PAPER

Check for updates

Preparation of a thermo-responsive drug carrier consisting of a biocompatible triblock copolymer and fullerene†

Kohei Kitano, [ID] [a] Kazuhiko Ishihara [ID] [b] and Shin-ichi Yusa [ID] *[a]

A triblock copolymer (PEG-*b*-PUEM-*b*-PMPC; EUM) comprising poly(ethylene glycol) (PEG), thermo-responsive poly(2-ureidoethyl methacrylate) (PUEM), and poly(2-(methacryloyloxy)ethyl phosphorylcholine) (PMPC) blocks was synthesized *via* controlled radical polymerization. PEG and PMPC blocks exhibit hydrophilicity and biocompatibility. The PUEM block exhibits an upper critical solution temperature (UCST). PMPC can dissolve hydrophobic fullerenes in water to form a complex by grinding PMPC and fullerene powders. Fullerene-C_{70} (C_{70}) and EUM were ground in a mortar and phosphate-buffered saline (PBS) was added to synthesize a water-soluble complex (C_{70}/EUM). C_{70}/EUM has a core–shell-corona structure, whose core is a complex of C_{70} and PMPC, the shell is PUEM, and corona is PEG. The maximum C_{70} concentration dissolved in PBS was 0.313 g L^{-1} at an EUM concentration of 2 g L^{-1}. The C_{70}/EUM hydrodynamic radius (R_h) was 34 nm in PBS at 10 °C, which increased due to the PUEM block's UCST phase transition with increasing temperature, and R_h attained a constant value of 38 nm above 36 °C. An anticancer drug, doxorubicin, was encapsulated in the PUEM shell by hydrophobic interactions in C_{70}/EUM at room temperature, which can be released by heating. The generation of singlet oxygen ($^1O_2$) from C_{70}/EUM upon visible-light irradiation was confirmed using the singlet oxygen sensor green indicator. Water-soluble C_{70}/EUM may be used as a carrier that releases encapsulated drugs when heated and as a photosensitizer for photodynamic therapy.

Introduction

Fullerenes such as C_{60} and C_{70} and their derivatives have been reported to function as enzyme inhibitors,[1–3] antivirals,[4–6] DNA scission agents,[7–9] and radical quenchers.[10] Furthermore, fullerenes such as C_{60} and C_{70} are expected to be used as photosensitizers for photodynamic therapy (PDT) since they generate singlet oxygen with high efficiency upon visible-light irradiation.[11–13] However, their biological applications are limited by their low water solubility.[14] Various fullerene derivatives with improved water solubility have been reported.[15–17] Fullerenes have been solubilized, forming a complex between fullerenes and host molecules such as cyclodextrins,[18,19] calixarenes,[20] micelles,[21,22] liposomes,[23,24] and poly(*N*-vinylpyrrolidone) (PVP).[25–27] Particularly, PVP can solubilize fullerenes with high concentrations to form a water-soluble complex due to charge-transfer interactions. Yamakoshi *et al.*[28] reported that

C_{60} and C_{70} can be solubilized in water at 0.4 and 0.2 g L^{-1}, respectively, using 50 g L^{-1} PVP.

Poly(2-(methacryloyloxy)ethyl phosphorylcholine) (PMPC) is water-soluble and biocompatible because the pendant phosphorylcholine group has the same chemical structure as the polar group in phospholipids from biological membranes.[29–31] The phosphorylcholine group is an electrically neutral zwitterionic structure. PMPC shows no biological reactions with biomolecules because it does not interact with plasma proteins.[32–34] We have reported that a water-soluble complex (C_{70}/PMPC) can be synthesized by mixing C_{70} and PMPC powders and grinding them.[35] 0.49 g L^{-1} C_{70} can be solubilized in water using 1 g L^{-1} PMPC.

The photosensitizer should be accumulated around the affected region for effective PDT with low side effects. There are tens to hundreds of nanometer-sized pores that are not observed in normal tissue at the vascular endothelium of cancer tissue. Particles of several nanometers in size are immediately excreted from the kidneys, and those larger than 400 nm are rapidly eliminated by macrophages. However, tens to hundreds of nanometers of particles accumulate in cancer tissue due to enhanced permeation and retention (EPR) effects.[36–38] The photosensitizer accumulated around the cancer tissue damages only cancer through the singlet oxygen

[a] Department of Applied Chemistry, Graduate School of Engineering, University of Hyogo, 2167 Shosha, Himeji, Hyogo 671-2280, Japan.
E-mail: yusa@eng.u-hyogo.ac.jp
[b] Department of Materials Engineering and Department of Bioengineering, School of Engineering, The University of Tokyo, 7-3-1, Hongo, Bunkyo-ku, Tokyo 113-8656, Japan
† Electronic supplementary information (ESI) available. See DOI: 10.1039/d1tb02183d

generated upon light irradiation. The light should be irradiated to the photosensitizer inside the body for PDT. PDT uses ordinary visible or near-infrared light; however, it is difficult for it to penetrate deep inside the body.[39] Thus, it should be combined with other PDT treatments for treating deep cancers inside the body. The photosensitizer C_{70} in a water-soluble C_{70}/PMPC complex can generate singlet oxygen upon visible-light irradiation. Therefore, the controlled release function of anti-cancer drugs upon heating was added to the C_{70}/PMPC complex. We focused on poly(2-ureidoethyl methacrylate) (PUEM), which exhibits an upper critical solution temperature (UCST).[40] The PUEM's pendant ureido groups combine with interpolymer chains below the UCST due to hydrogen bonds, and PUEM cannot dissolve in water. Alternatively, PUEM can dissolve in water due to an increase in molecular motion to break the hydrogen bonds above the UCST. UCST type polymers are applied for controlled release for drug delivery systems.[41] Deng et al.[42] synthesized a diblock copolymer (PEG-b-P(NAGA-co-AN)) composed of a hydrophilic poly(ethylene glycol) (PEG) and a thermo-responsive random copolymer (P(NAGA-co-AN)) of N-acryloyl glycolinamide and acrylonitrile. P(NAGA-co-AN) exhibits UCST due to hydrogen bonding interactions. PEG-b-P(NAGA-co-AN) forms core–shell micelles to associate with the P(NAGA-co-AN) blocks below the UCST. Hydrophobic drugs can be encapsulated in the core and released by heating.

In this study, we have synthesized a triblock copolymer (EUM; PEG_{45}-b-$PUEM_{101}$-b-$PMPC_{99}$) consisting of PEG for colloid stabilization in water, PUEM exhibiting UCST, and PMPC solubilizing C_{70} in water to synthesize a thermoresponsive drug carrier containing C_{70} photosensitizer. EUM was synthesized via reversible addition–fragmentation chain transfer (RAFT)-controlled radical polymerization using a PEG-macro chain transfer agent (CTA). The water-soluble complex (C_{70}/EUM) was synthesized by crushing powders of EUM and C_{70} together in a mortar and then adding water. The C_{70}/EUM complex formed a core–shell-corona micelle with a C_{70}/PMPC core, thermoresponsive PUEM shell, and hydrophilic PEG corona

(Fig. 1). C_{70}/EUM can encapsulate guest molecules such as hydrophobic drugs in the PUEM shell because the PUEM block becomes hydrophobic below the UCST. Water-soluble PEG exhibits biocompatibility.[43–45] C_{70}/EUM covered with biocompatible PEG coronas is expected to accumulate in the affected region due to the EPR effect in the body. Heating above the UCST of the PUEM shell can release the encapsulated hydrophobic drugs in C_{70}/EUM. C_{70}/EUM is thought to have two functions: singlet oxygen generation upon visible-light irradiation and a drug carrier that can release the drug when heated.

