RSC Advances

This is an *Accepted Manuscript*, which has been through the Royal Society of Chemistry peer review process and has been accepted for publication.

Accepted Manuscripts are published online shortly after acceptance, before technical editing, formatting and proof reading. Using this free service, authors can make their results available to the community, in citable form, before we publish the edited article. This *Accepted Manuscript* will be replaced by the edited, formatted and paginated article as soon as this is available.

You can find more information about *Accepted Manuscripts* in the Information for Authors.

Please note that technical editing may introduce minor changes to the text and/or graphics, which may alter content. The journal's standard Terms & Conditions and the Ethical quidelines still apply. In no event shall the Royal Society of Chemistry be held responsible for any errors or omissions in this *Accepted Manuscript* or any consequences arising from the use of any information it contains.

www.rsc.org/advances

AL SOCIETY
Chemistry

Journal Name

ARTICLE

Received 00th January 20xx, Accepted 00th January 20xx

DOI: 10.1039/x0xx00000x

www.rsc.org/

Fabrication of laminated and coated Nafion 117 membranes for reduced mass

transfer in microbial fuel cell

Vikash Kumar, Piyush Kumar, Arpita Nandy, Patit P. Kundu*

Abstract

Sulfonation of polyvinylidinefluoride (PVdF) with chlorosulfonic acid for 2 hours revealed a respective 33% degree of sulfonations (DS) in SPVdF membrane. The resulting sulfonated PVdF resins were used for Nafion-117 modification as coat and laminating material, where the modified Nafion-117 membranes (laminated and coated with SPVdF) were used as polymer electrolyte membranes in microbial fuel cell (MFC). Coating/lamination exhibited reduced oxygen diffusion across membranes by magnitude of less than two orders over pristine Nafion-117 membrane. This resulted in higher open circuit voltages (OCV) with increased coulombic efficiency (CE) in MFCs. With SPVdF coated and laminated nafion117 membranes, a respective IEC values of 0.57 and 0.46 meq g^{-1} and proton conductivity of 5.91×10⁻³ and 5.11×10⁻³ Scm⁻¹ were observed, indicating a maximum power and current density of $446.45 \pm 21 \text{mWm}^2$ & $1721.78 \pm 86 \text{mAm}^2$ and $413.79 \pm 21 \text{mWm}^2$ 20 mWm⁻² & 1657.57 \pm 82mAm⁻² in MFCs using mixed *firmicutes* as biocatalysts. The obtained coulombic efficiencies were higher (approx. 1-2%) over pristine Nafion 117, which were indicative of its reduced oxygen diffusion at anode. In effect, the study enumerates the efficiency of modified Nafion 117 with sulfonated PVdF coated and laminated membranes as separating barrier in single chambered MFCs for microbial bio-energy conversion.

INTRODUCTION

Microbial fuel cell (MFC) is a bacteria catalyzed electrochemical green energy device that not only reduces fossil fuel crisis, but also harnesses available bio-energy from the given substrate.^{1, 2} In the early 1980s, Bennetto et al were

one of the first groups which exploited microbial electrochemical potential in MFC researches.^{3, 4} This involved microbial substrate oxidation that generated potential at anode, ensuing liberation of protons and electrons in the system. In principle, electrons move towards cathode through

ARTICLE Journal Name

an external circuit, indicating current flow where, proton gets simultaneously transferred via polymer electrolyte membranes (PEM). This completes the circuit and ultimately allows conversion of chemical energy into electrical energy via microbes.

PEM serves as a decisive factor in MFCs, where, it alone contributes to more than 38% of the overall expense of the system.⁵ One good example is Nafion® that has been used widely because of its excellent chemical stability and higher proton conductivity. Relatively, in order to reduce the high cost, several other materials and its optimization have been conducted e.g., polystyrene, polyether ether ketone (PEEK), poly (arylene ether sulfone), phenylated polysulfone, polyphosphazenes, polyimides, polybenzimidazole(PBI) and polypropylene (PP) .⁶⁻⁹ As an alternate, membranes like sulfonated polystyrene-ethylene-butylene-polystyrene (SPSEBS) and sulfonated polyether ether ketone (PEEK) have been reported to be producing respectively approx. 106.9% and 55.2% higher power density over Nafion 117membrane. $10,11$

Recently, polyvinylidinefluoride (PVdF) and its co-polymer have been reported to enhance membrane property in Nafion. Its application as a blend/composite, coating and/or laminate material has been widely reported.¹²⁻¹⁶ Having a similar fluorinated-carbon backbone structure as Nafion, numerous studies on sulfonation, IPNs and varied blend compositions have been reported to enhance membrane's intrinsic properties.17-24 Generally, any ion permeable material with least mass transfer can function as efficient barrier in fuel cells. Nafion with its high cost and increased diffusion coefficient adds as a major drawback in fuel cell operation.25,26 This result in increased cross potentials in Nafion, which on coating/laminating with any mildhydrophobic polymer could be minimized to some extent. In a related study, Kim et al showed PVdF as a laminate material and concluded that the PEM consisting of 20% PVdF exhibits better cell performance at higher temperature $(>100^{\circ}C)^{16}$ Further, these laminated/coated membranes resulted in better membrane properties over their blend compositions in DMFC, due to their excellent inbuilt pore blocking efficiencies in the system.^{27, 28} However, being extremely hydrophobic, PVdF also reduces the resulting ion exchange and proton conductive capacity of the membranes. To minimize the effect, dehydrofluorination of PVdF with H_2SO_4 doping has been shown to enhance its ion conductivity as comparable to Nafion-115.²⁹ Further, electrophilic substitutions induced by sulfonation could also enhance PVdF intrinsic properties with adequate mechanical and chemical stability for membrane preparation.²⁴

