

JAAS

Novel method for measuring ultra-trace levels of U and Th in Au, Pt, Ir, and W matrices using ICP-QQQ-MS employing an O2 reaction gas

| Journal: | Journal of Analytical Atomic Spectrometry |
|----------------------------------|---|
| Manuscript ID | JA-ART-05-2020-000220.R1 |
| Article Type: | Paper |
| Date Submitted by the Author: | 28-Aug-2020 |
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| Table 1. Tuning and acquisition parameters applied for the mass shift method. Channels used for analyte |
|---|
| and tracer measurements are italicized. |

| parameter | condition |
|--|------------------------|
| single quad mode (interference chara | ucterization) |
| RF power (W) | 1600 |
| nebulizer gas (L/min) | 0.69 |
| makeup gas (L/min) | 0.17 |
| m/z | acq. time (s) |
| 227-239 | 1.0 |
| replicates | 3 |
| sweeps/replicate | 10 |
| MS/MS mode ^a (measurement accurac | cy and detection limit |
| O_2 gas flow (mL/min) | 0.38 |
| He gas flow (mL/min) | 0 |
| $m/z (O1 \rightarrow O2)$ | acq. time (s) |
| 227 -> 227 | 2.0 |
| $228 \rightarrow 228$ | 2.0 |
| $229 \rightarrow 229$ | 2.0 |
| $229 \rightarrow 245$ | 15.0 |
| $230 \rightarrow 230$ | 2.0 |
| $231 \rightarrow 247$ | 5.0 |
| $231 \rightarrow 263$ | 5.0 |
| $232 \rightarrow 232$ | 2.0 |
| $232 \rightarrow 248$ | 15.0 |
| $232 \rightarrow 240$ $233 \rightarrow 233$ | 2.0 |
| $233 \rightarrow 265$ | 15.0 |
| $233 \rightarrow 203$ $234 \rightarrow 234$ | 2.0 |
| $234 \rightarrow 254$ | 5.0 |
| $234 \rightarrow 266$ | 5.0 |
| $235 \rightarrow 235$ | 2.0 |
| $235 \rightarrow 255$ | 2.0 |
| $235 \rightarrow 231$ | 5.0 |
| 233 - 207 | 3.0 2.0 |
| 230 - 230 | 2.0 |
| $236 \rightarrow 252$ | 5.0 |
| $236 \rightarrow 268$ | 5.0 |
| $237 \rightarrow 237$ | 1.0 |
| $237 \rightarrow 253$ | 5.0 |
| $237 \rightarrow 269$ | 5.0 |
| $238 \rightarrow 238$ | 2.0 |
| $238 \rightarrow 270$ | 15.0 |
| 239 → 239 | 2.0 |

a – Reaction gas true flow rate calculated as the product of the fourth cell gas flowrate and the manufacturer provided conversion factor.

Table 2. Inferred polyatomic species detected in the SE solutions in single quad mode.

| mass | Interference species | |
|------|---|--|
| 227 | ¹⁹⁷ Au ¹⁴ N ¹⁶ O, minor Ir and Pt products | |
| 228 | ¹⁹⁶ Pt ¹⁶ O ₂ , ¹⁹² Pt ³⁶ Ar, ¹⁹³ Ir ³⁵ Cl, ¹⁹¹ Ir ³⁷ Cl | |
| 229 | ¹⁹⁴ Pt ³⁵ Cl, minor Pt, Ir, and W products | |
| 230 | ¹⁹⁴ Pt ³⁶ Ar, ¹⁸² W ¹⁶ O ₃ , ¹⁹³ Ir ³⁷ Cl | |
| 231 | ¹⁹¹ Ir ⁴⁰ Ar, ¹⁸³ W ¹⁶ O ₃ , ¹⁸² W ¹⁶ O ₃ H, ¹⁹⁵ Pt ³⁶ Ar | |
| 232 | ¹⁸⁴ W ¹⁶ O ₃ , ¹⁸³ W ¹⁶ O ₃ H, ¹⁹⁷ Au ³⁵ Cl, ¹⁹² Pt ⁴⁰ Ar, ¹⁹⁶ Pt ³⁶ Ar | |
| 233 | ¹⁹³ Ir ⁴⁰ Ar, ¹⁸⁴ W ¹⁶ O ₃ H, ¹⁹⁷ Au ³⁶ Ar | |
| 234 | ¹⁹⁴ Pt ⁴⁰ Ar, ¹⁹⁸ Pt ³⁶ Ar, ¹⁸⁶ W ¹⁶ O ₃ | |
| 235 | ¹⁹⁵ Pt ⁴⁰ Ar, ¹⁸⁶ W ¹⁶ O ₃ H | |
| 236 | ¹⁹⁶ Pt ⁴⁰ Ar, minor W products | |
| 237 | ¹⁹⁷ Au ⁴⁰ Ar, minor W products | |
| 238 | 198 Pt 40 Ar | |
| 239 | ¹⁹¹ Ir ¹⁶ O ₃ | |

