)080399 (NRCCRM, China) were used. Tetrabutyl titanate (TBOT),  
71 ethanol, acetic acid (HAc), HCl, NaOH and Tris(hydroxymethyl)aminomethane (Tris) were  
72 analytical grade (Sigma, USA). Cellulose filter paper (qualitative,  $\Phi$  7cm) was purchased from  
73 Xinhua Paper Industry (Hangzhou, China). River-derived humic acid was purchased from  
74 International Humic Substances Society (USA) and dissolved in 0.1 mol L<sup>-1</sup> NaOH solution. The  
75 desired pH was adjusted by HCl or Tris. The plasticware for storing reagent solutions and standards  
76 as well as water samples were Teflon PFA (Nalgene, Nalge, USA) or low-density polyethylene

77 (Nalgene, Nalge, USA) bottles. All containers were soaked in 10% HCl solution at least 24 h before  
78 use.

## 79 **2.2 Preconcentration and determination experiments**

80 The preparation of TCP and a enrichment of Pb(II) and Cd(II) on TCP in a circulating filtration  
81 system were according to a literature procedure [18]. After adsorption, the concentration of residual  
82 metals was measured by ICP-MS and TCP was washed by water and dried immediately, then the  
83 metal contents on TCP were determined by XRF. The adsorption ratio of metals on TCP was given  
84 as follows: Adsorption ratio= $(C_0-C_t)/C_0$ , where  $C_0$  and  $C_t$  were the initial and final concentration of  
85 each metal in the solution, respectively.

## 86 **2.3 Real samples analysis**

87 The water samples, including drinking water and river water, were collected from Zhangzhou,  
88 Fujian province, China. The insoluble particles in water samples were excluded by filtration  
89 through a 0.22  $\mu\text{m}$  pore (Millipore) cellulose acetate membrane. Adjusting the pH of the solution  
90 into 8.0, the filtrates were used for further preconcentration on TCP and determination by XRF.  
91 Meanwhile, the filtrates with 2.0 mL of concentrated  $\text{HNO}_3$  and 1.0 mL of  $\text{H}_2\text{O}_2$  (30%) were  
92 decomposed by microwaves under a pressure of 10 atm for 10 min. After being cooled naturally to  
93 room temperature, the decomposed solution was diluted to 25 mL and then used for metal  
94 determination by ICP-MS.

95

## 96 **3. Results and discussion**

### 97 **3.1 Characteristic of Nano-TiO<sub>2</sub>/Cellulose Paper**

98 The anatase TiO<sub>2</sub> possessed the highest chemisorption capacities among the crystal structures of  
99 anatase, rutile, mix crystal (Degussa P25) and amorphism, which could be attributed to its high

100 surface energy, rough surface and unsaturated oxygen bond [19]. Therefore, anatase TiO<sub>2</sub> was  
101 immobilized onto cellulose filter paper. Fig. 1 showed representative SEM and EDX images ( the  
102 previous report by us in Ref. 18 ) of the cellulose filter paper before and after the impregnation with  
103 TiO<sub>2</sub>. Compared the changes in the morphology, the surface of filter paper became rough because  
104 of depositing a TiO<sub>2</sub> film (shown in **Figure 1a, 1b and Figure 2**). This result was consistent with  
105 EDX images in Ref 16. Cellulose filter paper with large numbers of -OH and -C-O-O- groups could  
106 easily combine with Ti ion on its cellulose surface. The interaction between cellulose and TiO<sub>2</sub> was  
107 strong enough to keep the stability of TCP during its application [20]. The crystal structure of TiO<sub>2</sub>  
108 in TCP was anatase and its average crystallite size was 6.0 nm, according to the calculation on the  
109 anatase (101) diraction peaks with the Debye–Scherrer formula [18].