Based on the above ideas, we conducted a) the sulfonation of PVDF (SPVDF) with chlorosulfonic acid and b) used it as coat and laminate over Nafion 117 membrane, to study its effect on membrane water uptake, mass transfer, ion exchange and proton conductive capacities. Further, the casted SVPDF coated/laminated membranes have been analyzed in single chambered open air MFCs for bio-energy generation, using mixed electrogenic *firmicutes* as biocatalysts.

RESULT AND DISCUSSION

Fourier Transform Infrared (FTIR) analysis

Sulfonation of polyvinylidinefluoride (PVdF) was carried out with treatment of chlorosulfonic acid for 2 hours. The structural identities of various functional groups were characterized by FT-IR spectroscopy. Figure S1 (Supplementary Information) features the major peak differences in casted pure and sulfonated PVdF membranes. The corresponding IR peak intensities at 1164 cm^{-1} and 1399cm-1 were assigned to the symmetrical and asymmetrical stretching of the constituent $S=O$ bonds of $-SO₃H$ groups. Another peak at 1070 cm^{-1} revealed the presence of pendent sulfonic groups in the membrane structure. Relatively, the characteristic peak intensities at $3200-3390$ cm⁻¹ were observed in sulfonated membranes, corresponding to –OH group stretching. Since, all these peaks were found absent in pure PVdF, the latter ensured the effect of sulfonation on PVdF membrane structure. The peak intensities near 1458 cm-1 were found unchanged for both pure and sulfonated PVdF membranes, where, relatively different prominent peak intensities in S-PVdF ensured the incorporation of sulfonate groups within PVdF structure. The particular peak areas at 1164 cm^{-1} were used for comparing the obtained area (X) in sulfonated PVdF with that of pure PVdF membrane (Y). From the obtained X:Y ratio, the degree of substitution (DS) were calculated for the samples. In effect, with membrane darkening, the peak intensities revealed a maximum of 33% degree of substitution (DS) in sulfonated PVdF membrane, as obtained in earlier studies.³⁰

Water uptake and swelling ratio analysis

Using eq.1, water uptake (WU) values for the samples were obtained. Figure 1 depicts the respective water uptake values of the membranes. The sulfonation of the otherwise hydrophobic PVdF, showed subsequent increase in hydrophilicity, where increasing degree of sulfonation $(\sim$ 33%) indicated relatively an enhanced 8.7% water uptake in SPVdF membranes. Water absorption prevailed via hydrogen bond interactions between surface $-SO₃H$ groups and water molecules. As a result, the water retention capacity increased within sulfonated PVdF membranes.

Fig 1: Water uptake and swelling ratios of membranes Enhanced water uptake capacities were observed in Nafion 117 membranes with an approximate value of 17.1%. In cases of SPVdF- coated and laminated Nafion117 membranes, a respective water uptake of 14.2 and 12.6% were observed. These marked reductions were due to the incorporated SPVdF coats and laminates over Nafion117 membrane that resulted in less available sulfonic acids in the corresponding Nafion117 membranes to interact with water molecules. The

and Nafion117 membranes.

additional layer of SPVdF polymer constricted the straight meq g⁻¹ were observed from SPVdF, SPVdF laminated/coated channels in Nafion that slightly lowered its water affinity in the membranes.

A direct correlation existed between DS and water retention capacity of SPVdF membranes, where 2 hours sulfonation directly influenced the membrane structure with increased sulfonic group incorporation in the membranes. To rationalize the results, swelling ratio of the membranes were analyzed using eq. 2. This followed the same trend as that obtained for the water uptake analysis. A respective swelling ratio (SR) of 3%, 5.7%, 7.1% and 9.3% were observed in SPVdF, SPVdF laminated Nafion117, SPVdF coated Nafion117 and pristine Nafion117 membranes. The decline in the swelling ratios of SPVdF coated and laminated membranes, compared to pristine Nafion117 were due to the same aforementioned reason that reduced the water uptake capacities in the membranes. Nevertheless, reductions in both the parameters (water uptake and swelling ratios), obtained for SPVdF coated and laminated membranes, were considered better for reduced mass transfer over naive Nafion 117 membrane.