Experimental

Materials

2-(Methacryloyloxy)ethyl phosphorylcholine (MPC) was purchased from NOF Corp. (Tokyo, Japan) and used after recrystallization from acetonitrile.[31] Poly(2-(methacryloyloxy)ethyl phosphorylcholine) (PMPC, degree of polymerization (DP) = 100, number-average molecular weight (M_n) = 2.98 × 10^4 g mol^{-1}, molecular weight distribution (M_w/M_n) = 1.16) was synthesized via RAFT polymerization using 4-cyanopentanoic acid dithiobenzoate as a CTA. The CTA, α-methyltrithiocarbonate-S-phenylacetic acid (MTPA), was synthesized according to the literature.[46] 4,4′-Azobis(4-cyanovaleric acid) dihydrochloride (V-501, 95%), N,N′-dicyclohexylcarbodiimide (DCC, 95%), imidazole (98%), potassium cyanate (KOCN, 95%), fullerene-C_{70} (C_{70}, 99%), and doxorubicin (DOX) hydrochloride were purchased from Wako Pure Chemical (Osaka, Japan). 4-Dimethylaminopyridine (DMAP, 99%), poly(ethylene glycol) monomethyl ether (PEG_{45}, M_n = 2000, DP = 45), 2-aminoethyl methacrylate hydrochloride (AEM, 90%), phosphate-buffered saline (PBS), and deuterium oxide (D_2O) were purchased from Sigma-Aldrich (St. Louis, MO, USA). Singlet oxygen sensor green (SOSG) was purchased from Thermo Fisher Scientific (Waltham, MA, USA). Ion-exchanged water was used as pure water.
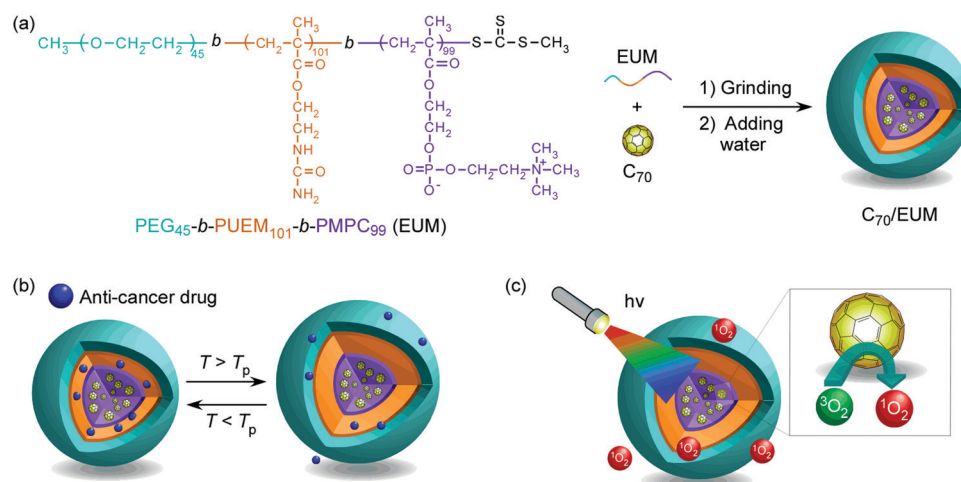

Fig. 1 (a) The chemical structure of PEG_{45}-b-$PUEM_{101}$-b-$PMPC_{99}$ (EUM) and a conceptual illustration of C_{70}/EUM micelle formation in water, (b) its thermo-responsive behavior, and (c) its light-responsive behavior upon visible-light irradiation.

Synthesis of PEG₄₅-MTPA

PEG$_{45}$ (22.4 g, 11.2 mmol), MTPA (3.47 g, 13.4 mmol), and DMAP (40.0 mg, 0.327 mmol) were dissolved in dichloromethane (100 mL). A dichloromethane solution (100 mL) of DCC (5.66 g, 27.4 mmol) was added dropwise to the solution for 30 min under an Ar atmosphere. After refluxing at 40 °C for 16 h, purification was performed on a silica-gel column using chloroform as the eluent. The solvent was evaporated. The residue was further dissolved in pure water, and PEG$_{45}$-MTPA was obtained by a freeze-drying technique (6.48 g, 25.1%). According to ^{1}H NMR, the number-average degree of polymerization (DP(NMR)) and number-average molecular weight (M_n(NMR)) determined from ^{1}H NMR were 45 and 2.24 × 10^3 g mol^{-1}, respectively (Fig. S1a, ESI†). According to gel-permeation chromatography (GPC) using tetrahydrofuran (THF) as an eluent, the number-average molecular weight M_n(GPC) and molecular weight distribution (M_w/M_n) were 4.40 × 10^3 g mol^{-1} and 1.03, respectively (Fig. S2a, ESI†).

Synthesis of PEG₄₅-b-PAEM₁₀₁-b-PMPC₉₉ (EAM)

The triblock copolymer (EAM; PEG$_{45}$-b-PAEM$_{101}$-b-PMPC$_{99}$) synthesis using PEG$_{45}$-MTPA as a macro CTA was performed in one pot as follows: AEM (2.09 g, 12.6 mmol), PEG$_{45}$-MTPA (274 mg, 0.121 mmol), and V-501 (13.9 mg, 0.0496 mmol) were dissolved in imidazole buffer ([imidazole] = 1.00 M, pH = 6.00, 12.1 mL) with a molar ratio of [AEM]/[PEG$_{45}$-MTPA]/[V-501] = 104/1/0.4. The solution was heated at 70 °C for 6 h under an Ar atmosphere. The conversion (PEG$_{45}$-b-PAEM$_{101}$) after polymerization obtained using ^{1}H NMR was 97.5%. DP(NMR) and M_n(NMR) obtained using ^{1}H NMR were 101 and 1.90 × 10^4 g mol^{-1}, respectively (Fig. S1b, ESI†). The M_n(GPC) and M_w/M_n determined by GPC measurement using acetate buffer as an eluent were 4.69 × 10^4 g mol^{-1} and 1.25, respectively (Fig. S2b, ESI†).

MPC (3.59 g, 12.2 mmol) and V-501 (14.1 mg, 0.0503 mmol) were dissolved in imidazole buffer (12.1 mL). The MPC solution was degassed with Ar and was added to the polymerization solution of PEG$_{45}$-b-PAEM$_{101}$ with a molar ratio of [MPC]/[PEG$_{45}$-b-PAEM$_{101}$]/[V-501] = 100/1/0.416. The solution was heated at 70 °C for 16 h under an Ar atmosphere. The polymer conversion after polymerization (PEG$_{45}$-b-PAEM$_{101}$-b-PMPC$_{99}$) obtained from ^{1}H NMR was 100%. Dialysis was performed overnight against pure water, and PEG$_{45}$-b-PAEM$_{101}$-b-PMPC$_{99}$ (EAM) was obtained via freeze-drying (5.51 g, 92.6%). DP(NMR) and M_n(NMR) obtained from ^{1}H NMR were 99 and 4.80 × 10^4 g mol^{-1}, respectively (Fig. S1c, ESI†). The M_n(GPC) and M_w/M_n determined using GPC were 5.29 × 10^4 g mol^{-1} and 1.61, respectively (Fig. S2b, ESI†).

Synthesis of PEG₄₅-b-PUEM₁₀₁-b-PMPC₉₉ (EUM)

Ureido groups were introduced in the pendant amino groups in the PAEM block in EAM as follows: EAM (4.00 g, 0.0828 mmol) and KOCN (808 mg, 9.94 mmol) were dissolved in imidazole buffer (16.0 mL) with molar ratio of [EAM]/[KOCN] = 1/120. The solution was heated at 50 °C for 24 h. PEG$_{45}$-b-PUEM$_{101}$-b-PMPC$_{99}$ (EUM) was obtained via freeze-drying (3.30 g, 70.6%)

after dialysis against pure water for three days. The conversion from the amine to ureido groups estimated using ^{1}H NMR was 100% (Fig. S3, ESI†). The M_n(GPC) and M_w/M_n obtained using GPC were 8.91 × 10^4 g mol^{-1} and 1.64, respectively (Fig. S2b, ESI†).