110 **Figure 1**

111 **Figure 2**

### 112 **3.2 Optimization of method**

113 To obtain the optimal conditions for the ppreconcentration of Pb(II) and Cd(II), the following  
114 parameters were optimized: (a) Sample pH value; (b) adsorption time; (c) dynamic adsorption; (d)  
115 adsorption capacity. Respective data and Figures were given in the Electronic Supporting Material.  
116 The following experimental conditions were found to give best results: (a) a sample pH value of 8.0  
117 (**Figure S1**); (b) an adsorption time of 3 h (**Figure S2**); (d) a sample volume of 1.0 L (**Figure S3**);  
118 (d) a dynamic adsorption of the second-order equation (**Table S1**).

### 119 **3.3 Adsorption Capacity of Nano-TiO<sub>2</sub>/Cellulose Paper for Pb(II) and Cd(II)**

120 The plots of adsorption capacity versus initial metal concentration were shown in **Figure 3**.  
121 Obviously, when initial concentrations of heavy metals were increased from 20.0 to 1000.0 µg L<sup>-1</sup>,  
122 positive correlation between the amount of each metals (Pb(II) and Cd(II)) adsorbed by TCP and

123 the equilibrium concentration of each metal in sample solution was obtained, and the adsorption  
124 rates of each metals were more than 90.0%. When metal concentration was  $1000.0 \mu\text{g L}^{-1}$ , TCP did  
125 not reach its saturation adsorption for Pb(II) and Cd(II), so the adsorption capacity of TCP for Pb(II)  
126 and Cd(II) were more than  $254 \mu\text{g}$  and  $259 \mu\text{g}$ , respectively.

127 **Figure 3**

### 128 **3.4 Evaluation of interference**

129 As the real environmental water samples always contain various organics and inorganics, some  
130 potentially interfering ions were investigated

131 In order to investigate the preconcentration and determination of Pb(II) and Cd(II) from their  
132 binary mixtures with diverse interference ions, an aliquot of solutions ( $1.0 \text{ L}$ ) containing  $20.0 \mu\text{g L}^{-1}$   
133 of Pb(II) and Cd(II) and interference ions were treated according to our recommended procedure.  
134 The results indicated that no interference was observed when Na(I), K(I)(1500 fold), Ca(II), Mg(II)  
135 (250 fold), Cu(II), Ni(II), Zn(II) (150 fold), Mn(II), Cr(III), Al(III) (100 fold), Fe(III) (50 fold),  
136 nitrate ion, chlorid ion (1000 fold) were added. All the concentrations of these potentially  
137 interfering ions were higher than the quality standard of surface water (GB3838-2002, China).

138 Humic acids (HA) were presented widely in natural waters. They were organic macromolecules  
139 with a large number of carboxylic (-COOH) and phenolic (-OH) groups, which could combine  
140 heavy metals [21,22]. The influences of humic acids addition ( $0\text{-}20.0 \text{ mg}\cdot\text{L}^{-1}$ ) on the adsorption  
141 ratio of Pb(II) and Cd(II) were so limited that they could be ignored (seen in **Figure 4**). HA had  
142 little effect on the surface properties of titanate although HA tended to affect competitive adsorption  
143 of Pb (II) and Cd (II) due to the formation of HA-metal coordination compounds [21,23], but the  
144 stability constant of the surface complex between metals and  $\text{TiO}_2$  (anatase) was more than that of  
145 HA-Metal complexes [18]. The presence of HA could not affect the XRF signal of Pb(II) and Cd(II)



146 at the concentration range of 3-21 mg L<sup>-1</sup>.