Analysis of IEC and proton conductivity

Membrane ion-exchange capacities (IEC) were calculated by displacing the initially attached/ incorporated ions by an oppositely charged ion present in the surrounding solution.³¹ Figure 2 illustrates the IECs obtained for the respective SPVdF, SPVdF laminated Nafion117, SPVdF coated Nafion117 and pristine Nafion117 membranes. A respective values of 0.13 meq g^{-1} , 0.46 meq g^{-1} , 0.57 meq g^{-1} , and 0.081

Fig 2: Ion exchange and proton conductive capacities of casted membranes

The values clearly signified the effect of 33% sulfonated PVdF incorporation as coat and laminate that showed approximately 30-45% reduced IEC over pristine Nafion117 membrane. With constriction, reduced $-SO₃H$ groups and lower water interaction, directly hindered the resulting IECs in SPVdF coated and laminated membranes. Similarly, the differences in IECs correspondingly influenced the proton conductivity of the membranes. Significant differences in

membrane proton conductivity (from 10^{-1} to 10^{-4} range) has been has been reported, indicating the effect of membrane nano-filler modification and compressions on transversal ion exchange. 32 Corresponding to that, a respective proton conductive values of 1.01×10^{-4} , 5.11×10^{-3} , 5.91×10^{-3} and 3.03×10^{-3} Scm⁻¹ were observed in SPVdF, SPVdF laminated Nafion117, SPVdF coated Nafion117 and naive Nafion117 membranes. The incorporation of SPVdF as coats and laminates reduced the availability of surface $-SO₃H$ groups within Nafion117 membranes. This, eventually aided in lowering of the transverse protonic conduction across SPVdF coated and laminated membranes. Nevertheless, it was expected as SPVdF layers on Nafion 117 membrane ensures the constriction of water channels, and thus, resulting in hampering the cross sectional ionic mobility of hydrogen ions in the form of hydronium ions. In effect, the obtained IEC and proton conductivity of SPVdF coated and laminated membranes corresponded towards its decreased WU and SR over Nafion117 with limited availability of free $-SO₃H$ groups, as observed earlier.

Oxygen diffusivity across membranes

Oxygen diffusivity across the membranes was analyzed, indicating the periodic variations in the dissolved oxygen content at anode. The increasing oxygen flux from cathode to anode ensued a respective mass transfer coefficient of 2.7 × 10^{-4} cm s⁻¹, 6.27×10^{-3} cm s⁻¹ and 8.73×10^{-2} cm s⁻¹ for Nafion 117, SPVdF-coated and laminated Nafion 117 membranes (Figure 3). Membrane compactness directly influences the mass transfers where, a similar study showed a

respective mass transfer values of \sim 5.0 \times 10⁻⁵ cm s⁻¹, 7.5 \times 10⁻⁵ ⁴ cm s⁻¹, 290 \times 10⁻⁵ cm s⁻¹ and 9.4 \times 10⁻⁵ cm s⁻¹ in glass fibers $(0.1-0.4, J$ -cloth and CEM as separators.³³ Membrane porosity was observed as the crucial factor in deciding the overall oxygen diffusion in the cell. Here, a direct relation with available sulfonated groups and oxygen permeability was observed. Nafion 117 with increased water upatke (-17.1%) , showed higher oxygen pervasion in comparison to SPVdFcoated and laminated membranes. The reason was attributed to the reduction of straight water channels due to blocked

pores by means of lamination and coating on Nafion membrane. In contrast, SPVdF-coated and laminated membranes with reduced pores and surface available $-SO_3H$ groups, showed limited mass transfer with an approximate 17% and 31% lower oxygen diffusions over pristine Nafion

117 membrane.

Fig 3: Oxygen diffusivity across the membranes

Increased mass transfer limits the overall cell efficiency, as direct anolyte oxidation generates cross-potential in the system that in turn, results in energy loss and limited energy

recovery from the system. SPVdF soaking and compression over Nafion117 membrane as coat and laminate showed least mass transport, which were considered as efficient qualities to be applied in MFCs for further analysis.

Open circuit voltages (OCVs)

Three fed batch MFCs were maintained under identical experimental conditions. The air facing side of cathode was kept covered with parafilm without employing any external load initially. This allowed favourable start-up conditions in MFCs, ensuing increased potentials at respective anode and cathode terminals. With random fluctuations, an average anode and cathode potentials of -225 mV and +171 mV were observed in MFCs using Ag/AgCl as reference electrode (data not shown). Ensuing stabilization, gradual increments in MFC open circuit voltages (OCV) were observed. A respective maximum OCV of 648±20 mV, 702±15mV and 748±12mV were observed in MFC-A, B and C, indicating the developed potentials at both ends (Figure 4).

Fig 4: Open circuit potential of MFCs

The corresponding lower OCV in MFC-A was attributed to the fitted Nafion 117 membrane, where its higher mass transfer resulted in direct anolyte exhaustion, leading to cross potential generation in the system. Further, this resulted in cell flooding and enhanced oxygen stoichiometry at anode that overall allowed lower OCVs in the system due to increased oxygen permeation. 34 In contrast, MFC-B and C (with SPVdF-coated and laminated) were observed with higher OCVs. Comparatively, the lower mass transfer and constricted inter-space volume in SPVdF-coated/laminated membranes showed increased OCVs over Nafion 117(in MFC-A).

Electrochemical impedance spectroscopy (EIS) analysis

Two probe configurations were employed for measuring the internal resistance (R_{in}) in MFCs.³⁵ Anode was considered as working electrode, whereas cathode was connected to the reference and counter terminals. The obtained Nyquist graphs were used to calculate the specific resistive components like activation resistance, ohmic resistance (Rm- attributed to electrode resistance, membrane resistance, etc.) and concentration resistance of the whole unit (Figure 5).

