

RSC Advances



This is an *Accepted Manuscript*, which has been through the Royal Society of Chemistry peer review process and has been accepted for publication.

Accepted Manuscripts are published online shortly after acceptance, before technical editing, formatting and proof reading. Using this free service, authors can make their results available to the community, in citable form, before we publish the edited article. This *Accepted Manuscript* will be replaced by the edited, formatted and paginated article as soon as this is available.

You can find more information about *Accepted Manuscripts* in the [Information for Authors](#).

Please note that technical editing may introduce minor changes to the text and/or graphics, which may alter content. The journal's standard [Terms & Conditions](#) and the [Ethical guidelines](#) still apply. In no event shall the Royal Society of Chemistry be held responsible for any errors or omissions in this *Accepted Manuscript* or any consequences arising from the use of any information it contains.

1 **Preparative separation of quaternary ammonium alkaloids from *Caulis***
2 ***Mahoniae* by conventional and pH-zone-refining counter-current**
3 **chromatography**

4 Heng Zhu ^{a,1}, Daijie Wang ^{b,1}, Lei Wen ^a, Jinqian Yu ^b, Yanling Geng ^b, Hengqiang
5 Zhao ^b, Ruixuan Zhao ^b, Xiao Wang ^{a,b*}

6 ^a College of pharmacy, Shandong University of traditional Chinese medicine, Jinan
7 250355, China;

8 ^b Shandong Key Laboratory of TCM Quality Control Technology, Shandong Analysis
9 and Test Center, Shandong Academy of Sciences, 19 Keyuan Street, Jinan 250014,
10 China

11

12

13

14 * **Corresponding author:** Dr. Xiao Wang. E-mail address: wangx@sdas.org. Tel.:+86
15 531 82605319; fax: +86 531 82964889.

16

17 ¹ These authors have equal contribution to this work. Both persons are the first
18 authors.

19 **Abstract**

20 In this work, the preparative separation of quaternary ammonium alkaloids from
21 *Caulis Mahoniae* by pH-zone-refining counter-current chromatography (PZRCCC)
22 was compared to a conventional high-speed counter-current chromatography

23 (HSCCC). A flow-rate changing strategy was performed in conventional CCC
24 separation with two-phase solvent systems composed of chloroform-methanol-0.5mM
25 HCl water solution (4:1.5:2, *v/v*). Compared to conventional CCC separation, 3.0 g
26 crude alkaloid extracts was carried out by PZRCCC with solvent system of
27 chloroform-methanol-water (4:3:3, *v/v*). The retainer acid and eluter base were
28 optimized by changing the concentration ratios, 60 mM HCl in upper aqueous phase
29 and 7.5 mM TEA in lower phase. From 3.0 g of the crude alkaloid extracts,
30 stepharanine (53.7 mg, 97.4%), columbamine (28.1 mg, 96.5%), jatrorrhizine (150.6
31 mg, 99.0%), palmatine (169.8 mg, 99.0%) and berberine (157.2 mg, 99.5%) were
32 obtained. The compounds were identified by ESI-MS and ¹H-NMR data. The results
33 indicated that PZRCCC is an excellent separation mode for separating quaternary
34 ammonium alkaloids compared with conventional CCC.

35 **1 Introduction**

36 Caulis Mahoniae, the dried stem of *Mahonia bealei* (Fort) Carr. or *Mahonia fortune*
37 (Lindl.) Fedde, is widely distributed in southeast of China. As a famous folk medicine,
38 Caulis Mahoniae is widely used for treating jaundice hepatitis, jaundice, skin ulcer,
39 etc ^{1, 2}. Caulis Mahoniae is also known for its effects of anti-cancer, anti-viral,
40 anti-inflammatory, anti-arrhythmia, anti-bacterial, and hypoglycemic. Major effective
41 compounds in Caulis Mahoniae are alkaloids which belong to quaternary ammonium
42 ³⁻⁶.

43 A number of traditional chromatography techniques are used for separating
44 alkaloids of quaternary ammonium. (e.g. pre-HPLC, alkaline silica gel and sephadex
45 LH-20). Although traditional methods are widely used, the disadvantages of them
46 could not be neglected, such as large solvent consumption, irreversible sample

47 adsorption and require complex multiple steps. It is necessary to establish an efficient
48 method to separate alkaloids of quaternary ammonium from *Caulis Mahoniae*.

49 High-speed counter-current chromatography (HSCCC) is a liquid-liquid partition
50 chromatography which can eliminate the irreversible adsorption without solid packing
51 support. Due to the advantages of HSCCC, such as large sample injection, high
52 recovery rate, simple sample preparation, and excellent repeatability ⁷, HSCCC has
53 been developed to separate and isolate various samples ⁸⁻¹⁶. pH-zone-refining
54 counter-current chromatography (PZRCCC), derived from HSCCC and initially
55 invented by Ito, can separate ionic compounds including alkaloids and organic acids.
56 The chromatography peaks are rectangular with highly concentrated sample according
57 to their hydrophobicities and pK_a values. Compared to conventional CCC, PZRCCC
58 is approximate 10-fold increase in sample loading, enrichment of minor impurities,
59 and high concentration of peaks.

60 In this paper, two CCC methods including conventional CCC and PZRCCC are
61 comparative employed for preparative separation alkaloids of quaternary ammonium
62 from *Caulis Mahoniae*. Fig. 1 shows the chemical structures of quaternary ammonium
63 alkaloids.

