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Advancements and Applications of Microwave-Assisted Deep Eutectic Solvent (MW-DES)
Lignin Extraction: A Comprehensive Review

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- 1 Nomenclature
- 2 MW-DES: microwave assisted-deep eutectic solvent
- 3 LCB: lignocellulosic biomass
- 4 ILs: ionic liquids
- 5 HBD: hydrogen bond donor
- 6 HBA: hydrogen bond acceptor
- 7 Kamlet-Taft (K-T)
- 8 ChCl: choline chloride
- 9 LA: lactic acid
- 10 AA: acetic acid
- 11 GA: glycolic acid
- 12 FA: formic acid
- 13 OA: oxalic acid
- 14 H-unit: paracoumaryl alcohol
- 15 S-unit: sinapyl alcohol
- 16 G-unit: coniferyl alcohol
- 17 LCCs: lignin-carbohydrate complexes
- 18 GBG: guaiacylglycerol- β -guaiacyl ether
- 19 Mn: average molecular weight
- 20 Mw: weight-average molecular weight
- 21 DESL: lignin extracted with DES
- 22 MWL: milled wood lignin

23 Abstract

24 Lignocellulosic-derived biorefineries present an environmentally friendly avenue for generating
25 biofuels and bioproducts, curbing emissions, and advancing environmental stewardship. Yet, the intricate
26 structure of biomass poses obstacles to its efficient conversion into bio-derived products while conserving
27 the native bonding motifs, especially for lignin. The limitations of conventional methodologies stimulated
28 the exploration of innovative solvents and processes. Deep eutectic solvents (DES), a class of solvents
29 comprised of mixtures of hydrogen-donor and hydrogen-acceptor compounds, boasting low toxicity,
30 remarkable solubility, adaptability in synthesis, and selectivity, have garnered increasing attention as
31 sustainable solvents for lignin extraction. The incorporation of microwave-assisted heating can further
32 enhance DES-based lignin extraction efficiency and potential for value creation.

33 This comprehensive review underscores the potential of the MW-DES technique in harnessing
34 lignin—a prized renewable resource—for the sustainable production of energy and materials. From
35 dissecting DES's elemental composition to elucidating DES-microwave-biomass interactions, scrutinizing
36 the MW-DES lignin extraction process factors, and delving into extracted lignin characterization, this
37 review aims to thoroughly evaluate the MW-DES approach's promise in leveraging lignin's potential across
38 diverse industrial sectors.

39

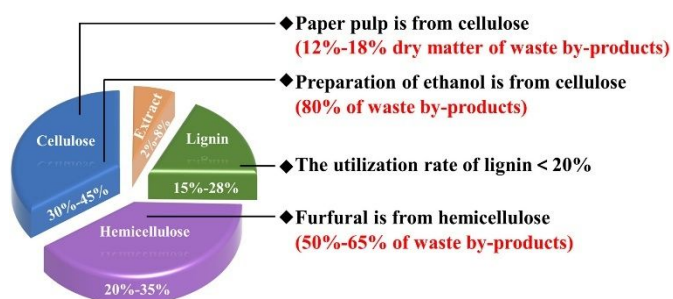
40 Keywords: microwave, deep eutectic solvents, DES, lignin, extraction, pulping

41 1. Introduction

42 Global energy consumption has expanded due to population expansion in the 20th century, which
43 has also encouraged the manufacture of fuels, chemicals, and materials that significantly rely on fossil
44 resources¹. The overconsumption of fossil resources has led to severe climate and environmental issues, as
45 well as concerns regarding energy security and sustainable development due to the vulnerability of oil prices
46 to fluctuations and dependence on imported fuels^{2,3}. To tackle these issues, the energy transition to
47 renewable resources like solar, wind, hydro, and geothermal is more imperative than ever. However, this
48 transition alone won't be enough as the demand for platform chemicals from oil refineries continues to rise
49 along with global economic expansion⁴. Lignocellulosic biomass (LCB) is a renewable organic material
50 sourced from vegetation, including forest residues, agricultural waste, energy crops, and organic fractions
51 of solid wastes. LCB has gained interest due to its sustainability, renewability, and carbon neutrality,
52 making it a potential alternative energy source. LCB-based biorefineries have emerged as a promising
53 approach, in which biomass is utilized to produce biofuels and valuable bioproducts to replace those
54 produced in traditional oil refineries. Biorefineries are also anticipated to decrease greenhouse gas
55 emissions and ease the energy crisis^{5,6}, while offering economic opportunities for agriculture and forestry
56 industries, organic waste management, and production of liquid fuels, platform chemicals, and carbon-
57 neutral energy⁷. Global annual production of LCB is estimated at 181.5 billion tons with potential supplies
58 being contributed by various sectors^{8,9}.

59 LCB is a heterogeneous organic matrix with a three-dimensional structure that is primarily composed
60 of lignin, cellulose, and hemicellulose, as well as a few other minor components, such as minerals and
61 extractives¹⁰. The three major components in LCB are inherently interconnected resulting in distinctive
62 chemical and physical properties, thereby establishing a sturdy structural matrix within the plant cell wall
63¹¹. In contrast to cellulose and hemicellulose, which primarily consist of sugars, lignin distinguishes itself
64 by its distinctive three-dimensional structure and highly heterogeneous characteristics. Lignin is a
65 heterogeneous polymer of various phenylpropanols, such as coniferyl, sinapyl, and p-coumaryl alcohols
66 depending on the plant species, type, or part. The cross-linked network and branched arrangement of the

67 phenylpropanols are the main reasons why LCB is challenging to break down during processes like
 68 conventional pulping or biomass conversion. Moreover, the heterogeneity of lignin in terms of both
 69 variability in bond chemistry and between plant species, not only limits upgrading and advanced
 70 applications for chemicals or materials, but also hinders further development of hemicellulose and cellulose
 71 applications¹². Lignin has always been industrially generated as a byproduct waste in biorefinery processes
 72 and had a relatively low utilization rate among the main components of LCB as Fig. 1 shows. As a natural
 73 aromatic biopolymer, lignin offers a potential as a renewable supplier for phenol, an aromatic precursor of
 74 polymers derived from petroleum¹³. Research development focused on harnessing lignin's distinctive
 75 properties for energy and chemicals is vital for sustainability, maximizing lignin utilization, and potentially
 76 enhancing the profitability and competitiveness of biorefinery.



77

78 **Fig. 1 The usage ratio of three major components in lignocellulosic biomass**¹⁴

79 To effectively use lignin, it is crucial to extract it with qualities that allow for further downstream
 80 conversions and upgrading. Widely used methods include organosolv fractionation, acid hydrolysis,
 81 alkaline pulping, steam explosion, dilute acid pretreatment, as well as enzymatic hydrolysis^{15,16}. These
 82 methods have been applied individually or in combination to achieve specific outcomes. However, each
 83 method has its limitations such as selectivity issues, degradation of lignin, high energy requirements,
 84 solvent cost and safety, lignin recovery efficiency, and so on. Common solvents used in the lignin extraction
 85 process including dilute acids, alkali solvents, and organic solvents, may reduce commercial profitability
 86 and the process's overall applicability because they are toxic, corrosive, volatile, or require high
 87 temperatures and pressures¹⁷. Although the capacity of ionic liquids (ILs) to extract lignin in high yields

88 has lately gained a lot of interest, its high cost, complex synthesis procedure, poor biodegradability, toxicity,
89 and poor recoverability prevent further application in lignin extraction ¹⁸.

90 To avoid the drawbacks of ILs while keeping their desirable properties, researchers have studied DES
91 as a distinctive and green designer solvent. DESs are non-aqueous solvents created by combining two or
92 more elements, commonly a hydrogen bond donor (HBD) and hydrogen bond acceptor (HBA), in a
93 predetermined ratio. These components undergo a eutectic phase transition at a relatively lower temperature
94 than each component's melting point ¹⁹. Because of low toxicity, broad liquid range, versatility, and
95 improved solubility, DESs have been explored in diverse areas like green chemistry, extraction, and
96 catalysis. Following the initial introduction of DESs by Abbott et al. as IL alternatives ²⁰, many studies have
97 investigated and succeeded in lignin extraction in DES ²¹⁻²³. Despite the potential of DES, several
98 challenges and limitations remain in its use for this purpose, particularly concerning extraction efficiency.
99 For example, DES-based lignin extraction often demands longer extraction times than the traditional
100 methods to achieve the same efficiency ^{24,25}. Microwave (MW)-assisted heating is a prominent solution to
101 enhance reaction efficiency and heating speed in the extraction medium. In comparison to conventional
102 heating, microwave-assisted heating provides swift and volumetric heating, energy efficiency, and precise
103 temperature control for DES-driven lignin extraction. This novel approach enables researchers to enhance
104 extraction efficiency, shorten processing time, and attain superior lignin extracts ²⁶.

105 The objective of this review is to provide a comprehensive overview of studies on the application of
106 DES in conjunction with the MW-DES method for lignin extraction. Our exploration is guided by three key
107 dimensions: (1) an examination of ongoing research on lignin extraction through MW-DES; (2) an
108 exploration of potential mechanisms associated with MW-DES; and (3) a systematic discussion on the
109 structural modifications of extracted lignin by MW-DES. Furthermore, this review presents insights into
110 the prospects and challenges associated with environmentally friendly and efficient lignin extractions,
111 highlighting the existing issues in the field.

112 2. The effects of different reactants (biomass and DES) for lignin extraction

113 2.1 Different DES for lignin extraction

114 The composition of DES plays a crucial role in defining the characteristics and effectiveness of these
115 solvents for various applications. DESs can be categorized based on their components and the interactions
116 that give rise to their eutectic properties. Broadly, DESs are classified into two main types: hydrogen bond
117 donor-acceptor DES and metal-containing DES. These primary categories can be further subdivided based
118 on the specific combinations of hydrogen donors, acceptors, and metal components as shown in Fig. 2.

