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Microwave-assisted synthesis of highly crystalline, multifunctional iron oxide nanocomposites for imaging applications†

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We report a reproducible single-step, microwave-assisted approach for the preparation of multifunctional magnetic nanocomposites comprising superparamagnetic iron oxide (Fe_3O_4) cores, a polyelectrolyte stabilizer and an organic dye with no requirement for post-processing. The stabilisers poly(sodium 4-styrenesulfonate (PSSS) and sodium polyphosphate (SPP) have been thoroughly investigated and from analysis of electron microscopy, dynamic light scattering measurements, magnetic hysteresis and magnetic resonance (MR) imaging, we show that the higher degree of Fe_3O_4 nanoparticle crystallinity achieved with the PSSS stabiliser leads to enhanced magnetic behaviour and thus better contrast agent relaxivity compared to the less crystalline, poorly defined particles obtained when SPP is employed as a stabiliser. We also demonstrate the potential for obtaining a multifunctional magnetic-fluorescent nanocomposite using our microwave-assisted synthesis. In this manner, we demonstrate the intimate link between synthetic methodology (microwave heating with polyelectrolyte stabilizer) and resulting properties (particle size, shape, magnetism) and how this underpins the functionality of the resulting nanocomposites as agents for biomedical imaging.

1 Introduction

The choice of synthetic approach employed for the preparation of nanoparticles is of crucial importance when designing materials for a specific function. Of growing recent interest has been the development of routes to nanoparticles which afford great control over particle shape and composition, vital in our efforts to realise an intimate understanding of their unique properties. One example is the field of magnetic nanoparticles, a class of materials whose size-dependent magnetic properties opens up their potential applications for hyperthermic cancer therapy, site-specific drug delivery and contrast enhancement in magnetic resonance (MR) imaging.[1–11] Control over the magnetic properties is desirable in order to tailor the candidate nanoparticle for a specific biomedical application.[12] Magnetic nanoparticles may be synthesised in a variety of ways,[13–15] including by co-precipitation,[16,17] hydrothermal methods,[18] and the decomposition of precursors at elevated temperatures.[19,20] Aqueous routes, such as co-precipitation, are advantageous in that the particles are prepared in biologically tolerated solvents, but often lead to polydisperse nanoparticles which may display some loss of particle crystallinity.[21] Advances made in the high temperature decomposition of organometallic precursors have led to highly crystalline, monodisperse nanoparticles, with a great degree of control over particle size.[22] These routes, where organic solvents are employed, generate hydrophobic nanoparticles, with additional work-up required to transfer particles to aqueous environments for subsequent biomedical use. A synthetic approach which produces nanoparticles with high crystallinity without the need for surface post-processing to induce hydrophilicity is therefore

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highly desirable.

With these concerns in mind, we have employed microwave-assisted methods for the preparation of functionalised iron oxide nanoparticles, which are becoming increasingly attractive in materials chemistry. Microwave approaches to particle synthesis have led to dramatic decreases in reaction times and greater control over product formation.[23–29] For example, the Niederberger group have developed microwave-assisted methods to prepare a range of nanoparticles of controlled sizes within minutes.[30] Since the first reports of the microwave-promoted hydrothermal synthesis of sub-micron haematite powders,[31] methods have been developed to prepare nanoparticles of a uniform size distribution. Nanoparticles of haematite (α-Fe_2O_3) have been reported from the irradiation of hydrolysed iron salts,[32] while more exotic α-Fe_2O_3 morphologies (cubes, rings and spindles) have been realised by Yu and co-workers by tailoring the reaction conditions to provide thermodynamic control over particle growth.[33,34] Superparamagnetic maghemite (γ-Fe_2O_3) nanoparticles may be prepared through a microwave-treated co-precipitation reaction with a typical particle size of 10 nm obtained.[35] Increases in nanoparticle crystallinity have been observed upon aging under microwave conditions, with post-processing using stearic acid resulting in stable organic ferrofluids.[36] Uptake in endothelial cells of superparamagnetic citrate-coated Fe_2O_3 nanoparticles prepared by microwave-assisted methods have also recently been reported.[37] Variation of reactant concentration in the microemulsion synthesis of Fe_3O_4 under microwave irradiation has resulted in monodisperse composites of variable sizes.[38] Reaction times for the polyol synthesis of cobalt ferrite nanoparticles has been decreased to just one hour by combining microwave heating,[39] while a variety of MFe_2O_4 ferrites (M = Zn, Ni, Mn, Co) have also been reported from microwave-hydrothermal methods.[40]

For their use as imaging agents, tailoring the surface chemistry of magnetic nanoparticles is essential.[22,41–44] For example, Liong *et al.* have reported the preparation of a multifunctional nanocomposite for imaging and targeted drug delivery where iron oxide nanoparticles are phase transferred from organic to aqueous solution before silica coating and co-condensation of a fluorescent moiety render them multifunctional.[45] Recently, Liu *et al.* have reported the use of polyacrylic acid in the synthesis of Fe_3O_4 nanoparticles to achieve tuneable particle cluster sizes of between 100 nm to 400 nm, which display good biocompatability.[46] We have previously reported the use of polyelectrolytes for the *in situ* stabilisation of magnetic nanoparticles during a co-precipitation reaction, which has led to new developments in magnetic fluid preparation.[47–49] Employing polyelectrolyte stabilisers *in situ* during nanoparticle nucleation and growth infers a high degree of stability induced by strong affinity of the anionic groups to the metal cations, with stable aqueous suspensions in the presence of a 0.5 T magnetic field obtained.