64 **2 Experimental**

65 **2.1 Reagents and materials**

66 Petroleum ether, chloroform, methanol, alcohol, triethylamine (TEA), and
67 hydrochloric acid (HCl) were analytical grade purchased from Siyou Chemical
68 Reagent Factory. Acetonitrile, phosphoric acid and TEA used for HPLC were of
69 chromatographic grade (Tedia Company, Inc., Fairfield, USA). Ultrapure water (18.2
70 M Ω) was purified by osmosis Milli-Q water system (Millipore, Bedford, MA, USA).
71 *Caulis Mahoniae* was purchased from Jinan Jian-lian TCM store and the specie was

72 identified by Professor Jia Li (Shandong University of Traditional Chinese Medicine,
73 Jinan, China). The voucher specimen (No. MD201509) was stored in Shandong
74 Analysis and Test Center, Jinan, Shandong, China.

75 **2.2 Apparatus**

76 HSCCC separation was performed by TBE-300A (Shanghai Tauto Biotechnology,
77 Shanghai, China) equipped with three multilayer coil separation PTFE columns
78 connected in series (total volume 300 mL, I.D. of the tube diameter 2.6 mm) with a 20
79 mL sample loop. The rotation speed was adjusted from 0 to 1000 rpm by speed
80 controller. During the separation process, the temperature of separation columns were
81 maintained at 25°C using a model HX series constant temperature circulator
82 instrument (Changcheng Company, Zhengzhou, China). TBP-5002 constant-flow
83 pump (Shanghai Tauto Biotechnology, Shanghai China) was used to pump the two
84 solvent systems. The chromatogram data of alkaloid extracts was collected using
85 Model 3057 portable recorder (Yokogawa, Sichuan Instrument Factory, Sichuan,
86 China). Spectra were generated from 8823A-UV detector (Beijing Emilion
87 Technology, Beijing, China) at the wavelength of 254 nm.

88 HPLC was performed on Waters e2695 equipment (Waters, Milford, MA, USA)
89 including a Waters 2998 diode assay detector (DAD) system, an automatic sample
90 injection, a Waters 2695 quaternary-solvent delivery system, and a millennium 32
91 workstation.

92 **2.3 Preparation of crude extracts from *Caulis Mahoniae***

93 The dried rhizomes of *Caulis Mahoniae* (4 kg, 40-60 mesh) were extracted under
94 reflux (70°C) with 2 L of 95% ethanol twice for two hours. After extraction, filtrated
95 extracts under decompression, then combined together and concentrated them by
96 rotary evaporation at 50°C.

97 The pH value of the liquid concentration was adjusted to 3 with 2% HCl. After that,
98 extracted the acid solution with petroleum ether for five times. The extraction solution
99 was slowly alkalized with 10% ammonia until the pH value was 10. After rotary
100 evaporation, 10.2 g of yellow precipitate was obtained and stored in refrigerator at
101 5°C.

102 **2.4 Preparation of solvent systems and sample solutions**

103 In conventional CCC, flow-rate changing strategy was used to separate total
104 alkaloids in *Caulis Mahoniae*. The two-phase solvent system used for conventional
105 CCC was chloroform-methanol-0.5mM HCl water (4:1.5:2, volume ratio, the same as
106 follows). After equilibrated in a separating funnel, the solvent system that used for the
107 following experiment was separated into two phases. 250 mg of crude alkaloid
108 extracts were dissolved in 5 mL stationary phase and 5 mL mobile phase.

109 In PZRCCC separation, a two phase solvent system consisted of
110 chloroform-methanol-water (4:3:3, v/v) was used. After equilibrated in a separating
111 funnel, the solution was divided into two phases for the following experiment. The
112 upper phase was acidified with 60 mM of HCl (stationary phase) and lower organic
113 phase was alkalified with 7.5 mM TEA (mobile phase). 3.0 g of crude alkaloid
114 extracts were dissolved in lower phase without alkalization and acidified upper phase.

115 **2.5 Separation procedure**

116 **2.5.1 Conventional CCC separation**

117 The multilayer-coiled columns were first filled with upper phase at 20.0 mL/min.
118 Then the HSCCC equipment was rotated at 800 rpm in a clockwise mode while the
119 lower mobile phase was pumped into the apparatus at 2.0 mL/min. After
120 hydrodynamic equilibrium established, sample solution with 250 mg total alkaloids
121 from *Caulis Mahoniae* was injected via the sample loop. The effluent was monitored

122 continuously with the UV detector at 254 nm. When elution time reached to 200
123 minutes, the mobile phase was changed into 10.0 mL/min until 310 min. The
124 stationary phase retention was defined as the ratio of stationary phase divided by the
125 total volume in column after separation.

126 **2.5.2 PZRCCC separation**

127 The PZRCCC was first filled with upper phase, and sample solution with 3.0 g total
128 alkaloids from *Caulis Mahoniae* was injected. The lower mobile phase was pumped
129 into the CCC column at 2.0 mL/min while rotated at 800 rpm in a clockwise mode.
130 The effluent was monitored at 254 nm and records were collected by portable recorder.
131 pH values of all fractions were tested at room temperature. The stationary phase
132 retention in PZRCCC was defined the same as conventional CCC.

133 **2.6 Analysis and identification of separation products**

134 Analysis used for HPLC was Waters Symmetry® C₁₈ column (4.6 mm×250 mm,
135 i.d., 5 µm) at 25°C. The mobile phase was acetonitrile-1% TEA (adjust the pH value to
136 3 with phosphoric acid) solution (25:75, v/v) and the flow-rate was 1.0 mL/min. And
137 the detector wavelength was 265 nm.