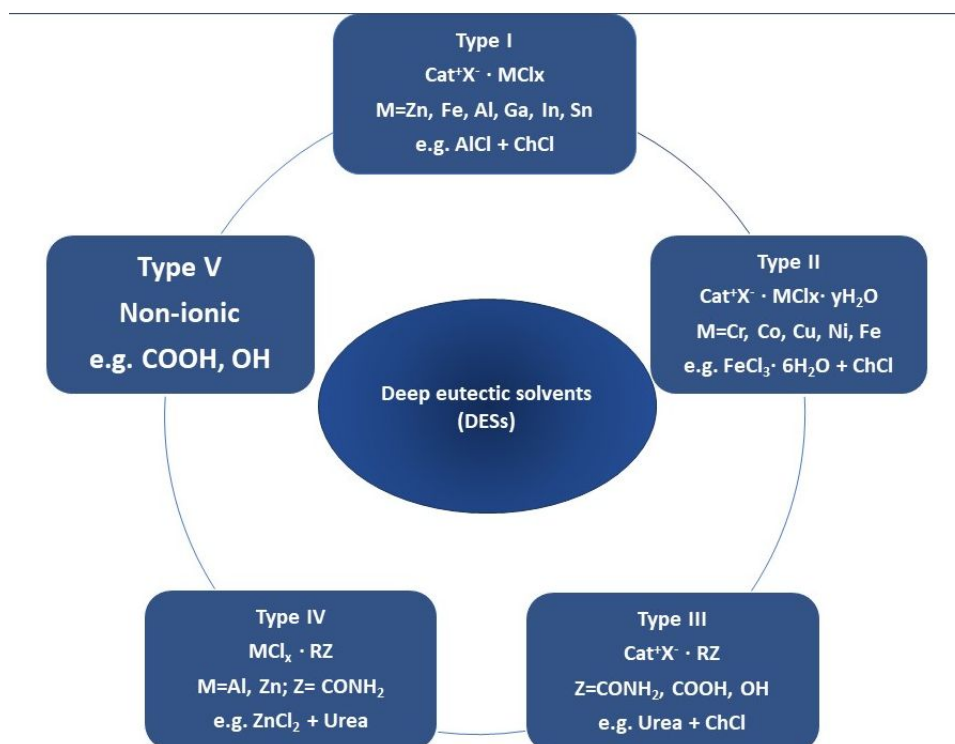
119 (1) Type I DESs: These DESs are produced from the combination of quaternary ammonium salt (e.g.,
120 choline chloride) and non-hydrated metal halide (e.g., AlCl_3). The use of Type I DES in the
121 processing of lignocellulose has not been widely adopted because of the high melting temperatures
122 of non-hydrated metal halides.

123 (2) Type II DESs: In contrast, one hydrated metal halide, such as $\text{AlCl}_3 \cdot 6\text{H}_2\text{O}$, and a quaternary
124 ammonium salt are the main components of Type II DESs, which make them more practical for
125 industrial applications since they are inherently resistant to air and moisture.

126 (3) Type III DESs: The main components of type III DESs are quaternary ammonium salt and other
127 HBDs, like carboxylic acids. Due to their affordable starting components, simple production,
128 interaction with water, low viscosity, and excellent biodegradability, these kinds of DESs are
129 frequently used in the processing of biomass ²⁷.

130 (4) Type IV DESs: Inorganic transition metals and HBDs (such as urea) are used in Type IV DESs
131 even though metal salts are often unable to ionize in non-aqueous conditions.

132 (5) Type V DESs: Frequently hydrophobic, Type V DESs are made entirely of non-ionic molecules.
133 Making the distinction between HBAs and HBDs for Type V DESs can be difficult. However, it
134 has been found that the preparation of Type V DESs relies heavily on an extraordinarily strong
135 interaction resulting from the differential in acidity between phenolic and aliphatic hydroxyl (OH)
136 groups ²⁸.



137

138 **Fig. 2 The formulation for categorizing deep eutectic solvents (DESs) (this figure has been**
 139 **adapted/reproduced from ref ²⁸ with permission from ELSEVIER copyright 2023).**

140 While numerous research endeavors have explored the realm of delignification and lignin extraction,
 141 it's worth noting that not all DESs prove equally effective in cleaving LCCs with lignocellulose and
 142 achieving efficient lignin fractionation. For efficient lignin extraction using DES, a synergistic interaction
 143 between HBDs and HBAs in the DES is essential. Typically, HBDs donate protons, facilitate hydrogen
 144 bonding, and interact with lignin's functional groups. HBAs, on the other hand, weaken the intermolecular
 145 forces holding lignin and other biopolymers in biomass. In Table 1, we summarized various DES and their
 146 effects on lignin extraction.

147 **Table 1 Lignin extraction with various DES**

| Feedstock | Temp (°C) | Time (h) | DES: feedstock | HBD | HBA | HBD: HBA | Yield (%) | Reference |
|-------------------|-----------|----------|----------------|------|------|----------|-----------|-----------|
| Douglas fir | 145 | 6 | 10:1 | LA | ChCl | 1:1 | 78.5 | 29 |
| Sugarcane bagasse | 160 | 14.1 | 17:1 | Urea | ChCl | 2:1 | 82 | 30 |
| Rice straw | 60 | 12 | 20:1 | LA | ChCl | 5:1 | 60 | 31 |
| Eucalyptus | 130 | 6 | 10:1 | LA | ChCl | 10:1 | 64 | 32 |
| Poplar wood | 130 | 6 | 49:1 | LA | ChCl | 9:1 | 61.5 | 33 |

| | | | | | | | | |
|------------------------------------|-----|----|------|---|------|------|-------|----|
| peach pit (Prunus persica) | 145 | 6 | 9:1 | LA | ChCl | 2:1 | 70.2 | 34 |
| walnut shell (Juglans nigra) | 145 | 6 | 9:1 | LA | ChCl | 2:1 | 64.3 | 34 |
| Akebia' herbal residues | 120 | 8 | 10:1 | FA | ChCl | 2:1 | 40.7 | 35 |
| Akebia' herbal residues | 120 | 8 | 10:1 | AA | ChCl | 2:1 | 33.8 | 35 |
| Akebia' herbal residues | 120 | 8 | 10:1 | GA | ChCl | 2:1 | 58.4 | 35 |
| Akebia' herbal residues | 120 | 8 | 10:1 | Levulinic acid | ChCl | 2:1 | 20.2 | 35 |
| Corncob | 150 | 15 | 16:1 | GA | ChCl | 2:1 | 59 | 36 |
| switchgrass | 110 | 1 | 10:1 | GA | ChCl | 2:1 | 18.73 | 37 |
| wheat straw | 90 | 12 | 20:1 | Monoethanolamide | ChCl | 6:1 | 81 | 38 |
| wheat straw | 90 | 12 | 20:1 | Diethanolamine | ChCl | 8:1 | 73.5 | 38 |
| wheat straw | 90 | 12 | 20:1 | Methyl diethanolamine | ChCl | 10:1 | 44.6 | 38 |
| wheat straw | 90 | 12 | 20:1 | Acetamide | ChCl | 2:1 | 3.4 | 38 |
| wheat straw | 90 | 12 | 20:1 | Urea | ChCl | 2:1 | 27.7 | 38 |
| wheat straw | 90 | 12 | 20:1 | Glycerol | ChCl | 2:1 | 24.7 | 38 |
| Corncob | 140 | 2 | n.d. | Benzyl trimethylammonium chloride | LA | 2:1 | 70 | 39 |
| Corncob | 140 | 2 | n.d. | Benzyl triethylammonium chloride | LA | 2:1 | 68 | 39 |

148

149 2.1.1 Different HBDs

150 Common HBDs applied for lignin extraction include carboxyl-containing HBDs, Hydroxyl-containing
 151 HBDs, and Amine/amide-containing HBDs. The functional groups in various HBDs have important effects
 152 on their interactions with lignin, affecting both lignin extraction efficiency and the resulting chemical
 153 structure.

154 According to Table 1, most research works have adopted ChCl-based DES and carboxyl-containing
 155 DES with acidic HBD such as LA, AA, FA, and levulinic acid. Carboxyl-containing HBDs remarkably
 156 perform well in biomass pretreatment and delignification, outperforming HBDs with other functional

157 groups. Their ability to remove over 90% of lignin from various lignocellulose sources ^{40,41} is attributed to
 158 the presence of active protons supplied by carboxylic acids, which catalyze the cleavage of chemical
 159 linkages in lignocellulose. The number of carboxyl groups within an HBD positively correlates with
 160 delignification efficiency, but an excessive number of carboxyl groups may impair interactions between
 161 DESs and lignin. Regarding acidic DES, its functional acidic sites can support proton-catalyzed bond
 162 cleavages, including those of glycosidic bonds, ether bonds, and carbohydrate/lignin interactions. In the
 163 lignin extraction process, these reactions are crucial processes. ⁴². Consequently, the pKa value of the DES
 164 can serve as a gauge for estimating its delignification capability. A lower pKa value indicates higher acidity,
 165 translating to enhanced performance in lignin extraction. Moreover, the Kamlet-Taft (K-T) solvatochromic
 166 parameters (α , β , π^*) can be employed to predict the attributes of the DES. In this context, α signifies H-
 167 bond acidity, β denotes basicity, and π^* reflects the solvent's polarity and polarizability. DES with elevated
 168 α and β values exhibit improved lignin extraction performance, where higher α values denote superior H
 169 proton donation capacity, and higher β values indicate enhanced H-bond acceptance capacity ⁴³. The values
 170 of some DES can be obtained in Table 2. Mankar et al. also found that DES (made with ChCl and OA)
 171 yielded the highest lignin extraction of 37.8%, mainly attributed to its high α value of 1.32 ⁴⁴. Mattonai et
 172 al. also compared four different HBDs including FA, AA, propanoic acid, and LA for their ability to extract
 173 lignin and found FA (followed by LA) provided the best yield (16% in weight, equivalent to 64% of the
 174 total Klason lignin) at 130 °C. Their rationale was rooted in the strength of the hydrogen bond network,
 175 with formic acid's small size and lactic acid's hydroxy group enhancing bond cleavage efficiency and DES
 176 solvation power, respectively ⁴⁵.