Preparation of the C₇₀/EUM Complex

C$_{70}$ (6.0 mg, 7.14 × 10^{-3} mmol) and EUM (20 mg, 4.14 × 10^{-4} mmol, M_n(theo) = 4.83 × 10^4 g mol^{-1}, M_w/M_n = 1.64) powders were ground in a mortar for 30 min. PBS (10 mL) was further added to adjust the polymer concentration (C_p) = 2 g L^{-1} to synthesize a water-soluble complex (C$_{70}$/EUM). Undissolved C$_{70}$ was eliminated by centrifugation at 6000 rpm for 30 min and then filtered through a 0.2 μm pore-sized filter. The C$_{70}$/PMPC complex, which comprises C$_{70}$ and PMPC, was synthesized in the same way as the C$_{70}$/EUM complex for comparison.

Measurements

^{1}H NMR spectra were obtained using a Bruker (Massachusetts, United States) DRX-500 and JEOL (Tokyo, Japan) ECZ400. GPC measurements using THF as the eluent were performed using a Shodex (Tokyo, Japan) DS-4 pump and a Shodex (Tokyo, Japan) RI-101 refractive index (RI) detector equipped with a Shodex (Tokyo, Japan) 10.0 μm bead size GF-7M column (exclusion limit ∼ 10^7) working at 40 °C, with a flow rate of 1.0 mL min^{-1}. The M_n(GPC) and M_w/M_n were calibrated using standard poly(styrene) samples. GPC measurements using acetate buffer as the eluent were performed using a JASCO (Tokyo, Japan) PU-8020 pump and a JASCO (Tokyo, Japan) RI-2031 Plus RI detector equipped with a Shodex (Tokyo, Japan) 10.0 μm bead size Ohpak SB-804 HQ column (exclusion limit of approximately 10^7) working at 40 °C, with a flow rate of 0.6 mL min^{-1}. A 0.5 M acetic acid aqueous solution containing 0.3 M sodium sulfate was used as the eluent. The M_n(GPC) and M_w/M_n were calibrated using standard poly(2-vinylpyridine) samples. Ultraviolet-visible (UV-vis) absorption spectra and percent transmittance (%T) at a wavelength of 700 nm were measured at varying temperatures using a JASCO (Tokyo, Japan) V-630BIO UV-vis spectrophotometer. Dynamic light scattering (DLS) measurements were performed with a Malvern (Worcestershire, UK) Zetasizer Nano ZS using a He–Ne laser (4 mW) with a wavelength of 632.8 nm as the light source. The hydrodynamic radius (R_h) and light scattering intensity (LSI) were obtained. Static light scattering (SLS) measurement was performed using an Otsuka Electronics Photal (Osaka, Japan) DLS-7000 HL light scattering spectrometer. A He–Ne laser (632.8 nm, 10.0 mW) was used as the light source. The weight-average molecular weight (M_w(SLS)) and radius of gyration (R_g) were estimated from the Zimm plots. The RI increment (dn/dC_p) values at 633 nm were determined using an Otsuka Electronics Photal (Osaka, Japan) DRM-3000 differential refractometer. Transmission electron microscopy (TEM) observations were performed using a JEOL (Tokyo, Japan) JEM-2100 microscope at an acceleration voltage of 160 kV. The TEM sample was prepared by placing one drop of aqueous solution on a copper grid coated with Formvar thin films. Excess water was blotted using filter

This journal is © The Royal Society of Chemistry 2022

J. Mater. Chem. B, 2022, **10**, 2551–2560 | **2553**

paper. The samples were stained with sodium phosphotungstate and dried under vacuum conditions. Fluorescence measurement was performed using a Hitachi F-2500 (Tokyo, Japan). Visible light at 590 nm was irradiated using OptCode (Tokyo, Japan) EX-590 with a power consumption of 3 W.

Characterization of C_{70}/EUM

The complex solution was diluted 100-fold and the UV-vis absorption spectrum was measured at 25 °C. The complex solution's absorbance before dilution was calculated by multiplying the absorbance of the complex solution after dilution by 100. The concentration of C_{70} ($[C_{70}]_s$) was calculated from its molar extinction coefficient in water at 384 nm (40.4 L g^{-1} cm^{-1})[28] and the absorbance.

Encapsulation and controlled release of DOX

DOX encapsulation in C_{70}/EUM was confirmed *via* fluorescence measurements. DOX (0.02 g L^{-1}) was dissolved in a C_{70}/EUM (C_p = 2 g L^{-1}, $[C_{70}]_s$ = 0.313 g L^{-1}) PBS solution and stirred at 25 °C for 3 days to synthesize DOX-encapsulating C_{70}/EUM (DOX@C_{70}/EUM). Dialysis was performed against a substantial excess of PBS at 25 °C for 3 days with a polycarbonate dialysis membrane (Harvard MFP, Massachusetts, USA) with a pore size of 10 nm to remove the DOX that could not be encapsulated. It was confirmed that the free DOX in the C_{70}/EUM solution was eliminated by comparing the fluorescence intensity inside the dialysis membrane of the DOX@C_{70}/EUM solution with the DOX-only blank solution. The concentration of DOX was calculated from the fluorescence intensity to estimate the amount of DOX in C_{70}/EUM. The release of DOX from DOX@C_{70}/EUM by heating was confirmed by fluorescence. The DOX@C_{70}/EUM ([DOX] = 0.0099 g L^{-1}) and DOX (0.01 g L^{-1}) blank solutions were dialyzed against PBS using a polycarbonate dialysis membrane at 25 °C and 50 °C. The fluorescence intensity of DOX emitted to the outside of the dialysis membrane was used to estimate the release rate.

Generation of singlet oxygen from C_{70}/EUM upon visible-light irradiation

SOSG was used to confirm $^1O_2$ generation. The fluorescence intensity increases when SOSG is oxidized with $^1O_2$ (Scheme S1, ESI†). The C_{70}/EUM PBS solutions at C_p = 2 g L^{-1} and $[C_{70}]_s$ = 0.313 g L^{-1} were diluted 10-fold with PBS, and SOSG was added to make it 2 μM. The solution was irradiated upon 590 nm visible light using an OptCode (Tokyo, Japan) EX-590 LED light, and the fluorescence spectra of SOSG were obtained. A quartz

cell with a 1 cm optical path length was used. The SOSG fluorescence was measured at an excitation wavelength of 420 nm with excitation and emission slight widths at 20 nm.

Results and discussion

Preparation of PEG$_{45}$-*b*-PAEM$_{101}$-*b*-PMPC$_{99}$ (EUM)

A macro CTA, PEG$_{45}$-MTPA, was synthesized *via* condensation reaction of mono-methoxy PEG$_{45}$ having an OH group at the chain end with MTPA having a carboxylic acid (Scheme S2, ESI†). DP(NMR) of PEG$_{45}$-MTPA estimated from the integrated intensity ratio of the peaks attributed to PEG methylene protons at 3.4 ppm and the MTPA methyl proton at 2.7 ppm was 45 (Fig. S1a, ESI†). AEM and MPC were continuously polymerized in one pot *via* RAFT polymerization using PEG$_{45}$-MTPA. The PAEM block's DP(NMR) estimated from the integrated intensity ratio of the peaks attributed to PEG methylene proton at 3.6 ppm to the pendant methylene protons in the PAEM block at 3.3 ppm was 101 (Fig. S1b, ESI†). The PMPC block's DP(NMR) estimated from the integrated intensity ratio of the peaks of the pendant methylene protons in the PAEM block at 3.3 ppm to the pendant methyl protons in the PMPC block at 3.1 ppm was 99 (Fig. S1c, ESI†). GPC measurements were performed for PEG$_{45}$-MTPA using THF as the eluent (Fig. S2a, ESI†), and for PEG$_{45}$-*b*-PAEM$_{101}$, EAM, and EUM using aqueous acetic acid solution (Fig. S2b, ESI†). M_w/M_n of PEG$_{45}$-MTPA, PEG$_{45}$-*b*-PAEM$_{101}$, EAM, and EUM were 1.03, 1.25, 1.61, and 1.64, respectively. Theoretical DP (DP(theo)) and number-average molecular weight (M_n(theo)) values were calculated from the following equations

$$DP(theo) = \frac{[M]_0}{[CTA]_0} \times \frac{x}{100} \qquad (1)$$

$$M_n(theo) = DP(theo) \times M_m + M_{CTA} \qquad (2)$$

where x is the monomer's percent conversion, $[M]_0$ and $[CTA]_0$ are the initial concentrations of the monomer and CTA, respectively, and M_m and M_{CTA} are the molecular weights of the monomer and CTA, respectively. Table 1 summarizes the results.