Here, we report the reproducible, repeatable preparation of bare Fe_3O_4 nanoparticles and the first use of microwave synthesis to obtain multi-coordinating polyelectrolyte-stabilised Fe_3O_4 nanoparticles and polyelectrolyte stabilised magnetic fluids functionalised with a fluorescent dye according to Scheme 1. We are particularly interested in what effect, if any, the choice of polyelectrolyte stabiliser has on the resulting nanoparticle shape, size and crystallinity. The polyelectrolytes used were poly(sodium 4-styrenesulfonate (PSSS), which has been employed previously as a stabiliser in a traditional co-precipitation reaction,[48] and sodium polyphosphate (SPP) employed as an *in situ* stabiliser for the first time, where the sulfonate and phosphate groups, respectively, bind to the iron ions in solution prior to particle precipitation. In this manner, the polyelectrolytes act as stabilisers preventing the further growth of the nanoparticles, while at the same time promoting colloidal stability of the particles in water.

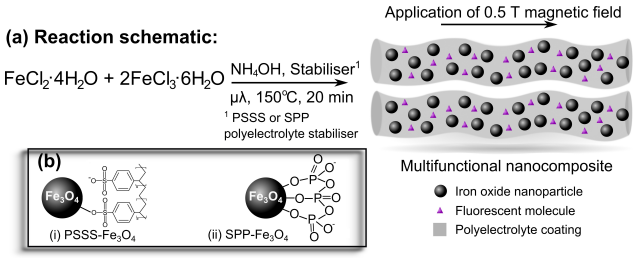

Fig. 1 (a) Schematic representation of one-pot synthesis of multifunctional nanocomposite material, using microwave irradiation. Application of an external 0.5 T magnetic field may induce linear assemblies. Legend shows iron oxide nanoparticles, fluorescent molecule (in this case, Rhodamine B), and polyelectrolyte coating which make up this nanocomposite. (b) Binding of (i) PSSS and (ii) SPP stabilisers to iron oxide nanoparticle surface.

2 Results and discussion

A modified co-precipitation technique has been employed, wherein a precursor solution of ferric and ferrous chlorides in the presence of polyelectrolyte solution was first prepared. The co-precipitation method involves several processes: nucleation, seed formation and growth.[50,51] Rapid particle nucleation follows the addition of ammonia base, after which the particle suspension is transferred to a microwave cavity and particle growth occurs under microwave irradiation at 150 °C for 20 mins. All samples were washed until neutral. In the case of the bare particles, the sample was dried for further analysis. For the polyelectrolyte samples, the final, neutral fifth washings were highly stable colloidal suspensions, which have been characterised by dynamic light scattering (DLS), transmission electron microscopy (TEM), MR imaging and their cellular interactions analysed. The remaining solids were dried and analysed by X-ray diffraction (XRD), X-ray absorption spectroscopy (XAS), SQUID magnetometry, thermogravimetric analysis (TGA) and IR.

XRD patterns collected on dried powders of each sample are shown in Figure 2(a). For comparison, a diffraction pattern was also collected of magnetite nanoparticles prepared using the traditional co-precipitation route without any microwave treatment (red line). The patterns obtained for all samples may be indexed to the cubic spinel, magnetite (Fe_3O_4). The broad peaks observed are typical for nanoparticles and the particle sizes, which may be obtained from the Scherrer equation, are included in Table 1. These are on the order of 10 nm. Also shown in Figure 2 are the patterns obtained for polyelectrolyte-stabilised samples, labelled PSSS-Fe_3O_4 and SPP-Fe_3O_4. Given that the XRD patterns for mag-

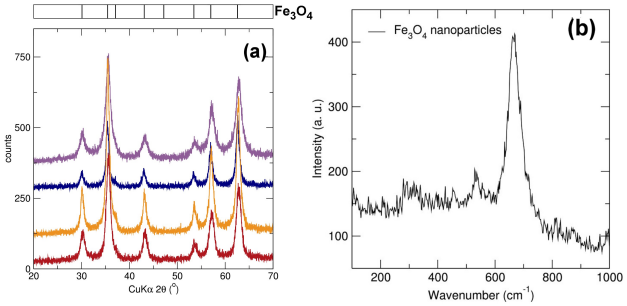

Fig. 2 (a) Powder XRD patterns for samples prepared using microwave-assisted methods: bare magnetite (orange), PSSS-Fe_3O_4 (navy) and SPP-Fe_3O_4 (purple). For comparison, a pattern of magnetite nanoparticles prepared using traditional co-precipitation synthesis is included (red) together with a reference ICSD pattern. Raman spectrum of pure Fe_3O_4 nanoparticles prepared using microwave-assisted method. Peak at 664 cm^{-1} assigned to A_{1g} mode of magnetite, with E_g and T_{2g} modes centered at 301 and 527 cm^{-1} respectively. No peaks for haematite or maghemite were observed.