| commlol | ²²⁹ Th (| (fg·g ⁻¹) | ²³³ U (| fg·g⁻¹) |
|---|--|--|--|--|
| sample" | target | measured ^b | target | measured |
| blanks | | | | |
| Sp 0-1 | 0 | 0.4 ± 0.4 | 0 | 2.6 ± 1.2 |
| Sp 0-2 | 0 | 0.1 ± 0.2 | 0 | 2.7 ± 0.4 |
| Sp 0-3 | 0 | 0.1 ± 0.2 | 0 | 4.1 ± 1.7 |
| Sp 1-1 | 0.989 ± 0.005 | 0.7 ± 0.8 | 1.09 ± 0.01 | 1.7 ± 1.2 |
| Sp 1-1 | 0.989 ± 0.005 | 0.7 ± 0.8 | 1.09 ± 0.01 | 1.7 ± 1.2 |
| Sm 1 0 | 1.15 ± 0.01 | 1.4 ± 0.5 | 1 776 1 77 771 | 21 + 12 |
| Sp 1-2 | 1.15 ± 0.01 | 1.4 ± 0.5 | 1.26 ± 0.01 | 2.1 ± 1.3 |
| Sp 1-2 Sp 1-3 | 1.15 ± 0.01 1.14 ± 0.01 | 1.4 ± 0.5 0.7 ± 0.9 | 1.26 ± 0.01 1.25 ± 0.01 | 2.1 ± 1.3 1.6 ± 1.2 |
| Sp 1-2 Sp 1-3 Sp 10-1 | 1.15 ± 0.01 1.14 ± 0.01 11.6 ± 0.1 | 1.4 ± 0.5 0.7 ± 0.9 9.6 ± 1.7 | 1.26 ± 0.01 1.25 ± 0.01 12.8 ± 0.1 | 2.1 ± 1.3 1.6 ± 1.2 13.8 ± 1.8 |
| Sp 1-2 Sp 1-3 Sp 10-1 Sp 10-2 | 1.15 ± 0.01 1.14 ± 0.01 11.6 ± 0.1 11.6 ± 0.1 | $1.4 \pm 0.5 \\ 0.7 \pm 0.9 \\ 9.6 \pm 1.7 \\ 9.3 \pm 1.8$ | 1.26 ± 0.01 1.25 ± 0.01 12.8 ± 0.1 12.8 ± 0.1 | 2.1 ± 1.3 1.6 ± 1.2 13.8 ± 1.8 13 ± 2 |
| Sp 1-2 Sp 1-3 Sp 10-1 Sp 10-2 Sp 10-3 | 1.15 ± 0.01 1.14 ± 0.01 11.6 ± 0.1 11.6 ± 0.1 11.6 ± 0.1 | 1.4 ± 0.5 0.7 ± 0.9 9.6 ± 1.7 9.3 ± 1.8 10.0 ± 1.6 | 1.26 ± 0.01 1.25 ± 0.01 12.8 ± 0.1 12.8 ± 0.1 12.8 ± 0.1 12.8 ± 0.1 | 2.1 ± 1.3 1.6 ± 1.2 13.8 ± 1.8 13 ± 2 14.1 ± 1.8 |
| Sp 1-2 Sp 1-3 Sp 10-1 Sp 10-2 Sp 10-3 Sp 100-1 | 1.15 ± 0.01 1.14 ± 0.01 11.6 ± 0.1 11.6 ± 0.1 11.6 ± 0.1 10.1 ± 1 | 1.4 ± 0.5 0.7 ± 0.9 9.6 ± 1.7 9.3 ± 1.8 10.0 ± 1.6 101 ± 6 | 1.26 ± 0.01 1.25 ± 0.01 12.8 ± 0.1 12.8 ± 0.1 12.8 ± 0.1 101 ± 1 | $2.1 \pm 1.3 \\ 1.6 \pm 1.2 \\ 13.8 \pm 1.8 \\ 13 \pm 2 \\ 14.1 \pm 1.8 \\ 99 \pm 4$ |
| Sp 1-2 Sp 1-3 Sp 10-1 Sp 10-2 Sp 10-3 Sp 100-1 Sp 100-2 | 1.15 ± 0.01 1.14 ± 0.01 11.6 ± 0.1 11.6 ± 0.1 11.6 ± 0.1 101 ± 1 101 ± 1 | 1.4 ± 0.5 0.7 ± 0.9 9.6 ± 1.7 9.3 ± 1.8 10.0 ± 1.6 101 ± 6 104 ± 4 | 1.26 ± 0.01 1.25 ± 0.01 12.8 ± 0.1 12.8 ± 0.1 12.8 ± 0.1 101 ± 1 101 ± 1 | 2.1 ± 1.3 1.6 ± 1.2 13.8 ± 1.8 13 ± 2 14.1 ± 1.8 99 ± 4 91 ± 3 |

Table 3. Summary of the method accuracy analyses conducted in ME solutions using the natural isotopes of U and Th as the tracer.

a –Samples (or Sp) are labeled as Sp-x-y, where 'x' is the intended U and Th concentration and 'y' is the replicate number

b – The error is the standard deviation of a triplicate measurement

| sample | replicate | weight (g) | ²³² Th (pg·g⁻¹) | LOD (pg·g⁻¹) | ²³⁸ U (pg·g⁻¹) | LOD (pg·g⁻¹) |
|-----------------------------|-----------|------------|----------------------------|--------------|---------------------------|--------------|
| | | | | | | |
| direct crimp | 1 | 0.3142 | 35 ± 7 | 5 | 41 ± 5 | 1 |
| (Axon'Cable) | 2 | 0.2945 | 63 ± 8 | 6 | 56 ± 7 | 1 |
| board mount (Axon'Cable) | 1 | 0.1873 | 99 ± 10 | 10 | 120 ± 11 | 2 |
| and handing wire | 1 | 0.0997 | 18 ± 14 | 17 | 52 ± 6 | 8 |
| | 2 | 0.2295 | 27 ± 7 | 7 | 190 ± 15 | 3 |
| (Ametek Inc) | 3 | 0.2572 | 34 ± 5 | 7 | 54 ± 8 | 3 |
| | 1 | 0.0162 | 12.8 ± 0.7 | 1 | 9.9 ± 1.0 | 2 |
| SQUID arrays (NIST) | 2 | 0.0173 | 14.1 ± 0.7 | 1 | 8.6 ± 0.9 | 1 |
| | 3 | 0.0155 | 19 ± 2 | 1 | 15 ± 2 | 2 |

Novel method for measuring ultra-trace levels of U and Th in Au, Pt, Ir, and W matrices using ICP-QQQ MS employing an O₂ reaction gas