The ^{1}H NMR spectra for EAM and EUM were measured in D$_2$O at 25 °C and 80 °C (Fig. S3, ESI†). The peak of pendant methylene protons adjacent to the primary amine in the PAEM block at 3.3 ppm (*g*) disappeared in the ^{1}H NMR for EUM. This suggests that the pendant primary amine groups in the PAEM block were replaced with 100% ureido groups in the PUEM

Table 1 Molecular characteristics of the synthesized polymers

Polymers	M_n (theoretical) × 10^{-4} (g mol^{-1})	DP (theoretical)	M_n (NMR) × 10^{-4} (g mol^{-1})	DP (NMR)	M_n (GPC) × 10^{-4} (g mol^{-1})	M_w/M_n
PEG$_{45}$-MTPA	—	—	0.224	45[a]	0.44	1.03
PEG$_{45}$-*b*-PAEM$_{101}$	1.90	101[b]	1.90	101[b]	4.69	1.25
PEG$_{45}$-*b*-PAEM$_{101}$-*b*-PMPC$_{99}$	4.82	100[c]	4.80	99[c]	5.29	1.61
EUM	5.24	101[d]	—[e]	—[e]	8.91	1.64

[a] DP of PEG. [b] DP of PAEM. [c] DP of PMPC. [d] DP of PUEM. [e] The signals were overlapped.

2554 | *J. Mater. Chem. B*, 2022, **10**, 2551–2560

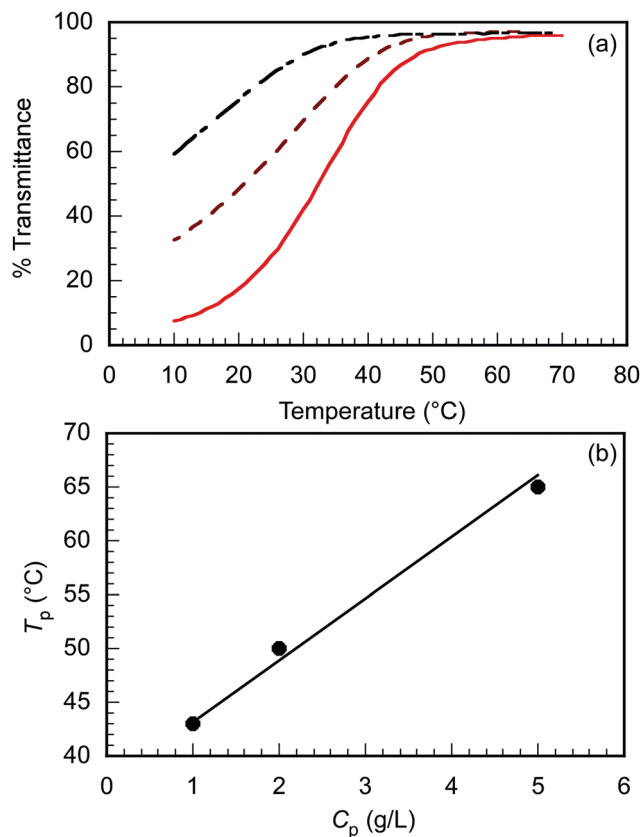This journal is © The Royal Society of Chemistry 2022

Fig. 2 (a) Percent transmittance (%T) of EUM in PBS at various C_p = 1.0 (—·—·—·), 2.0 (— — —), and 5.0 g L^{-1} (———) as a function of temperature with the cooling process, and (b) phase transition temperature (T_p) of EUM in PBS as a function of C_p.

block. Although the peak intensity of pendant methylene protons adjacent to the ureido in the PUEM block in EUM at 3.4 ppm (i) was barely observed at 25 °C (Fig. S3b, ESI†), the peak intensity at 3.4 ppm (i) increased at 80 °C (Fig. S3c, ESI†). The NMR peak becomes broad when the proton mobility decreases due to a decrease in the spin–spin relaxation time.[47] Therefore, this result suggests that the PUEM block's motion in EUM was restricted in D$_2$O at 25 °C.

The triblock copolymer, EUM, which contains the thermoresponsive PUEM block bearing pendant ureido groups exhibits UCST behavior in water. The polymer concentration (C_p) dependence on the thermo-responsive behavior of EUM was investigated. EUM was dissolved in PBS at various C_p values. The percent transmittance (%T) at 700 nm was measured as a function of temperature at the cooling process (Fig. 2a). The phase transition temperatures (T_p) at C_p = 1.0, 2.0, and 5.0 g L^{-1} were 43 °C, 50 °C, and 65 °C, respectively. T_p increased linearly with C_p (Fig. 2b). This phenomenon suggests that the frequency of collisions between polymer chains increased with increasing C_p to easily form hydrogen bonds between the pendant ureido groups in the PUEM blocks.

Preparation of C$_{70}$/EUM

The maximum C$_{70}$ concentration ([C$_{70}$]$_s$) to be solubilized by the formation of a water-soluble C$_{70}$/EUM complex was

determined. EUM and various amounts of C$_{70}$ ([C$_{70}$]$_f$) powders were ground and mixed in a mortar for 30 min. PBS was added to the mixture to synthesize C$_{70}$/EUM aqueous solution with C_p = 2.0 g L^{-1}. A water-soluble C$_{70}$/PMPC complex was prepared using C$_{70}$ and PMPC powders by the same method for comparison. From the UV-vis absorption spectra of C$_{70}$/EUM (Fig. 3) and C$_{70}$/PMPC (Fig. S4, ESI†), maximum absorptions are observed at 250 and 384 nm for C$_{70}$, suggesting that C$_{70}$ could be solubilized in PBS. The absorbance of C$_{70}$ increased for C$_{70}$/EUM and C$_{70}$/PMPC with an increase in [C$_{70}$]$_f$. The [C$_{70}$]$_s$ values were calculated using the molar extinction coefficient of C$_{70}$ in water at 384 nm (40.4 L g^{-1} cm^{-1})[28] and the absorbance of the complex aqueous solution. The maximum [C$_{70}$]$_s$ values using EUM and PMPC were 0.31 g L^{-1} (Fig. 3b) and 0.83 g L^{-1} (Fig. S4b, ESI†), respectively. The maximum EUM [C$_{70}$]$_s$ was lower than that of PMPC at C_p = 2 g L^{-1} because the EUM's PEG and PUEM blocks cannot contribute to solubilizing C$_{70}$. C$_{70}$ powder was mixed with PEG$_{45}$ or PUEM$_{100}$ powders at [C$_{70}$]$_f$ = 0.10 g L^{-1}; however, C$_{70}$ could not be solubilized in PBS (Fig. S5, ESI†). This observation suggests that the C$_{70}$/EUM core was formed from the C$_{70}$ and PMPC block. The maximum [C$_{70}$]$_s$ (0.313 g L^{-1}) obtained using EUM with C_p = 2 g L^{-1} was higher than that ([C$_{70}$]$_s$ = 0.2 g L^{-1}) obtained using PVP (MW = 4.00 × 10^4 g mol^{-1}) with C_p = 50 g L^{-1}.[28] All subsequent experiments were conducted using C$_{70}$/EUM with [C$_{70}$]$_s$ = 0.313 g L^{-1}.
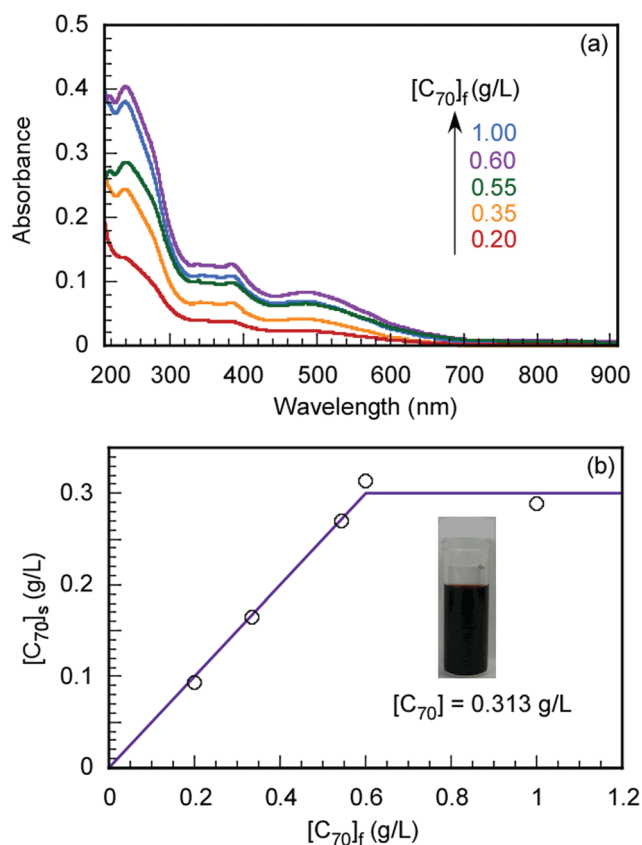