Fig 5: Electrochemical impedance spectroscopy (EIS) of MFCs

Ohmic resistances (Rm), as calculated from the equivalent circuit (S2, supplementary information), were observed as \sim 5.7Ω, \sim 9.42Ω and \sim 14.7Ω for MFC-A, B and C respectively. Minimal ionic resistance with fitted Nafion 117 was observed in MFC-A, which was mainly due to its increased proton conduction and ion exchange capacity. Further, higher water retention in Nafion 117 allowed increased transverse ionic conduction across MEA. Significant increments in ohmic resistances were observed in MFC-B and C, revealing their higher membrane impedance with increased charge transfer hindrances. Nevertheless, it was expected, as constricted inter-space volume in SPVdF-coated and laminated membranes allowed least proton and ionic conduction, relative to higher charge transfer resistance. Comparatively, this higher internal resistance in SPVdF-coated and laminated membranes provided increased ionic hindrance in MFC-B and C, ensuing in their lower performance over naive Nafion117 membrane.

Polarization

Despite of reduced OCV, MFC-A showed enhanced cell efficiency over other employed units, with increased voltage drops and currents. A respective maximum current of 1.07 mA, 1.03mA and 0.99mA were observed from MFC-A, B and C, indicating Nafion 117 progressive hike over SPVdFcoated and laminated membranes (S3, Supplementary Information). The reason was attributed to the higher proton and ionic conductivity of Nafion 117 that aided in enhanced MFC-A performance. Applying multiple resistances $(10⁷ - 10)$ $Ω$), polarization curves for individual MFCs were obtained (Figure 6). Here, MEA was an inclusive factor, where close electrode spacing resulted in increased cell efficiency that in turn, reduced the overlaying electrolyte resistance between the electrodes.³⁶

Fig 6: Polarization curves of MFCs (v-voltage, p-power density)

At higher resistances, frequent voltage drops were observed, indicating the increased activation losses in the system. This was indicative of the energy lost in initiating the charge transfer redox reaction, from microbes to the anode surface. In addition, higher voltage drops were observed at lower external resistances, indicating the electron flow ease within the circuit. In effect, MFC-A (with Nafion117) showed a highest power and current density of 481.55 ± 27 mWm⁻² and 1788.18 \pm 84 mAm⁻² at 251 Ω. Comparatively, a respective power and current density of 446.45 ± 21 mWm⁻² & 1721.78 \pm 86mAm⁻² and 413.79 \pm 20 mWm⁻² & 1657.57 \pm 82mAm⁻² were observed from MFC-B and C. In overall, the relative higher polarity, IEC and proton conductivity of Nafion117, showed an approximate 7% and 14% enhanced MFC-A performance over MFC-B and C. Although, the differences

were marginal, the results indicated the effectiveness of SPVdF coating/lamination over Nafion 117 membrane. The obtained power output was compared with other relevant studies, to indicate the effectiveness of SPVdF coated/laminated membrane in MFC (T1, Supplementary Information). Different lower cost materials (e.g., SPEEK/PES, sulfonated PE/ poly styrene-DVB) showed reduced power densities with respect to SPVdF coated/laminated membranes used here, however, other parameters like anolyte, microbes and electrode configurations (such as MEAs) were altogether different in these cases. Nevertheless, the reduced mass transfer in SPVdF coated/laminated membranes could well be enumerated as efficient membrane property that showed increased OCV with relatively comparable (with \sim 7-14% lower) performance with respect to pristine Nafion 117 membrane.

ARTICLE Journal Name

were observed. Electron transfer from biofilm to electrode indicated microbial oxidation, whereas reduction peaks corresponded towards the microbial reduction (charge transfer from electrode to biofilm). This redox activity was attributed to the microbial cell surface proteins that evidently ensured its biocatalytic activity, resulting in subsequent substrate utilization from the employed *firmicutes* on repeated potential cycling (S4, Supplementary Information).

Fig 7: SEM images of bio-films at electrode surface.

Bio-electrochemical analysis

EPS (exo-polysaccharide) present on the microbial cell wall plays crucial role in its surface attachment as support. This thereby results in biofilm formation that electro-actively oxidizes substrate and allows potential development at anode. Figure 7 shows the SEM images of predominant microbial colonies associated at electrode's surface (carbon cloth fibers), which were evidently engaged in the electrochemical oxidation of substrates. Cyclic voltammogram (CV) analysis, showed prominent redox activities at 856.5 mV (at 10.79 μ A) vs. ref as oxidation peak), with two distinct reduction peaks at 256.6 mV and -236.7 mV vs. ref at -10.7 µA and -7.7 µA

Substrate removal analysis

To analyze the relative substrate exhaustions (as anolyte), chemical oxygen demand (COD) removals were measured periodically in the system. Figure 8 depicts the respective % COD removals in MFCs in 30 days run. A respective COD removal of \sim 93.97%, \sim 90.27% and \sim 84.15% were observed in MFC-A, B and C, where these variations were primarily indicative of the employed membranes in the unit. Increased substrate exhaustion in MFC-A corresponded with its enhanced cell performance, as Nafion 117 bestowed highest polarity amongst all fitted membranes.