138 Agilent 5973N mass selective detector was used to detect the molecular weight of
139 pure compounds with ESI interface. The NMR spectrum was tested by Varian-600
140 spectrometer with TMS as internal standard and DMSO as the solvent (Varian, Palo
141 Alto, CA, USA).

142 **3. Results and discussion**

143 **3.1 Conventional CCC separation**

144 According to the alkaloid properties and the previous test on HSCCC separation,
145 the solvent systems composed petroleum ether-ethyl acetate-methanol-water
146 (1:1:1:1.1, v/v), ethyl acetate-*n*-butanol-0.5mM HCl water solution (4:1:5, v/v), and

147 chloroform-methanol-0.5mM HCl water solution (4:1.5:2, *v/v*) were designed to get
148 an efficient separation of the alkaloid compounds. The K_D values of solvent systems
149 of HSCCC are presented in Table 1. Table 1 shows that the K_D values were too small
150 and unsuitable to separate alkaloids from *Caulis Mahoniae* when using petroleum
151 ether-ethyl acetate-methanol-water (1:1:1:1, *v/v*) and ethyl acetate-*n*-butanol-0.5mM
152 HCl solution (4:1:5, *v/v*) as the solvent system. In view of alkaloids were soluble in
153 chloroform, the solvent system composed of chloroform-methanol-0.5mM HCl water
154 solution were chosen for the separation. When chloroform-methanol-0.5mM HCl
155 water solution (4:1.5:2, *v/v*) was used, compounds A, D and E afforded a suitable K_D
156 value for separating but the elution time of compounds B and C were too long. In
157 consideration of increasing the flow-rate had little effect on the stationary phase
158 retention in chloroform series solvent systems, a flow-rate changing strategy was used
159 in compounds B and C separation.

160 It was then assayed the HSCCC separation with chloroform-methanol-0.5mM HCl
161 water solution (4:1.5:2, *v/v*). The lower phase of chloroform-methanol-0.5mM HCl
162 water solution (4:1.5:2, *v/v*) were used as mobile phase in flow-rate changing strategy
163 (0-200 min, 2.0 mL/min; 200-240 min, 10.0 mL/min), a good separation result could
164 be obtained. After that, five compounds were successfully isolated from 250 mg crude
165 alkaloid extracts of *Caulis Mahoniae* (shown in Fig. 3) including stephananine
166 (compound A, 13.2 mg), columbamine (compound B, 6.6 mg), jatrorrhizine
167 (compound C, 17.3 mg), palmatine (compound D, 13.4 mg), and berberine
168 (compound E, 14.7 mg) with the purities of 97.5%, 96.0%, 98.2%, 99.0%, and 99.5%
169 as determined by HPLC, and the compound recoveries were 98%, 97%, 98%, 93%,
170 and 97%, respectively. The productivities of the organic solvent were also calculated.
171 In conventional CCC, the productivities per hour of the five compounds (compound A,

172 B, C, D, E) were 2.6, 1.3, 3.4, 2.6, and 2.9 mg/h, respectively, while 9.4, 4.7, 12.4, 9.6,
173 and 10.5 mg/L (per liter of the solvent), respectively.

174 **3.2 PZRCCC Separation**

175 Through preliminary experiment, the solvent system composed of
176 chloroform-methanol-water (4:3:3, v/v) was selected with different concentration of
177 retainer acid and eluter base in PZRCCC separation. When 1.0 g crude alkaloid
178 extracts were separated with 30 mM HCl as retainer acid (upper phase) and 10 mM
179 TEA as eluter base (lower phase), compound A was successfully separated with a
180 purity over 98% (Fig. 4I) while compounds B, C and D, E were not fully separated.
181 When 60 Mm HCl and 10 mM TEA was used, the peak resolution was improved with
182 elution time longer. As shown in (Fig. 4II), 1.0 g crude alkaloid extracts were
183 completely separated. Then, the sample amount increased to 3.0 g, compound A was
184 separated successfully while compounds B, C and D, E partly separated. When the
185 stationary phase was acidified with 60 mM HCl and the mobie phase was alkalified
186 with 7.5 mM TEA, 3.0 g crude alkaloid extracts were successful separation as shown
187 in (Fig. 4III). Ultimately, five compounds were isolated and were identified as
188 stepharanine (compound A, 53.7 mg), columbamine (compound B, 28.1 mg),
189 jatrorrhizine (compound C, 150.6 mg), palmatine (compound D, 169.8 mg), and
190 berberine (compound E, 157.2 mg) with the purities of 97.4%, 96.5%, 99.0%, 98.7%,
191 and 98.5%, respectively, while they were determined by HPLC. Recoveries of five
192 compounds were 33%, 34%, 71%, 98%, and 86%, respectively. In PZRCCC, the
193 productivities per hour of the five compounds (compound A, B, C, D, E) were 7.4, 3.9,
194 20.6, 23.3, and 21.5 mg/h, respectively. And the productivities per liter of the organic
195 solvent of the five compounds were 44.8, 23.4, 125.5, 141.5, and 131.0 mg/L,
196 respectively.

