



Nanoscale

**DNA Conformational Equilibrium Enables Continuously Changing of Curvatures**

Journal:	<i>Nanoscale</i>
Manuscript ID	NR-COM-09-2022-005404.R1
Article Type:	Communication
Date Submitted by the Author:	05-Dec-2022
Complete List of Authors:	Mao, Dake; Purdue University, Department of Chemistry Paluzzi, Victoria; Purdue University, Department of Chemistry Zhang, Cuizheng; Purdue University, Department of Chemistry Mao, Chengde; University of Purdue, Department of Chemistry

SCHOLARONE™  
Manuscripts

# DNA Conformational Equilibrium Enables Continuously Changing of Curvatures

Dake Mao, Victoria E. Paluzzi, Cuizheng Zhang, and Chengde Mao\*

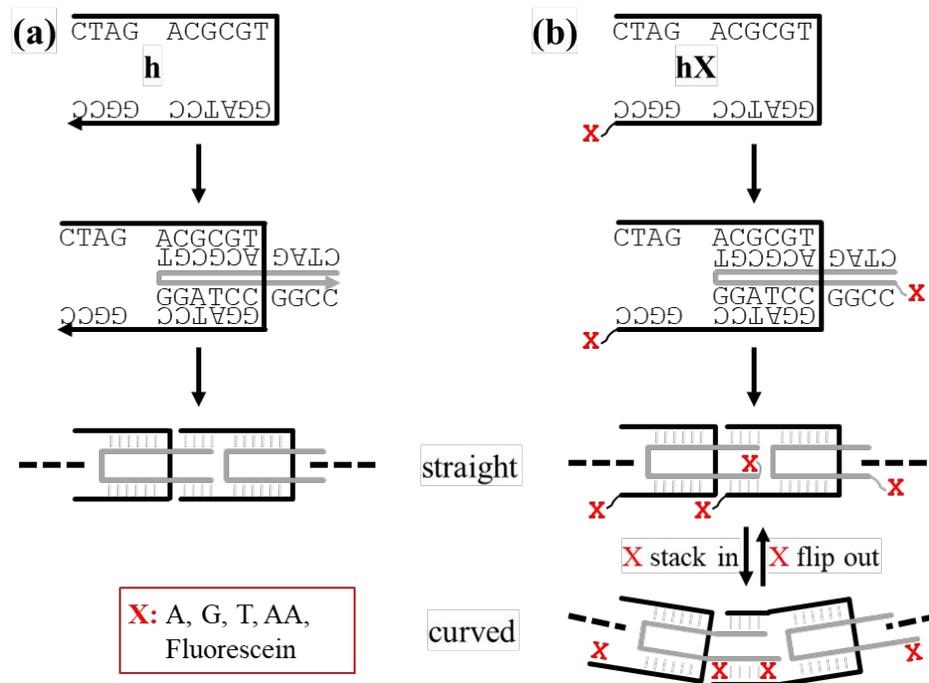
Purdue University, Department of Chemistry, West Lafayette, IN 47907, USA

\* mao@purdue.edu

## Abstract

Assembly of complex structures from a small set of tiles is a common theme in biology. For example, many copies of identical proteins make up polyhedral-shaped, viral capsids and tubulin can make long microtubules. This inspired the development of tile-based DNA self-assembly for nanoconstruction, particularly for structures with high symmetries. In the final structure, each type of motif will adopt the same conformation, either rigid or with defined flexibility. For structures that have no symmetry, their assembly remains a challenge from a small set of tiles. To meet this challenge, algorithmic self-assembly has been explored driven by computational science, but it is not clear how to implement this approach to one-dimensional (1D) structures. Here, we have demonstrated that a constant shift of a conformational equilibrium could allow 1D structures to evolve. As shown by atomic force microscopy imaging, one type of DNA tile has successfully assembled into DNA spirals and concentric circles, which becomes less and less curved from the structure's center outward. This work points to a new direction for tile-based DNA assembly.