3 Khadouja Harouaka, Eric W. Hoppe, Isaac J. Arnquist

5 Abstract

Increased demand for improving ultra-low background detection capabilities for rare-event fundamental physics applications has resulted in the need for fast, convenient and clean assay methodologies that either preclude or reduce chemical sample pre-processing. A novel method for the measurement of ultratrace concentrations (fg·g⁻¹ level) of natural ²³²Th and ²³⁸U and non-natural tracer isotopes ²²⁹Th and ²³³U were measured in a solution of 10 µg·g⁻¹ each of Au, Pt, Ir, and W in 2% HNO₃ using an ICP-QQQ-MS. Polyatomic interferences across a m/z range of 227-239 were characterized: the major interferents with ²²⁹Th⁺ is ¹⁹⁴Pt³⁵Cl⁺; with ²³²Th⁺ are ¹⁸⁴W¹⁶O₃⁺, ¹⁸³W¹⁶O₃H⁺, ¹⁹²Pt⁴⁰Ar⁺, ¹⁹⁶Pt³⁶Ar⁺, ¹⁹⁵Pt³⁷Cl⁺, and ¹⁹⁷Au³⁵Cl⁺; with 233 U⁺ are 193 Ir⁴⁰Ar⁺, 197 Au³⁶Ar⁺, 184 W¹⁶O₃H⁺; and with 238 U⁺ is 198 Pt⁴⁰Ar⁺. Scanning the selected *m/z* range of 227-270 showed that higher oxide polyatomic species from the matrix elements either did not form or did not create significant background on the target analyte masses. All measured concentrations in standard solutions matched the target values within the 98% confidence interval. The Th measurements were 80% accurate or better at the 10 $\text{fg}\cdot\text{g}^{-1}$ level and above, and the U measurements were 90% accurate or better at the 10 $fg \cdot g^{-1}$ level and above. Measurements at the 1 $fg \cdot g^{-1}$ level were consistent with target values within 1 standard deviation, although the standard deviations of all three replicates were greater than 20% of the measured concentration value. Method detection limits in the matrix solutions were calculated to be 2.74 fg Th and 12.9 fg U. In an electronic sample, which typically have 0.1% precious metal content, our method would give detection limits of 274 fg Th and 1291 fg U given a maximum of 10 μ g/g⁻ ¹ coinage metal matrix. This method is but one example of how state of the art quadrupole mass spectrometry and collision reaction cell technology can be leveraged to develop novel analytical capability at ultra-trace levels.

28 1. Introduction

The advent of commercial *elemental* tandem mass spectrometry (MS/MS) instrumentation has given rise to multiple opportunities for method development aimed at analytes in challenging matrices that either reduce or eliminate the need for extensive sample pre-processing procedures that are both time-consuming and prone to contamination. Such methodologies are of great importance to fundamental high energy and nuclear physics next-generation "rare event" experiments (e.g., dark matter detection, neutrinoless double beta decay, and solar neutrino detection) that require ultra-high purity, low-background detector materials to reach their sensitivity target goals e.g., (1-3). Materials used in detectors undergo high sensitivity assays to accurately model background levels from inherent and/or imparted radiocontamination e.g., (4–10). The main contributors to backgrounds arise from naturally 39 occurring ²³⁸U and ²³²Th contaminants (the focus of this study) and their progeny, but also ⁴⁰K and 40 cosmogenic activation products, *e.g.*, ⁶⁰Co. Typically, the highest sensitivity chemical assays are conducted 41 with inductively coupled plasma mass spectrometry (ICP-MS), which, with meticulous sample 42 preparations and tailored tuning, can measure as low as $fg \cdot g^{-1}$ levels of ²³⁸U and ²³²Th in a variety of 43 materials such as electroformed copper *e.g.*, (10–13).

Some target materials are often comprised of metals that make achieving ultrasensitive measurements using ICP-MS problematic as polyatomic interferences may overlap with the target analyte masses of ²³²Th and ²³⁸U as well as ²²⁹Th and ²³³U that are used as isotope dilution spikes for quantitation of ²³²Th and ²³⁸U. For example, electric contacts, pins, and printed circuit boards (PCBs) in electronic components, metallization on application-specific integrated components (ASICs) and silicon photomultipliers (SiPMs), as well as common resistors, diodes, and superconducting quantum interference device (SQUID) arrays oftentimes contain coinage metals such as Au, Pt, Ir, and W which create polyatomic interferences within the *m/z* range of 227-238 *e.g.*, (14–18).

Polyatomic interferences can sometimes be avoided by diluting the sample sufficiently, applying instrumental tuning strategies (e.q., use of dry or cool plasma), or by removing the offending elements during sample preparations (e.g., via ion exchange chromatography, precipitation, etc.). Dilution is typically not viable when attempting to achieve $pg \cdot g^{-1}$ to $fg \cdot g^{-1}$ level measurements as the analyte can be diluted to below detection limits. Chromatographic separations are time-consuming, require a suite of reagents, and are not straightforward for removing Au, Pt, and Ir as they have the same retention as U and Th in most ion exchange resins due to their similar chemical affinities (19-21). While a two-column method could be used for total removal (22), such methods are further time-consuming and create multiple avenues for contamination given the significant effort required for removing backgrounds from ion exchange resins to achieve fg·g⁻¹ levels of detection (11). Furthermore, digestions of samples containing these metals often requires the need for HCl or aqua regia, which can produce increased Th

and U background when introduced to the ion exchange resin, even in trace quantities, and/or create
matrices with insufficient resin affinity. Furthermore, the use of HCl and HCl containing acids is also the
source of chloride polyatomic species that overlap with the target analytes. Minimizing sample
preparation procedures is critical for eliminating sources of contamination and increasing sample
throughput.

To overcome these challenges, we instead developed a mass shift method where an O₂ reaction gas is reacted with ²³³U, ²³⁸U, ²²⁹Th, and ²³²Th to form monoxide (at *m/z* 245, 248, 249, and 254) and/or dioxide species (at *m/z* 261, 264, 265, 270). The synthetic isotopes of ²³³U and ²²⁹Th in the tracer (at *m/z* 245 and 261 for the single and dioxide of Th, and *m/z* 249 and 265 for the monoxide and dioxide of U, respectively) are used to quantify the oxides of the natural ²³⁸U and ²³²Th in the sample following standard isotope dilution methods.