177 **Table 2 The Kamlet-Taft (K-T) solvatochromic parameters (α , β , π^*) of some DES ⁴⁴**

| DES | α | β | π |
|----------|----------|---------|-------|
| ChCl: OA | 1.32 | 0.62 | 1.09 |
| ChCl: LA | 1.17 | 0.51 | 1.12 |
| ChCl: AA | 0.92 | 0.54 | 1.10 |

| | | | |
|----------|------|------|------|
| ChCl: BA | 0.81 | 0.53 | 0.96 |
|----------|------|------|------|

178

179 2.1.2 Different HBAs

180 HBAs form a synergy with HBDs in the DES. Hou et al. proved that the introduction of ChCl to HBDs
181 (lactic acid, acetic acid, glycolic acid) helped enhance the lignin dissolution compared to pure acid ³⁵.
182 Common HBAs are different forms of sulfonium, phosphonium, and ammonium salts and ChCl is one of
183 the most often used HBAs for creating DESs because of its nontoxicity, affordability, and biodegradability.
184 According to earlier studies, lignin removal was enhanced by the highly electronegative Cl⁻ in ChCl's
185 tendency to form H-bonds with the hydroxyl groups in lignin ⁴⁶. Additional investigations have
186 demonstrated that the halide counterion present in the HBA promotes β -O-4' bond breaking and inhibits
187 lignin self-condensation ^{17,47}. Hou et al. also conducted research on rice straw with DES composed of
188 different HBAs and HBDs and found that when ChCl was chosen as the HBA, a weaker pretreatment
189 efficiency was observed for most of the DESs, regardless of the kinds of HBDs ¹⁷ compared to the acidic
190 Lac-based DESs. However, regardless of the HBD used, the biomass dissolving efficiency of the DESs was
191 higher when lactic acid was used as the HBA than when ChCl-based DESs were used. This demonstrates
192 that the degree of DES pretreatment and biomass fractionation efficiency are mostly determined by the
193 appropriate choice of HBA and HBD. Therefore, the first step in effectively utilizing DESs is to determine
194 the best possible combinations.

195 Hydroxyl-containing groups are also commonly used for DES synthesis including polyalcohol-based
196 HBDs (e.g., ethylene glycol, propylene glycol, and glycerol) and carbohydrate-based HBDs (e.g., glucose,
197 fructose, and xylitol). When combined with a harmless HBA (such as betaine or ChCl), this kind of DES
198 often shows high biocompatibility with enzymes and microorganisms at nearly neutral pH degrees ⁴⁸. Most
199 studies have demonstrated that, even under harsh pretreatment conditions, polyalcohols, or carbohydrate-
200 based DESs have poor efficiency during lignin dissolving and separation because of the neutrality of the
201 hydroxyl-containing HBDs. Grass, with its weaker covalent link, is a good candidate for a hydroxyl-
202 containing HBD pretreatment.³⁷ To illustrate, the ChCl/glycerol DES stands as an instance where potent

203 lignin fractionation during biomass pretreatment isn't readily apparent. This lack of efficacy is attributed to
204 the DES's limited presence of active protons and acidic sites. In the ChCl–glycerol system, chloride ions
205 (Cl^-) are encapsulated by an arrangement of mutual anionic hydrogen bonds ($[\text{Cl}(\text{glycerol})]^-$) and cationic
206 H-bonds ($[\text{Ch}(\text{glycerol})]^+$). This structure thus reduces the occupied-site anions' capacity to absorb H-bonds,
207 which results in a dearth of protons and inactive acidic sites and, ultimately, subpar performance⁴³. A higher
208 number of hydroxyl groups in HBDs can reduce lignin removal due to the formation of extensive hydrogen
209 bonds between HBA and HBD, weakening the ability of HBA to compete with intra-molecular bonding in
210 biomass components⁴³. The position and type of hydroxyl groups in HBDs can influence their performance
211 in lignin extraction⁴⁹. To improve the DES, some researchers also tried to introduce the third constitute in
212 DES to form a ternary system. Xia et al. integrated $\text{AlCl}_3 \cdot 6\text{H}_2\text{O}$ into the ChCl/glycerol DES using the
213 framework of acidic multisite coordination theory. This novel DES was then employed for lignin extraction.
214 They discovered that supramolecular complexes were created by the synchronized competition of anionic
215 H-bonds and unidentate aluminum ligands, which allowed multisite bridging ligands to cleave both H-
216 bonds and ether bonds in LCCs. Therefore, a remarkable 95.46% increase in lignin fractionation efficiency
217 was achieved. The lignin's purity also impressively reached $94 \pm 0.45\%$. A unique ternary DES system was
218 developed by Wang et al. in a different investigation⁶³. It consists of ChCl and glycol, plus OA or
219 trifluoromethanesulfonic acid (HOTf). The results showed that both solvent solutions successfully
220 integrated EG (80–85%) into the remaining enzyme lignin. The recovered lignin fragments also showed an
221 impressive linkage concentration, consisting of 60-80% aryl ether bonds.

222 HBDs with amine/amide groups, such as urea, imidazole, and ethanolamine, were explored for their
223 lignin fractionation effects, too. More intermolecular H-bonds between HBDs and HBAs can be formed by
224 HBDs with more amines, amides, or hydroxyls. This decreases the amount of free and active groups that
225 can interact with lignocellulose and lowers the efficiency of lignin formation during DES processing.

226 The process for removing lignin from biomass when utilizing chlorine-based DES is based on the
227 development of an H-bond (hydrogen bond) between the lignin and the halogen anions, such as Cl^- , of the

228 DES. With the -OH groups found in lignin; those halogen anions can form hydrogen bonds. This contact
229 encourages the extraction of aromatic chemicals as well as the lignin's breakdown. The ability of the
230 electronegative halogen anion in the DES to establish an H-bond with lignin is therefore intimately related
231 to the effectiveness of lignin extraction ⁵⁰.

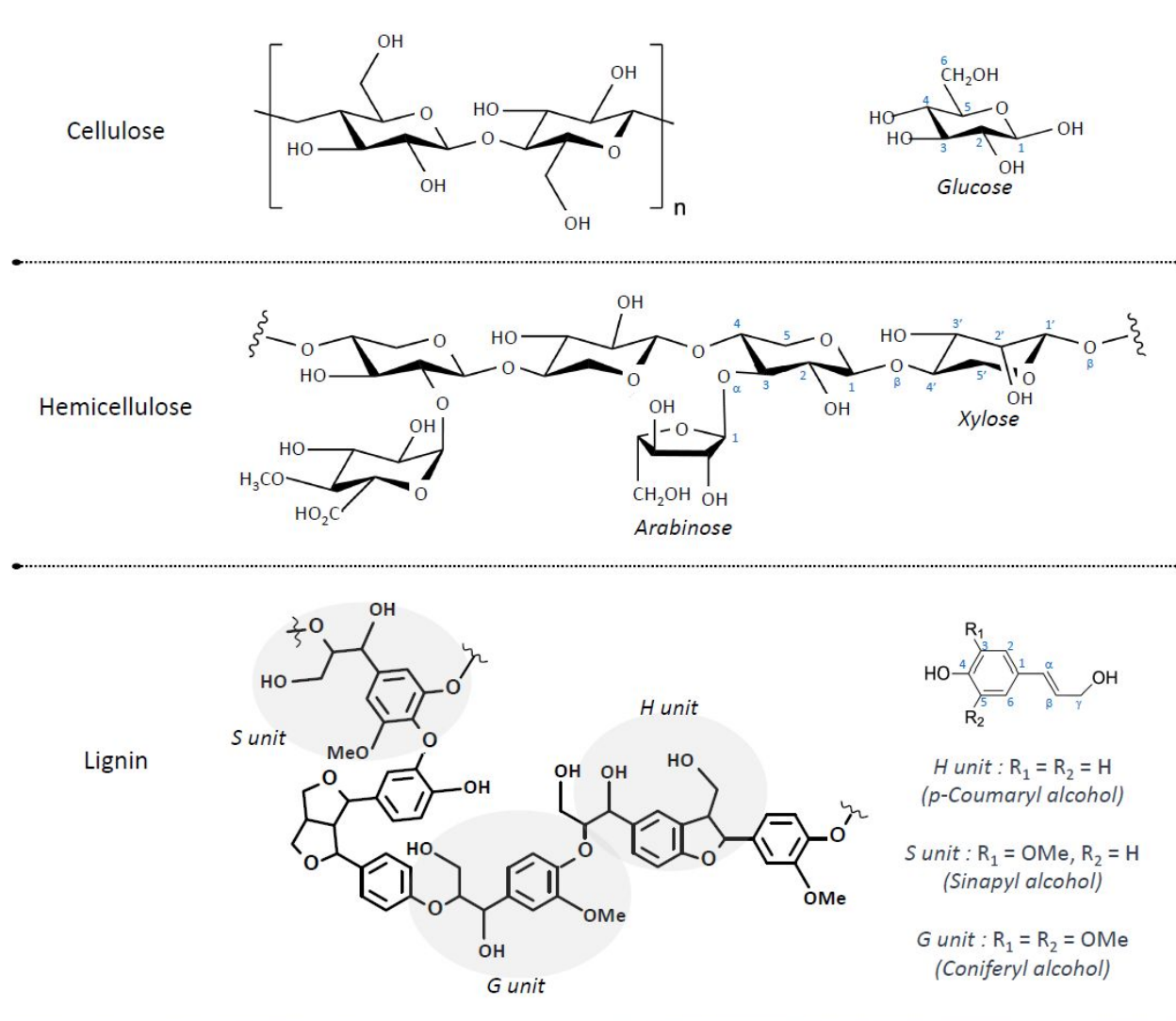
232 2.2 Different biomass for lignin extraction

233 2.2.1 Interactions between main biomass fractions and DES

234 In the lignin-carbohydrate complexes (LCCs) found in LCB, natural lignin is closely bound to
235 cellulose and hemicellulose ⁵¹. To effectively use DES in lignin extraction, it is crucial to comprehend the
236 basic interactions between it and the key components of biomass.

237 Lignin is structured from phenolic monomers (H, G, and S units) interconnected predominantly by
238 aryl ether bonds such as β -O-4 and α -O-4, and carbon-carbon bonds (C-C bonds) such as 5-5 or β - β . The
239 strength of C-C bonds, owing to their elevated bond dissociation energies, usually results in greater
240 resistance to cleavage when contrasted with C-O bonds, as depicted in Fig. 3. Alongside aryl ether and C-
241 C linkages, DES also breaks down lignin via various chemical reactions, encompassing hydroxylation,
242 demethoxylation, and crosslinking ^{52,53}.