Fig 8: COD removal from MFCs

However, despite of its relatively enhanced proton conductive and ion exchange capacity, lower coulombic efficiency (CE) was observed in MFC-A over SPVdF-coated and laminated membranes (Figure 9).

The reason was the reduced oxygen diffusion in SPVdFcoated and laminated membranes that resulted in higher coulombic efficiencies in MFC-B and C. Generally, COD removal is largely influenced by the employed microbes and

it is necessary to keep them electrogenetically active without oxygen intrusion. Here, air cathode influenced the cell efficiency with configured cell electrodes, as MEA reduces the cell overall impedance with sustained energy recovery from the system. $37, 38$ Direct substrate exhaustion showed higher COD removal, but lower CE in Nafion 117, establishing an inverse relation between COD removal and CE in MFCs. A clear reduction of approx. \sim 1- 2% CE was observed in Nafion 117 over SPVdF coated and laminated Nafion 117 membranes. The generated cross potentials showed lower CE in MFC-A with relatively higher COD removal because of its increased mass transfer through Nafion 117 membrane. In contrast, SPVdF-coated and laminated Nafion 117 membranes with lower oxygen diffusion showed higher coulombic efficiency (CE) over pristine Nafion 117 membrane. Minimal ionic conduction resulted in lower current extracts with higher cell impedance in MFC-B and C but, the reduced oxygen diffusion across SPVdF-coated and laminated membranes enhanced its OCV and relative CE, in comparison to Nafion 117 membrane.

Microbial sustenance gets adversely affected with other fermentative and methanogenic reactions that predominate in the system. This further aids in limiting microbial metabolism with increased oxygen diffusion.³⁹⁻⁴³

Nevertheless, microbial adequacy with successive COD removals corresponded with the energy performance of the units, where SPVdF laminated/coated Nafion 117 membrane showed highest energy recovery (in terms of CE) from the system. In addition, this lower mass transfer and comparable power generation ability enabled them as promising

ARTICLE Journal Name

coat/laminate separator for harvesting increased energy from bio-electrochemical systems.

MATERIALS AND METHODS

General Conditions

All chemicals used were of analytical and biochemical grade. PVdF (Avg. M_w -530,000) and Nafion resin were obtained from Sigma Aldrich. Chlorosulfonic acid was bought from Merck Millipore, India. Under strict sterile conditions, all microbial experiments were performed.

Preparation of membranes

PVdF Sulfonation: PVdF granules (3 gm) were sulfonated using chlorosulfonic acid as sulfonating agent at 60°C for 2 hrs.³⁰ The resulting dark brown pellets (on sulfonation) were neutralized with 1, 2 dichloroloethane and rinsed thoroughly with de-ionized water. Further, this was dissolved at NMP at 80°C and casted as sulfonated PVdF (SPVdF) membranes.

The obtained pure and sulfonated SPVdF membranes were subjected for Fourier transform infra red spectroscopy (FT-IR), for reference. Degree of substitution (DS) as calculated from IR peak intensities were calculated from the characteristic x: y peak ratios. 24

Coating of SPVdF on Nafion 117 membrane: The dissolved sulfonated PVdF solution was used to coat Nafion 117 membrane. Nafion 117 was dipped in viscous SPVdF solution for 15 minutes, followed by air drying. This was repeated 4-5 times and further allowed to soak overnight at 80°C.

Nafion 117 lamination: The obtained sulfonated PVdF solution (10% w/v each) were used to cast two $\sim 50 \mu m$ thick SPVdF membranes. Further, Nafion 117 were sandwiched between the casted SPVdF membranes and hot pressed at 130°C under 30 Mpa pressure for 3 mins. The resulted in a compressed SPVdF laminated Nafion117 membrane of \sim 213 µm thickness. Further, these membranes were subjected for water uptake, ion exchange and proton conductivity measurements as described earlier.⁴⁴

Water uptake and swelling study

Small piece of membranes were used for water uptake (WU) and swelling ratio (SR) analysis. Water uptakes and swelling ratios were measured using the following equations:

Water uptake $(\%)=(W_{wet} - W_{dry}) (100)/W_{dry}$ (1)

Swelling Ratio (
$$
\degree
$$
) = $(T_{wet} - T_{dry}) (100)/T_{dry}$ (2)

Where, W_{wet}/T_{wet} represented the wet weight/thickness of the membranes after soaking in water for 24 hrs, and $W_{\text{dry}}/T_{\text{dry}}$ were the respective dry weight/thickness of the membranes.

Ion exchange capacity (IEC)

Using the conventional titration method, ion exchange capacities (IECs) of the membranes were determined. Membranes were allowed to soak in 1 M $H₂SO₄$ solution overnight and later rinsed with DI water. The samples were again soaked in 50 mL (1M NaCl solution) overnight, in order to allow replacement of protons with sodium ions. The remaining solution was titrated with 0.01 N NaOH solution, using phenolphthalein as indicator. The IEC value (in meq g-¹) was calculated using the following equation:

$$
IEC = (VNaOH) (SNaOH)/Wdry
$$
 (3)

Where, V_{NaOH} is the volume of NaOH used in the titration, and W_{dry} is the dry weight of the membrane in gm. S_{NaOH} is the strength of NaOH used in the experiment for determination of IEC.