DNA has been exploited to build nanostructures and nanomachines by programmed self-assembly.<sup>1-6</sup> A key event of structural DNA nanotechnology is the development of a series of rigid DNA nanomotifs,<sup>7-10</sup> which must satisfy various, precise geometric requirements. In contrast, structural flexibility in nanomotifs would lead to uncontrolled association among motifs; and thus, is generally avoided. Consequently, all nanomotifs will have the same, predefined conformation in the final structure. This is a powerful strategy but fails in assembling continuously changing structures. For example, spirals are simple structures, topologically equivalent to 1D chains, and could, in principle, be assembled from tiles in a 1D sense. In the spiral, individual tiles will have similar, but different conformations. Tiles become less and less curved from the structure's center outward. Strategies and nanomotifs reported previously could not be easily adapted to assemble such continuously changing structures.<sup>11-15</sup> Thus, we ask a question: can we design an assembly strategy/nanomotif in such a way that the tiles accommodate the necessary flexibility to continuously change their curvature? Would these assemble into structures that require the tiles to adapt to continuously changing conformations? Herein, we have developed an approach to address this problem. As a demonstration, a series of DNA spirals have been assembled by homo-polymerization of single component, 21-nucleotide (nt)-long DNA strands. The structures have been thoroughly characterized with polyacrylamide gel electrophoresis (PAGE) and atomic force microscopy (AFM) imaging.



**Figure 1.** DNA homopolymerization. (a) Assembly from tiles with one defined conformation. Two copies of DNA strand **h** consisting of four palindromes can dimerize

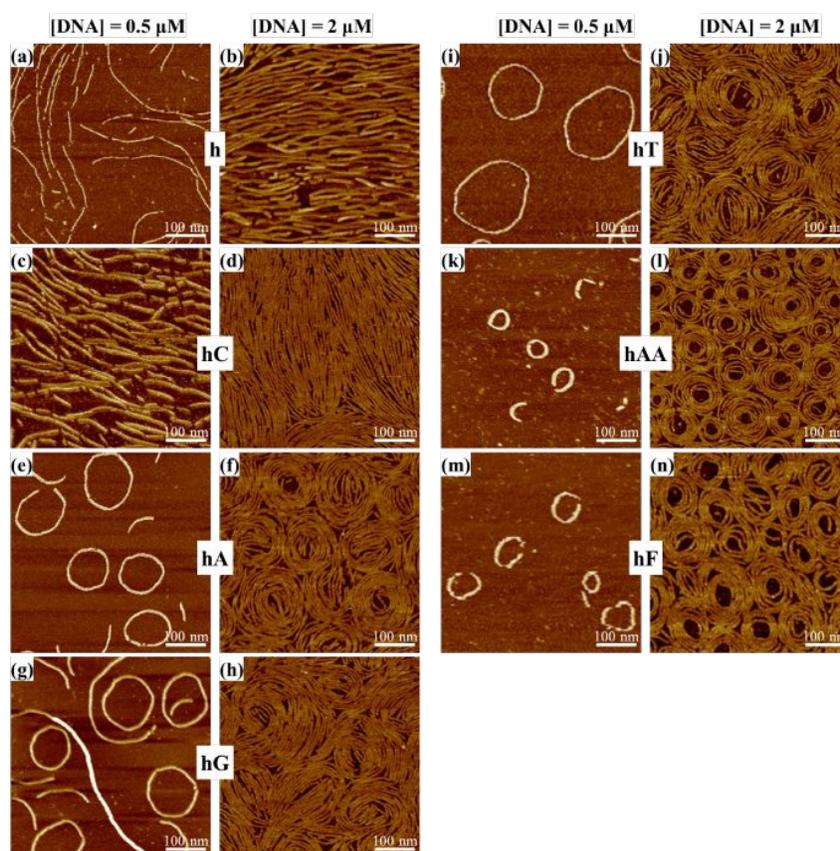
and further homopolymerize to 1D arrays. (b) Assembly from tiles under conformational equilibrium. The DNA strand **hX** has an extra chemical group X. When stacking into DNA duplexes, X will cause 1D arrays to bend because of the length difference between the duplexes in top and bottom. When X flips out of the helical domain, the 1D arrays will be straight. By different combinations of the two conformational states, the 1D arrays could continuously change their curvatures in a wide range.