243



244

245 **Fig. 3 Chemical compositions of cellulose, lignin, and hemicellulose.**

246 According to previous studies^{40,55}, the DES-based lignin extraction is summarized in three steps as

247 illustrated in Fig. 4. The initial phase involves the solubilization of lignin, wherein the lignin present in the

248 lignocellulosic biomass is dissolved within the DES solution. To engage in composition with the links in

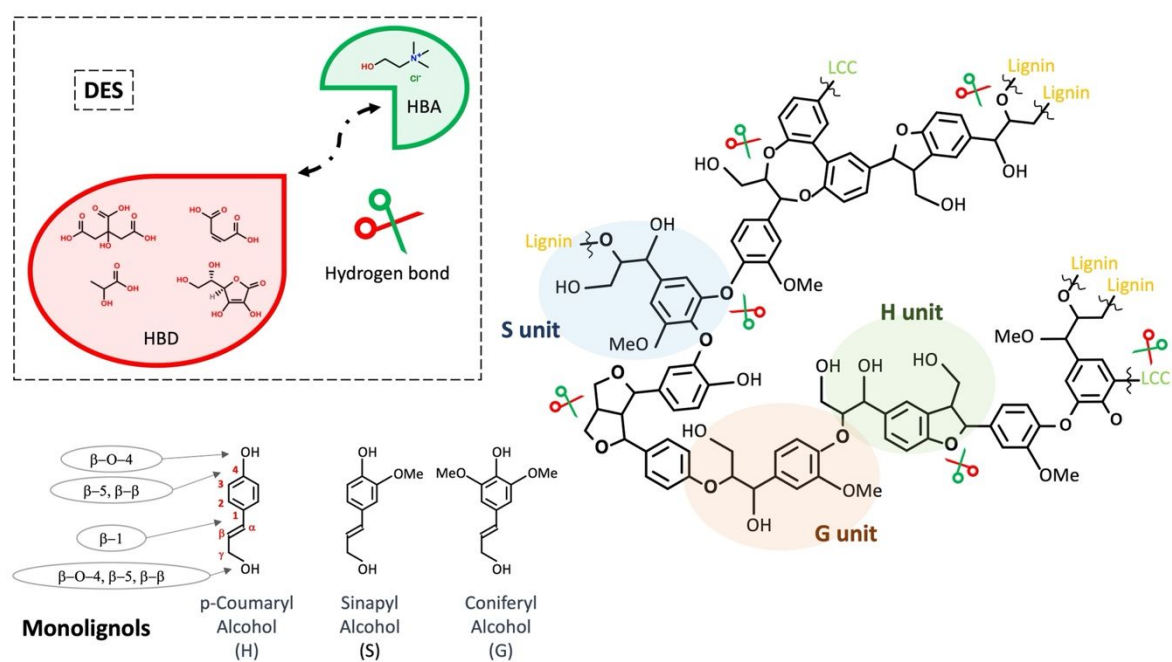
249 the LCC network, the ions in the DES form strong hydrogen bonds with the hydroxyl (OH) groups in the

250 polysaccharides. As a result, the ester and ether connections within the inherent lignin break down, leading

251 to the separation of the lignin macromolecules from the LCC structure. Because a substantial proportion of

252 lignin remains insolubilized at this stage, the lignin yield is low but results in a relatively high molecular

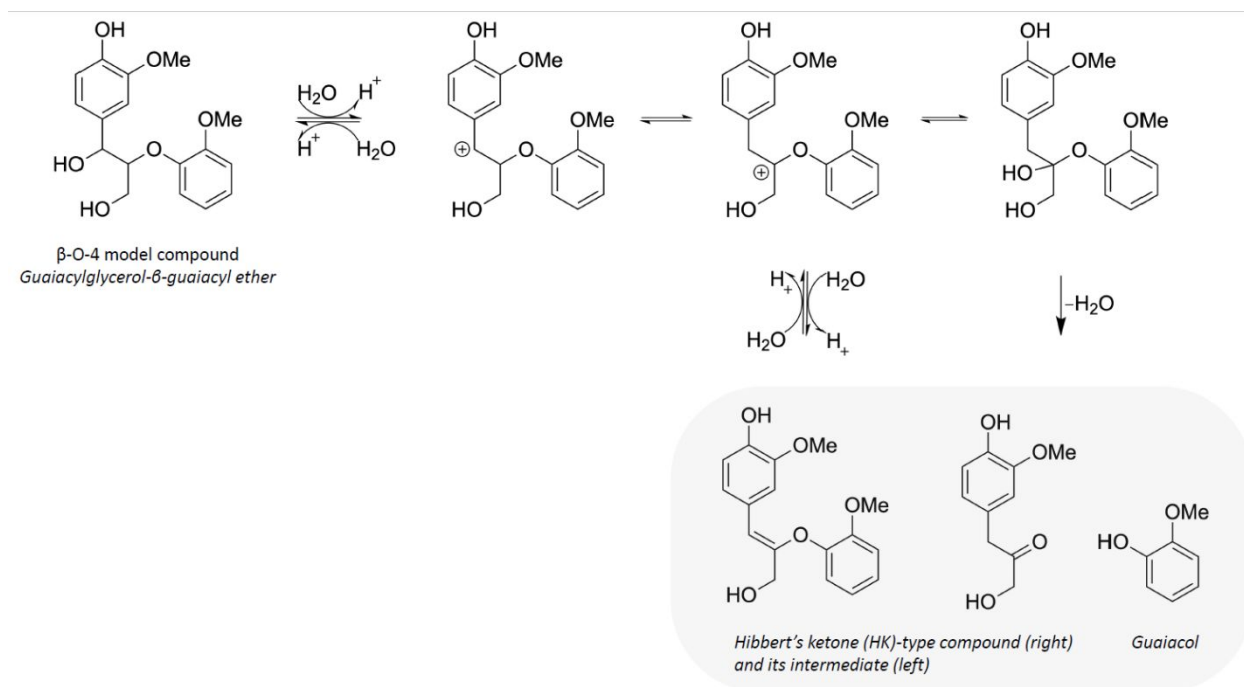
253 weight, comprising a notable quantity of breakable ether connections. The subsequent phase involves the
 254 depolymerization of lignin, wherein further fractionation occurs, leading to the breakdown of lignin
 255 macromolecules into oligomers characterized by reduced molecular weight. Since ether links account for a
 256 sizable fraction (45–65%) of the interunit bonds in natural lignin, they are specifically the β -O-4
 257 connections that break during depolymerization. The exploration of the DES-driven lignin extraction
 258 mechanism involved subjecting a lignin dimer model compound, guaiacylglycerol- β -guaiacyl ether (GBG),
 259 to ChCl-Lac DES at a temperature of 145°C. Complete hydrolysis of GBG resulted in the generation of
 260 guaiacol (G1) and a Hibbert's ketone (HK)-type compound (G2), with the presence of an intermediate, G3,
 261 observed (as shown in Fig. 5). This process resembles lignin acidolysis catalyzed by HCl. Notably, DES
 262 treatment exhibited limited formation of by-products or phenolic re-condensation products compared to
 263 lignin acidolysis. Interestingly, DES treatment of hydroxymatairesinol (HMR), a dimeric model compound
 264 linked by a β - β (C-C bond), revealed no fragmentation products even after a three-hour reaction. This
 265 underscores DES's capacity for selective ether bond cleavage without affecting the C-C linkages in lignin.



266

267 **Fig. 4 The extraction of lignin and the predicted breakage sites for common intramolecular linkages**
 268 **and LCC benzyl ester structures using DES processing.**

269 The elucidation of the mechanism steps during this stage of the acidic cleavage of aryl ether bonds in
 270 lignin model compounds offers insights into tuning processing methods. These steps encompass: (i)
 271 protonation of the α -carbon hydroxyl group; (ii) α -carbon dehydration and β -carbon deprotonation; (iii) the
 272 formation of carbocation and/or enol-ether intermediate; (iv) β -carbon hydroxylation; and (v) cleavage of
 273 β -O-4 ether bond cleavage^{56,57}. Lastly, the concluding phase involves lignin condensation. Lignin
 274 oligomers may interact with unsaturated reactive intermediates post-hydrolysis, leading to the formation of
 275 higher molecular weight complexes, particularly under severe reaction conditions. The repolymerized
 276 lignin oligomers generated in this stage are anticipated to be unreactive due to the formation of stable C-C
 277 bonds⁵⁸.



279 **Fig. 5 The main mechanisms driving the breakdown of β -O-4 linkages in lignin during DES**
 280 **treatment were identified via investigations involving the model compound (this figure has been**
 281 **adapted/reproduced from ref²⁹ with permission from ROYAL SOCIETY OF CHEMISTRY,**
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283 The most common biopolymer in the world, cellulose, is a linear polysaccharide made of D-glucose
 284 subunits joined together by -(1,4)-glycosidic linkages, as seen in Fig. 3. Hydrogen bond connection between

285 the parallel chains increases the durability of the glycosidic link in cellulose, making it less soluble in
286 common solvents.

287 Due to its disorganized structure, DES has a more limited capacity to dissolve cellulose than ILs. This
288 reduced capacity limits the rise in entropy that results from interacting with cellulose, which results in lower
289 solubility toward cellulose^{59,60}. To target lignin extraction while reducing the dissolution of cellulose and
290 hemicellulose, it is best to choose a DES with low attachment to cellulose.

291 The dissolution of cellulose entails a significant disruption of both its intermolecular and
292 intramolecular hydrogen-bond networks. Like ILs, DES works to dissolve cellulose by breaking the original
293 hydrogen connections between molecules and forming new ones between the -OH groups of cellulose and
294 the DES. The intermolecular and intramolecular hydrogen bonds in cellulose are weakened by a solvent's
295 capacity to serve as a hydrogen bond acceptor. A significant link between cellulose solubility and the
296 hydrogen bond basicity (β) of ILs or DESs has been shown in earlier studies⁶¹. It was suggested that β can
297 be used as an indicator to estimate cellulose dissolution⁶². Higher β values are associated with better
298 cellulose dissolution, and the type of anions strongly influences β . Anions like Cl⁻, OAc⁻, HCOO⁻,
299 (MeO)₂PO₂⁻, morpholine, or imidazole, which act as favorable hydrogen bond acceptors, are promising
300 options for forming DESs that facilitate cellulose dissolution⁶³.