2. Experimental

75 2.1 Reagents and labware

Optima grade HNO₃ (Fisher Scientific; Pittsburgh, PA) and 18.2 MΩ·cm deionized water was used
to make all reagents used in this study. Elemental standards from HPS (North Charleston, SC) and Inorganic
Ventures (Christiansburg, VA) were used to make the matrix solutions. The single element (SE) standards
used were 1000 µg·g⁻¹ each of Au in 2% HNO₃, Pt in 5% HCl, Ir in 2% HCl, and W in 2% HNO₃ and 1% HF.
The standard solutions were prepared gravimetrically by diluting the 1000 µg·g⁻¹ stock to 10 µg·g⁻¹ with
2% HNO₃. Laboratory work was conducted in a class 10000 cleanroom located at Pacific Northwest
National Laboratory (Richland WA, USA).

In order to ensure cleanliness and achieve ultralow background detection limits for Th and U, PFA
vials (Savillex, Eden Prairie, MN) were used both for sample digestions, dilutions, and as autosampler vials.
Vials were initially cleaned by subsequent leaching in 25% HCl and 40% HNO₃ overnight at 80°C. The vials
were validated by adding 1.8 mL of 5% HNO₃ to each vial, allowing them to leach overnight at 80°C, and

then measuring the natural and synthetic Th and U background in the leachate. Vials were considered suitable for use in this study when the measured leachate solutions were at background levels for pure 5% HNO₃ blanks.

2.2 Instrumentation

An Agilent 8900 inductively coupled plasma triple quadrupole mass spectrometer (ICP-QQQ-MS; Agilent Technologies, Santa Clara, CA, USA) with integrated autosampler, Ni skimmer and sampler cones, s-lens, and a standard electron multiplier detector was used for all measurements conducted in this study. The instrument was equipped with a quartz double-pass spray chamber and a 100 μL·min⁻¹ microflow PFA nebulizer (Elemental Scientific, Omaha, NE, USA). The tuning parameters for optimal signal sensitivity and stability were first determined in single quad mode by tuning the mass analyzer and ion optics with a 0.1 $ng \cdot g^{-1} = 205 Tl$ standard. The instrument was then switched to MS/MS mode with O₂ gas enabled, and the ion optics and collision cell parameters were refined for maximum oxide generation by observing the relative signals of YO and Y (at m/z 105 and 89) and of CeO and Ce (at m/z of 156 and 140) in a 0.1 ng·g⁻¹ standard. Once optimized, a 500 fg·g⁻¹ solution of natural ²³⁸U and ²³²Th was used to double-check the maximum sensitivity of both the monoxide and dioxide product species. All m/z collected in single quad mode had 1 second integration times. The selected m/z collected in MS/MS mode capture both the highest sensitivity analyte species of UO_2 and ThO. Additional m/z were selected in order to monitor for overlapping polyatomic interferences and their oxides in the range of the U and Th analytes. Sample acquisition times varied by the signal intensity at a given m/z to ensure higher quality measurements; 15 s was allotted to the target analytes UO₂ and ThO, 5 s was allotted to the m/z of the oxides of the polyatomics that directly interfere with the target analytes, and 1-2 s was allotted to the m/z of the unreacted polyatomics. A summary of the instrumental parameters is described in Table 1.

Table 1. Tuning and acquisition parameters applied for the mass shift method. Channels used for the analyte and tracer are italicized.

condition

parameter

| single quad mode (interference chard | acterization) | |
|---|---|-------------|
| RF power (W) | 1600 | |
| nebulizer gas (L/min) | 0.69 | |
| makeup gas (L/min) | 0.17 | |
| m/z | acq. time (s) | |
| 227-239 | 1.0 | |
| replicates | 3 | |
| sweeps/replicate | 10 | |
| MS/MS mode ^a (measurement accura | cy and detection limit) | |
| Ω_2 gas flow (mI /min) | 0.38 | |
| He gas flow (mL/min) | 0 | |
| $m/z (\Omega_1 \rightarrow \Omega_2)$ | acq_time(s) | |
| $227 \rightarrow 227$ | 20 | |
| $227 \rightarrow 227$ | 2.0 | |
| $220 \rightarrow 220$ | 2.0 | |
| $229 \rightarrow 245$ | 15.0 | |
| $227 \rightarrow 243$ | 2.0 | |
| $230 \rightarrow 247$ | 5.0 | |
| $231 \rightarrow 263$ | 5.0 | |
| $231 \rightarrow 203$ | 2.0 | |
| $232 \rightarrow 248$ | 15 0 | |
| $232 \rightarrow 240$ $233 \rightarrow 233$ | 2.0 | |
| $233 \rightarrow 265$ | 15.0 | |
| $233 \rightarrow 203$ | 2.0 | |
| $234 \rightarrow 250$ | 5.0 | |
| $234 \rightarrow 266$ | 5.0 | |
| $237 \rightarrow 235$ | 2.0 | |
| $235 \rightarrow 251$ | 5.0 | |
| $235 \rightarrow 267$ | 5.0 | |
| $235 \rightarrow 236$ | 2.0 | |
| $236 \rightarrow 252$ | 5.0 | |
| $236 \rightarrow 268$ | 5.0 | |
| $237 \rightarrow 237$ | 1.0 | |
| $237 \rightarrow 253$ | 5.0 | |
| $237 \rightarrow 269$ | 50 | |
| $238 \rightarrow 238$ | 20 | |
| $238 \rightarrow 270$ | 15.0 | |
| $230 \rightarrow 230$ | 2.0 | |
| 2577257 | culated as the product of the fourth cell | as flowrate |
| manufacturer provided conversion for | tor | bus nowrate |

A stock multi-element standard solution (ME solution) containing 10 µg·g⁻¹ of Au, Pt, Ir, and W was prepared from single element ICP standards. Single element 10 µg·g⁻¹ stock solutions (SE solutions) of the same elements were also prepared. In order to characterize the matrix-derived polyatomic interfering species, both the 10 µg·g⁻¹ ME and SE solutions were measured individually in single quad mode by scanning across m/z from 227 to 239. A series of ME solutions at 0.1, 1, 5, and 10 µg·g⁻¹ were measured to determine the signal sensitivity of the polyatomic species as well as the required washout time. In the end, 10 µg·g⁻¹ was selected as the maximum concentration for our experiments in order to ensure a high enough signal such that all polyatomic species formed could be detected but also low enough in concentration to not cause significant detriment to the instrument (e.g., coating cones/optics, long carryover).