147 **Figure 4**

### 148 **3.5 Methods performance and analysis of real samples**

149 According to the above mentioned procedure, a series of experiments were conducted to obtain  
150 the calibration graph, precision and detection limit for the determination of Pb (II) and Cd (II). The  
151 analytical curves consisted of seven points and good correlation coefficients were shown in **Figure**  
152 **5**. The calibration curves presented linear behavior in the concentration range from 1.0 to 100 µg L<sup>-1</sup>,  
153 for Pb (II) and Cd (II), the curves could be described as  $M=0.920C-1.992$  ( $R^2=0.997$ ,  $n=6$ ), and  
154  $M=1.055C-1.998$  ( $R^2=0.998$ ,  $n=6$ ), respectively, where M was the amount of metals on TCP, and C  
155 was the elemental concentration in the water sample. The detection limits (based on  $3\sigma$  of the blank  
156 determinations,  $n=11$ ) were 0.69 and 0.51 µg L<sup>-1</sup> for Pb (II) and Cd (II), respectively, and the levels  
157 of quantification were 2.30 and 1.71 µg L<sup>-1</sup>, respectively. Meanwhile, the certified reference water  
158 samples, GBW(E)080401 (certified Cd concentration: 0.10 mg L<sup>-1</sup>) and GBW(E)080398 (certified  
159 Pb concentration: 0.50 mg L<sup>-1</sup>) were used to validate the above mentioned procedure, and the  
160 relative standard error ( $n=5$ ) were 4.1% and 4.4% for Pb (II) and Cd (II), respectively. As shown in  
161 **Table 1**, compared the characteristic data of present method with those reported in Ref. 27-29, the  
162 detection limits was notably improved. Though the methods in Ref. 24-26 had lower detection limits,  
163 they were not suitable for field monitoring because of their large instruments, expert operators, and  
164 sophisticated sample pretreatment.

165 **Figure 5**

166 **Table 1**

167 The real water samples were collected from drinking water and Jiulong River in Zhangzhou,  
168 Fujian province, China. The concentration of Pb (II) and Cd (II) in water samples was determined

169 by our procedure. The comparisons between our method and ICP-MS method were summarized in  
170 **Table 2** and **Table 3**. The results of this method The results were in agreement with ICP-MS  
171 determinations.

172 **Table 2**

173 **Table 3**

#### 174 **4. Conclusion**

175 In this study, an integration of TCP and field-portable XRF for the preconcentration and field  
176 determination of Pb (II) and Cd (II) in natural water was studied. This method showed high  
177 adsorption capacity, inexpensive, simple pretreatment and detection for Pb (II) and Cd (II) without  
178 the interference of coexisted metals and humic substances, i.e., the major obstacle of XRF was  
179 overcome.

180

#### 181 **Acknowledgement**

182 This work is supported by the National Natural Science Foundation of China (21475055, and  
183 21175115, S.X. L), the Program for New Century Excellent Talents in University (NCET-11 0904,  
184 S.X. L), and the Science & Technology Committee of Fujian Province, China (2012Y0065, F.Y. Z).

185 **References**

- 186 1 J. R. Koduru, K. D. Lee, *Food Chem.*, 2014, **150**, 1-8.
- 187 2 M. Hua, Y. Jiang, B. Wu, B. Pan, X. Zhao, Q. Zhang, *ACS Appl. Mater. Interfaces*, 2013, **5**,
- 188 12135-12142.
- 189 3 C. Duran, A. Gundogdu, V. N. Bulut, M. Soylak, L. Elci, H. B. Senturk, M. Tufekci, *J. Hazard.*
- 190 *Mater.*, 2007, **146**, 347-355.
- 191 4 M. Ezoddin, F. Shemirani, K. Abdi, M. Khosravi Saghezchi, R. JamaliM, *J. Hazard. Mater.*, 2010,
- 192 **178**, 900-905.
- 193 5 A. Milne, W. Landing, M. Bizimis, P. Morton, *Anal. Chim. Acta*, 2010, **665**, 200-207.
- 194 6 Q. Hu, G. Yang, J. Yin, Y. Yao, *Talanta*, 2002, **57**, 751-756.
- 195 7 A. Manivannan, R. Kawasaki, D. A. Tryk, A. Fujishima, *Electrochim. Acta*, 2004, **49**, 3313-3318.
- 196 8 L. S. G. Teixeira, E. S. Santos, L. S. Nunes, *Anal. Chim. Acta*, 2012, **722**, 29-33.
- 197 9 M. Alcalde-Molina, J. Ruiz-Jimenez, M. D. Luque de Castro, *Anal. Chim. Acta*, 2009, **652**,
- 198 148-153.
- 199 10 P. T. Palmer, R. Jacobs, P. E. Baker, K. Ferguson, S. Webber, *J. Agric. Food Chem.*, 2009, **57**,
- 200 2605-2613.
- 201 11 C. Fontàs, I. Queralt, M. Hidalgo, *Spectrochim. Acta Part B*, 2006, **61**, 407-413.
- 202 12 S. Özdemir, V. Okumuş, A. Dündar, E. Kılınç, *Microchim. Acta*, 2013, **180**, 719-739.
- 203 13 J. H. Chen, H. T. Xing, H.X. Guo, W. Weng, S.R. Hu, S.X. Li, Y. H. Huang, X. Sun, Z. B. Su, *J.*
- 204 *Mater. Chem. A*, 2014, **2**, 12561-12570.
- 205 14 W. Liang, Y. Chen, F. Zheng, S. X. Li, *Microchim. Acta*, 2014, 1-7.
- 206 15 X. Lin, Y. Chen, S. Li, *Analytical Methods*, 2013, **5**, 6480-6485.
- 207 16 F. Y. Zheng, S. X. Li, L. X. Lin, L. Q. Cheng, *J. Hazard. Mater.*, 2009, **172**, 618-622.