197 3.4 Identification of the isolated compounds

198 Compound A (peak A in Fig. 3, peak A in Fig. 4III): light yellow crystal in
199 chloroform methanol, bismuth potassium iodide reaction was positive. UV ($\lambda_{\max}^{\text{MeOH}}$):
200 227, 282, 348 nm. Positive ESI-MS (m/z): 324.1 $[\text{M}+\text{H}]^+$. $^1\text{H-NMR}$ (125 MHz,
201 $\text{DMSO-}d_6$) δ_{ppm} : 9.70 (1H, s, H-8), 8.73 (1H, s, H-13), 7.89 (1H, d, $J = 9.0$ Hz, H-12),
202 7.85 (1H, d, $J = 9.0$ Hz, H-11), 7.54 (1H, s, H-1), 7.03 (1H, s, H-4), 4.89 (2H, t, $J =$
203 6.0 Hz, H-6), 3.17 (2H, t, $J = 6.0$ Hz, H-5), 4.04 (3H, s, 9-OCH₃), 3.88 (3H, s,
204 3-OCH₃). Compared to literature²⁷, this compound was identified as stepharanine.

205 Compound B (peak B in Fig. 3, peak B in Fig. 4III): light yellow spiculas in
206 chloroform methanol, bismuth potassium iodide reaction was positive. UV ($\lambda_{\max}^{\text{MeOH}}$):
207 263, 345 nm. Positive ESI-MS (m/z): 338.4 $[\text{M}+\text{H}]^+$. $^1\text{H-NMR}$ (125 MHz, $\text{DMSO-}d_6$)
208 δ_{ppm} : 9.87 (1H, s, H-8), 8.80 (1H, s, H-13), 8.21 (1H, d, $J = 8.4$ Hz, H-11), 8.03 (1H, d,
209 $J = 8.4$ Hz, H-12), 7.58 (1H, s, H-1), 7.10 (1H, s, H-4), 4.96 (2H, t, $J = 5.4$ Hz, H-6),
210 4.10 (3H, s, 10-OCH₃), 4.08 (3H, s, 9-OCH₃), 3.91 (3H, s, 3-OCH₃), 3.20 (2H, t, $J =$
211 5.4 Hz, H-5). Compared to literature²⁸, this compound was identified as
212 columbamine.

213 Compound C (peak C in Fig. 3, peak C in Fig. 4III): light red spiculas in
214 chloroform methanol, bismuth potassium iodide reaction was positive. UV ($\lambda_{\max}^{\text{MeOH}}$):
215 265, 345 nm. Positive ESI-MS (m/z): 338.4 $[\text{M}+\text{H}]^+$. $^1\text{H-NMR}$ (125 MHz, $\text{DMSO-}d_6$)
216 δ_{ppm} : 9.85 (1H, s, H-8), 9.01 (1H, s, H-13), 8.11 (1H, d, $J = 8.4$ Hz, H-12), 8.00 (1H, d,
217 $J = 8.4$ Hz, H-11), 7.29 (1H, s, H-1), 6.96 (1H, s, H-4), 4.91 (2H, t, $J = 5.4$ Hz, H-6),
218 4.08 (3H, s, 10-OCH₃), 4.06 (3H, s, 9-OCH₃), 3.95 (9H, s, 2-OCH₃), 3.15 (2H, t, $J =$
219 5.4 Hz, H-5). Compared to literature²⁹, this compound was identified as jatrorrhizine.

220 Compound D (peak D in Fig. 3, peak D in Fig. 4III): Yellow needle crystal in
221 chloroform methanol, bismuth potassium iodide reaction was positive. UV ($\lambda_{\max}^{\text{MeOH}}$):

222 273, 346 nm. Positive ESI-MS (m/z): 352.4 $[M+H]^+$. 1H -NMR (125 MHz, DMSO- d_6)
223 δ_{ppm} : 7.72 (1H, s, H-1), 7.09 (1H, s, H-4), 9.89 (1H, s, H-8), 8.22 (1H, d, $J = 8.4$ Hz,
224 H-11), 8.02 (1H, d, $J = 8.4$ Hz, H-12), 9.04 (1H, s, H-13), 4.95 (2H, t, $J = 5.4$ Hz,
225 H-6), 4.12 (3H, s, 10-OCH₃), 4.07 (3H, s, 9-OCH₃), 3.96 (3H, s, 2-OCH₃), 3.87 (3H, s,
226 3-OCH₃), 3.24 (2H, t, $J = 5.4$ Hz, H-5). Compared to literature ³⁰, this compound was
227 identified as palmatine.

228 Compound E (peak E in Fig. 3, peak E in Fig. 4III): Yellow needle crystal in
229 chloroform methanol, bismuth potassium iodide reaction was positive. UV (λ_{max}^{MeOH}):
230 263, 346 nm. Positive ESI-MS (m/z): 336.4 $[M+H]^+$. 1H -NMR (125 MHz, DMSO- d_6)
231 δ_{ppm} : 9.91 (1H, s, H-8), 8.95 (1H, s, H-13), 8.20 (1H, d, $J = 8.4$ Hz, H-11), 8.01 (1H, d,
232 $J = 8.4$ Hz, H-12), 7.78 (1H, s, H-1), 7.07(1H, s, H-4), 6.16 (2H, s, 2, 3-OCH₂O), 4.93
233 (2H, t, $J = 5.4$ Hz, H-6), 4.08 (3H, s, 10-OCH₃), 4.06 (3H, s, 9-OCH₃), 3.22 (2H, t, $J =$
234 5.4 Hz, H-5). Compared to literature ³¹, this compound was identified as berberine.