Fig. 3 (a) UV-vis absorption spectra of C$_{70}$/EUM at C_p = 2 g L^{-1} and (b) the amount of solubilized C$_{70}$ in PBS ([C$_{70}$]$_s$) as a function of the amount of C$_{70}$ ([C$_{70}$]$_f$) used.
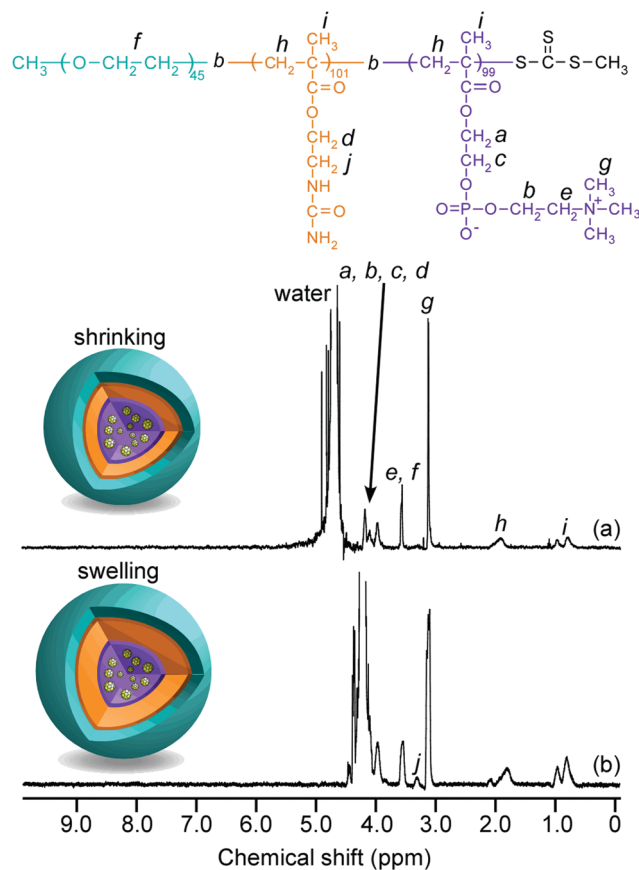
This journal is © The Royal Society of Chemistry 2022

J. Mater. Chem. B, 2022, 10, 2551–2560 | 2555

Fig. 4 ^{1}H NMR spectra for C_{70}/EUM in D_2O at (a) 25 °C and (b) at 80 °C.

Characterization of C_{70}/EUM

^{1}H NMR spectra for C_{70}/EUM were obtained in D_2O at 25 °C and 80 °C (Fig. 4). Although the NMR signal of the pendant methylene protons adjacent to the ureido group in the PUEM block was observed at 3.4 ppm at 80 °C, the signal disappeared at 25 °C. This observation suggests that 80 °C was higher than the phase transition temperature (T_p) of the PUEM block in C_{70}/EUM. Alternatively, the NMR signal for the PUEM shells in C_{70}/EUM became broad at 25 °C due to the decreasing mobility caused by the hydrophobic interactions of the PUEM shells below T_p.

SLS measurements were performed in PBS to characterize the association state of the complex (Fig. S6 and Table 2, ESI†). The theoretical expanded polymer chain lengths calculated from the DP values of EUM (245) and PMPC (100) were

61.3 and 25.0 nm, respectively, based on the C–C bond chain length. The R_g values of C_{70}/EUM and C_{70}/PMPC were 58.4 and 40.0 nm, respectively. The R_g value of C_{70}/EUM was similar to the theoretical expanded polymer chain length of EUM. This result suggests that C_{70}/EUM formed the core–shell-corona micelle structure without intermicellar aggregation. The R_g value of C_{70}/PMPC was larger than the theoretical chain length of 25.0 nm. This suggests that C_{70}/PMPC cannot form a clear core–shell structure. The aggregate's shape is a rigid sphere when the R_g/R_h value is 0.8, a random coil when it is 1, and a rod when it is larger than 2.[48] The R_g/R_h values of C_{70}/EUM and C_{70}/PMPC were 1.60 and 0.885, respectively. Therefore, C_{70}/EUM's shape was almost spherical, and C_{70}/PMPC formed aggregates similar to a rigid sphere. The inside of the C_{70}/PMPC complex was tightly packed because it does not have the shell and corona layers. Therefore, the R_g/R_h value of C_{70}/PMPC was similar to that of a rigid sphere.

The aggregation number (N_{agg}), which is the number of polymer chains forming one aggregate, can be calculated from the following equation

$$N_{agg} = \frac{M_w(SLS)}{M_{wpoly} + rM_{wC70}} \tag{3}$$

where M_{wpoly} and M_{wC70} are the weight-average molecular weights of the polymer and C_{70}, respectively, and r is the molar ratio of C_{70} to the polymer. N_{agg} of C_{70}/EUM was calculated to be 70.9. The number of C_{70} (N_{C70}) contained in one C_{70}/EUM complex was calculated to be 691 from r and N_{agg}. A single EUM polymer chain solubilized about 10 C_{70} molecules based on the N_{agg} and N_{C70} of C_{70}/EUM. Similarly, a single PMPC chain can solubilize 15 C_{70} molecules in the case of C_{70}/PMPC. PMPC can solubilize a greater amount of C_{70} than EUM in water because the PEG and PUEM blocks in EUM cannot contribute to C_{70} solubilization. The density (d) of the complex was calculated from the following formula

$$d = \frac{M_w(SLS)}{4/3\pi R_g^3 N_A} \tag{4}$$

where N_A represents Avogadro's number. The d values of C_{70}/EUM and C_{70}/PMPC were 0.0133 and 0.0144 g/cm^3, respectively. The d value of C_{70}/EUM was lower than that of C_{70}/PMPC, because C_{70}/EUM contains the PUEM shell and PEG corona layers, which are well-hydrated.