- 208 17 S. X. Li, F. Y. Zheng, S. J. Cai, T. S. Cai, *J. Hazard. Mater.*, 2011, **189**, 609-613.
- 209 18 S. X. Li, X. F. Lin, F. Y. Zheng, W. J. Liang, Y. X. Zhong, J. B. Cai, *Anal. Chem.*, 2014, **86**,  
210 7079-7083.
- 211 19 X. Xie, L. Gao, *Curr. Appl. Phys.*, 2009, **9**, S185-S188.
- 212 20 B. Zhao, Z. Shao, *J. Phys. Chem. C*, 2012, **116**, 17440-17447.
- 213 21 W. W. Tang, G. M. Zeng, J. L. Gong, J. Liang, P. Xu, C. Zhang, B. B. Huang, *Sci. Total*  
214 *Environ.*, 2014, **468**, 1014-1027.
- 215 22 M. L. Pacheco, E. M. Peña-Méndez, J. Havel, *Chemosphere*, 2003, **51**, 95-108.
- 216 23 T. Wang, W. Liu, L. Xiong, N. Xu, J. R. Ni, *Chem. Eng. J.*, 2013, **215-216**, 366-374.
- 217 24 M. Amjadi, A. Samadi, J. L. Manzoori, *Microchim. Acta*, 2015, 1-7.
- 218 25 S. K. Behzad, A. Balati, M. M. Amini, M. Ghanbari, *Microchim. Acta*, 2014, **181**, 1781-1788.
- 219 26 M. Ghazaghi, H. Shirkhanloo, H. Z. Mousavi, A. M. Rashidi, *Microchim. Acta*, 2015, **182**,  
220 1263-1272.
- 221 27 Y. Ding, W. Zhu, Y. Xu, X. Qian, *Sensor. Actuat. B Chem.*, 2015, **220**, 762-771.
- 222 28 N. Promphet, P. Rattanarat, R. Rangkupan, O. Chailapakul, N. Rodthongkum, *Sensor. Actuat. B*  
223 *Chem.*, 2015, **207**, 526-534.
- 224 29 K. Kocot, B. Zawisza, E. Marguá, I. Queralt, M. Hidalgo, R. Sitko, *J. Anal. Atom. Spectrom.*,  
225 2013, **28**, 736-742.

226 **Table 1** Comparison of the published methods with the developed method in this work

227 **Table 2** Determination of Cd(II) by this method and ICP-MS

228 **Table 3** Determination of Pb(II) by this method and ICP-MS

229 **Table 1** Comparison of the published methods with the developed method in this work