Proton conductivity

To measure the conductivity of membrane, AC impedance spectroscopy was employed in transverse direction at a frequency range of 1 Hz to 10^5 Hz of 10 mV amplitude (Gamry Reference-600) (S5, Supplementary Information). The conductivities of the samples (σ) were calculated from the nyquist data, using the following equation:

$$
\sigma = T/R.A \tag{4}
$$

where, T is the thickness of the sample, A is the crosssectional area of the sample, R is the resistance derived from the lower intercept of the high frequency semi-circle on a complex impedance plane with the real (Z) axis.

Cell configuration

Membrane electrode assemblies (MEAs): For MEA preparation, carbon cloths (6 cm^2) (Zoltek pvt. Ltd, USA) were used as electrodes, where the membranes (Nafion 117, SPVdF coated and SPVdF laminated Nafion 117) were sandwiched between carbon cloth electrodes (as anodes and cathodes). These assemblies were further hot pressed in compression moulding at 130 °C for 25 seconds at 6.84 MPa pressure. This condition was optimized for perfect assembling, as on higher ranges, the fabric became brittle and carbon cloth lost its texture. Further, a total of 3 mgcm-2

catalyst mixture of 10:90 wt% (Pt/C) with 5% Nafion resin was loaded on the cathode side of the carbon cloth by paint coating.

MFC Fabrication: Three single chambered identical MFC units of 150mL liquid volume (anode chamber) containing Nafion 117, SPVdF coated and laminated Nafion 117 membranes as MEAs were named as MFC-A, B and C respectively (S6, Supplementary information). For oxygen reduction, the catalyst loaded air facing side of cathode was kept facing outward. Other requisite fabrications e.g. inlet/outlet sealing, electrode fixing, electrical connections etc were done accordingly.

Oxygen Diffusivity Measurement: A dissolved oxygen probe (Horiba Pvt. Ltd, Kyoto Japan) was fixed in the anode chamber. In order to remove the dissolved oxygen content at anode, purified N_2 gas was purged in the chamber. The mass transfer coefficient k (cm s^{-1}), as characterized by oxygen diffusivity was calculated from cathode to anode chamber over time, using the mass balance equation

$$
k\frac{1}{4} = -V/At \ln [C_s - C/C_s]
$$
 (5)

Where, V is the anode chamber volume, A is membrane cross-sectional area, C is the anode oxygen concentration, and C_s is cathode oxygen concentration (assumed to be the saturation concentration of oxygen in water, or 7.8 mg L^{-1}). Oxygen diffusivity was recorded periodically (in triplicate) to monitor the diffused oxygen content in the chamber.

Microbes

DNA isolation and PCR amplification: Microbial genomic DNA isolation was done using standard phenol-chloroform method.⁴⁵ Using universal primers Y1Forward (40th) 5'- TGGCTC AGAACGAACGCGGCGGC-3' and Y2Reverse (337th) 5'-CCCACTGCTGCCTCC CGTAGGAGT-3' 16S rRNA gene amplification was conducted in polymerase chain reaction (PCR) (Applied biosystem, US). For microbial identification, sequencing and BLAST tools were employed to allot accession numbers to the strains (EMBL accession no. HE648059, HE648060, HF548664). These were *Firmicute* class *lysinibacillus* species that were found facultative anaerobes when tested for viability in gas pack jar.

Anolyte Feed: Mixed microbial strains were suspended in 50 mM phosphate buffer (50ml volume) and subsequently transferred to 100 ml synthetic wastewater (pH~6.9). The COD composition of the feed wastewater was 1500±210 mg l⁻¹ (total nitrogen: 117±24 mg l⁻¹, PO₄-P: 27±3 mg l⁻¹, MgSO₄:45 mg l⁻¹). A final volume of 150 ml (with microbe) anolyte was used as feed in MFCs.

Start-up and measurements

Initially, MFC units were assembled with sterilized deionized water at anode. This was later replaced with anolyte using peristaltic pump. Using multimeter (Keithley Instruments, Cleveland, OH, USA), and a potentiometer (G600; Gamry Instrument Inc., Warminster, PA, USA), connected to a personal computer, periodic monitoring of MFCs (at 24 h intervals) were conducted. Current (I) and

ARTICLE Journal Name

potential (V) measurements were recorded after allowing the circuit to stabilize for 8-10 minutes. Power (W) was calculated using relation P=IV, where I and V represented the recorded respective current and voltages. Normalizing the power with anode surface area (6 cm^2) , power densities $(mW/m²)$ of the respective MFCs were calculated.