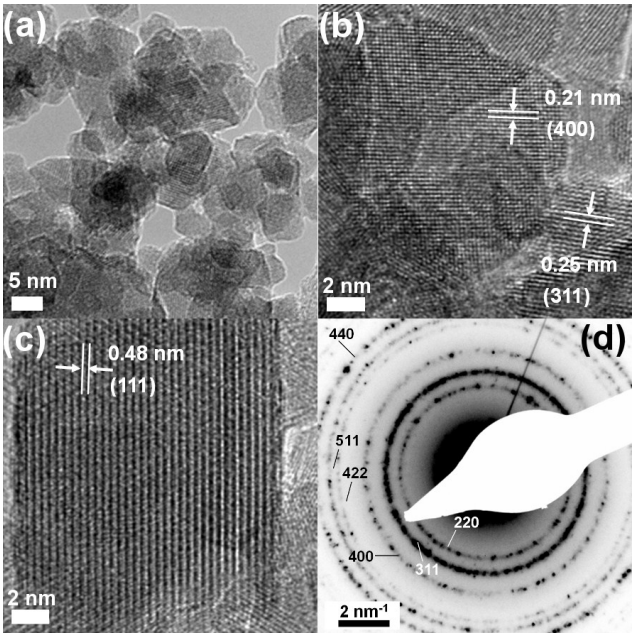

Fig. 3 (a) HRTEM images of bare Fe_3O_4 nanoparticles prepared by microwave routes reveal aggregation of discrete nanoparticles. Each individual particle is highly crystalline, with lattice fringes clearly visible. (b, c) The interplanar spacings are indexed to the corresponding magnetite *(hkl)* reflection. (d) SAED pattern from an agglomerate of particles, with the pattern indexed to magnetite.

netite and maghemite are similar, Raman spectroscopy measurements were performed. Using Raman spectroscopy, it is possible to assign characteristic peaks for magnetite (which contains both Fe^{3+} and Fe^{2+} ions) and the oxidised iron oxides maghemite (γ-Fe_2O_3) and haematite (α-Fe_2O_3). Figure 2(b) shows the resulting Raman spectrum for the pure uncoated nanoparticles, displaying a strong peak at 664 cm^{-1} which can be attributed to the A_{1g} mode of magnetite. Also visible are the E_g and T_{2g} modes, centered at 301 and 527 cm^{-1} respectively. All peaks observed are in excellent agreement with previous assignments for magnetite in the literature.[4,52,53] There are no peaks for haematite or maghemite observed.

The comparatively broader peak shapes for the Fe_3O_4-SPP sample are reflected in the decreased particle size values obtained from the Scherrer equation for these particles (10.9 nm for SPP-coated particles, 11.2 nm for uncoated nanoparticles). HRTEM images for pure Fe_3O_4 samples reveal single-crystal nanoparticles (see Figure 3). The Fe_3O_4 particles are aggregated, with a typical particle size of 12±2 nm (measured for N=100 particles). Analysis of lattice spacing and SAED patterns, shown in Figure 3(d), confirm the nanoparticles to be magnetite (Fe_3O_4, JCPDS index card number 19-629). It is clear from these measurements that the microwave iron oxide sample contains both Fe^{2+} and Fe^{3+} ions, in good agreement with the Rietveld profile analysis of this sample to Fe_3O_4.

Interestingly, the nanoparticle shape, size and aggregate nature are significantly affected by the polyelectrolyte stabilisers employed under otherwise identical reaction conditions. Electron microscopy images of the polyelectrolyte-stabilised samples are shown in Figure 4, where the presence of the PSSS-polyelectrolyte is evidenced by the core-shell appearance of the nanocomposite (Figure 4(a)). The presence of polyelectrolyte on the PSSS- and SPP-Fe_3O_4 nanoparticle surface has also been confirmed by IR spectroscopy (see Supplemental Figure S1). Both samples give a Fe-O stretch at $\sim$ 530 cm^{-1} and a broad stretch at 3400 cm^{-1} for physically adsorbed water on the particle surface. In the case of PSSS-Fe_3O_4, an Fe-O-S stretch is noted at 669 cm^{-1}, which indicates that the mode of binding is through the sulfonate group to the surface iron atoms of the nanoparticles. There are also stretches noted for the sulfonate groups at 775, 830, 1115, 1160 and 1405 cm^{-1}. In the case of the SPP-Fe_3O_4 sample, a Fe-O-P stretch is observed at 992 cm^{-1}, indicating the mode of binding is via the phosphate groups to the surface iron atoms. Stretches relating to the phosphate groups are also observed at 869 and 1255 cm^{-1}. TGA plots for bare magnetite, PSSS- and SPP-Fe_3O_4 nanoparticles are shown in Supplemental Figure S2a. The mass loss for pure magnetite is low at 3.4% and likely represents the loss of strongly adsorbed water and dehydration of surface hydroxyl groups. The mass losses are greater for the PSSS- and SPP-Fe_3O_4 samples, at 9.3% and 7.8% respectively, due to the removal of the polyelectrolyte surfactant at increasing temperatures. For both polyelectrolytes, the particles appear aggregated in the electron microscopy images (Figure 4), with higher magnification images clearly showing each agglomerated region consisting of numerous discrete nanoparticles, which are clustered together. The shape of pure Fe_3O_4 and PSSS-Fe_3O_4 nanoparticles appear better defined than the SPP-Fe_3O_4 sample, whose particles appear smaller on average and irregular in shape (Figure 4(f)). A high degree of crystallinity of these polyelectrolyte-stabilised nanoparticles is observed, where lattice spacings consistent with magnetite are identified. SAED patterns are in excellent agreement with these observations (shown in Supplemental information Figure S3).