Figure 1 illustrates the DNA assembly route. Each DNA architecture is homo-polymerized from a two-stranded, double-crossover (DX)-like (DXL) motif, which, in turn, is assembled via homodimerization from a short, single DNA strand. The basic sequence **h** is 20 nucleotides (nts) long and all other strands, **hXs**, contain a chemical group X (a single base A, G, T, C, or two bases AA, or a fluorescein, Figure S1) at the 3' end in addition to the basic sequence **h**. Those strands are named according to the chemical group X as **hX**. Strand **h** includes four palindromic segments: two 6-nt-long segments in the middle and two 4-nt-long segments at the two ends. During the polymerization, two **h** strands will first hybridize and associate with each other by the two central 6-nt-long palindromes to form a DXL motif. The four remaining palindromic tails could further hybridize with each other, resulting in one-dimensional (1D) arrays, exhibiting well-defined straight morphology (Figure 1a). The 1D arrays could be viewed as two, long, interconnected, parallel, pseudo-continuous DNA duplexes. For strands **hXs**, the extra chemical groups (X) will fundamentally change the array morphologies (Figure 1b). X will remain un-paired and can either be out of the duplex as a spare group, or stack in the helical domains. The two conformations consist of a dynamic and reversible equilibrium. Its balance is determined by the chemical structure of X and the system's environment. When X is out of the duplex domain, the resulting 1D arrays have straight morphology. When X stacks in the duplex, the two parallel duplexes will have different lengths and the mismatched lengths will force the 1D arrays to bend into curved shapes.<sup>16, 17</sup> For purines (G or A) and fluorescein, X has a large aromatic plane, thus, can stack strongly with DNA base pairs. For pyrimidine (T and C), X has a small aromatic plane; thus can only weakly stack with DNA base pairs and is more likely to remain outside of the DNA duplex.<sup>18</sup> Overall, the existence of X will have two effects: (1) 1D arrays can bend because of the length mismatch; (2) the bending extent along the 1D array could widely vary as the conformational equilibrium of the motifs could be easily changed. In DNA ladder, the equilibrium is also influenced by the surrounding environment, for example, the bending angle could be decrease and the circular structure could be more straight by more X base flipping out.

We first investigated DNA assembly in solution by native PAGE. DNA was dissolved in TAE/Mg<sup>2+</sup> buffer and directly analyzed with native PAGE at room temperature after thermal annealing. At 20  $\mu$ M (Figure S2), strand **h** assembled into large polymers, which appeared as a band near the well. In contrast, DNA strands (**hX**) with extra chemical groups appeared as smears with increased electrophoretic mobilities,

indicating they are smaller polymers. This observation suggested that the sticky-end cohesion was weakened by the extra chemical group intercalation. This caused the DNA polymer to dissociate into oligomers in solution. Judging by the native PAGE data, the destabilizing capability of X followed this order:  $C < G < A \ll T \sim AA \sim F$ . At  $2 \mu\text{M}$ , no appreciated polymerization in native PAGE was observed for all DNA strands except **h** (Figure S3).

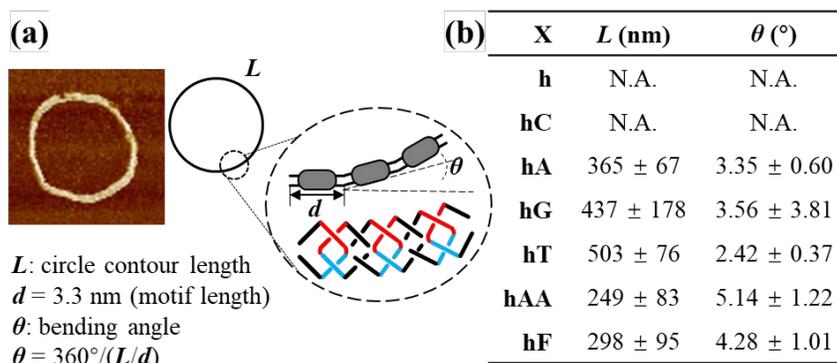
Then we used AFM to directly visualize the morphology of the assembled DNA 1D arrays. DNA solutions in TAE/Mg<sup>2+</sup> buffer were deposited on the fresh-cleaved mica surface and incubated for 10 min. After removing the excess solution, a TA/Mg<sup>2+</sup>/Ni<sup>2+</sup> imaging buffer was added onto the mica surface and the DNA samples were imaged in tapping mode.<sup>19, 20</sup>



**Figure 2.** AFM study of the morphologies of DNA homopolymers assembled from different DNA strands at two different DNA concentrations. (a) & (b) **h**; (c) & (d) **hC**; (e) & (f) **hA**; (g) & (h) **hG**; (i) & (j) **hT**; (k) & (l) **hAA**; (m) & (n) **hF**.