301 Hemicellulose is a heterogeneous branched polymer comprising diverse sugar units. It generally
302 exhibits higher solubility in DES compared to cellulose. However, hemicellulose extraction is regarded as
303 less attractive in biomass conversion compared to cellulose or lignin extractions, and the studies on the
304 reaction mechanism between hemicellulose and DES are limited. Xylan is a type of hemicellulose
305 commonly used to study hemicellulose in lignocellulosic biomass. Researchers found that xylan exhibits
306 good solubility in DES as a result of hydrogen bonding interactions between DES molecules and the
307 polysaccharide⁶⁴. This interaction disrupts the intermolecular hydrogen bonding of the xylan solute.
308 Investigations have indicated that DES, with potent electron-withdrawing groups, aids in the efficient

309 removal of xylan. Furthermore, an inversely proportional correlation between the removal of xylan and the
 310 pKa value (acid dissociation constant) of the HBD was observed, with temperature dependence ⁴⁹.

311 2.2.2 Different biomass types

312 Although the ideal biomass for lignin extraction seems to be biomass with high lignin content, high
 313 lignin content is also correlated with the degree of recalcitrance in feedstock since it represents the amount
 314 of lignin and carbohydrate linkage. In the study of Wan et al. ultrafast microwave-assisted ChCl:LA
 315 pretreatment was developed for fractionating switchgrass, corn stover, and *Miscanthus* and corn stover
 316 showed the highest lignin removal (about 80%) because the structure of corn stover is less recalcitrant ⁶⁵.
 317 Kohli et al. investigated six DESs for lignin extraction from miscanthus and birchwood, they discovered
 318 that ChCl-FA and ChCl-OA DESs, respectively, extracted the most lignin from the miscanthus and
 319 birchwood. Furthermore, it takes longer to remove lignin from birchwood than from miscanthus.
 320 Additionally, the structure of lignin differs depending on the feedstock; lignin from birchwood displayed
 321 greater structural alterations ⁶⁶. This can be due to the more complex and denser structure of birchwood
 322 than miscanthus.

323 Besides, different feedstock also requires different solvents for lignin extraction. In the study of Lynam,
 324 the AA: ChCl was the most effective DES in lowering molecular weight and raising PDI of lignin extracted
 325 from rich husks and sugarcane bagasse, while LA: ChCl enhanced these characteristics for coffee chaff,
 326 and the best DES for lignin extraction from corn stover is FA: ChCl⁶⁷. When DES with acetic acid is chosen
 327 to extract lignin, it may be able to dissolve components that can be acetylated in biomass which affects the
 328 extracting result of different biomass ⁶⁸.

329 **Table 3 lignocellulose and lignin composition for softwoods, hardwoods, and grasses (⁶⁹).**

| | Lignocellulose composition (wt.%) | | | Monolignol distribution in lignin (wt.%) | | |
|-----------|-----------------------------------|---------------|--------|--|-------|-------|
| | Cellulose | hemicellulose | Lignin | H | G | S |
| Softwoods | 46-50 | 19-22 | 21-29 | <5 | >95 | 0 |
| Hardwoods | 40-46 | 17-23 | 18-25 | 0-8 | 25-50 | 45-75 |

| | | | | | | |
|---------|-------|-------|-------|------|-------|-------|
| Grasses | 28-37 | 23-29 | 17-20 | 5-35 | 35-80 | 20-55 |
|---------|-------|-------|-------|------|-------|-------|

330

331 3. The influence of process parameters for lignin extraction with DES

332 3.1 Common process for lignin extraction with DES

333 While specific details may vary depending on the DES formulation and the kind of biomass being

334 treated, the general steps are shown in Fig. 6. (1) First, preparing the biomass and DES, usually biomass

335 should be pretreated to remove impurities and decrease the biomass particle size. At the same time, DES is

336 prepared by mixing the appropriate HBD and HBA in a specific ratio. The selection of the DES components

337 can influence the efficiency and selectivity of lignin extraction. (2) The pretreated biomass is then mixed

338 with the DES to ensure proper contact and interaction between the DES and the lignocellulosic material.

339 The mixture is typically heated to an appropriate temperature to enhance lignin solubility and improve

340 extraction efficiency. Microwave-assisted heating may be used for faster and more uniform heating. (3)

341 During the heating and mixing process, the DES penetrates the biomass and solvates the lignin, causing it

342 to dissociate from the lignocellulosic matrix. The DES selectively extracts lignin from the biomass while

343 leaving cellulose and hemicellulose largely intact, as long as the type of DES does not have a strong

344 solubilization effect on them. (4) Once the extraction process is complete, the lignin-DES solution is

345 separated from the remaining biomass components. Various separation methods can be used, such as

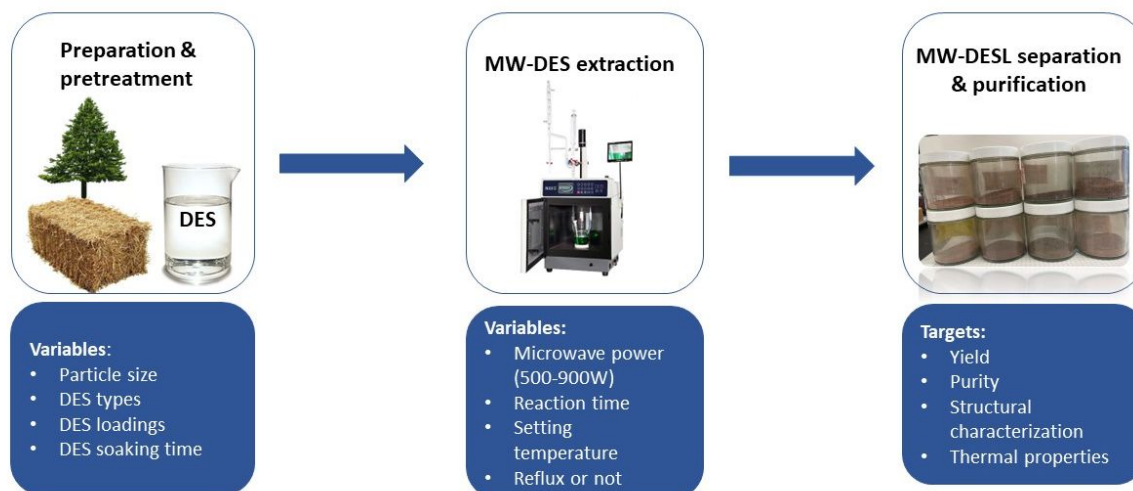
346 filtration, centrifugation, or decantation. (5) Following this separation, an antisolvent is usually added to

347 the lignin-DES mixture that changes the partition coefficients and allows the solubilized lignin to precipitate.

348 The recovered lignin is then characterized to determine its purity, molecular structure, and properties. DES

349 is recovered and potentially reused in subsequent extraction processes, which helps reduce the overall

350 solvent consumption and waste generation.



351

352

Fig. 6 Common process for lignin extraction from LCB with DES.

353

354 3.2 Severity of extraction reactions

355 Reaction time and temperature are the common factors affecting lignin extraction. The severity of the
 356 reaction can be measured by the H factor (Equation 1), which is a measurement that evaluates the severity
 357 of a certain chemical pulping pretreatment technique for lignin removal by taking temperature and duration
 358 into consideration.

$$359 \quad H = \int_0^t \exp\left(43.2 - \frac{16115}{T}\right) dt \quad (1)$$

360 where T is the temperature in Kelvin (K) and t is the time in hours (h). Common extraction of lignin
 361 needs a temperature range from 100°C to 150°C (212°F to 302°F) as Table 1 shows. Although some studies
 362 can perform the extraction at temperatures lower than 100 °C , a long reaction time is required as
 363 compensation ⁷⁰. At the appropriate temperature range, the lignin extraction efficiency increases with the
 364 elevated temperature. For all DESs, higher temperatures weaken the attractive forces by promoting
 365 molecular dynamics and increasing the kinetic energy, therefore, the viscosity of DES can be decreased and
 366 the heat and mass transfers get improved. For instance, it was found that as the temperature climbed from
 367 50 to 100 °C, the shear viscosity of DES Gly:ChCl (1:1) decreased by a factor of ten ⁷¹. However the best
 368 temperature varies from DES types and biomass types.

369 Reaction time is another factor that matters for lignin extraction efficiency. Longer DES
370 soaking/reaction time will decrease the ether bonds and certain monomers. There is a greater reduction in
371 the number-average molecular weight (M_n) and weight-average molecular weight (M_w) of lignin with
372 longer treatment times at higher temperatures⁵¹. Usually, a higher temperature or longer reaction time will
373 lead to a much lower percentage of β -O-4 bonds in the extracted lignin, and mild treatment conditions give
374 a higher content of preserved ether bonds in DES lignin⁷².

375 High pressure is not necessary for lignin extraction with DES, normally, the extraction is conducted
376 under atmospheric pressure. However, due to some closed extraction systems, the vapor pressure of DES
377 itself can form during the extraction process. In general, DESs showed higher vapor pressure compared to
378 common ionic liquids. The vapor pressure-temperature relationship of DESs seems to be more complex
379 than that of molecular liquids as well as that of most ionic liquids⁷³. The change in pressure can influence
380 the density, viscosity, phase behavior, and dissolution capacity, which results in the lignin extraction.
381 However, there are not enough studies on the effect of pressure on lignin extraction with DES. Future
382 studies are needed to understand the extraction system and the DES properties. 4 The introduction of the
383 microwave (MW) heating method in lignin extraction

384 While the effectiveness of DES in lignin extraction has been demonstrated in many studies, one
385 significant challenge is the extended duration of reaction, often spanning hours or even days, resulting in
386 substantial energy consumption and diminished economic gains. The introduction of MW heating is a good
387 method due to its heating mechanism, and due to several compelling advantages of the combination of MW
388 heating and DES:

389 (1) Rapid and Efficient Heating: MW heating offers fast and efficient heating compared to
390 conventional heating methods. The microwaves directly interact with the polar molecules in the DES,
391 causing rapid molecular rotation and agitation, leading to efficient energy transfer to the solvent. This rapid
392 volumetric heating can significantly reduce the extraction time and increase the overall efficiency of the
393 process. The H-factor can be greatly reduced with MW heating in comparison to conventional heating. At

394 150 °C for 20 minutes of pre-treatment, with a slightly lower H factor of 55.5, Mankar et al.'s trials produced
395 the highest lignin extraction yield of 73.9% ^{21,44}. Liu et al. found that the combination of microwave
396 irradiation and DES effectively breaks down LCCs and permits speedy separation of wood lignin. MW
397 heating proved crucial for achieving efficient lignocellulose fractionation with the solvent, as it only
398 required 3 minutes of heating at 80°C under 800 W microwave irradiation. In contrast, achieving similar
399 results without MW heating would take significantly longer, approximately 9 hours, using a conventional
400 bath oil heating method ⁴⁰. Muley et al. [12], also found that FA-based DES can efficiently fractionate lignin
401 from biomass in as little as 1 min at a temperature of 150°C, with 85.8% yield.