Measurement accuracy and method detection limits were determined by spiking the 10 $\mu g \cdot g^{-1} ME$ solution with a known amount of non-natural ²³³U and ²²⁹Th as isotope dilution tracers and using the natural ²³⁸U and ²³²Th inherent in the ME solution as the spike for isotope dilution quantitation. This method was selected given the expectation that the standard solutions would have natural ²³⁸U and ²³²Th well above the fg·g⁻¹ concentration levels we wished to interrogate, which was confirmed through measurement as the ME solution had 3.06 \pm 0.05 and 1.60 \pm 0.03 pg·g⁻¹ for ²³²Th and ²³⁸U, respectively (error is 1SD, n=3) after verifying that the polyatomic interferences were sufficiently avoided with the mass shift method. A portion of the stock ME solution was spiked with 100 fg·g⁻¹ of ²²⁹Th and ²³³U tracer and was measured in triplicate in order to quantify the concentration of natural ²³²Th and ²³⁸U inherent to the ME stock. Quantitation followed the standard isotope dilution method (Eqn. 1).

 $C_{analyte} = \frac{I_{analyte}C_{tracer}}{I_{tracer}}$ Eqn. 1

Here, 'C' is the concentration and 'I' is the measured signal intensity of the oxide of the analyte and
tracer isotope. Once determined with the aforementioned high degree of certainty, the concentration of

| was used to quantify the synthetic ²³³ U and ²²⁹ Th in subsequent mined by comparing the measured concentration of ²³³ U and ²²⁹ Th orepared samples. The samples were prepared in triplicate, by h 1, 10, and 100 fg·g ⁻¹ of synthetic ²³³ U and ²²⁹ Th gravimetrically. ³ U and ²²⁹ Th were used as blanks to determine the detection limits. culated as 3 times the standard deviation of the triplicate easurement accuracy is determined simply by comparing the blank |
|--|
| mined by comparing the measured concentration of ²³³ U and ²²⁹ Th orepared samples. The samples were prepared in triplicate, by h 1, 10, and 100 fg·g ⁻¹ of synthetic ²³³ U and ²²⁹ Th gravimetrically. ³ U and ²²⁹ Th were used as blanks to determine the detection limits. culated as 3 times the standard deviation of the triplicate easurement accuracy is determined simply by comparing the blank |
| mined by comparing the measured concentration of ²³³ U and ²²⁹ Th prepared samples. The samples were prepared in triplicate, by h 1, 10, and 100 fg·g ⁻¹ of synthetic ²³³ U and ²²⁹ Th gravimetrically. ³ U and ²²⁹ Th were used as blanks to determine the detection limits. culated as 3 times the standard deviation of the triplicate easurement accuracy is determined simply by comparing the blank |
| prepared samples. The samples were prepared in triplicate, by h 1, 10, and 100 fg·g ⁻¹ of synthetic ²³³ U and ²²⁹ Th gravimetrically. ³ U and ²²⁹ Th were used as blanks to determine the detection limits. culated as 3 times the standard deviation of the triplicate easurement accuracy is determined simply by comparing the blank |
| h 1, 10, and 100 fg·g ⁻¹ of synthetic ²³³ U and ²²⁹ Th gravimetrically. ³ U and ²²⁹ Th were used as blanks to determine the detection limits. culated as 3 times the standard deviation of the triplicate reasurement accuracy is determined simply by comparing the blank |
| ³ U and ²²⁹ Th were used as blanks to determine the detection limits. culated as 3 times the standard deviation of the triplicate reasurement accuracy is determined simply by comparing the blank |
| culated as 3 times the standard deviation of the triplicate as a stimes the standard deviation of the triplicate |
| asurement accuracy is determined simply by comparing the blank |
| |
| ents with the target value following Eqn. 2. |
| $\left \frac{C_m-C_t}{C_t}\right $) Eqn. 2 |
| easured concentration and C_t is the target concentration. The 95% |
| e triplicate measurements were also calculated and compared with |
| |
| |
| |
| n from matrix elements across the m/z range of 227 to 239 in single |
| intensity of the background interferences on m/z 229, 232, 233 and |
| ne sum of the backgrounds from the SE solutions is in good |
| al of the ME solution, indicating that the formation of the |
| ntly affected by the presence of the other metal ions in the matrix. |
| nces was deduced by determining the major products formed from |
| andards. In most cases, the product species can be inferred from the |
| paring the ratio of the signal of the isotopes with the natural |
| |
| |

| ~ | |
|--|--------------------------|
| 3 4 | 162 |
| 5 6 | 163 |
| 7 8 | 164 |
| 9 10 | 165 |
| 11 12 12 | 166 |
| 13 14 15 | 167 |
| 16 17 | 168 |
| 18 19 | 169 |
| 20 21 | 170 |
| 24 25 26 27 28 29 30 31 32 33 43 5 36 37 38 39 40 41 42 43 44 50 51 52 53 54 55 56 7 8 9 | 171 172 173 174 |