In order to probe the behaviour of these polyelectrolyte nanocomposites in the presence of a magnetic field, samples were

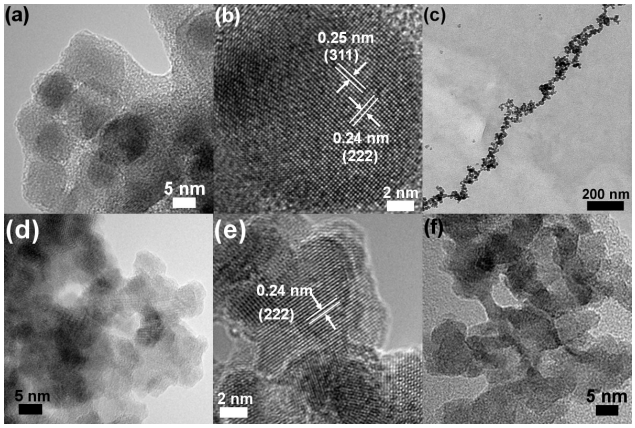

Fig. 4 HRTEM images of (a, b) PSSS- and (d, e) SPP-Fe_3O_4 nanoparticles. The major difference here is the irregular particle shape of the SPP-Fe_3O_4 (f) compared to PSSS-Fe_3O_4 which has a core-shell appearance . The formation of regular, linear assemblies is noted for the PSSS-Fe_3O_4 samples (c) upon application of a 0.5T magnetic field.

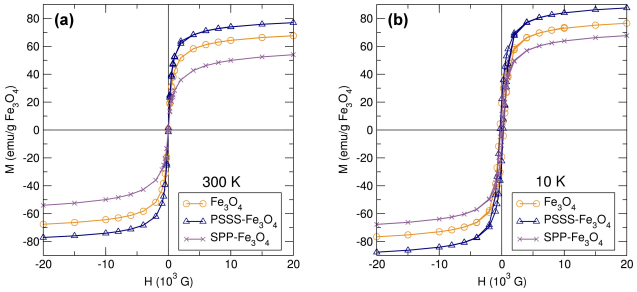

Fig. 5 Magnetic hysteresis loops of pure Fe_3O_4, PSSS-Fe_3O_4 and SPP-Fe_3O_4 at (a) 300 K and (b) 10 K. Data are shown per gram of Fe_3O_4 which was determined from TGA results.

dried in a 0.5 T magnetic field and analysed using microscopy. In the case of the PSSS-stabilised nanoparticles, linear assemblies are observed [Figure 4(c)]; reminiscent of previous reports for samples prepared by co-precipitation alone.[48] Currently, we are focussing attention on understanding how the nature of the polyelectrolyte and the chain length affects the formation of these linear assemblies and the resulting implications on imaging behaviour of nanoparticulate contrast agents.

The aggregation of these stabilised nanoparticles has been investigated further by measuring the average hydrodynamic radii at 298 K using DLS [Supplemental Figure S2(b)]. For both samples and for a range of reproduced reactions, this average is found to be close to 100 nm. The polydispersity of the SPP-Fe_3O_4 sample is greater than that for the PSSS-Fe_3O_4 sample, which is evidenced by the broader peak shape and the slightly higher polydispersity index (PDI) (0.158 for SPP-Fe_3O_4 and 0.097 for PSSS-Fe_3O_4). The low values obtained for the PDI values (<0.2) indicate a unimodal distribution of monodisperse clusters for both samples. Upon exposure to a 0.5 T magnetic field these values do not change, confirming the excellent water stability these polyelectrolyte stabilisers infer on the Fe_3O_4 nanoparticles prepared using microwave methods. Table 1 summarises the particle sizes as calculated from Scherrer broadening (XRD), average particle size (TEM), hydrodynamic radius and zeta potential (DLS). The observed zeta potentials are below -30 mV for both samples, which confirms the negative surface charge we postulate in Scheme 1 and reaffirms the excellent water stability of these suspensions, which was the case over a six month period as confirmed by DLS analysis.[54]