402 (2) Selective Heating: Microwaves selectively heat polar molecules, such as water, which is often
403 present in biomass and DES formulations. By targeting the polar parts of the biomass with selective heating
404 given by microwaves, lignin can be extracted and dissolved more easily while being mostly unaffected by
405 other parts of the biomass. The inherently charged nature of the DES mixture also allows it to directly
406 interact with the oscillating electric field component of the microwaves, leading to a non-random increased
407 mobility (as opposed to the random movement in conventional heating).

408 (3) Enhanced Solvent-Solute Interactions: The biomass's DES and lignin interactions are improved by
409 MW heating. The better extraction efficiency is produced by the enhanced agitation and molecular mobility
410 inside the DES, which creates a more suitable environment for lignin solubilization and increases the
411 chances of solvents' active moieties encountering a target bond for breakage.

412 (4) Improved Mass Transfer: The microwave-induced agitation and mixing at the molecular scale
413 within the DES-biomass mixture improve mass transfer rates, enabling better contact between the solvent
414 and lignin. This efficient mass transfer enhances the extraction of lignin from the biomass matrix.

415 (5) Lower Energy Consumption and Environmental Sustainability: Due to its ability to bypass the
416 reactor wall and interact directly with the biomass-DES substrate, MW heating is often more energy-
417 efficient compared to conventional heating methods. It can lead to reduced energy consumption, making

418 the extraction process more environmentally friendly. The thermal lags associated with conventional
419 heating are eliminated, as microwaves simply turn on and off on demand, and, as an electric-heating method,
420 its energy source can be produced via solar, wind, or nuclear sources, leading to a de-carbonized processing
421 method and reducing the associated environmental costs.

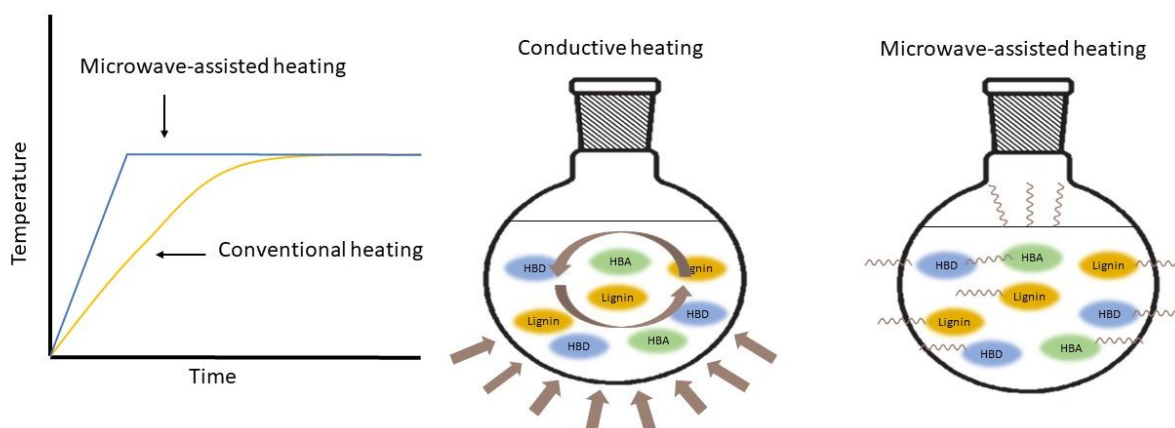
422 (6) Versatility: MW heating with DES is a versatile approach that can be applied to various
423 lignocellulosic biomass sources and different DES formulations. It can be optimized and tailored for
424 specific biomass types, making it applicable to a wide range of lignin extraction processes.

425 4.1 MW-assisted heating mechanism

426 Microwaves are part of the electromagnetic spectrum, ranging mainly from 300 MHz to 300 GHz in
427 frequency ⁷⁴. Microwave-induced heating effect in materials is due primarily to two mechanisms: dipolar
428 polarization and ionic conduction ^{75,76}. When a high-frequency electric field (the electric field components
429 of the microwaves) interacts with charged particles or ions within a material, such as dissolved salts (e.g.,
430 sodium, potassium, and calcium chlorides), the ions experience an alternating force that alternates billions
431 of times per second (depending on the microwave frequency) in a direction parallel to the electric field lines
432 ⁷⁴. The oscillating electric field accelerates these ions, leading to collisions with neighboring particles and
433 the transfer of kinetic energy, resulting in increased agitation and heat within the material. This process
434 converts energy from the microwave field into heat, facilitating microwave-induced heating ⁷⁷. The second
435 interaction that contributes to MW heating involves polar molecules, particularly water, commonly found
436 in liquid materials ⁷⁸. In polar molecules like water, the separation of positive and negative charges creates
437 a dipole, which tends to align itself with the direction of the electric field component of the microwaves.
438 As the field direction changes rapidly, the dipolar molecules either lag behind the field changes, dissipating
439 heat as it always trying to catch up with its resting state, or, if the molecule's relaxation times are within a
440 certain range vs the period of field oscillations, the dipoles start rotating rapidly, generating molecular
441 friction and subsequent heat. The combined effects of ionic interaction (in the presence of charged particles

442 in solvents) and dipolar rotation serve as the fundamental heating mechanisms that make microwave-
 443 assisted extraction rapid and efficient.

444 The uniform and rapid heating facilitated by these mechanisms make this process an exceptional
 445 choice for lignin extraction, particularly when employed alongside DES. Because of the way in which they
 446 transfer heat, MW heating is generally quicker and more effective than traditional heating, which reduces
 447 the amount of energy and reaction time needed. In terms of the lignin extraction process, studies also prove
 448 the high efficiency of microwave-assisted heating. Sun et al. adopted microwave-based heating for
 449 switchgrass pretreatment, followed by hydrolyzing with cellulase enzymes, and found that microwave
 450 pretreatment resulted in a greater sugar yield than normal heating due to the disruption of recalcitrant
 451 structures ⁷⁹. Kumar et al. ⁸⁰ also obtained higher glucose yields from switchgrass biomass pre-treated with
 452 DES in the presence of microwaves for 40 min (5.06 g/L) compared to conventional heating for 3 hours
 453 (4.66 g/L).



454

455 **Fig. 7 Comparison between conventional heating and microwave heating mechanisms.**

456 4.2 Interactions between microwave, DES, and biomass

457 When determining if a material is suitable for MW heating, dielectric parameters including the
 458 dielectric constant (DC), loss factor (LF), and loss tangent ($\tan \delta$) are very important. DC refers to the
 459 material's ability to maintain charge, whereas LF refers to the electrical energy loss in dielectrics. $\tan \delta$, or
 460 the ratio of LF to DC, is a material's ability to convert electromagnetic energy into heat at a particular
 461 frequency and temperature. The efficiency of MW heating greatly depends on the material's dielectric

462 properties, with low dielectric loss materials exhibiting poor MW absorption characteristics and high
463 dielectric loss materials permitting surface heating. Therefore, it is crucial to understand how dielectric
464 properties affect MW processing in order to build an efficient system.⁸¹.

465 As discussed above, microwave-assisted processes using DESs have gained popularity in research due
466 to the unique characteristics of both MW heating and DESs' physicochemical properties which are
467 conducive to bond breaking. Recent investigations have revealed that DESs demonstrate a stronger thermal
468 heating response to microwaves compared to water. Published literature indicates that DESs' dielectric
469 properties are very favorable to microwave absorption and conversion into heat [12], with $\tan \delta$ values at
470 2450 MHz (one of the FCC allotted frequencies for industrial applications) of 0.9 for a 2:1 mixture of
471 formic acid: ChCl, 0.54 for 1:1 mixture of oxalic acid: ChCl, and 0.505 for 1:1 mixture of lactic acid: ChCl.
472 For reference, the $\tan \delta$ for water at the same frequency is approximately 0.085⁷⁸. This response is
473 influenced by various factors, such as the ability to form hydrogen bond networks, the presence of hydroxyl
474 groups, the ionic nature of the HBA component (including ChCl), and the increased polarity of DESs upon
475 exposure to an electromagnetic field. Thermal decomposition represents a crucial constraint for the MW
476 heating response, as it disrupts the hydrogen bond network and leads to the breakdown of the HBD. In
477 addition, in such applications, other substances in the reaction mixture may absorb microwave energy and
478 initiate desired reactions. Thus, DES plays a crucial role not only in the heating process, but also in
479 enhancing the reaction rate and overall efficiency by providing a suitable environment for the reactants and
480 facilitating their interaction.

481 Temperature, frequency⁸², moisture content, density⁸³, and composition affect the dielectric
482 characteristics of the biomass itself^{84,85}. Among the three main components of biomass, hemicellulose
483 generally has the best dielectric properties. Due to the amorphous nature and the presence of polar
484 functional groups, such as hydroxyl groups, hemicellulose is more susceptible to dielectric heating.
485 Compared to hemicellulose, the strong hydrogen bonding and crystalline structure of cellulose limit the
486 interaction with electromagnetic waves, including microwaves. While lignin can be a good microwave

487 absorber, its dielectric properties may vary depending on its chemical composition and degree of
488 polymerization. The presence of aromatic rings and polar functional groups contributes to its microwave-
489 absorbing ability to some extent, but it may not match the dielectric properties of hemicellulose ⁸⁶.
490 Additionally, when biomass is treated, the dielectric characteristics of the material change continually. As
491 the biomass is relatively intact (in terms of cellulose-hemicellulose-lignin network) at the beginning of the
492 process, there is a lack of mobility of these compounds and therefore the biomass components themselves
493 contribute relatively little to the overall heating process. However, as the dissolution process progresses
494 and lignin and hemicellulose are being liberated from the matrix, they can contribute to the dielectric
495 heating effects. If small particles are charred as well in the process (or are included on purpose to act as
496 additional microwave absorbers), these dielectric properties also change. For example, the process of
497 pyrolyzing biomass, either in microwave cavities⁸⁷ or conventional reactors might result in the formation
498 of some char or tar, which can alter the dielectric characteristics of the biomass and function as a catalyst
499 ⁸⁸.