| | pecies, which are arbitrarily defined for identification purposes as species that form at an | | | | | |
|--|---|--|--|--|--|--|
| intensity great | ter than 100 cps, are listed in Table 2. The determination of these interferences was not | | | | | |
| always straigh | tforward in cases where there is multiple overlap between polyatomic species (especially | | | | | |
| with respect to oxides and hydrides). In these cases, the combined signal of what are likely to be several | | | | | | |
| minor products produce a detectable signal that can be significant (<i>i.e.,</i> 100 cps or greater). It is | | | | | | |
| important to consider the characterization of interferences to at least these levels (100 cps) given that | | | | | | |
| ypical sensitiv | vities in ultra-trace measurements can reach around 2000 cps/pg \cdot g \cdot 1 for U and Th in our | | | | | |
| instrument. | | | | | | |
| Table 2. Inferr | red polyatomic species detected in the SE solutions in single quad mode. | | | | | |
| mass | Interference species | | | | | |
| 227 | ¹⁹⁷ Au ¹⁴ N ¹⁶ O, minor Ir and Pt products | | | | | |
| 228 | ¹⁹⁶ Pt ¹⁶ O ₂ , ¹⁹² Pt ³⁶ Ar, ¹⁹³ Ir ³⁵ Cl, ¹⁹¹ Ir ³⁷ Cl | | | | | |
| 229 | ¹⁹⁴ Pt ³⁵ Cl, minor Pt, Ir, and W products | | | | | |
| 230 | ¹⁹⁴ Pt ³⁶ Ar, ¹⁸² W ¹⁶ O ₃ , ¹⁹³ Ir ³⁷ Cl | | | | | |
| 231 | ¹⁹¹ Ir ⁴⁰ Ar, ¹⁸³ W ¹⁶ O ₃ , ¹⁸² W ¹⁶ O ₃ H, ¹⁹⁵ Pt ³⁶ Ar | | | | | |
| 232 | ¹⁸⁴ W ¹⁶ O ₃ , ¹⁸³ W ¹⁶ O ₃ H, ¹⁹⁷ Au ³⁵ Cl, ¹⁹² Pt ⁴⁰ Ar, ¹⁹⁶ Pt ³⁶ Ar | | | | | |
| 233 | ¹⁹³ Ir ⁴⁰ Ar, ¹⁸⁴ W ¹⁶ O ₃ H, ¹⁹⁷ Au ³⁶ Ar | | | | | |
| 234 | ¹⁹⁴ Pt ⁴⁰ Ar, ¹⁹⁸ Pt ³⁶ Ar, ¹⁸⁶ W ¹⁶ O ₃ | | | | | |
| 235 | ¹⁹⁵ Pt ⁴⁰ Ar, ¹⁸⁶ W ¹⁶ O ₃ H | | | | | |
| 236 | ¹⁹⁶ Pt ⁴⁰ Ar, minor W products | | | | | |
| 237 | ¹⁹⁷ Au ⁴⁰ Ar, minor W products | | | | | |
| 238 | ¹⁹⁸ Pt ⁴⁰ Ar | | | | | |
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their hydrides, PtO₂ and their hydrides, PtNO, and PtCl, as well as WNO, WFO, and WO₃H₂ across the

m/z range of 227-238.



Figure 1. Matrix-derived interferences across *m/z* of 227 to 239 measured in single quad mode.

When operating in MS/MS mode, the overall signal intensity drops by roughly a factor of 3, as seen in (Fig. 2). The oxide interferences are not affected by the presence of O_2 . While the argides of Pt and Au appear to react with O_2 to form argon oxide species, only the most abundant species are even detectable at m/z 250-253 with intensities of <10 cps (Fig. 2). The interference signals at m/z 250-253, and lack of signal at m/z 266-269 suggest that the formation of argon *monoxide* species is favored compared to the argon *dioxide* species.

Upon optimizing the instrument optics and reaction cell parameters for maximum oxide formation of U and Th species in the reaction cell, it was found that the monoxide for Th (ThO) and dioxide for U (UO₂) gave the highest intensity signal. It is unlikely that there are significant polyatomic interferences with our target analytes of ThO and UO₂. The main interferent on ²²⁹Th¹⁶O would be

 194 Pt³⁵Cl¹⁶O. However, it is not likely to have formed given the absence of signal at m/z 247 where the

almost equally abundant ¹⁹⁶Pt³⁵Cl¹⁶O species would be detected. Similarly, the main interference species

on ²³²Th¹⁶O at *m/z* 248 would be ¹⁸⁴W¹⁶O₄, ¹⁸³W¹⁶O₄H, ¹⁹⁷Au³⁵Cl¹⁶O, ¹⁹⁵Pt³⁷Cl¹⁶O, and/or ¹⁹²Pt⁴⁰Ar¹⁶O. We

can preclude the presence of tetraoxides of W due to the absence of signal at mass 247, which would

correspond to the abundant species of ¹⁸³W¹⁶O₄ and ¹⁸²W¹⁶O₄H. The presence of ¹⁹⁵Pt³⁷Cl¹⁶O overlapping

with 232 Th¹⁶O is also precluded by the absence of 196 Pt³⁵Cl¹⁶O at m/z 247. While PtArO species are clearly

formed on m/z 250-252, it is unlikely that ¹⁹²Pt⁴⁰Ar¹⁶O is a significant interferent on ²³²Th¹⁶O as ¹⁹²Pt is a

minor isotope of Pt (0.78% natural abundance) and the major PtArO species are only detectable at 5-6

cps. We cannot directly infer if the ¹⁹⁷Au³⁷Cl¹⁶O species forms on *m/z* 248 by looking for ¹⁹⁷Au³⁷Cl¹⁶O at

m/z 250 due to overlap with ¹⁹⁴Pt⁴⁰Ar¹⁶O. We can, however, surmise that if it does form, it would be of a

lower intensity than the more abundant $^{197}Au^{40}Ar^{16}O$, which forms at m/z 253 with an intensity of just 7

cps. Finally, we can also determine from the data that IrArO species do not form under our experimental

No signal was observed at m/z 266-269, which precludes the significant formation of any

polyatomic dioxide interference species in the range of the UO_2 analytes. This result points to the clear

advantage of employing a mass shift method where the analyte species is in the dioxide form when

instrumentation and can therefore serve as incentive to the production of elemental instrumentation

that can analyze up to an m/z of 275 or higher. This especially applies to all mass shift methods with

analytes that react to form particularly large adducts when reacted with reactive gases (e.g., BiCS₂; (24))

We have demonstrated that the O_2 reaction gas method sufficiently removes overlapping

polyatomic interferences from our analytes and is therefore a possible method for ultra-trace Th and U

determinations. The following section addresses the viability of such an ultra-trace analysis.

possible. The utility in measuring the dioxide of the analyte species is limited by commercial

conditions due to the absence of signal at m/z 247.