Substrate removals were analyzed by periodically measuring the chemical oxygen demand (COD) at 420 nm (Anatech Labs India Pvt. Ltd.) in triplicate. The coulombic efficiency (CE) of the fed-batch MFCs were calculated from the following equation:

$$
CE = M \int_0^t I dt / FbV_{an} \Delta COD \tag{6}
$$

where 'M' represents the molecular weight of $oxygen(M =$ 32), 'F' is Faraday's constant, 'b' denotes the number of electrons exchanged per mole of oxygen ($b = 4$), 'V_{an}' is the liquid volume in anode, and ∆COD is the change in the chemical oxygen demand over time t.⁴⁴

Electrochemical impedance spectroscopy (EIS)

In order to measure the internal resistance of the units, potentiostatic EIS at a frequency range of 10^3 kHz to 1 mHz were performed employing 10 mV AC voltage. The internal resistances (R_{in}) were determined from the plotted Nyquist graphs of all MFCs.³⁵

Cyclic Voltammetry (CV)

To assess microbial electrochemical activity, cyclic voltammograms at a scan rate of 1-2mV/s were analyzed (G600; Gamry Instrument, USA). Three electrode setup

containing, anode as the working, cathode as the counter, and Ag ⁄ AgCl as reference electrode were used. Reference electrode (Ag ⁄ AgCl) was placed near anode terminal to minimize iR drop at solution-electrode interface. To ensure microbial activity, control experiments (without microbes) were conducted separately.

Scanning Electron Microscopy (SEM)

For SEM analysis, biofilms were fixed on the electrode using 2.5% gluteraldehyde with 0.1 M phosphate buffer solution. Subsequently, further dehydration with 30% - 100% ethanol was done following sputter coating the samples under vacuum with thin gold layer.⁴⁴ To analyze microbial adhesion at electrode's surface, scanning electron microscope (Carl Zeiss EVO® 18 electron microscope) were employed with an acceleration voltage of 15 kV.

CONCLUSION

In summary, modification of Nafion 117 with sulfonated PVdF membranes as coat and laminates were conducted in order to minimize the increased mass transfer across pristine Nafion membrane. In comparison to Nafion 117, approximately ~31% reduced oxygen diffusions in coated/laminated membrane were observed. When compared as MEA, this reduced mass transfer contributed towards increased OCVs in MFCs using mixed electrogenic *firmicutes* as biocatalysts. However, the power outputs were slightly lower which was obvious due to the increased polarity of naive Nafion 117 that ensued minimal ionic hindrance but, higher oxygen diffusion over coated/laminated SPVdF

membranes. Comparing the performance, approximately 7- 14% reduced power outputs were observed in SPVdF coated/laminated membranes over pure Nafion membrane. Despite these reductions, SPVdF coated/laminated membranes showed increased couloumbic efficiencies (approx ~1-2%) over Nafion 117 membrane. The reason was attributed to their reduced mass transfer that prevented direct anolyte exhaustion at anode. In effect, with reduced cross potentials, a maximum power and current density of $446.45 \pm$ $21 \text{mWm}^2 \& 1721.78 \pm 86 \text{mAm}^2$ and $413.79 \pm 20 \text{ mWm}^2 \&$ 1657.57 ± 82 mAm⁻² were observed in SPVdF coated/laminated membranes, indicating its efficiency as comparable to pristine Nafion 117 membrane in usage. MFC being a future application, demands more profound investigations in such diversified areas of membrane technology for practical higher efficient alternatives.

ACKNOWLEDGEMENTS

The financial research aid provided from Council of Scientific and Industrial Research (CSIR) and Department of Science and Technology (DST), Govt. of India is duly acknowledged.

Notes and References

Notes

*Advanced Polymer Laboratory, Department of Polymer Science &Technology, 92 A. P. C. Road, University of Calcutta, Kolkata-700009. *Corresponding author Fax & Tel: 91-33-2352-510; E-mail: ppk923@yahoo.com* Authors declare no competing financial interest.