500 The MW-DES extraction system for lignin involves interactions between microwaves and both biomass
501 and DES. Recent studies showed that DES pretreatment with MW assistance performed better and
502 consumed less energy. With ChCl:LA and ChCl:OA and MW-assistant heating, more than 70% lignin had
503 been extracted within minutes ^{40,65}. Microwave irradiation in this process enhances the ion properties and
504 raises the molecular polarity of DESs, which might be one route for improved lignin extraction efficiency.

Table 4 Literature review of lignin extraction with the MW-DES method

| Feedstock | Reaction condition | | | | DES composition | | | Results | | Reference |
|---------------------------|--------------------|------------|-----------|----------------|------------------|------|----------|-----------|------------|-----------|
| | Temp (°C) | Time (min) | Power (W) | DES: feedstock | HBD | HBA | HBD: HBA | Yield (%) | Purity (%) | |
| Fir (<i>Abies alba</i>) | 110 | 30 | 600 | 20 : 1 | FA | ChCl | 5 : 1 | 10 | n.d. | 45 |
| | 120 | 30 | 600 | 20 : 1 | FA | ChCl | 5 : 1 | 11 | n.d. | |
| | 130 | 30 | 600 | 20 : 1 | FA | ChCl | 5 : 1 | 16 | n.d. | |
| | 110 | 30 | 600 | 20 : 1 | AA | ChCl | 5 : 1 | 2 | n.d. | |
| | 120 | 30 | 600 | 20 : 1 | AA | ChCl | 5 : 1 | 3 | n.d. | |
| | 130 | 30 | 600 | 20 : 1 | AA | ChCl | 5 : 1 | 7 | n.d. | |
| | 110 | 30 | 600 | 20 : 1 | propanoic acid | ChCl | 5 : 1 | 1.5 | n.d. | |
| | 120 | 30 | 600 | 20 : 1 | propanoic acid | ChCl | 5 : 1 | 2 | n.d. | |
| | 130 | 30 | 600 | 20 : 1 | propanoic acid | ChCl | 5 : 1 | 8.5 | n.d. | |
| | 110 | 30 | 600 | 20 : 1 | LA | ChCl | 5 : 1 | 3.5 | n.d. | |
| | 120 | 30 | 600 | 20 : 1 | LA | ChCl | 5 : 1 | 15 | n.d. | |
| | 130 | 30 | 600 | 20 : 1 | LA | ChCl | 5 : 1 | 12.5 | n.d. | |
| | Southern pine | 150 | 15 | n.d. | 6: 1 | FA | ChCl | 2: 1 | 89.6 | n.d. |
| 130 | | 15 | n.d. | 6: 1 | oxalic dihydrate | acid | ChCl | 1: 1 | 83.8 | n.d. |
| Rice straw | 155 | 30 | n.d. | 10: 1 | FA | ChCl | 2: 1 | 36.1 | n.d. | 80 |
| Poplar wood flour | 81 | 3 | 800 | 20 : 1 | oxalic dihydrate | acid | ChCl | 1 : 1 | 15.4 | 80 |

| | | | | | | | | | | | |
|--------------|-----|------|-----|--------|------------------|------|------|-------|-------|-------|----|
| | 80 | 5 | 800 | 20 : 1 | oxalic dihydrate | acid | ChCl | 1 : 1 | 15.8 | 93 | 40 |
| | 80 | 10 | 800 | 20 : 1 | oxalic dihydrate | acid | ChCl | 1 : 1 | 15.1 | 96 | |
| | 80 | 9h | n.d | 20 : 1 | oxalic dihydrate | acid | ChCl | 1 : 1 | ~5 | 85 | |
| | 110 | 9h | n.d | 20 : 1 | oxalic dihydrate | acid | ChCl | 1 : 1 | 17.5 | 90 | |
| | 125 | 20 | n.d | 30 : 1 | Butyric acid | | ChCl | 2 : 1 | 7.5 | n.d. | 44 |
| | 125 | 20 | n.d | 30 : 1 | AA | | ChCl | 2 : 1 | 16 | n.d. | |
| | 125 | 20 | n.d | 30 : 1 | LA | | ChCl | 2 : 1 | 36 | n.d. | |
| | 125 | 20 | n.d | 30 : 1 | oxalic dihydrate | acid | ChCl | 2 : 1 | 37 | n.d. | |
| | 90 | 20 | n.d | 30 : 1 | LA | | ChCl | 2 : 1 | 10.9 | n.d. | |
| | 110 | 20 | n.d | 30 : 1 | LA | | ChCl | 2 : 1 | 28.5 | n.d. | |
| | 130 | 20 | n.d | 30 : 1 | LA | | ChCl | 2 : 1 | 35.1 | n.d. | |
| | 150 | 20 | n.d | 30 : 1 | LA | | ChCl | 2 : 1 | 73.9 | n.d. | |
| coconut coir | 170 | 20 | n.d | 30 : 1 | LA | | ChCl | 2 : 1 | 71.2 | n.d. | |
| | 150 | 5 | n.d | 30 : 1 | LA | | ChCl | 2 : 1 | 35.1 | n.d. | |
| | 150 | 10 | n.d | 30 : 1 | LA | | ChCl | 2 : 1 | 42.4 | n.d. | |
| | 150 | 30 | n.d | 30 : 1 | LA | | ChCl | 2 : 1 | 56 | n.d. | |
| | 150 | 40 | n.d | 30 : 1 | LA | | ChCl | 2 : 1 | 53 | n.d. | |
| | 150 | 20 | n.d | 30 : 1 | LA | | ChCl | 1:00 | 16.9 | n.d. | |
| | 150 | 20 | n.d | 30 : 1 | LA | | ChCl | 4 : 1 | 82 | n.d. | |
| | 150 | 20 | n.d | 30 : 1 | LA | | ChCl | 6 : 1 | 50 | n.d. | |
| | 150 | 20 | n.d | 30 : 1 | LA | | ChCl | 8 : 1 | 40 | n.d. | |
| switchgrass | 152 | 0.75 | n.d | 10 : 1 | LA | | ChCl | 2 : 1 | 48.39 | 87.42 | 65 |

| | | | | | | | | | | | |
|-------------|------|-------|-----|------|-----|--------|----------|------------|-------|-------|----|
| corn stover | | | 152 | 0.75 | n.d | 10 : 1 | LA | ChCl 2 : 1 | 70.84 | 84.96 | |
| miscanthus | | | 152 | 0.75 | n.d | 10 : 1 | LA | ChCl 2 : 1 | 42.67 | 86.01 | |
| | | | 90 | 30 | 200 | 20 : 1 | PTSA-Gly | ChCl 2:1:1 | 6.4 | n.d. | |
| | | | 110 | 30 | 200 | 20 : 1 | PTSA-Gly | ChCl 2:1:1 | 29.2 | n.d. | |
| Cocoa | Bean | Shell | 130 | 30 | 200 | 20 : 1 | PTSA-Gly | ChCl 2:1:1 | 95.5 | n.d. | 89 |
| (CBS) | | | 90 | 30 | 200 | 10 : 1 | PTSA-Gly | ChCl 2:1:1 | 3.5 | n.d. | |
| | | | 110 | 30 | 200 | 10 : 1 | PTSA-Gly | ChCl 2:1:1 | 16.3 | n.d. | |
| | | | 130 | 30 | 200 | 10 : 1 | PTSA-Gly | ChCl 2:1:1 | 57.5 | n.d. | |

n.d.: not defined in the literature, LA: lactic acid, AA: acetic acid, FA: formic acid

5 Exploration pathways for chemical transformation of lignin

Except for determining the yield and purity of lignin, the characterization of lignin is crucial for unraveling the mechanism and optimizing the process of lignin extraction with DES. It provides detailed insights into the structural and chemical changes such as the molecular weight, polydispersity, degree of polymerization, and functional groups during the extraction process.

Common methods for lignin characterization include elemental analysis, Fourier Transform Infrared Spectroscopy (FTIR), and Nuclear Magnetic Resonance (NMR) Spectroscopy for determining functional groups and lignin's molecular structure, Gel Permeation Chromatography (GPC) for identifying the molecular weight distribution, Ultraviolet-Visible (UV-Vis), Spectroscopy, Pyrolysis-Gas Chromatography-Mass Spectrometry (Py-GC-MS), Thermogravimetric Analysis (TGA) for the thermal stability, Scanning Electron Microscopy (SEM), Transmission Electron Microscopy (TEM), and X-ray Diffraction (XRD).

5.1 Lignin modification with DES extraction

Pan et al indicated that lignin with more phenolic hydroxyl groups, less aliphatic hydroxyl groups, low molecular weight, and narrow polydispersity showed high antioxidant activity⁹⁰. Lignin extracted with DES often displays a narrower molecular weight distribution, improved purity, better structural integrity, and more functional groups. Those samples that underwent this procedure showed improved thermal stability and exceptional antioxidant activity⁹¹. Zhang et al. compared the Douglas Fir lignin (DESL) extracted with DES, milled wood lignin (MWL), and other typical software lignin and they found that DESL had a narrower and lower molecular weight range, ranging from 490-2600 g/mol with a peak molecular weight of roughly 890 g/mol²⁹. Wen et al. conducted a series of characterizations of lignin extracted with DES (lactic acid- ChCl) under 60-140°C. They discovered that during DES treatment, the primary change involved the selective dissociation of the aryl ether linkage (β -O-4) without affecting the C-C bonds. This conclusion is consistent with our previous work that when subjected to DES treatment (ChCl/lactic acid) at 145°C, the β -O-4 dimeric model (guaiacylglycerol- β -guaiacyl ether) was completely converted, leading to the stoichiometric release of Hibbert's ketone and guaiacol, with no cleavage observed

for the β - β -linked dimeric model (hydroxymatairesinol, HMR) ²⁹. The cleavage of ether linkage resulted in the increased phenolic hydroxyl groups in lignin, while aliphatic hydroxyl groups decreased, possibly due to DES-induced dehydration or acylation reactions. Besides, in the aromatic region (Fig. 8), DES primarily catalyzed the degradation of G-type lignin units at lower temperatures (60-100°C) and, at higher temperatures (120-140°C), S-type lignin was more readily removed than G-type lignin ⁹². Besides, the oxidation and demethoxylation reactions also occurred during the DES treatment process, leading to the condensations of G and S-types lignin. This will cause an increase in the S/G ratio. ^{93,94}The oxidation and demethoxylation reactions that occurred during the DES treatment process, led to the condensations of G and S subunit lignin, which also contributed to an increase in the S/G ratio (Ma et al., 2021, Ma et al., 2020).