TEM observation confirmed the spherical shape of the C_{70}/EUM complex (Fig. 5). The average R_{TEM} obtained from TEM was 33.5 nm, which was lower than the R_h = 36.3 nm calculated

Table 2 DLS and SLS data for the C_{70}/EUM and C_{70}/PMPC complexes in PBS at 25 °C

Samples	r^a	$M_w(SLS)^b \times 10^{-6}$ (g mol^{-1})	R_g^c (nm)	R_h^d (nm)	R_g/R_h	d^e (g cm^{-3})	N_{agg}^f	N_{C70}^g	dn/dC_p^h (mL g^{-1})
C_{70}/EUM	9.8	6.67	58.4	36.3	1.60	0.0133	70.9	691	0.068
C_{70}/PMPC	15	2.32	40.0	45.2	0.885	0.0144	46.7	686	0.780

^{a} Molar ratio of C_{70} and polymer ([C_{70}]/[polymer]). ^{b} Weight-average molecular weight estimated from SLS. ^{c} Radius of gyration estimated from SLS. ^{d} Hydrodynamic radius estimated from DLS. ^{e} Density of complex calculated from eqn (4). ^{f} Aggregation number of the polymer chains calculated from eqn (3). ^{g} Number of C_{70} in one complex calculated from r and N_{agg}. ^{h} Refractive index increment.
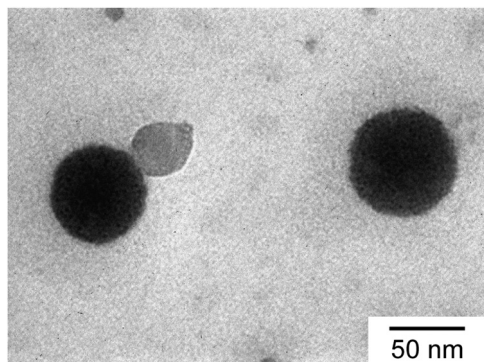
Fig. 5 TEM image of C_{70}/EUM.

by DLS at 25 °C due to shrinkage of the TEM sample during the drying process.

Thermo responsive behavior of C_{70}/EUM

R_h and LSI were investigated as a function of temperature to confirm the swelling of C_{70}/EUM in PBS due to the PUEM shell's UCST behavior upon heating (Fig. 6). T_p was defined as the temperature at which R_h became constant during a heating process. Below 36 °C, R_h increased with temperature. However, R_h became constant above 36 °C at about 38 nm. Therefore, C_{70}/EUM's T_p was determined to be 36 °C. The T_p of EUM without C_{70} was 50 °C (Fig. 2); however, the T_p of C_{70}/EUM decreased to 36 °C. The PUEM block in C_{70}/EUM is anchored to the hydrophobic core containing C_{70}. The T_p of C_{70}/EUM was decreased because the PUEM polymer chain was affected by the core's hydrophobicity. LSI is sensitive to the formation of large aggregates due to intermicellar aggregations. No intermicellar aggregation of C_{70}/EUM occurred because the LSI was constant regardless of temperature. The R_h values were repeatedly measured at 25 °C and 70 °C to confirm the reversibility of swelling and shrinking of C_{70}/EUM by temperature (Fig. S7, ESI†). The R_h values at 25 °C and 70 °C were *ca.* 36.5 and *ca.* 38.2 nm, respectively, for five temperature cycles. Thus, the C_{70}/EUM complex phase transition is reversible.
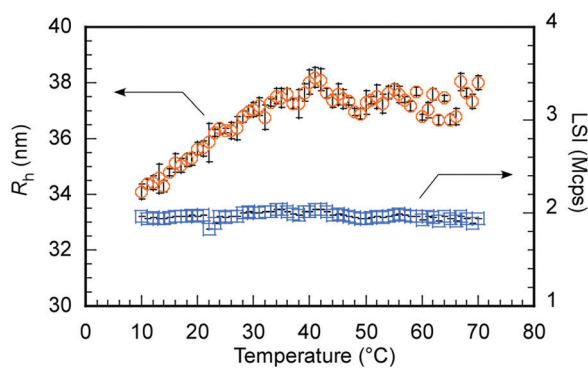
Generation of singlet oxygen from C_{70}/EUM upon visible-light irradiation

The C_{70}/EUM solution with C_p = 2 g L^{-1} and $[C_{70}]_s$ = 0.313 g L^{-1} was diluted 10-fold with PBS, and SOSG was added to make it 2 μM. The solution was irradiated with light at 590 nm and SOSG's fluorescence spectra were measured for the irradiation time, t (Fig. S8, ESI†). The ratios (I/I_0) of SOSG's maximum fluorescence intensity (I) at t to that before irradiation (I_0) were plotted against t (Fig. 7). SOSG is oxidized by $^1O_2$ and the fluorescence intensity at 500–550 nm increased. The I/I_0 ratios were almost constant regardless of light irradiation when the light was irradiated towards SOSG PBS solution and SOSG PBS solution with EUM. Alternatively, when the SOSG PBS solution with the C_{70}/EUM complex containing C_{70} was irradiated, I/I_0 of SOSG increased with the irradiation time. This observation indicates that C_{70} in C_{70}/EUM's core can generate $^1O_2$ under light irradiation. There is almost no error for the samples that cannot generate $^1O_2$. The plots of C_{70}/EUM contains up to 1.5% error.

Encapsulation and controlled release of doxorubicin (DOX)

The following method was used to encapsulate a guest molecule, DOX, in C_{70}/EUM: C_{70}/EUM and DOX were stirred in PBS for 3 days at room temperature, and any DOX that could not be encapsulated was removed by dialysis. The removal of free DOX from DOX@C_{70}/EUM solution was confirmed by comparing the fluorescence intensity inside the dialysis membrane of the DOX@C_{70}/EUM solution with that of the DOX-only solution (Fig. S9, ESI†). The fluorescence intensity of DOX@C_{70}/EUM was almost constant after 24 h dialysis, whereas that of the DOX-only solution decreased. These results show that DOX can be encapsulated in C_{70}/EUM. From eqn (5) and (6), DOX encapsulation efficiency (EE) and DOX encapsulation content (EC) were estimated after 72 h dialysis. EE is the ratio of the encapsulated DOX concentration ([DOX]) to the used DOX concentration ($[DOX]_0$). [DOX] can be estimated from the
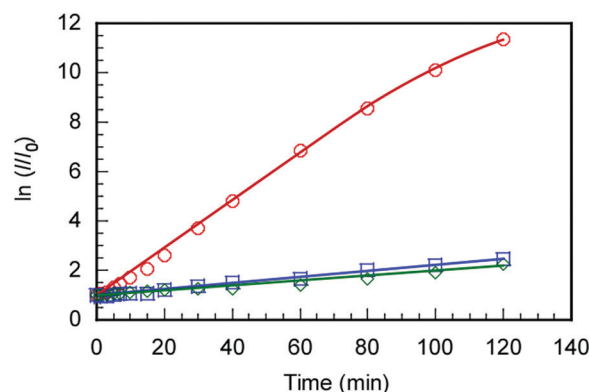

Fig. 7 Changes in SOSG fluorescence intensity (2 μM) with C_{70}/EUM at $[C_{70}]_s$ = 0.03 g L^{-1} (○), without C_{70} (◇), and without EUM and C_{70} (□) as a function of irradiation time at 25 °C; I and I_0 were the fluorescence intensities at the maximum wavelength (525 nm) after and before 590 nm light irradiation.

Fig. 6 Hydrodynamic radius (R_h, ○) and light scattering intensity (LSI, □) of C_{70}/EUM at C_p = 2 g L^{-1} and $[C_{70}]_s$ = 0.313 g L^{-1} in PBS as a function of temperature.