235 4. Conclusions

236 In this paper, two separation models conventional HSCCC and PZRCCC were
237 successful used to preparative separation quaternary ammonium alkaloids from *Caulis*
238 *Mahoniae*.. Five compounds were obtained in one-step separation with two CCC
239 separation models and were identified as stepharanine, columbamine, jatrorrhizine,
240 palmatine and berberine with the purities over 96%. The results demonstrated that in
241 order to save elution time, flow-rate changing strategy may be employed in
242 conventional HSCCC separation. Due to quaternary ammonium alkaloids are high
243 polarity, reverse elution PZRCCC can be used for the low organic phase as mobile
244 phase. And the concentration of the eluate (mobile phase) and retainer (stationary
245 phase) are optimized by alkali and acid. Compared with the conventional CCC,
246 PZRCCC is an efficient and rapid method for separation alkaloids of quaternary

247 ammonium because of the high concentration of fractions and large sample loading
248 capacity.

249 Acknowledgments

250 The article was financially supported by National Natural Science Foundation of
251 China (21506119).

252 References

- 253 1. Y. Cong, Y. Wang, X. T. Wang and Q. Li, *Chin. Tradit. Pat. Med.*, 2011, **33**,
254 1008-1010.
- 255 2. J. Liu, Y. S. Chen, J. Li, L. H. Zhu and Y. Cong, *J. Henan Univ.*, 2014, **33**, 170-174.
- 256 3. X. T. Wang, M. Li, J. K. Lei and W. Zhao, *Chin. J. Gerontol.*, 2008, **28**, 1143-1144.
- 257 4. Z. W. Yan, A. X. Bao, L. Ge, Z. F. Tian, K. D and Yang, *J. Guangxi. Univ.*, 2015,
258 **40**, 528-531.
- 259 5. X. T. Wang and M. Li, *J. Henan Univ.*, 2007, **26**, 16-18.
- 260 6. M. Li and X. T. Wang, *J. Henan Univ.*, 2007, **26**, 24-26.
- 261 7. Y. Ito, *J. Chromatogr. A*, 2005, **1065**, 145-168.
- 262 8. J. Kang, D. Y. Gu, T. Wu, M. Wang, H. Zhang, H. Guo, Y. X. Yin, Y. Yang and J.
263 Tian, *Sep. Purif. Technol.*, 2016, **162**, 142-147.
- 264 9. X. Y. Huang, S. Ignatova, P. Hewitson and D. L. Di, *Trends in Analytical Chemistry*,
265 2016, **77**, 214-225.
- 266 10. D. B. Ren, B. S. Han, Z. Q. Xin, W. B. Liu, S. S. Ma, Y. Z. Liang and L. Z. Yi, *Sep.*
267 *Purif. Technol.*, 2016, **165**, 160-165.
- 268 11. Y. F. He, X. Y. Wang, Y. R. Suo, C. X. Ding and H. L. Wang, *J. Chromatogr. Sci.*,
269 2016, **54**, 479-485.
- 270 12. H. H. Lv, W. N. Zhou, X. Y. Wang, Z. H. Wang, Y. R. Suo and H. L. Wang, *J.*
271 *Chromatogr. Sci.*, 2016, **54**, 744-751.

- 272 13. P. L. Zhang, N. Xie, K. W. Tang, X. M. Chen and W. F. Xu, *Sep. Purif. Technol.*,
273 2016, **164**, 41-48.
- 274 14. X. L. Ye, D. Cao, F. Y. Song, G. R. Fan and F. H. Wu, *Sep. Sci. Technol.*, 2016, **51**,
275 807-815.
- 276 15. Z. L. Gan, Z. Liang, X. S. Chen, X. Wen, Y. X. Wang, M. Li and Y. Y. Ni, *J.*
277 *Chromatogr. B*, 2016, **1011**, 99-107.
- 278 16. S. J. Zhang, W. Q. Wu, D. Y. Li and Q. G. Zheng, *Sep. Sci. Technol.*, 2016, **51**,
279 673-680.
- 280 17. H. J. Dong, Y. Q. Zhang, L. Fang, W. J. Duan, X. Wang and L. Q. Huang, *J.*
281 *Chromatogr. B*, 2011, **879**, 945-949.
- 282 18. Q. Yu, S. Q. Tong, J. Z. Yan, C. Q. Hong, W. F. Zhai and Y. Q. Li, *J. Sep. Sci.* 2011,
283 **34**, 278-285.
- 284 19. R. L. Hu, X. J. Dai, Y. B. Lu and Y. J. Pan, *J. Chromatogr. B*, 2010, **878**,
285 1881-1884.
- 286 20. Z. J. Zheng, M. L. Wang, D. J. Wang, W. J. Duan, X. Wang and C. C. Zheng, *J.*
287 *Chromatogr. B*, 2010, **878**, 1647-1651.
- 288 21. Y. Li, F. F. Cai, M. Zhang, H. Y. Zhang, Y. R. Wang and P. Hu, *J. Chromatogr. A*,
289 2015, **1378**, 58-64.
- 290 22. C. L. Sun, J. Li, X. Wang, W. J. Duan, T. Y. Zhang and Y. Ito, *J. Chromatogr. A*,
291 2014, **1370**, 156-161.
- 292 23. A. Kotland, S. Chollet, J. M. Autret, C. Diard, L. Marchal and J. H. Renault, *J.*
293 *Chromatogr. A*, 2015, **1391**, 80-87.
- 294 24. C. L. Sun, F. Liu, J. Sun, J. Li and X. Wang, *J. Chromatogr. A*, 2016, **1427**,
295 96-101.
- 296 25. M. Bakri, Q. Chen, Q. L. Ma, Y. Yang, A. Abdukadir and H. A. Aisa, *J.*