DNA samples from every individual DNA strand (**h** or **hX**) assembled into large nanostructures, which exhibited a rich variety of morphologies depending on the identity of the chemical group (X) and the DNA concentration (Figure 2 and S4). Poly(**h**), the polymer assembled from strand **h** without any extra chemical group at the 3' end,

appeared as straight fibers. Poly(**hC**) behaved similarly, indicating the extra base (C) at the 3' end flipped outside of the helical domain. When X is chemical group (A, G, T, AA, and Fluorescein) other than C, the assembled polymers accompanied with obvious, but different degrees of curvature. At a low DNA concentration (0.5  $\mu\text{M}$ ), well-separated, discrete circles or arcs were observed. The circles of poly(**hAA**) and poly(**hF**) were substantially smaller than the circles of poly(**hA**), poly(**hG**), and they all were smaller than poly(**hT**) circles. At a high DNA concentration (2  $\mu\text{M}$ ), closely packed spirals or multi-layered, concentric circles completely covered the mica surfaces. The  $\text{Mg}^{2+}$  concentration had a similar effect on the DNA assembly as the DNA concentration did (Figure S5). Divalent cations  $\text{Mg}^{2+}$  could promote the homopolymerization and DNA adsorption onto the mica surface, thus increasing the effective DNA concentration on the mica surface for DNA assembly. Please note that the two exact DNA concentrations (0.5 and 2  $\mu\text{M}$ ) were chosen from our experience based on the strength of the inter-motif interaction (sticky-end interaction). For different sticky-ends (in terms of length and base composition), the interaction strength would vary and the DNA concentration would be adjusted accordingly. But the overall phenomenon should be similar.

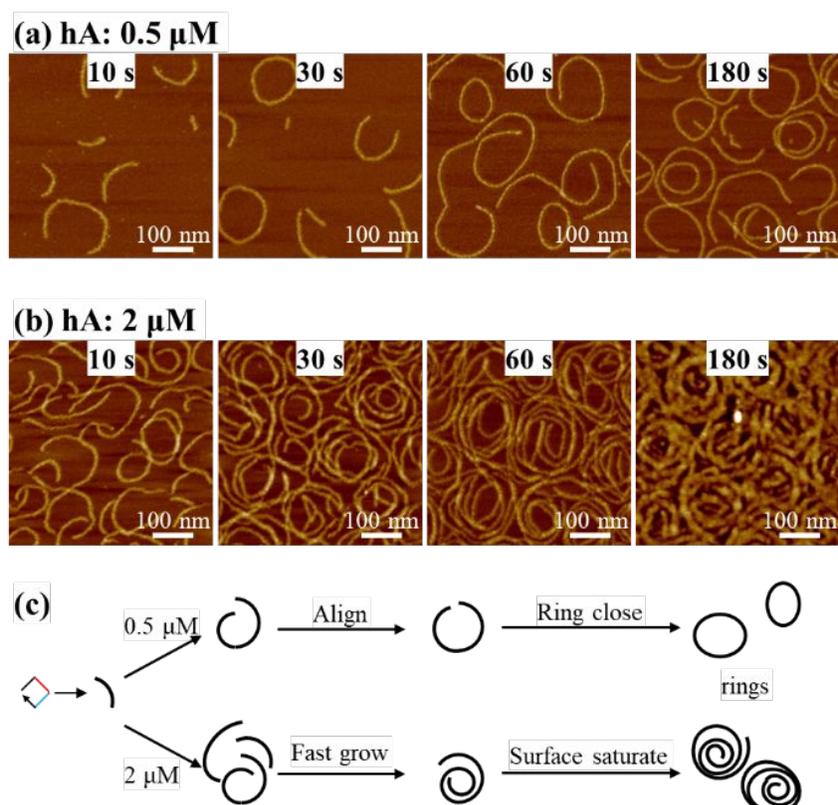


**Figure 3.** The bending angle of curved DNA structure with different modifications. (a) Schematic drawing of bending angle calculation from single closed circles. (b) The calculated bending angles motifs from different DNA strands.