This journal is © The Royal Society of Chemistry 2022

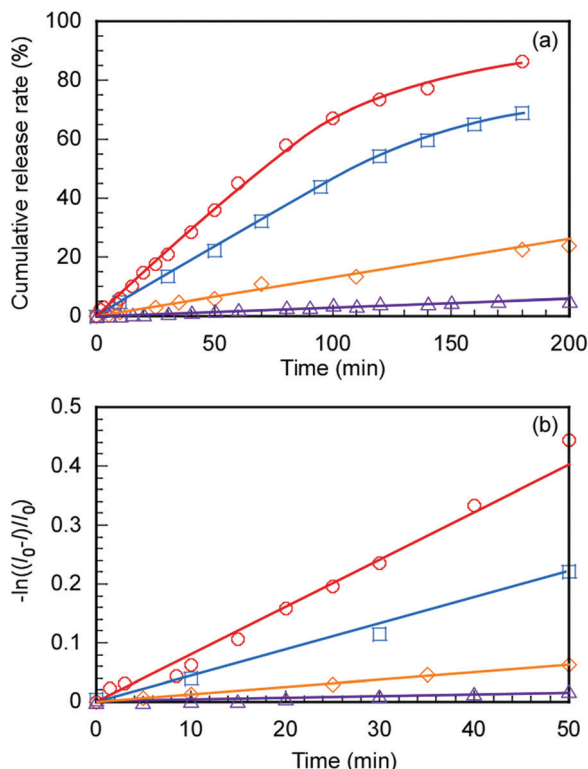*J. Mater. Chem. B*, 2022, **10**, 2551–2560 | **2557**

Fig. 8 (a) DOX release from DOX@C$_{70}$/EUM at 25 °C (△) and 50 °C (◇) and a blank solution without C$_{70}$/EUM at 25 °C (□) and 50 °C (○) in PBS. (b) $-\ln((I_0 - I)/I_0)$ was plotted against dialysis time; I is the fluorescence intensity and I_0 is the initial intensity.

fluorescence intensity. EC is the ratio of [DOX] to C_P. EE(%) and EC(%) were calculated using the following equations:

$$EE(\%) = \frac{[DOX]}{[DOX]_0} \times 100 \qquad (5)$$

$$EC(\%) = \frac{[DOX]}{C_P} \times 100 \qquad (6)$$

EE(%) and EC(%) for DOX@C$_{70}$/EUM were 49.5% and 0.495%, respectively.

The controlled release of DOX from DOX@C$_{70}$/EUM with increasing temperature was evaluated. The DOX@C$_{70}$/EUM solutions were dialyzed against PBS at 25 °C and 50 °C. The dialysis membrane with a 10 nm pore size was used. DOX can permeate the membrane, whereas large-sized C$_{70}$/EUM cannot. DOX fluorescence intensity released to the outside of the membrane was investigated with respect to the dialysis time (Fig. 8a). The DOX-only solution was studied using the same method for comparison. DOX release amounts at 50 °C and 25 °C after 200 min were 90% and 70%, respectively, for the DOX-only solution. The significant increase at 50 °C was attributed to an increase in DOX thermal motion due to high temperature. The amounts of DOX released from DOX@C$_{70}$/EUM at 50 °C and 25 °C after 200 min were 20% and 5.0%, respectively. The plots contain up to 1.2% error. At 50 °C the release amount of DOX@C$_{70}$/EUM solution was lower than that of the DOX-only solution. This suggests that not only is DOX encapsulated in

the PUEM shell in C$_{70}$/EUM due to hydrophobic interactions but it also interacts with the hydrophobic core containing hydrophobic C$_{70}$. Assuming that DOX release follows the first-order mechanism,[49,50] the release rate constant (k) can be calculated from the following equation:

$$\ln((I_0 - I)/I_0) = -kt \qquad (7)$$

where I_0 indicates the initial fluorescence intensity of DOX before dialysis, I indicates the fluorescence intensity outside of the membrane at t, and t is the dialysis time. The slope of the plots in Fig. 8b shows that k can be calculated. The k values for the comparative DOX-only solution at 25 °C and 50 °C were 0.264 s^{-1} and 0.481 s^{-1}, respectively. Therefore, the release rate increased by 1.82 times at 50 °C compared to 25 °C due to the increase in DOX thermal motion. The k values for the DOX@C$_{70}$/EUM solution at 25 °C and 50 °C were 0.0180 s^{-1} and 0.0792 s^{-1}, respectively. The rate of release rose by 4.40 times when the temperature was raised to 50 °C from 25 °C. The increase in k due to heating in the presence of C$_{70}$/EUM was larger than in the absence of C$_{70}$/EUM. This suggests that DOX release could be controlled by increasing the temperature rather than just a simple change in thermal motion. The affinity between DOX and the PUEM block of C$_{70}$/EUM was changed drastically at 25 °C and 50 °C due to the UCST behavior of the PUEM block. At 50 °C the affinity of DOX with the PUEM block was low. Therefore, DOX can be released from C$_{70}$/EUM at 50 °C.

The effect of DOX encapsulation and release on C$_{70}$/EUM size was evaluated using DLS (Fig. S10, ESI†). The R_h value of C$_{70}$/EUM without DOX was 36.3 nm. The R_h value of DOX@C$_{70}$/EUM was 39.8 nm. The R_h value of C$_{70}$/EUM decreased to 32.7 nm after DOX release. The LSIs of DOX@C$_{70}$/EUM before and after the release of DOX were 2160 and 1830 kcps, respectively. From these results, the influence of DOX encapsulation and release on the size and structure of the complex was minimal.

Conclusions

A thermo-responsive triblock copolymer, EUM, was synthesized *via* controlled radical polymerization. EUM can dissolve C$_{70}$ in PBS to form a C$_{70}$/EUM complex. The maximum solubilized C$_{70}$ concentration in PBS by EUM was 0.313 g L^{-1}, which was superior to a previously reported solubilization method using PVP. The T_p value of C$_{70}$/EUM in PBS was 36 °C according to DLS measurements. C$_{70}$/EUM can generate $^1O_2$ by visible-light irradiation at 590 nm in PBS. C$_{70}$/EUM can encapsulate the anticancer agent, DOX, at 25 °C by hydrophobic interactions between the PUEM block in EUM and DOX. The encapsulated DOX can be released when heated. C$_{70}$/EUM may be expected to function as both a thermo-responsive drug carrier and a photosensitizer for PDT.

Author contributions

Conceptualization, S. Y.; data curation, K. K. and S. Y.; formal analysis, K. K. and S. Y.; investigation, K. K., K. I., and S. Y.;

2558 | *J. Mater. Chem. B*, 2022, **10**, 2551–2560

This journal is © The Royal Society of Chemistry 2022

methodology, K. I. and S. Y.; project administration, S. Y.; supervision, S. Y.; visualization, K. K.; writing – original draft, K. K. and S. Y.; writing – review and editing, K. K. K. I. and S. Y. All authors have read and agreed to the published version of the manuscript.

Conflicts of interest

There are no conflicts of interest to declare.

Acknowledgements

References

1 S. Durdagi, T. Mavromoustakos, N. Chronakis and M. G. Papadopoulos, *Bioorg. Med. Chem.*, 2008, **16**, 9957–9974.

2 G. Pastorin, S. Marchesan, J. Hoebeke, T. Da Ros, L. Ehret-Sabatier, J. P. Briand, M. Prato and A. Bianco, *Org. Biomol. Chem.*, 2006, **4**, 2556–2562.

3 A. Innocenti, S. Durdagi, N. Doostdar, T. Amanda Strom, A. R. Barron and C. T. Supuran, *Bioorg. Med. Chem.*, 2010, **18**, 2822–2828.

4 Á. Serrano-Aroca, K. Takayama, A. Tuñón-Molina, M. Seyran, S. S. Hassan, P. Pal Choudhury, V. N. Uversky, K. Lundstrom, P. Adadi, G. Palù, A. A. A. Aljabali, G. Chauhan, R. Kandimalla, M. M. Tambuwala, A. Lal, T. M. Abd El-Aziz, S. Sherchan, D. Barh, E. M. Redwan, N. G. Bazan, Y. K. Mishra, B. D. Uhal and A. Brufsky, *ACS Nano*, 2021, **15**, 8069–8086.