ARTICLE Journal Name

References

- 1. D. L. Hoskins, X. Zhang, M. A. Hickner and B. E. Logan, Bioresour. Technol., 2014, 172, 156–161
- 2. Y. Hou, K. Li, H. Luo, G. Liu, R. Zhang, B. Qin and S. Chen, Environ. Sci. Eng., 2014, 8, 137–143.
- 3. H.P. Bennetto, J.L. Stirling, K. Tanaka and C.A. Vega, Biotechnol Bioeng, 1983, 25, 559-568.
- 4. R.M.Allen and H.P.Bennetto, Appl Biochem Biotechnol, 1993, 39, 27-40.
- 5. M. Ghasemi, W.R.W. Daud, S.H.A. Hassan, S. E. Oh, M. Ismail, M. Rahimnejad, J.M. Jahim, J. Alloys Compd., 2013, 580, 245-255.
- 6. H. Zhang and P. K. Shen, Chem. Rev., 2012, 112, 2780– 2832.
- 7. J. Yang, P. K. Shen, J. Varcoe and Z. Wei, J. Power Sources, 2009, 189, 1016–1019.
- 8. L. A. Diaz, G. C. Abuin and H. R. Corti, J. Membr. Sci., 2012, 411–412, 35–44.
- 9. L. A. Neves, J. Benavente, I. M. Coelhoso and J. G. Crespo, J.Membr. Sci., 2010, 347, 42–52.
- 10. S. Ayyaru, P. Letchoumanane, S. Dharmalingam and A. Stanislaus, J. Power Sources, 2012, 217, 204–208.
- 11. S. Ayyaru and S. Dharmalingam, Bioresour Technol, 2011,102, 11167–11171.
- 12. K. Y. Cho, J. Y. Eom, H. Y. Jung, N. S. Choi, Y.M. Lee, J. K. Park, J. H. Choi, K. W. Park and Y. E. Sung, Electrochim Acta, 2004, 50, 583–588.
- 13.J. Wootthikanokkhan and N. Seeponkai, J. Appl. Polymer Sci., 2006, 102, 5941–5947.
- 14. W.H. Choi and W.H. Jo, J. Power Sources, 2009, 188,127–131.
- 15. K.Y. Cho, H.Y. Jung, K.A. Sung, W.K. Kim, S.J. Sung, J. K. Park, J.H. Choi and Y.E. Sung, J. Power Sources, 2006, 159, 524–528.
- 16. H. J. Kim, H. J. Kim, Y. G. Shul and H. S. Han, J. Power Sources, 2004, 135, 66–71.
- 17. K. Dutta, S. Das, P. Kumar and P. P. Kundu, Appl. Energy,2014, 118, 183–191.
- 18. C. W. Lin, R. Thangamuthu and C. J. Yang, J. Membr. Sci.,2005, 253, 23–31.
- 19. P. Kumar, S. K. Jagwani and P. P. Kundu, Materials Today Communications, 2015, 2, e1–e8.
- 20. F. Liu, N. A. Hashim, Y. Liu, M. R. M. Abed and K. Li, J.Membr. Sci., 2011, 375, 1–27.
- 21. P. Kumar, K. Dutta and P. P. Kundu, Int. J. Energy Res., 2014,38, 41–50.
- 22. P. P. Kundu, B. T. Kim, J. E. Ahn, H. S. Han and Y. G. Shul, J.Power Sources, 2007, 171, 86–91.
- 23. M. K. Song, Y. T. Kim, J. F. Fenton, H. R. Kunz and H. W. Rhee, J. Power Sources, 2003, 117, 14–21.
- 24. S. Das, P. Kumar, K. Dutta and P. P. Kundu, Appl. Energy,2014, 113, 169–177.
- 25. S. Slade, S.A. Campbell, T.R. Ralph and F.C. Walsh, J. Electrochem. Soc., 2002, 149, A1556.
- 26. C. Heitner-Wirguin, J. Membr. Sci., 1996, 120, 1.
- 27. D. Xu, G. Zhang, N. Zhang, H. Li, Y. Zhang, K. Shao, M. Han, C.M. Lew and H. Na, J. Mater. Chem., 2010, 20, 9239-9245.
- **Journal Name ARTICLE ARTICLE**
- Energy, 2010, 35,10482-10488.
- 29. M. K. Song, Y. T. Kim, J.F. Fenton, H. R. Kunz and H. W. Rhee, J. Power Sources, 2003, 117,14-21.
- 30. P. Kumar and P. P. Kundu, Electrochim Acta, 2015, 173, 124–130.
- 31.J.D. Jeon and S.Y. Kwak, J Phys Chem B. 2007, 111, 9437–43.
- 32. S. Moghaddam, E. Pengwang, Y. B. Jiang, A.R. Garcia, D.J. Burnett, C. J. Brinker, R. I. Masel and M. A. Shannon, Nat Nanotechnol, 2010, 5, 230-6.
- 33. X. Zhang, S. Cheng, X. Wang, X. Huang and B. E. Logan, Environ. Sci. Technol, 2009, 43, 8456–8461.
- 34. P. Kumar, K. Dutta and P.P. Kundu, Int. J. Energy Res., 2014, 38, 41-50.
- 35. Z.He and F.Mansfeld, Energy Environ Sci, 2009, 2, 215– 9.
- 36. A. ElMekawy, H. Hegab, X. Dominguez-Benetton and D. Pant, Bioresour Technol. 2013, 142, 672–82.
- 37. V. Kumar, A. Nandy, S. Das, M. Salahuddin and P. P. Kundu, Appl. Energy,2015, 137, 310–321.
- 38. R. Rudra, V. Kumar and P. P. Kundu, RSC.Adv., 2015, 5, 83436-47.
- 39. A. ElMekawy, H. Hegab and D. Pant, Energy Environ Sci. 2014, 7, 3921-33.
- 40. B.Min , J.Kim , S.Oh , J.M.Regan and B.E.Logan, Water Res 2005, 39, 4961–8.
- 41. Z.He, S.D.Minteer and L.T.Angenent, Environ Sci Technol, 2005, 39, 5262–7.
- 28. C. Zhao, H. Lin, Q. Zhang and H. Na, Int. J. Hydrogen 42.J.Greenman, A.Ga´lvez, L.Giusti and I.Ieropoulos, Enzyme Microb Technol, 2009, 44,112–9.
	- 43.J. J. Fornero, M. Rosenbaum, L. T. Angenent, Electroanal, 2010, 22, 832 – 843.
	- 44. V. Kumar, P. Kumar, A. Nandy and P. P. Kundu, RSC.Adv., 2015, 5, 30758-67.
	- 45. A. Ghosh, B. Maity, K. Chakrabati and D. Chattopadhyay, Microb Ecol, 2007, 54, 452-459.

21x10mm (300 x 300 DPI)