- 297 *Chromatogr. B*, 2015, **1006**, 138-145.
- 298 26. C. L. Sun, J. Li, D. J. Wang, J. Q. Yu, X. Wang, and L. Q. Huang, RSC Advances,
299 2015, 92, 75831-75837.
- 300 27. Z. M. Lv, Q. J. Zhang, R. Y. Chen and D. Q. Yu., *Chin. J. Chin. Mater. Med.*, 2011,
301 **36**, 1024-1027.
- 302 28. H.M. Zhang, X.M. Cheng, C.H. Wang and Z.T. Wang, *Chin. J. Pharm.* 2008, **39**,
303 588-590.
- 304 29. T.J. Hsieh, Y.C. Chia, Y.C. Wu and C.Y. Chen, *J. Chin. Chem. Soc.* 2004, **51**,
305 443-446.
- 306 30. Y.H. Yang, C.L. Gan, *Heilongjiang Med. J.* 2009, **22**, 480-481.
- 307 31. G. H. Lu, J. M. Chen, L. W. Wang, et al. *West China J. Pharm. Sci.*, 1995, **4**, 202.
- 308
- 309
- 310
- 311
- 312
- 313
- 314
- 315
- 316
- 317
- 318
- 319
- 320
- 321

322

Captions to the figures

323 **Fig. 1** Chemical structures of compounds separated from *Caulis Mahoniae*.

324

325 **Fig. 2** The HPLC chromatogram of crude alkaloids from *Caulis Mahoniae*.

326 Experimental conditions: column, Waters Symmetry® C₁₈ column (4.6 mm×250 mm,

327 i.d., 5 µm); mobile phase, acetonitrile-1% TEA (adjust the pH value to 3 with

328 phosphoric acid) solution (25:75, v/v); detection, 265 nm; flow-rate, 1.0 mL/min.

329

330 **Fig. 3** Conventional CCC separation and HPLC analysis to the HSCCC peak fractions.

331 Conditions: stationary phase, the upper phase of chloroform-methanol-0.5mM HCl

332 water solution (4:1.5:2, v/v); 0-200 min, flow-rate 2.0 mL/min, 200-240 min,

333 flow-rate 10.0 mL/min; sample size, 250 mg; detection, 254 nm; stationary phase

334 retention, 66.7% (flow-rate 2.0 mL/min), 52.8% (flow-rate 10.0 mL/min); revolution

335 speed, 800 rpm. HPLC conditions are as in Fig. 2.

336

337 **Fig. 4** PZRCCC separation and HPLC analysis to the HSCCC peak fractions.

338 Conditions: stationary phase, the upper phase of chloroform-methanol-water (4:3:3,

339 v/v); detection, 254 nm; flow-rate, 2.0 mL/min; revolution speed, 800 rpm. I: 30 mM

340 HCl in upper stationary phase and 10 mM TEA in lower phase, sample loading 1.0 g,

341 stationary phase retention 40%; II: 60 mM HCl in upper stationary phase and 10 mM

342 TEA in lower phase, sample loading 1.0 g, stationary phase retention 41.7%; III: 60

343 mM HCl in upper stationary phase and 10 mM TEA in lower phase, sample loading

344 3.0 g, stationary phase retention 36.9%; IV: 60 mM HCl in upper phase and 7.5 mM

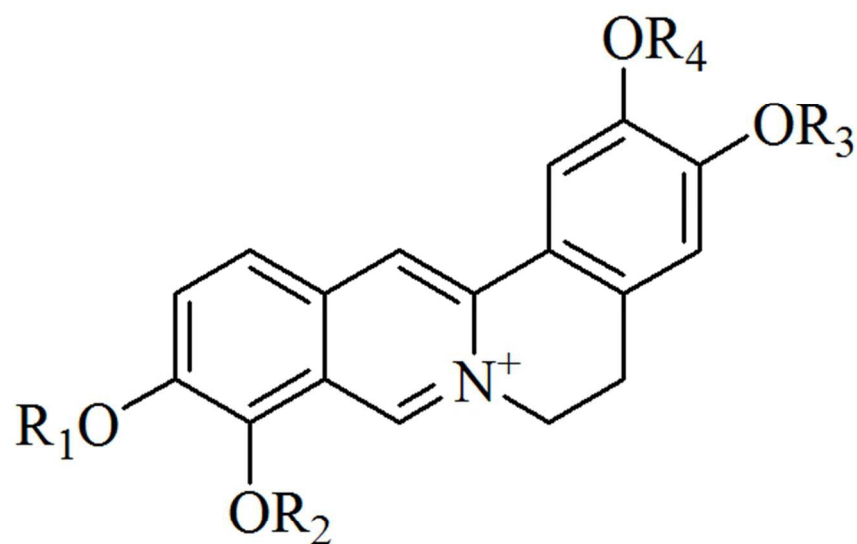
345 TEA in lower phase, sample loading 3.0 g, retention of stationary phase 36.4%.

346

347 **Table 1** The K_D -values of target compounds in different solvent systems

Solvent system	K_D -values of compounds A-E				
	A	B	C	D	E
petroleum ether-ethyl acetate-methanol-water 1:1:1:1.1	0.24	0.32	0.22	0.19	0.33
ethyl acetate- <i>n</i> -butanol-0.5mM HCl water solution 4:1:5	0.07	0.12	0.16	0.11	0.29
chloroform-methanol-0.5mM HCl water solution 4:1.5:2	1.27	4.75	5.50	0.40	0.57

348



	R ₁	R ₂	R ₃	R ₄
stepharanine	H	CH ₃	CH ₃	H
columbamine	CH ₃	CH ₃	CH ₃	H
jatrorrhizine	CH ₃	CH ₃	CH ₃	H
palmatine	CH ₃	CH ₃	CH ₃	CH ₃
berberine	CH ₃	CH ₃	— CH ₂ —	

Fig. 1 Chemical structures of compounds separated from *Caulis Mahoniae*.