To understand the driving force of the flexible bending in the homopolymer, we measured the average bending angle between two adjacent DXL motifs for all curved homopolymers (Figure 3). In a closed DNA homopolymer circle, the sum of all the bending angles ( $\theta$ ) was  $360^\circ$ . The number of DXL motifs in the circle could be calculated by the circle contour length ( $L$ ) divided by the motif length ( $d$ ), which is 10 base pairs, or 3.3 nm assuming the rise of a DNA duplex is 0.33 nm/base pair. Thus, the bending angle ( $\theta$ ) could be calculated as:

$$\theta = 360^\circ / (L/d) = 360^\circ / (L/3.3 \text{ nm})$$

By tuning the incubation conditions, we prepared poly(**hX**) as single circles for all curved designs and imaged them by AFM (Figure S6-S10). Note that no conformational constraints were imposed on inter-motif interactions during the cyclization of the single circles. We used TopoStats<sup>21</sup>, an imaging processing software, to filter AFM images and measure contour lengths of the circles (, Figure 3b, Supporting information). For **hT**, the average bending angle between motifs was  $2.42 \pm 0.37^\circ$ , while the angles for **hA** and **hG** were  $3.35 \pm 0.60^\circ$  and  $3.56 \pm 3.81^\circ$ , respectively. This phenomenon is because purines have a larger aromatic plane than pyrimidines, thus can more effectively stack into a DNA duplex and cause the array to bend. For **hAA**, the bending angle further increased to  $5.14 \pm 1.22^\circ$  due to the insertion of two bases (compared with one base) into a duplex which would introduce a larger length mismatch (Figure 1b). Interestingly, the bending effect was not limited to natural DNA bases. Other chemical moieties with a flat, aromatic plane could stack into DNA duplex and induce arrays to bend as well.<sup>22, 23</sup> When a fluorescein was incorporated at the 3' end (strand **hF**), an average bending angle of  $4.28 \pm 1.01^\circ$  was observed.



**Figure 4.** The assembly kinetics of curved structures under different DNA concentrations exemplified by **hA**. AFM images of curved structure formation at DNA concentration of (a) 0.5 μM and (b) 2 μM (c) Proposed schemes of assembly under thermodynamic or kinetic control.

We further conducted kinetic studies to understand the effects of inter-motif flexibility and assembly conditions on the final morphologies of the DNA homopolymers (Figure 4 and S11-S16). To capture intermediate states at specific time-points during assembly, DNA samples were rapidly dried and immediately AFM imaged in air. Such a protocol prevented the DNA samples from any further change that could be associated with aqueous AFM imaging. At a high DNA concentration (2.0  $\mu\text{M}$ ), **hX** molecules quickly adsorbed onto the mica surface and grew fast, continuous 1D arrays adjacent to existing 1D arrays (Figure 4a), which initially formed single rings and long arcs at 10 s and then transformed spirals or concentric ring structures after 30 s. At a low DNA concentration (0.5  $\mu\text{M}$ ), DNA molecules slowly adsorbed onto the mica surface and more distinct circles formed (Figure 4b). Under this concentration, the DNA ladder formed single rings at 30 s and further grew into  $\sim 50\%$  single rings and  $\sim 50\%$  two-layer concentric rings, indicating the continuous DNA deposition. Generally, the DNA molecules adsorbed onto the mica surface and oligomerized on the substrate with surface templating, and simultaneously self-aligned or joined with other oligomers. The final morphology was determined by the kinetic competition between adsorption (thus the effective surface DNA concentration) and refiguration (circle-closing alignment) (Figure 4c). For the slow adsorption at a low DNA concentration, the homopolymer had sufficient time to align itself in an intra-polymer fashion into a closed circle. The random stacking in/flipping out of X rendered a range of circle size distribution. In contrast, the fast adsorption at a high DNA concentration saturated the surface. Thus, a large number of polymers would simultaneously form and align in an inter-polymer fashion to form concentric rings or continuous spirals to maximize coverage of the mica surface. The flexibility of X stacking in/flipping out allowed the motifs flexibility to adopt the necessary curvature to form those final morphologies. According to native PAGE (Fig. S2), the sticky ends were weaker for **hT**, **hAA** and **hF**. It explained the slow growth rate of corresponding DNA homopolymers (Figure S14-16). Furthermore, inter-motif assembly is a function of DNA concentration. Thus, the formation of circular structures took longer time at 0.5  $\mu\text{M}$  DNA concentration than at 2  $\mu\text{M}$  DNA concentration. However, if the incubation was through the thermodynamically, slow annealing process with a high DNA concentration, the final morphology turned into only concentric circles. This demonstrated that the circle-close alignment would minimize the system's free energy (Figure S17). With the inflexible chemical groups, only the semi-straight fibers' density change was observed during the changes in time and DNA concentration.