Methods	Materials	Linear range		Detection limit		Ref.
		$(\mu\text{g L}^{-1})$		$(\mu\text{g L}^{-1})$		
		Cd(II)	Pd(II)	Cd(II)	Pd(II)	
Flame atomic absorption spectrometry	Halloysite nanotubes / $\text{Fe}_3\text{O}_4$	0.5-50	-	0.27	-	24
Flame atomic absorption spectrometry	3-aminopropyltriethoxysilane-2,4-bis(3,5-dimethylpyrazol)-triazine / $\text{Fe}_3\text{O}_4$ nanoparticle	1-100	3-100	0.01	0.7	25
Electrothermal atomic absorption spectrometry	graphene / the zeolite clinoptilolite	0.24-10.3	0.011-0.48	0.004	0.07	26
Fluorescence spectrometry	2,2-dipicolylamine	0-1124	0-2072	4.0	8.1	27
Electrochemical sensor	graphene/polyaniline/polystyrene nanoporous fiber	10-500	10-500	4.43	3.30	28
X-ray fluorescence spectrometry	multiwalled carbon nanotubes	0-50	0-50	1.0	2.1	29
X-ray fluorescence spectrometry	Nano- $\text{TiO}_2$ /cellulose paper	1.0-50	1.0-50	0.69	0.51	This work

230

231 **Table 2** Determination of Cd(II) by this method and ICP-MS

Water samples	Add ( $\mu\text{g L}^{-1}$ )	Found ( $\mu\text{g L}^{-1}$ )	Recovery (%)	Found by ICP-MS ( $\mu\text{g L}^{-1}$ )
Drinking water	0	$0.65 \pm 0.07$	-	0.70
	1.0	$1.58 \pm 0.21$	93.0	1.61
	2.0	$2.58 \pm 0.14$	96.5	2.72
Jiulongjiang water	0	$0.91 \pm 0.09$	-	0.86
	1.0	$1.86 \pm 0.22$	95.0	1.79
	2.0	$2.80 \pm 0.16$	94.5	2.89

232 **Table 3** Determination of Pb(II) by this method and ICP-MS

Water samples	Add ( $\mu\text{g L}^{-1}$ )	Found ( $\mu\text{g L}^{-1}$ )	Recovery (%)	Found by ICP-MS ( $\mu\text{g L}^{-1}$ )
	0	-	-	0.07
Drinking water	1.0	$1.09 \pm 0.19$	109.0	1.11
	2.0	$1.95 \pm 0.13$	97.5	2.09
	0	-	-	0.29
Jiulongjiang water	1.0	$1.16 \pm 0.16$	116.0	1.32
	2.0	$1.88 \pm 0.13$	94.0	2.43



233 **Figure Captions**

234 **Figure 1** SEM images of cellulose filter paper (a) and TiO<sub>2</sub>/cellulose paper (b)

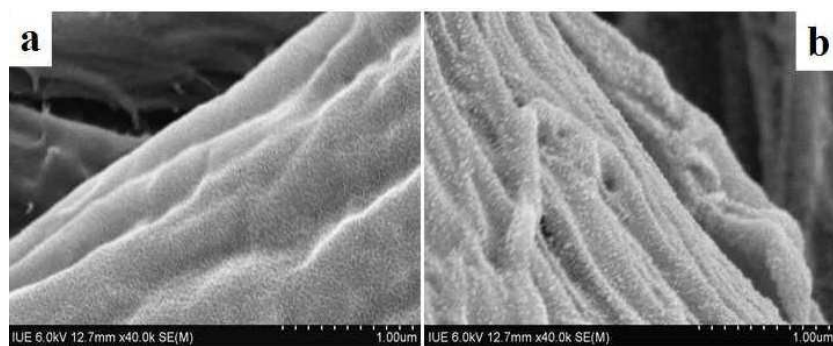
235 **Figure 2** EDX images of cellulose filter paper and TiO<sub>2</sub>/cellulose paper

236 **Figure 3** Adsorption capacity of Pb(II) and Cd(II) by TiO<sub>2</sub>/cellulose paper

237 **Figure 4** Effect of humic acid on the adsorption of Cd (II) and Pb (II) ( $C_{\text{metal}}=200 \mu\text{g L}^{-1}$ , pH=8.0;

238 t=3 h; V=1.0 L)

239 **Figure 5** Calibration curves of Cd (II) and Pb (II)