59x61mm (300 x 300 DPI)

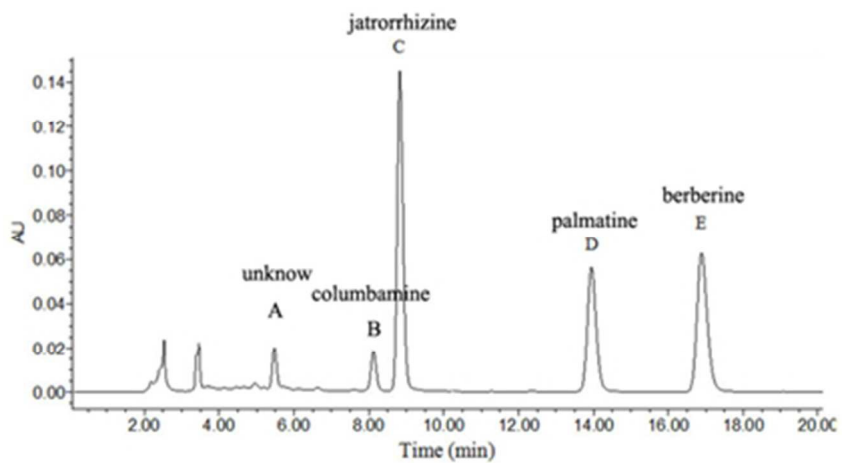


Fig. 2 The HPLC chromatogram of crude alkaloids from *Caulis Mahoniae*. Experimental conditions: column, Waters Symmetry® C18 column (4.6×250 mm, i.d., 5µm); mobile phase, acetonitrile-1% TEA (adjust the pH value to 3 with phosphoric acid) solution (25:75, v/v); detection, 265 nm; flow-rate, 1.0 mL/min.

36x19mm (300 x 300 DPI)

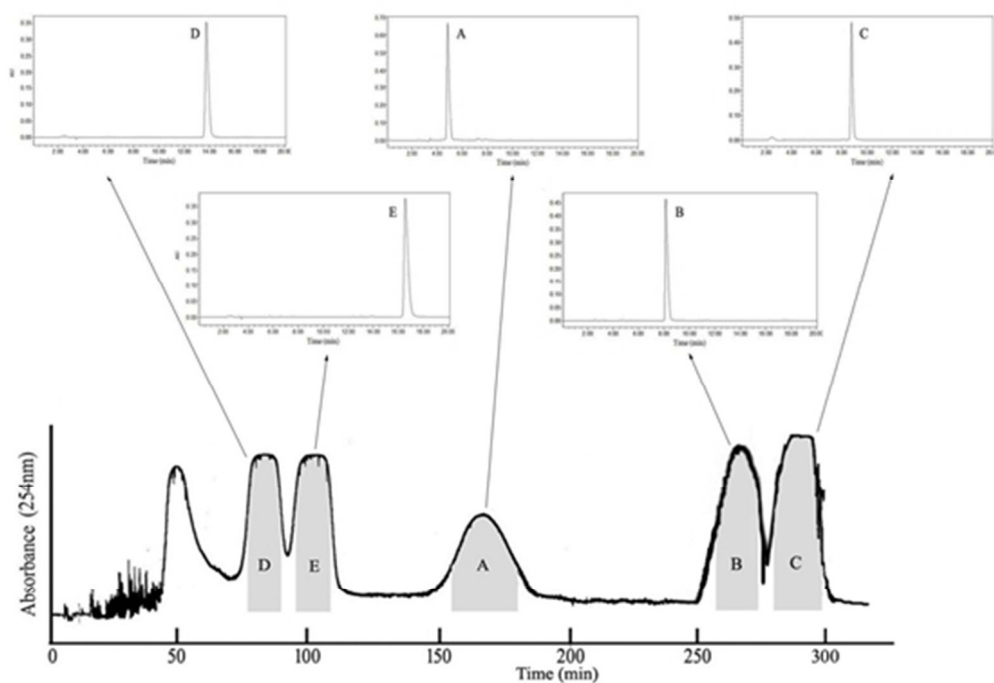


Fig. 3 Conventional HSCCC separation and HPLC analysis to the HSCCC peak fractions. Conditions: stationary phase, the upper phase of chloroform-methanol-0.5mM HCl water solution (4:1.5:2, v/v); 0-200 min, flow-rate 2.0 mL/min, 200-240 min, flow-rate 10.0 mL/min; sample size, 250 mg; detection, 254 nm; stationary phase retention, 66.7%; revolution speed, 800 rpm. HPLC conditions are as in Fig. 2.