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| | Sp 0-1 | 0 | 0.4 ± 0.4 | 0 | 2.6 ± 1.2 | |
| | Sp 0-2 | 0 | 0.1 ± 0.2 | 0 | 2.7 ± 0.4 | |
| | Sp 0-3 | 0 | 0.1 ± 0.2 | 0 | 4.1 ± 1.7 | |
| | blank subtracted s | amples | | | | |
| | Sp 1-1 | 0.989 ± 0.005 | 0.7 ± 0.8 | 1.09 ± 0.01 | 1.7 ± 1.2 | |
| | Sp 1-2 | 1.15 ± 0.01 | 1.4 ± 0.5 | 1.26 ± 0.01 | 2.1 ± 1.3 | |
| | Sp 1-3 | 1.14 ± 0.01 | 0.7 ± 0.9 | 1.25 ± 0.01 | 1.6 ± 1.2 | |
| | Sp 10-1 | 11.6 ± 0.1 | 9.6 ± 1.7 | 12.8 ± 0.1 | 13.8 ± 1.8 | |
| | Sp 10-2 | 11.6 ± 0.1 | 9.3 ± 1.8 | 12.8 ± 0.1 | 13 ± 2 | |
| | Sp 10-3 | 11.6 ± 0.1 | 10.0 ± 1.6 | 12.8 ± 0.1 | 14.1 ± 1.8 | |
| | Sp 100-1 | 101 ± 1 | 101 ± 6 | 101 ± 1 | $99 \hspace{0.1in} \pm 4$ | |
| | Sp 100-2 | 101 ± 1 | 104 ± 4 | 101 ± 1 | 91 ± 3 | |
| | Sp 100-3 | 100 ± 1 | 108 ± 6 | 100 ± 1 | 96 ± 4 | |
| r t | a -samples (of sp) are labeled as sp-x-y, where x is the intended of and in concentration and y is replicate number b –The error is the standard deviation of a triplicate measurement All measurements matched the target value within the 98% confidence limit. The meth | | | | | |
| accuracy results indicate that the mass shift method can determine | | | | | | |
| accuracy at 100 fg·g ⁻¹ levels, with 83% accuracy at 10 fg·g ⁻¹ levels, and with 73% accuracy at the 1 f | | | | | | |
| C | concentration rang | e. Similarly, ²³³ U can b | e measured with 9 | 5% accuracy at 100 f | g·g⁻¹ levels, with | |
| ĉ | accuracy at 10 fg g | ¹ levels, and with 51% | accuracy at 1 fg⋅g ⁻¹ | levels (Table 3). Our | demonstration c | |
| S | atisfactory remova | l of the relevant interfe | erence species (section | on 3.1) gives us a high | degree of confic | |
| with regards to the characterization of natural ²³² Th and ²³⁸ U inherent to the ME solution and subsequ | | | | | | |
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particularly at 10 fg·g⁻¹ and above, show the viability of our mass shift method for challenging ultra-trace
 measurements.
 The LOD using the ME matrix in this study was 3 and 13 fg of Th and U, respectively, which is about

an order of magnitude higher than detection limits that are typically achievable in 2% HNO₃ blank solutions measured in single quad mode with our instrument. While the U signal had a higher sensitivity, the Th signal had better short-term stability (i.e., had smaller deviations between replicate measurements) and therefore had lower LODs. Our method accuracy and detection limit results are specific to a coinage metal background of 10 μ g·g⁻¹. Given the propensity for metals such as Au, Pt, Ir, and W to create carryover effects in the instrument, it is not desirable to measure solutions with coinage metal matrices greater than 10 μ g·g⁻¹. In many electronic components such as ASICs, SiPMs, capacitors, diodes, PCBs, resistors, and SQUID arrays, the coinage metal content is mainly contained within coatings or electrical contacts that often make up $\leq 0.1\%$ of the total weight *e.g.*, (14,16,25). Assuming a 1 g sample has 1 mg of coinage metal content, dilution down to 10 µg coinage metals g sample solution⁻¹ via our method would yield detection limits of 300 and 1300 fg of Th and U, respectively. It is useful to note that the complete digestion of an electronic sample with minor amounts of coinage metals typically involves use of HNO₃, HCl, and HF, which involves multiple dry down steps in order for the final measured solution to have a matrix of 2% HNO₃. It is, therefore, feasible to reduce the dissolved coinage metal content during the dry down steps through precipitation when using an isotopic tracer, which would limit the need for dilution and improve the detection limits (table 4). This could also enable the measurement of samples with a much greater coinage metal content, or even solid metal samples, with similar detection limits. Examples of such analyses are described in the following section.

262 3.3 Determination of U and Th in electronic samples

collaborations (1–3). Samples include board mounts and crimps (Axon' Cable; Montmirail, France), gold bonding wire (Ametek Inc; Berwyn, PA), and superconducting quantum interference device (SQUID) arrays (fabricated by NIST; Boulder, CO). All the samples contain proprietary concentrations of some or all of the coinage metal interferants with U and Th: The Axon' board mount and crimps contain some quantity of Au and other precious metals. The bonding wire is made of gold and the NIST fabricated SQUIDs contained ~1-5 wt% Au as well as trace amounts of W, Pt and Ir.

The samples were spiked with the synthetic ²²⁹Th and ²³³U tracer and digested in a mixture of concentrated acids (Optima grade nitric acid, hydrochloric acid, hydrofluoric acid) either at room temperature, or in a microwave digestion system (Mars 6; CEM corporation, Charlotte, NC)) as needed. Aliquots of the digested samples were diluted/resuspended in 2% HNO₃ as necessary and analysed following the O₂ mass shift method described above. The ²³²Th and ²³⁸U concentrations were determined following isotope dilution methods (eqn. 1). The results of the measurements are reported in table 4.