In summary, this study provides a novel and straightforward strategy to introduce the dynamic flexibility to DNA homopolymers and bend the DNA nanostructures by adding a chemical modification (natural bases or other aromatic moieties) at ends of DNA strands. This dynamic bending allows for the assembly of DNA nanostructures with constantly changing curvatures from single component strands. Such capability was not accessible previously and could be achieved by a large number of DNA components.<sup>16, 24,</sup>

<sup>25</sup> This study has expanded the toolbox for structural DNA nanotechnology in two aspects. In large and complicated nanostructures, such as DNA origami, the extra base could be applied to weaken the connection strength or to fine tune the partial curvature. Additionally, this strategy could enable the flexible and multiple curvature in DNA tile-based design with limited DNA components. Conceivably, the dynamic flexibility of DNA homopolymers can be used as soft or elastic building unit for nanomachines and nanorobots.<sup>26-32</sup>

**Acknowledgement:** We thank NSF for financial support this work.

**Reference:**

1. Nadrian C. Seeman and Hanadi F. Sleiman. DNA nanotechnology. *Nat. Rev. Mater.* **3**, 17068 (2017).
2. Constantine G. Evans and Erik Winfree. Physical principles for DNA tile self-assembly. *Chem. Soc. Rev.* **46**, 3808–3829 (2017).
3. Matthew R. Jones, Nadrian C. Seeman and Chad A. Mirkin. Programmable materials and the nature of the DNA bond. *Science*. **347**, 1260901 (2015).
4. Andre V. Pinheiro, Dongran Han, William M. Shih and Hao Yan. Challenges and opportunities for structural DNA nanotechnology. *Nat. Nanotechnol.* **6**, 763–772 (2011).
5. Fei Zhang, Jeanette Nangreave, Yan Liu and Hao Yan. Structural DNA nanotechnology: State of the art and future perspective. *J. Am. Chem. Soc.* **136**, 11198–11211 (2014).
6. Qinqin Hu, Hua Li, Lihua Wang, Hongzhou Gu and Chunhai Fan. DNA Nanotechnology-Enabled Drug Delivery Systems. *Chem. Rev.* **119**, 6459–6506 (2019).
7. Erik Winfree, Furong Liu, Lisa A Wenzler and Nadrian C Seeman. Design and self-assembly of two-dimensional DNA crystals. *Nature*. **394**, 539–544 (1998).
8. Paul W.K. Rothemund. Folding DNA to create nanoscale shapes and patterns. *Nature*. **440**, 297–302 (2006).
9. Yonggang Ke, Luvena L. Ong, William M. Shih and Peng Yin. Three-dimensional structures self-assembled from DNA bricks. *Science*. **338**, 1177–1183 (2012).
10. Mengxi Zheng, Zhe Li, Longfei Liu, Mo Li, Victoria E. Paluzzi, Jong Hyun Choi and Chengde Mao. Kinetic DNA Self-Assembly: Simultaneously Co-folding Complementary DNA Strands into Identical Nanostructures. *J. Am. Chem. Soc.* **143**, 20363–20367 (2021).
11. Carlos E. Castro, Hai Jun Su, Alexander E. Marras, Lifeng Zhou and Joshua Johnson. Mechanical design of DNA nanostructures. *Nanoscale*. **7**, 5913–5921 (2015).
12. Yang Yang, Jing Wang, Hideki Shigematsu, Weiming Xu, William M. Shih, James E. Rothman and Chenxiang Lin. Self-assembly of size-controlled liposomes on DNA nanotemplates. *Nat. Chem.* **8**, 476–483 (2016).