Magnetization curves were measured at 300 K and 10 K in magnetic field of up to 2×10^4 G and are shown in Figure 5. There is negligible coercivity and remanence noted at 300 K, indicative of superparamagnetic, single-domain iron oxide particles. The magnetization is unsaturated up to 2 T, even at 10 K. Uncoated magnetite prepared using our microwave-assisted method gives a saturation magnetisation of 67.6 emu g^{-1} at 300 K. While this is lower than the theoretical value for bulk magnetite of 98 emu g^{-1} (most likely due to spin disorder on the particle surface[55]), this value is higher than previously reported saturation magnetisation values for aqueous routes to iron oxide nanoparticles (40-50 emu g^{-1}).[56,57] By employing polyelectrolytes as stabilisers we have found that primary particle sizes and morphologies change and this, in turn, has a marked effect on the resulting magnetic properties. The SPP-stabilised particles are smaller in size, with ill-defined shape, and this is reflected in the reduced M_s value of 53.9 emu g^{-1} at 300 K. The PSSS-stabilised samples, on the other hand, have an M_s value of 77.1 emu g^{-1} at 300 K and appear highly crystalline in HRTEM. Interestingly, the magnetisation value here is significantly higher that M_s values previously obtained from NMRD data for PSSS-stabilised Fe_3O_4 (30-50 emu g^{-1}) prepared without the additional microwave synthesis step.[58]

The saturation magnetisation value is greatly affected by crystallinity of the sample. For high temperature decomposition routes using organic surfactants, for example, the highly crystalline and uniform nature of the particles is manifested in similarly higher M_s values ($\sim$ 80 emu g^{-1}).[59] For biological use, organic-surfactant coated particles require further post-processing to transfer into aqueous solutions to obtain stable suspensions.[26,60] The microwave-assisted synthesis reported here achieves both properties in a single step: excellent magnetic properties combined with long-term aqueous stability. From these data, we have shown the combination of the PSSS stabiliser and microwave irradiation results in a water-stable crystalline material, without compromising the magnetic properties.

To evaluate the MR efficacy of the PSSS-Fe_3O_4 and SPP-Fe_3O_4 samples, T1, T2 and T2*-weighted images were collected at varying Fe concentrations using a 3 T clinical MRI scanner. Relaxivity values are reported in Table 2. Increasing Fe concentrations ($\sim$0 - 2 mM in H_2O) were imaged and are displayed in Figure 6. Higher relaxation rates are noted upon increasing Fe concentrations, because PSSS-Fe_3O_4 and SPP-Fe_3O_4 shorten the T2 relaxation time, which reduces the signal intensity, i.e. negative contrast. Interestingly, the contrast displayed by the PSSS-Fe_3O_4 sample is greater than that of the SPP-Fe_3O_4 particles. The increased crystallinity of these particles and their enhanced magnetic properties are the major factors for this behaviour. Since it is the magnetic moments of the particles interacting with the water protons that result in

Table 1 Average particle sizes calculated from the Scherrer equation using XRD patterns collected, TEM (N = 100 particles, size given in nm with standard deviation) and DLS where Z-ave is the hydrodynamic radius and PDI the polydispersity index.

Sample	XRD (size nm)	TEM (size nm)	DLS Z-ave (PDI)	Zeta potential (mV)
Fe_3O_4	11.2	12 ± 2	N/A (N/A)	N/A[a]
Fe_3O_4–PSSS	16.1	13.4 ± 1.5	94.74 (0.097)	-41.5
Fe_3O_4–SPP	10.9	10.1 ± 1.5	104.93 (0.158)	-46.9

[a] No measurements were carried out on pure Fe_3O_4 nanoparticles prepared using microwave methods as no stable suspension resulted.

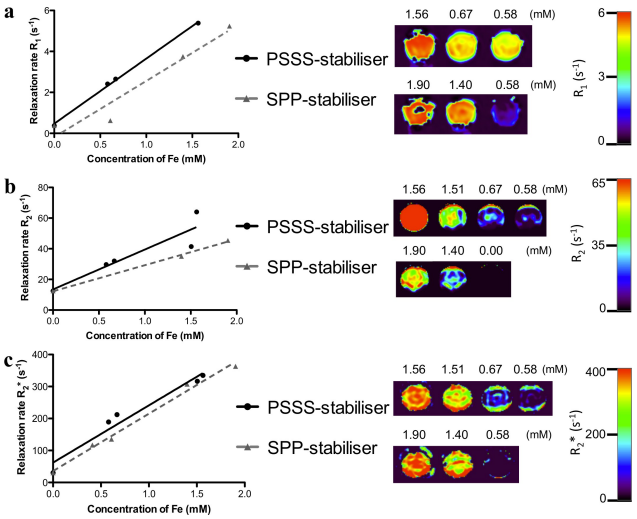

Fig. 6 Relaxivities of the PSSS (straight line) and SPP (dashed line) nanocomposites measured at 3 T and 20 °C. Scatter plots show correlations between measured R1 (a), R2 (b), and R2* (c) values of the nanocomposites and iron concentrations measured using ICP-MS. The relaxation rates (R1, R2 and R2*) were determined at 3 T using T1, T2 and T2* mapping sequences, respectively, and aqueous solutions between 0 and 2 mM of the contrast agents. Pearson's correlation coefficient values ranged from 0.92 to 0.99. Phantom MRI images of the formulations show R1, R2 and R2* maps in colour scale. R1, R2 and R2* values increase with increasing concentrations of contrast agents (highest concentration on the left).