| - | . spiloate | weight (g) | 232111 (pg·g -) | LOD (pg·g·) | 23°U (pg·g -) | LOD (pg·g· |
|---------------------|---|---|--|---|--|---|
| | | | | | | |
| direct crimp | 1 | 0.3142 0.2945 | 35 ± 7 63 ± 8 | 5 6 | 41 ± 5 56 ± 7 | 1 1 |
| (Axon'Cable) | 2 | | | | | |
| | | | | | | |
| board mount | 1 | 0.1873 | 99 ± 10 | 10 | 120 ± 11 | 2 |
| (Axon'Cable) | | | | | | |
| | | | | | | |
| | 1 | 0.0997 | 18 ± 14 | 17 | 52 ± 6 | 8 |
| | 2 | 0.2295 | 27 ± 7 | 7 | 190 ± 15 | 3 |
| (Ametek Inc) | 3 | 0.2572 | 34 ± 5 | 7 | 54 ± 8 | 3 |
| | | | | | | |
| | 1 | 0.0162 | 12.8 ± 0.7 | 1 | 9.9 ± 1.0 | 2 |
| SQUID arrays (NIST) | 2 | 0.0173 | 14.1 ± 0.7 | 1 | 8.6 ± 0.9 | 1 |
| | 3 | 0.0155 | 19 ± 2 | 1 | 15 ± 2 | 2 |
| | direct crimp (Axon'Cable) board mount (Axon'Cable) gold bonding wire (Ametek Inc) SQUID arrays (NIST) | direct crimp (Axon'Cable)1board mount (Axon'Cable)1gold bonding wire (Ametek Inc)1 2 3SQUID arrays (NIST)2 2 3 | direct crimp (Axon'Cable) 1 0.3142 board mount (Axon'Cable) 1 0.1873 gold bonding wire (Ametek Inc) 1 0.0997 3 0.2295 0.2295 0.2572 SQUID arrays (NIST) 1 0.0162 3 0.0155 | $\begin{array}{c c} \text{direct crimp} & 1 & 0.3142 & 35 \pm 7 \\ (\text{Axon'Cable}) & 2 & 0.2945 & 63 \pm 8 \end{array}$ $\begin{array}{c c} \text{board mount} \\ (\text{Axon'Cable}) & 1 & 0.1873 & 99 \pm 10 \\ (\text{Axon'Cable}) & 1 & 0.0997 & 18 \pm 14 \\ 2 & 0.2295 & 27 \pm 7 \\ (\text{Ametek Inc}) & 3 & 0.2572 & 34 \pm 5 \end{array}$ $\begin{array}{c c} \text{SQUID arrays (NIST)} & 1 & 0.0162 & 12.8 \pm 0.7 \\ 2 & 0.0173 & 14.1 \pm 0.7 \\ 3 & 0.0155 & 19 \pm 2 \end{array}$ | $ \begin{array}{c cccc} direct crimp & 1 & 0.3142 & 35 \pm 7 & 5 \\ (Axon'Cable) & 2 & 0.2945 & 63 \pm 8 & 6 \\ \hline board mount \\ (Axon'Cable) & 1 & 0.1873 & 99 \pm 10 & 10 \\ \hline gold bonding wire \\ (Ametek Inc) & 1 & 0.0997 & 18 \pm 14 & 17 \\ 2 & 0.2295 & 27 \pm 7 & 7 \\ 3 & 0.2572 & 34 \pm 5 & 7 \\ \hline SQUID arrays (NIST) & 1 \\ 2 & 0.0173 & 14.1 \pm 0.7 & 1 \\ 3 & 0.0155 & 19 \pm 2 & 1 \end{array} $ | $ \begin{array}{ccccc} direct \ crimp \\ (Axon'Cable) & 1 \\ 2 & 0.2945 & 63 \pm 8 \\ 6 & 56 \pm 7 \\ \hline \\ board \ mount \\ (Axon'Cable) & 1 \\ 0.1873 & 99 \pm 10 \\ 10 & 120 \pm 11 \\ \hline \\ case & 10 \\ c$ |

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4. Conclusions

The work in this study demonstrates a novel method for the determination of fg·g⁻¹ levels of U and Th in a matrix of Au, Pt, Ir, and W. The method employs an O_2 reaction gas to produce analyte ions of

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interest free from or with minor contributions from polyatomic interferences. Some of the polyatomic interferences produced by the matrix elements were shown to form oxides at intensities that were too low to cause a significant interference with the target masses. The polyatomic interferences at m/z of 234-237 that formed monoxide species at m/z of 250-253 did not form dioxide species at m/z 266-269. The technique was able to measure standards solutions as low as 10 fg·g⁻¹ with >80% accuracy.

288 The method is valuable because it provides a simple and convenient alternative for the 289 measurement of ultra-trace U and Th in coinage metal matrices for which no viable chemical separation 290 exists. This method will allow the high energy and nuclear physics communities to meet the growing demand for lower thresholds of radio contaminants in components with challenging matrices (e.g., 291 292 electronics). This method may also allow us to revisit the electrowinning technique to assay ultra-pure 293 electroformed copper using Ir electrodes, where, prior to this method, Ir contamination at concentrations 294 10^5 times greater than the inherent U made the ultra-trace assay untenable (Matthews et al., 2016). 295 Finally, this method will also be useful to other fields of research, such as the semiconductor industries 296 where ultra-low levels of radio-contaminants are also of increasing importance *e.g.*, (17).

4 ₅ 297 Acknowledgements:

The authors acknowledge standard procurement assistance from May-Lin Thomas at PNNL. The authors 298 299 also acknowledge assay samples provided by the following high energy physics and nuclear physics 300 collaborations: MAJORANA, LEGEND, nEXO, and SuperCDMS. The Axon' samples were supplied by Matthew Busch (MAJORANA/LEGEND), the bonding wire was supplied by Dave Moore (nEXO) and the 301 302 SQUIDs were supplied by Bruce Hines (SuperCDMS). The preparation of the manuscript has benefited 303 from the review of two anonymous reviewers. This work was funded by PNNL Laboratory Directed 304 Research and Development funds under the Nuclear Physics, Particle Physics, Astrophysics, and Cosmology Initiative. The Pacific Northwest National Laboratory is a multi-program national laboratory 305 306 operated for the U.S. Department of Energy (DOE) by Battelle Memorial Institute under contract number 307 DE-AC05-76RL01830.

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