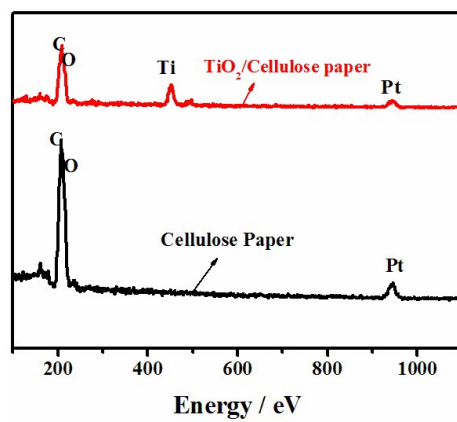


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**Figure 1** SEM images of cellulose filter paper (a) and TiO<sub>2</sub>/cellulose paper (b)



243

244

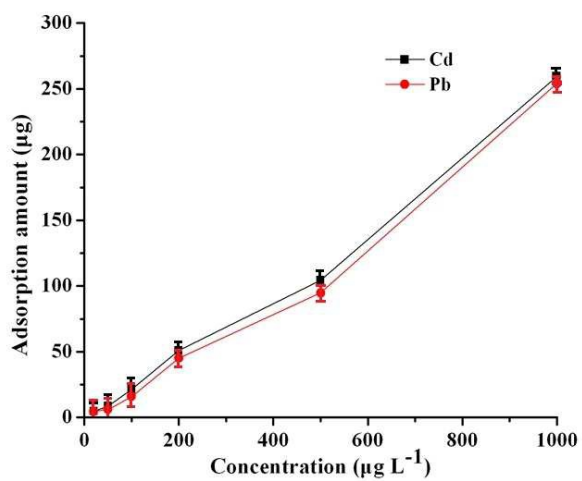
**Figure 2** EDX images of cellulose filter paper and TiO<sub>2</sub>/cellulose paper

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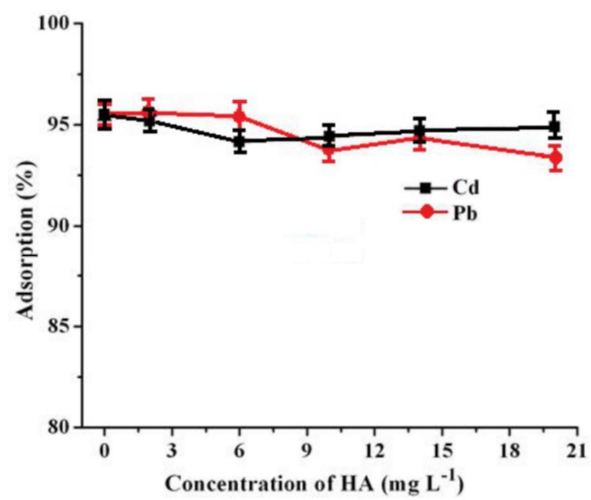
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**Figure 3** Adsorption capacity of Pb(II) and Cd(II) by TiO<sub>2</sub>/cellulose paper



251

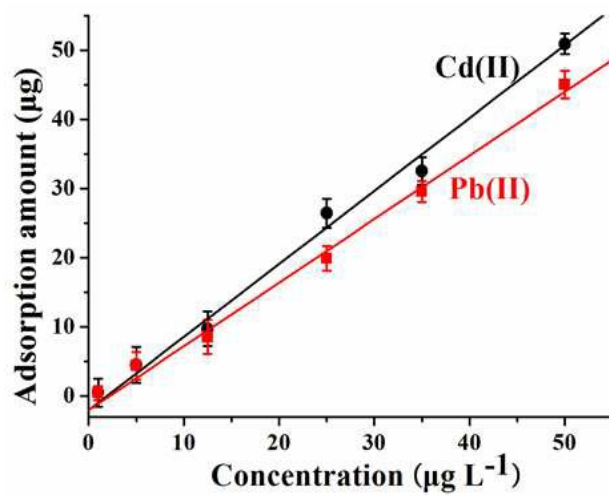
252

**Figure 4** Effect of humic acid on the adsorption of Cd (II) and Pb (II)

253

( $C_{\text{metal}}=200 \mu\text{g L}^{-1}$ ,  $\text{pH}=8.0$ ;  $t=3 \text{ h}$ ;  $V=1.0 \text{ L}$ )

254



255

256

**Figure 5** Calibration curves of Cd (II) and Pb (II)