46x33mm (300 x 300 DPI)

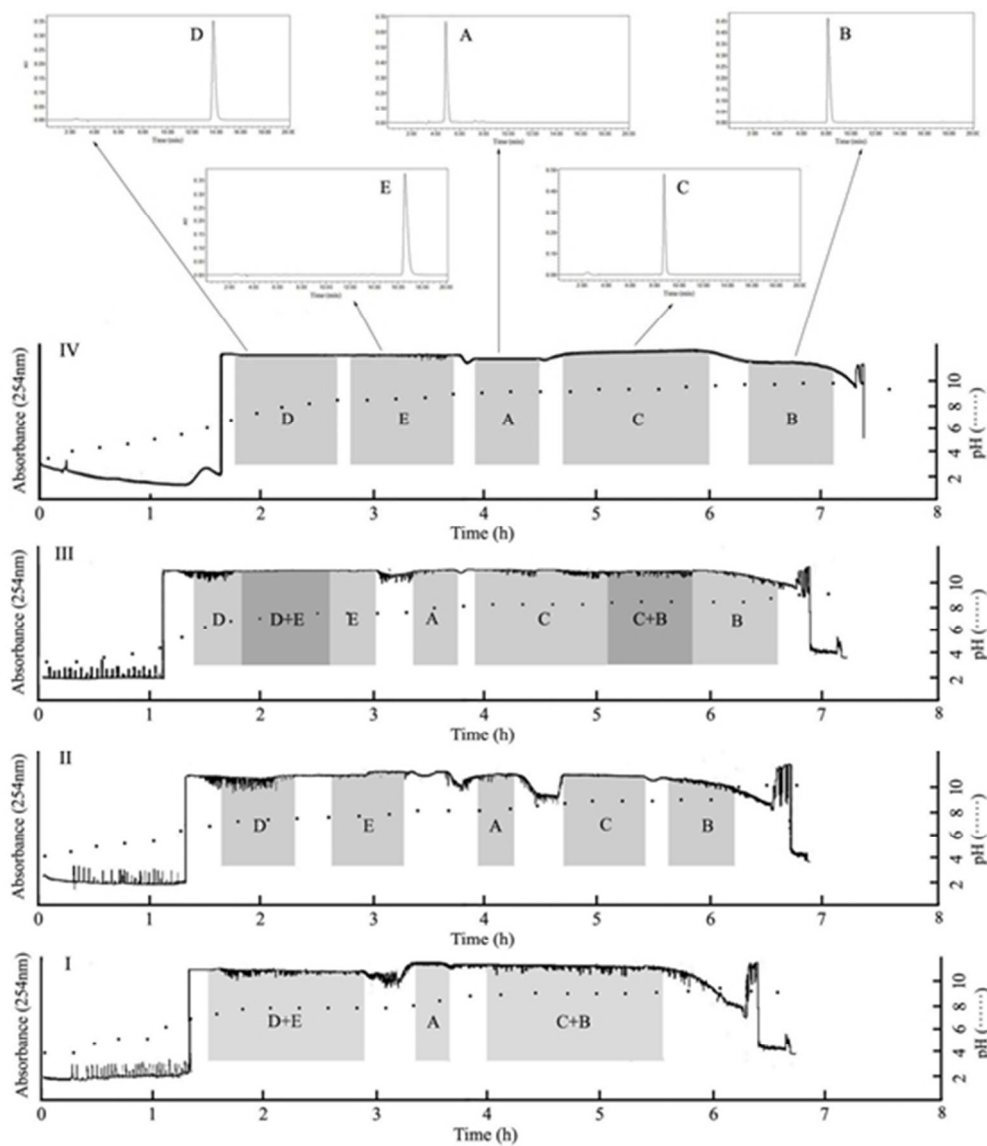
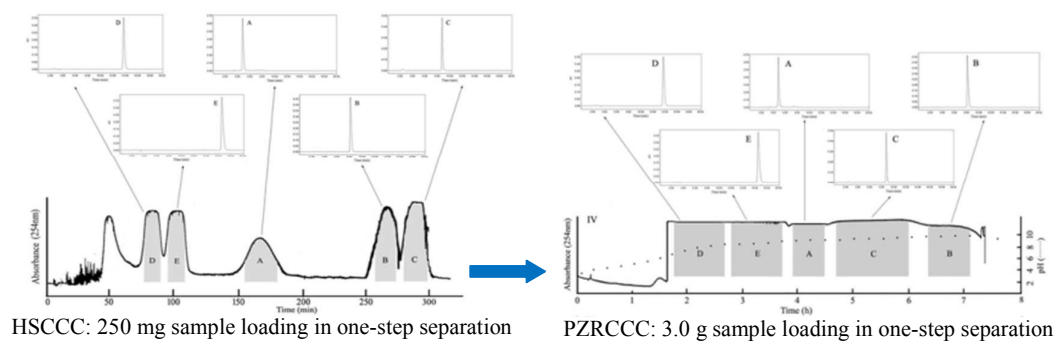


Fig. 4 PZRCCC separation and HPLC analysis to the HSCCC peak fractions. Conditions: stationary phase, the upper phase of chloroform-methanol-water (4:3:3, v/v); detection, 254 nm; flow-rate, 2.0 mL/min; revolution speed, 800 rpm. I: 30 mM HCl in upper stationary phase and 10 mM TEA in lower phase, sample loading 1.0 g, stationary phase retention 40%; II: 60 mM HCl in upper stationary phase and 10 mM TEA in lower phase, sample loading 1.0 g, stationary phase retention 41.7%; III: 60 mM HCl in upper stationary phase and 10 mM TEA in lower phase, sample loading 3.0 g, stationary phase retention 36.9%; IV: 60 mM HCl in upper phase and 7.5 mM TEA in lower phase, sample loading 3.0 g, retention of stationary phase 36.4%.

47x53mm (300 x 300 DPI)

Graphical Abstracts



Compared with the conventional HSCCC, PZRCCC is an efficient and rapid method for separation alkaloids of quaternary ammonium because of the high concentration of fractions and large sample loading capacity