13. Henri G. Franquelim, Alena Khmelinskaia, Jean Philippe Sobczak, Hendrik Dietz and Petra Schwille. Membrane sculpting by curved DNA origami scaffolds. *Nat. Commun.* **9**, 811 (2018).
14. Constantine G. Evans and Erik Winfree. Physical principles for DNA tile self-assembly. *Chem. Soc. Rev.* **46**, 3808-3829 (2017).
15. Sungjin Lee, Suyoun Park, Muhammad Tayyab Raza, Thi Bich Ngoc Nguyen, Thi Hong Nhung Vu, Anshula Tandon and Sung Ha Park. Multirule-Combined Algorithmic Assembly Demonstrated by DNA Tiles. *ACS Appl. Polym. Mater.* **4**, 5441–5448 (2022).
16. Hendrik Dietz, Shawn M. Douglas and William M. Shih. Folding DNA into twisted and curved nanoscale shapes. *Science.* **325**, 725–730 (2009).
17. Mengxi Zheng, Qian Li, Qian Li, Victoria E. Paluzzi, Jong Hyun Choi and Chengde Mao. Engineering the Nanoscaled Morphologies of Linear DNA Homopolymers. *Macromol. Rapid Commun.* **42**, 2100217 (2021).
18. R. Erik Holmlin, Peter J. Dandliker and Jacqueline K. Barton. Charge Transfer through the DNA Base Stack. *Angew. Chem. Int. Ed.* **36**, 2714–2730 (1997).
19. M. Bezanilla, B. Drake, E. Nudler, M. Kashlev, P. K. Hansma and H. G. Hansma. Motion and enzymatic degradation of DNA in the atomic force microscope. *Biophys. J.* **67**, 2454–2459 (1994).
20. Longfei Liu, Yulin Li, Yong Wang, Jianwei Zheng and Chengde Mao. Regulating DNA Self-assembly by DNA–Surface Interactions. *ChemBioChem.* **18**, 2404–2407 (2017).
21. Joseph G. Beton, Robert Moorehead, Luzie Helfmann, Robert Gray, Bart W. Hoogenboom, Agnel Praveen Joseph, Maya Topf and Alice L.B. Pyne. TopoStats – A program for automated tracing of biomolecules from AFM images. *Methods.* **193**, 68–79 (2021).
22. Xiang D. Liu, Hong Y. Diao and Norio Nishi. Applied chemistry of natural DNA. *Chem. Soc. Rev.* **37**, 2745–2757 (2008).
23. Amparo Ruiz-Carretero, Pim G.A. Janssen, Adrien Kaeser and Albertus P.H.J. Schenning. DNA-templated assembly of dyes and extended  $\pi$ -conjugated systems. *Chem. Commun.* **47**, 4340–4347 (2011).
24. Dongran Han, Suchetan Pal, Jeanette Nangreave, Zhengtao Deng, Yan Liu and Hao Yan. DNA origami with complex curvatures in three-dimensional space. *Science.* **332**, 342–346 (2011).
25. Keyao Pan, Do Nyun Kim, Fei Zhang, Matthew R. Adendorff, Hao Yan and Mark Bathe. Lattice-free prediction of three-dimensional structure of programmed DNA assemblies. *Nat. Commun.* **5**, 5578 (2014).
26. Jonathan Bath and Andrew J. Turberfield. DNA nanomachines. *Nat. Nanotechnol.* **2**, 275–284 (2007).
27. Huajie Liu and Dongsheng Liu. DNA nanomachines and their functional evolution. *Chem. Commun.*, 2625–2636 (2009).

28. Jiang Li, Chunhai Fan, Hao Pei, Jiye Shi and Qing Huang. Smart drug delivery nanocarriers with self-assembled DNA nanostructures. *Adv. Mater.* **25**, 4386–4396 (2013).
29. Hyungmok Joh and Donglei Emma Fan. Materials and Schemes of Multimodal Reconfigurable Micro/Nanomachines and Robots: Review and Perspective. *Adv. Mater.* **33**, 2101965 (2021).
30. Jong Bum Lee, Songming Peng, Dayong Yang, Young Hoon Roh, Hisakage Funabashi, Nokyoung Park, Edward J. Rice, Liwei Chen, Rong Long, Mingming Wu and Dan Luo. A mechanical metamaterial made from a DNA hydrogel. *Nat. Nanotechnol.* **7**, 816–820 (2012).
31. Shogo Hamada, Kenneth G. Yancey, Yehudah Pardo, Mingzhe Gan, Max Vanatta, Duo An, Yue Hu, Thomas L. Derrien, Roanna Ruiz, Peifeng Liu, Jenny Sabin and Dan Luo. Dynamic DNA material with emergent locomotion behavior powered by artificial metabolism. *Sci. Robot.* **4**, eaaw3512 (2019).
32. Yuanyuan Guo, Qiushuang Zhang, Qiwen Zhu, Jing Gao, Xinyuan Zhu, Haijun Yu, Yuehua Li, Chuan Zhang. Copackaging photosensitizer and PD-L1 siRNA in a nucleic acid nanogel for synergistic cancer photoimmunotherapy. *Sci. Adv.* **8**, eabn2941 (2022).