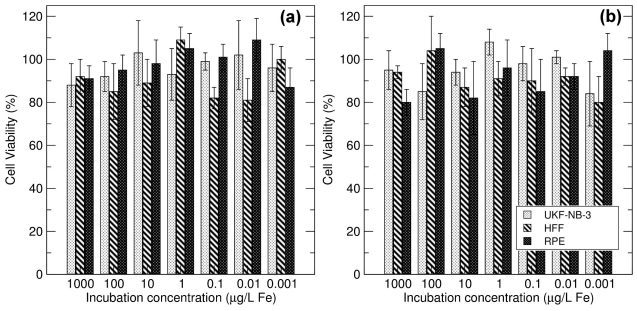

Fig. 7 Cytotoxicity profiles of (a) PSSS-Fe_3O_4 and (b) SPP-Fe_3O_4 in three different cell lines of UKF-NB-3 neuroblastoma cells, primary human retinal pigment epithelial (RPE) cells and primary human foreskin fibroblasts (HFF) in concentrations up to 1 mg iron/L.

image contrast, the crystalline nature of the particles is vital in determining how efficiently this may occur. In the case of the SPP-stabilised nanoparticles, the irregular shape and decrease in saturation magnetization point to greater disorder of the surface spins in these particles which will play a role in decreasing the imaging efficacy. This is in good agreement with an extensive study reported by Vuong *et al.*, who have proposed a method for predicting T_2 relaxation based on the nanocrystal size and magnetization values.[61] For our PSSS-Fe_3O_4 particles, the excellent magnetic properties and the crystalline nature of the primary particle indicates the promising potential for these contrast agents for MR imaging. MR contrast properties for several other reports of functionalised iron oxide nanoparticles are also included in Table 2.[48,62,63]

Enhanced MR contrast has been previously noted for linear assemblies of iron oxide nanoparticles.[48,58,64] In the current case, we observe relaxivities on the order of commercially available contrast agents and nanoparticle suspensions prepared by high temperature decomposition routes, with the added advantage here of no additional work-up required to transfer to aqueous conditions.[65–67] Recently, Zboril and co-workers reported extraordinarily high relaxivities of 735 mM s^{-1} for iron oxide nanoparticles with a 1 nm terephthalic acid coating, postulating that effective spin-transfer to surrounding water protons is mediated via π-conjugation pathways through the organic surfactant.[68] It is enticing to consider the combined use of active spin-transfer surfactants with microwave processing to further optimise relaxivity behaviour.

The effect of stabiliser on cell toxicity was examined by cell viability studies on suspensions of the polyelectrolyte-stabilised particles co-incubated at increasing concentrations (0.001 μg–1000μg) with a range of cell lines and these results are depicted in Figure 7. Regardless of the stabiliser employed, all nanoparticle suspensions tested were found to be non-toxic to a variety of mammalian cell lines, demonstrating the non-toxic robustness of these materials. Iron concentration ranges were similar to previous reports.[69] This is determined by the fact they did not affect the viability of UKF-NB-3 neuroblastoma cells, primary human retinal pigment epithelial (RPE) cells, or primary human foreskin fibroblasts (HFF) even in concentrations up to 1 mg iron/L.

To demonstrate uptake into mammalian cells, fluorescently-labelled samples were prepared by the addition of Rhodamine B to the PSSS polyelectrolyte solution before particle precipitation, where association is driven by the electrostatic interactions between the dye and the polyelectrolyte. The fluorescent nature of the nanocomposite was confirmed by fluorescence spectroscopy (λ_{ex} = 545 nm) and an enhanced aggregation of the composite is noted in DLS, where electrostatic interactions drive the formation of larger cluster sizes and is reflected by the larger Z-ave (108.2 nm; PDI 0.169) [see Supplemental Figure S4]. The sample spent several days over a 0.5 T magnetic field before DLS measurement to remove any larger aggregates and to ensure a stable suspension remained. Removal of any excess dye was confirmed by continuous washing of the sample and evaluation by

Table 2 Samples and corresponding relaxation properties in H_2O at 3 T.

Sample	Field	r_1 ($mM^{-1}s^{-1}$)	r_2 (mM^{-1} s^{-1})	r_2^* (mM^{-1} s^{-1})	r_2/r_1
Fe_3O_4–PSSS	3 T	3.18 ± 0.10	26.02 ± 6.54	179.00 ± 22.75	8.18
Fe_3O_4–SPP	3 T	2.74 ± 0.51	17.04 ± 0.91	179.31 ± 10.66	6.21
Fe_3O_4–PSSS (ref 48)[a]	1.5 T	7.2	89.4	-	0.08
Fe_3O_4–PEG-BP (ref 62)	3 T	9.5	28.2	-	2.97
Fe_3O_4–PEG (ref 63)	3 T	4.77	29.2	-	6.12

[a] Note these measurements performed in 1.5 T field.