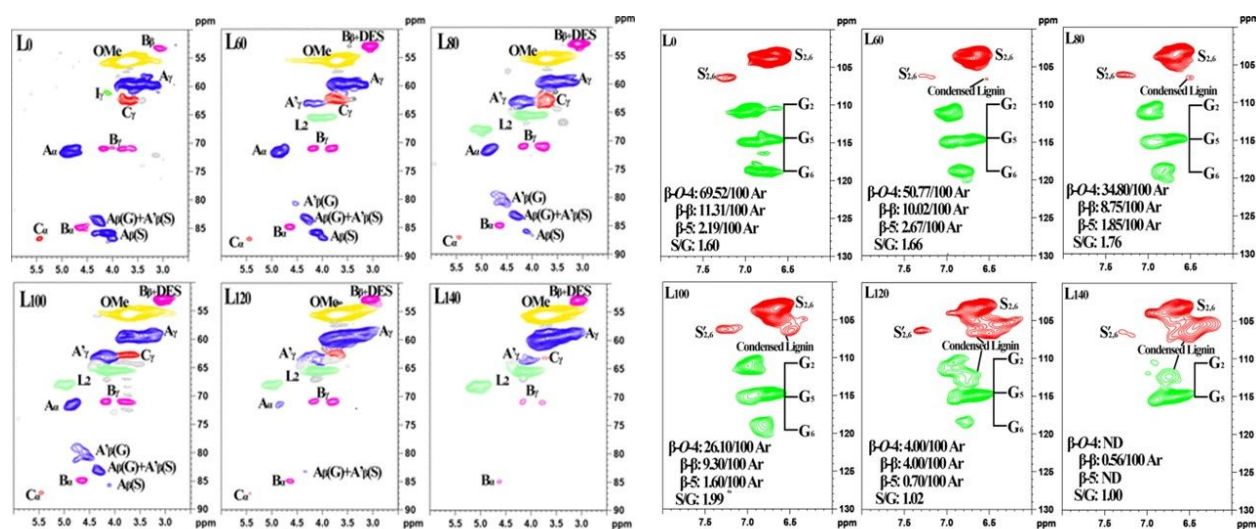


Fig. 8 2D-HSQC NMR spectra of the lignin side-chain region (left) and aromatic region (right) ⁹².

Apart from the depolymerization of lignin, the recondensation of lignin also occurred and resulted in more complex lignin compounds. But Pant's research showed that DES-extracted lignin compared with Kraft lignin had less condensed C-C linkages, which is favorable for further valorization to aromatics ⁴⁴.

5.2 Differences of lignin with MW-heating and other heating methods

When utilizing DES under strict reaction circumstances, conventional heating that relies on conduction/convection-based methods causes heat dissipation because of temperature gradients [21]. In contrast, MW heating employs an electromagnetic field that induces rapid oscillation and rotation of polar

molecules and charged moieties within the DES-lignocellulosic biomass mixture, accelerating heat generation [29]. This intensified heating facilitates lignin dissolution by breaking weak intermolecular forces, consequently enhancing lignin solubility in the solvent [12].

Research suggests that MW heating may also generate localized heat spots within the complex biomass structure, potentially leading to burst-like effects within particles. As a result, the biomass structure is disrupted, leading to a higher yield in lignin extraction. Molecular dynamic simulations also indicate that microwaves could stretch certain bonds within real lignin more compared to conventional heating, increasing the likelihood of bond breakage, and thus accelerating polymer breakdown [12]. Microwave-enhanced heating thus may lead to stressor effects in the microstructure, which breaks weak intermolecular forces and enhances lignin solubility in the solvent and can trigger specific reactions like additional dielectric heating and ionic conduction from the liberated small molecules. These reactions can selectively cleave crucial lignin bonds, such as ether linkages (β -O-4 bonds)⁹⁵, pivotal for lignin depolymerization and extraction. The cleavage of these linkages can elevate the phenolic hydroxyl content in the extracted lignin, potentially synergize with DES, and contribute to ether linkage cleavage during extraction. Multiphysics simulations of microwave heating of biomass [68] and lignin⁸⁷, the same one as above Nde et al] also indicate that the sample placement in the reactor also heavily influences the heating performance.

Furthermore, the combination of microwave and DES has been shown to efficiently cleave LCCs, yielding lignin with a low molecular weight (913), low polydispersity (1.25), and high purity (96%)⁴⁰. The bound carbohydrate content and lignin molecular weight decreased when the severity of microwave-assisted extraction was increased. Increased extraction temperature and duration led to a greater release of OCH₃ groups in lignin as a result of more severe demethylation reactions⁹⁶. Li et al. found that a lignin yield of 17.98% was reached by microwave-assisted extraction at 109°C for 60 minutes. The resultant lignin fraction has a low weight-average molecular weight of 6070 g/mol and a low bound sugar concentration of 1.81%. It also showed significant antioxidant activity (RSI 1.15), exceeding that of the common commercial antioxidant BHT but falling short of BHA⁹⁵. Additionally, Zhou et al. demonstrated that the lignin

recovered via MW-heating is a combination of the GS-type with the predominant inter-unit connection being the -O-4' ether bond, but it is expected that the contributions of individual lignin monomeric subunits would be heavily dependent on the source of biomass as well. The radical scavenging index (RSI) of MW-assisted lignin was greater than that of milled wood lignin (0.53) in terms of antioxidant activity against DPPH, indicating that ML displayed significantly stronger antioxidant activity than milled wood lignin.⁹⁷

However, the characteristics of extracted lignin can also vary depending on the composition of DES and the extracting conditions. There is a significant knowledge gap with respect to a wide range of DES types, microwave conditions, and extracted lignin properties, such as functional groups, molecular weight, and purity, and more investigations are needed to properly design unit operations targeting lignin separation via microwave methods.

6 Prospects and challenges

In summary, DES application in lignin extraction boasts appealing attributes: eco-friendliness, efficient extraction, selective cleavage, and synergy with MW heating for accelerated reaction rates and improved energy efficiency. However, integrating MW-DES into lignin extraction faces challenges such as the MW-assisted DES realm lacks comprehensive insights into mechanisms, MW-DES interplay, recovery strategies, and scalability summarized in Table 5. Future challenges include: (1) Complexity: Despite the vast range of uses and variety of DES types, insufficient study has been done on its characteristics, particularly its thermodynamic properties⁹⁸. On this basis, the extraction of lignin involves multiple components, and multiple reactions, and is a complex system. Many experiments with different DESs, feedstocks, and conditions need to be conducted.

(2) Affordability: Although MW-assisted heating decreased the overall cost of the lignin extraction process, DES itself is not an economic-effective solvent. The effective recycling method for DES and the use of less expensive DES components to synthesize DES can be the potential solutions for reducing the cost. Besides, for a collaborative biorefinery, people also work on developing biomass-derived DES (bio-DES/green DES), which can add value to the whole industry. Further detailed investigation on these

research areas will provide the versatile uses of bio-DES for significantly lower cost and eco-friendly processes.

(3) **Scaling Challenges:** Lab-scale MW-DES success doesn't guarantee industrial scalability for lignin extraction. In larger-scale reactors, achieving uniform microwave heating throughout the entire volume can be difficult. Hotspots and temperature variations may occur, making it challenging to ensure consistent and controlled reactions. Besides, the potential for microwave leakage, especially in high-power systems, needs to be carefully managed to ensure the safety of personnel and the environment.

(4) **Consistency:** There are many factors that can affect the consistency of final lignin products. MW-DES process can be more sensitive to the biomass sample size and geometry exposed to microwave energy. Although addressing these challenges mandates concerted research initiatives, it is pivotal in fully harnessing lignin's potential as a valuable and renewable resource spanning diverse industrial sectors.

Table 5 Comparison between lignin extracted with MW heating and other heating methods.

| | MW heating | Conventional heating |
|--|--|---|
| Extraction yield | Higher extraction yield (80-90%) | Lower extraction yield (50-70%) |
| Reaction condition | Lower temperature and shorter time (130-150°C , 15-60 min) | Higher temperature and longer time (>140°C, hours) |
| Efficiency | Efficient and rapid, volumetric heating | Slower, temperature gradient-driven heating |
| Lignin properties | <ul style="list-style-type: none"> • Lower moisture content. • Lower Mw, Mn, and PDI (PDI (polydispersity (Mw/Mn) index) • Less char residue. • Minimal to no ether linkages. • Higher antioxidant activity. Formation of cross-peaks resembling Hibbert's ketone (HK). | <ul style="list-style-type: none"> • Varying molecular weights. Varying polydispersity, and chemical properties |
| Lignin properties Solvent consumption | <ul style="list-style-type: none"> • Lower moisture content. • Lower Mw, Mn, and PDI (PDI (polydispersity (Mw/Mn) index) • Less char residue. • Minimal to no ether linkages. | <ul style="list-style-type: none"> • Varying molecular weights. • Varying polydispersity, and chemical properties • Larger volumes of solvent to ensure thorough penetration and |

| | | |
|--------------------|---|---|
| | <ul style="list-style-type: none"> • Higher antioxidant activity. • Formation of cross-peaks resembling Hibbert's ketone (HK). • Less solvent consumption | extraction from the lignocellulosic biomass. |
| Solvent recycles | The purity required for the recycled DES is important. MW heating might affect the purity differently from conventional methods. | <ul style="list-style-type: none"> • Thermal Degradation: conventional heating methods involve exposing the DES to high temperatures for extended periods. • Side Reactions: the elevated temperatures during conventional heating can trigger undesired side reactions between the DES and other components in the biomass matrix. • Solvent Loss: the use of high temperatures and prolonged heating times in conventional methods can lead to solvent loss through evaporation. <p>Purity Issues: Exposure to high temperatures can cause the DES to pick up impurities or contaminants from the biomass.</p> |
| Scale up potential | The infrastructure required for MW heating might differ from that of conventional heating methods. This could impact the feasibility of implementing microwave-assisted recycling on an industrial scale. | <ul style="list-style-type: none"> • Conventional heating methods that work well at small scales might face challenges when scaled up to industrial levels. Issues related to heat transfer, equipment design, and process control can arise. |

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