5 A. B. Kornev, A. S. Peregudov, V. M. Martynenko, J. Balzarini, B. Hoorelbeke and P. A. Troshin, *Chem. Commun.*, 2011, **47**, 8298–8300.

6 E. Nakamura and H. Isobe, *Acc. Chem. Res.*, 2003, **36**, 807–815.

7 S. Samal and K. E. Geckeler, *Macromol. Biosci.*, 2001, **1**, 329–331.

8 A. Kumar, M. V. Rao and S. K. Menon, *Tetrahedron Lett.*, 2009, **50**, 6526–6530.

9 F. Paquin, J. Rivnay, A. Salleo, N. Stingelin and C. Silva, *J. Mater. Chem. C*, 2015, **3**, 10715–10722.

10 Z. Markovic and V. Trajkovic, *Biomaterials*, 2008, **29**, 3561–3573.

11 M. Wang, S. Maragani, L. Huang, S. Jeon, T. Canteenwala, M. R. Hamblin and L. Y. Chiang, *Eur. J. Med. Chem.*, 2013, **63**, 170–184.

12 S. K. Sharma, L. Y. Chiang and M. R. Hamblin, *Nanomedicine*, 2011, **6**, 1813–1825.

13 P. Mroz, A. Pawlak, M. Satti, H. Lee, T. Wharton, H. Gali, T. Sarna and M. R. Hamblin, *Free Radical Biol. Med.*, 2007, **43**, 711–719.

14 V. Milic, M. Posa, B. Srdjenovic and A. Luísa, *Colloids Surf., B*, 2011, **82**, 46–53.

15 C. F. Richardson, D. I. Schuster and S. R. Wilson, *Org. Lett.*, 2000, **2**, 1011–1014.

16 Z. Yao and K. C. Tam, *Macromol. Rapid Commun.*, 2011, **32**, 1863–1885.

17 S. Bosi, L. Feruglio, T. Da Ros, G. Spalluto, B. Gregoretti, M. Terdoslavich, G. Decorti, S. Passamonti, S. Moro and M. Prato, *J. Med. Chem.*, 2004, **47**, 6711–6715.

18 H. M. Wang and G. Wenz, *Beilstein J. Org. Chem.*, 2012, **8**, 1644–1651.

19 K. Komatsu, K. Fujiwara and T. Braun, *J. Chem. Soc., Perkin Trans. 1*, 1999, 2963–2966.

20 S. Kunsági-Máté, G. Vasapollo, K. Szabó, I. Bitter, G. Mele, L. Longo and L. Kollár, *J. Incl. Phenom. Macrocycl. Chem.*, 2008, **60**, 71–78.

21 T. Metanawin, T. Tang, R. Chen, D. Vernon and X. Wang, *Nanotechnology*, 2011, **22**, 235604–235613.

22 C. C. Cheng, W. L. Lin, Z. S. Liao, C. W. Chu, J. J. Huang, S. Y. Huang, W. L. Fan and D. J. Lee, *Polym. Chem.*, 2017, **8**, 7469–7474.

23 Y. Doi, A. Ikeda, M. Akiyama, M. Nagano, T. Shigematsu, T. Ogawa, T. Takeya and T. Nagasaki, *Chem. – Eur. J.*, 2008, **14**, 8892–8897.

24 A. Ikeda, T. Sato, K. Kitamura, K. Nishiguchi, Y. Sasaki, J. Kikuchi, T. Ogawa, K. Yogo and T. Takeya, *Org. Biomol. Chem.*, 2005, **3**, 2907–2909.

25 L. Xiao, K. Matsubayashi and N. Miwa, *Arch. Dermatol. Res.*, 2007, **299**, 245–257.

26 H. Takada, K. Kokubo, K. Matsubayashi and T. Oshima, *Biosci., Biotechnol., Biochem.*, 2006, **70**, 3088–3093.

27 H. Aoshima, S. Yamana, S. Nakamura and T. Mashino, *J. Toxicol. Sci.*, 2010, **35**, 401–409.

28 Y. Yamakoshi, T. Yagami, K. Fukuhara, S. Sueyoshi and N. Miyata, *Chem. Commun.*, 1994, 517–518.

29 L. Chrit, P. Bastien, B. Biatry, J. T. Simonnet, A. Potter and A. M. Minondo, *Biopolymers*, 2007, **85**, 359–369.

30 O. Tairy, N. Kampf, M. J. Driver, S. P. Armes and J. Klein, *Macromolecules*, 2015, **48**, 140–151.

31 K. Ishihara, T. Ueda and N. Nakabayashi, *Polym. J.*, 1990, **22**, 355–360.

32 A. Lewis, Y. Tang, S. Brocchini, J. W. Choi and A. Godwin, *Bioconjugate Chem.*, 2008, **19**, 2144–2155.

33 Y. Iwasaki and K. Ishihara, *Sci. Technol. Adv. Mater.*, 2012, **13**, 064101–064115.

34 T. Konno, J. Watanabe and K. Ishihara, *J. Biomed. Mater. Res., Part A*, 2003, **65**, 209–214.

35 T. Ohata, K. Ishihara, Y. Iwasaki, A. Sangsuwan, S. Fujii, K. Sakurai, Y. Ohara and S. Yusa, *Polym. J.*, 2016, **48**, 999–1005.

36 H. Hatakeyama, H. Akita and H. Harashima, *Adv. Drug Delivery Rev.*, 2011, **63**, 152–160.

This journal is © The Royal Society of Chemistry 2022

J. Mater. Chem. B, 2022, **10**, 2551–2560 | **2559**

37 H. Maeda, *J. Controlled Release*, 2012, **164**, 138–144.

38 V. Torchilin, *Adv. Drug Delivery Rev.*, 2011, **63**, 131–135.

39 M. Ethirajan, Y. Chen, P. Joshi and R. K. Pandey, *Chem. Soc. Rev.*, 2011, **40**, 340–362.

40 N. Shimada, H. Ino, K. Maie, M. Nakayama, A. Kano and A. Maruyama, *Biomacrimolecules*, 2011, **12**, 3418–3422.

41 G. Huang, H. Li, S. T. Feng, X. Li, G. Tong, J. Liu, C. Quan, Q. Jiang, C. Zhang and Z. Li, *Macromol. Chem. Phys.*, 2015, **216**, 1014–1023.

42 Y. Deng, F. Käfer, T. Chen, Q. Jin, J. Ji and S. Agarwal, *Small*, 2018, **14**, 1802420.

43 L. Douziech-Eyrolles, H. Marchais, K. Hervé, E. Munnier, M. Soucé, C. Linassier, P. Dubois and I. Chourpa, *Int. J. Nanomed.*, 2007, **2**, 541–550.

44 T. Il Kim, H. J. Seo, J. S. Choi, H. S. Jang, J. U. Baek, K. Kim and J. S. Park, *Biomacromolecules*, 2004, **5**, 2487–2492.

45 G. N. Grover, J. Lam, T. H. Nguyen, T. Segura and H. D. Maynard, *Biomacromolecules*, 2012, **13**, 3013–3017.

46 S. Yusa, T. Endo and M. Ito, *J. Polym. Sci., Part A: Polym. Chem.*, 2009, **47**, 6827–6838.

47 S. Yusa, K. Fukuda, T. Yamamoto, K. Ishihara and Y. Morishima, *Biomacromolecules*, 2005, **6**, 663–670.

48 A. Z. Akcasu and C. C. Han, *Macromolecule*, 1979, **12**, 276–280.

49 F. Meng, G. H. M. Engbers and J. Feijen, *J. Controlled Release*, 2005, **101**, 187–198.

50 S. Dash, P. N. Murthy, L. Nath and P. Chowdhury, *Acta Pol. Pharm.*, 2010, **67**, 217–223.