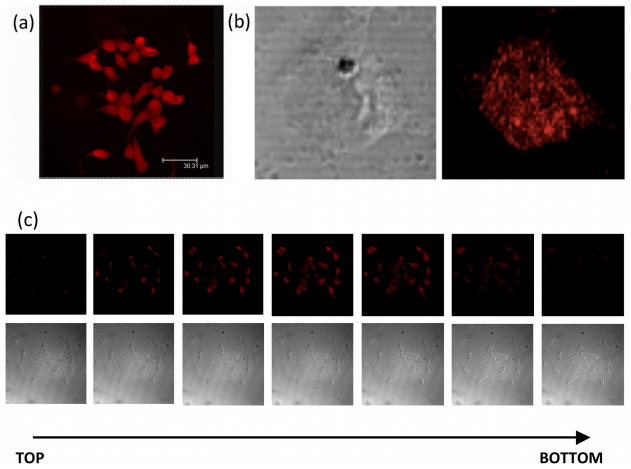

Fig. 8 Immunofluorescence confocal microscopy imaging of UKF-NB-3 neuroblastoma cells treated with nanoparticles for 24 h. In all cells, the nanoparticle suspension was internalised (a) and particles appear punctated (b). Internalisation is confirmed and when viewing slices through the cell (c).

fluorescence spectroscopy before redispersion into water and also by dialysis experiements of the resulting nanocomposite (see Supplemental Figure S5). Figure 8 shows the resulting nanoparticle uptake by UKF-NB-3 neuroblastoma cells, which can be clearly located throughout the cytoplasm. Of importance here is that this internalisation process had no effect on viability as described above. Complementary Z-scan images confirm the location of the functionalised nanoparticles throughout the cell cytoplasm [Figure 8(c)], rather than on the cell surface.

3 Conclusions

In conclusion, we have demonstrated a reproducible and reliable single step route to multifunctional magnetic nanocomposite materials, using a microwave-assisted synthesis, in order to provide stable magnetic fluids for use as MR contrast agents. Here, the underpinning structure-property relationship in these materials is highlighted by the synthetic approach taken and the resulting outcome this has on the functional properties of the nanocomposites. The choice of stabilizer - polysulfate versus polyphosphate - has a substantial effect on the magnetic properties, which translates to their ability to enhance the relaxation mechanism of surrounding water protons in MR imaging. Whilst not the highest relaxivities reported for iron oxide nanocomposites, the comprehensive analysis presented here using a full range of characterisation techniques demonstrates PSSS as an excellent stabiliser for the preparation of multifunctional magnetic nanocomposite materials and our microwave-assisted synthetic approach can be applied to a full range of ferrite-based nanoparticles and polyelectrolyte-stabilised systems. Previous reports have shown that effective contrast agents can be obtained using PSSS as a stabiliser for iron oxide nanoparticles without any microwave treatment.[48,58] In the case of the nanocomposites prepared without microwave heating, the hydrodynamic radius and polydispersity index is greater (136 nm and 0.21, respectively). Most interestingly though is that the saturation magnetisation (obtained from NMR dispersion measurements) is consistently lower for samples prepared without the application of microwave heating (30 âĂŞ 50 emg g^{-1}). This has a resulting implication on the contrast agent efficacy (values of r_1 and r_2 obtained are of the same order for both samples, but for the sample prepared without microwave treatment, the relaxation properties were measured only at 1.5 T). Therefore, the addition of this microwave heating step in the case of the PSSS stabiliser positively affects the subsequent properties and functionality of the nanocomposite. The direct link between particle crystallinity and resulting magnetic behaviour and the governing effect this has on MR imaging capability is particularly interesting. We show that these nanocomposites are non-toxic to a range of mammalian cells, where their uptake can be confocally imaged. The ease of this approach allows for the preparation of extremely stable magnetic fluids for combined MR contrast efficacy and optical imaging and paves the way for a synthetic methodology which allows for greater control over final functionality.

4 Experimental

4.1 Materials

All materials were obtained from Sigma Aldrich. Millipore water was deoxygenated by boiling and then cooling under nitrogen gas. A CEM Discover SP system was used for microwave heating. FTIR spectra (400–2000 cm^{-1}) were recorded using a Shimadzu IR Affinity-1 spectrophotometer. X-ray diffraction was performed on a Bruker D8 X-ray diffractometer and powder patterns were analysed using Rietveld refinement as embodied in the Fullprof suite. X-ray absorption spectroscopy was performed on the B18 beamline at Diamond Light Source. Pellets of samples and standards were prepared by ball milling the materials and cellulose binder for 10 min, followed by pressing at 8 tonnes for 5 min to produce 1.3 cm^2 discs. The discs were then irradiated with synchrotron X-ray radiation to excite the Fe K-edge. Thermogravimetric analysis was obtained with a Netzsch STA 409 PC Luxx TGA machine. All samples were heated in air to 700 °C. Dynamic light scattering measurements were performed using a Malvern Zetasizer nano-ZS. Transmission electron microscopy (TEM) images were taken on a JEOL JEM (200-FX) operating at 120 kV.

Samples were prepared on a formvar coated copper grid. Some grids were dried over a 0.5 T magnet. High resolution TEM and selected area electron diffraction (SAED) were performed on a FEI Tecnai TF20 instrument fitted with a field emission gun, operated at 200 keV. TEM samples were prepared by dispersing the sample in deionised water and dropping the solution onto an amorphous holey carbon coated grid. TEM data were obtained and processed using either Digital Micrograph or IMAGEJ 1.41 software.

4.2 Preparation of magnetic nanocomposites

$FeCl_3.6H_2O$ (2.70 g; 10 mmol) and $FeCl_2.4H_2O$ (0.99 g; 5 mmol) were dissolved in 10 mL deoxygenated water. Polyelectrolyte stabiliser (0.2 g of either PSSS or SPP) was dissolved in the iron solution. The solution was heated to 80 °C. Ammonia solution (10 mL; 28-30%) was injected at a rate of approximately 2.5 mL/s and the solution was stirred for 20 minutes before transferring to the microwave cavity to be heated at 150 °C for 20 minutes. For fluorescently-labelled samples, Rhodamine B (2×10^{-4} g; 10 mmol) was dissolved in 2 mL deoxygenated water and the PSSS polyelectrolyte (0.2 g) was added to this and stirred for two hours. This was transferred to 10 mL of Fe^{3+}/Fe^{2+} solution (2.70 g; 10 mmol and 0.99 g; 5 mmol respectively), before addition of 10 mL ammonia solution at 80 °C. The resulting black precipitates were washed with Millipore water (5×20 mL) with the final two washings being used in TEM, DLS, FTIR, MRI and confocal measurements. The solid precipitate was analysed with XRD, TGA, HRTEM and SQUID measurements.

4.3 MR imaging

The tubes of different PSSS-Fe_3O_4 and SPP-Fe_3O_4 concentrations were placed in a rack in the centre of the magnet. MR imaging was performed with a standard extremity flex coil on a clinical 3 T MRI scanner (Achieva, Philips Healthcare, Best, The Netherlands). T_2 was determined with a 2D multi-spin-echo sequence (FOV = 120×120 mm^2, matrix = 432×432, measured slice thickness = 3 mm, echo train length = 5, TE = 10 ms, TR = 725 ms, flip angle = 90°). The acquired imaging data was transferred to a computer running Matlab and analysed using an in-house Matlab tool to receive the relaxation time T_2 for each Fe concentration. Excel was used to plot the relaxation rate R_2 over the concentration and the relaxivity value was determined using linear regression. Iron concentrations of all MRI scanned serial dilutions of SPIONs were determined by Inductively Coupled Plasma Mass Spectrometry (ICP-MS). Briefly, samples were digested in 70% nitric acid overnight at room temperature, followed by dilution in deionized water. A standard curve was acquired with each sample set for iron concentration determination.

4.4 Investigating the Effect of Nanoparticles on Cell Viability

The effects of the nanoparticles on cell viability were determined in UKF-NB-3 neuroblastoma cells, primary human retinal pigment epithelial cells, and primary human foreskin fibroblast cells. UKF-NB-3 cells were cultivated in IMDM supplemented with 10% foetal calf serum (FCS), 100 IU/mL penicillin, and 100 mg/mL streptomycin. Retinal pigment epithelial cells were cultured in DMEM supplemented with 20% FCS, 100 IU/mL penicillin, and 100 mg/mL streptomycin. Fibroblasts were cultured in DMEM supplemented with 10% FCS, 100 IU/mLpenicillin, and 100 mg/mL streptomycin. All cells were cultivated at 37 °C in humidified 5% CO_2 atmosphere as previously described.[70,71] Cell viability upon the addition of nanoparticle preparations at different concentrations was determined by the 3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyltetrazolium bromide (MTT) dye reduction assay after 120 h of incubation as described previously.[70,71]

4.5 Determination of Cellular Location of Nanoparticles by Fluorescent Microscopy

UKF-NB3 cells were seeded at 2×10^5 cells/well in a 24 well plate, containing a coverslip in each well, and grown at 37 °C for 48 hours. Rhodamine conjugated nanoparticles were diluted to 0.099 mg/L iron concentration in media, 1 mL added to appropriate wells and incubated for 3 hours before aspirating and fixing with methanol at -20 °C for 5 minutes. Coverslips were mounted using mowiol and anti-fade and slides examined using a Leica confocal laser scanning (TCS 4; ×63 oil lens) microscope.

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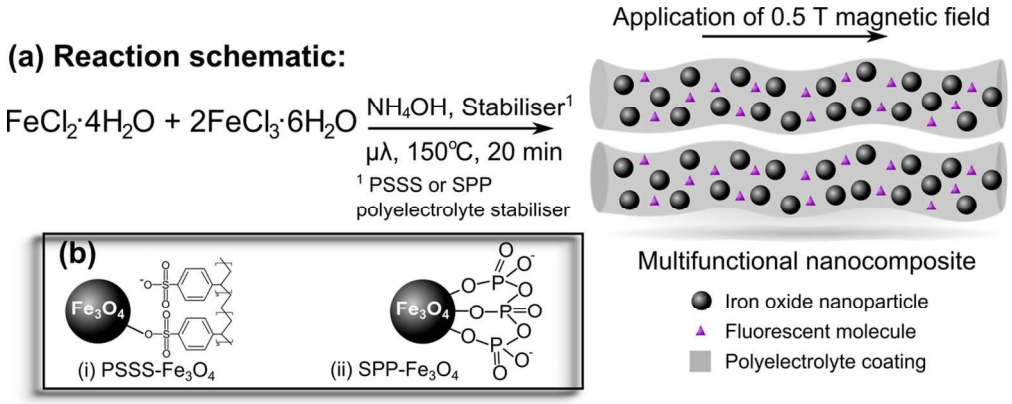

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