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Changes of labile, stable and water-soluble fractions of biochar after two years in a vineyard soil†

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The long-term stability of biochar in soil is a key factor for carbon storage and agro-environmental benefits in soil applications. Therefore, to better use biochar as both a sustainable soil organic amendment and a carbon sequestration technique, it is necessary to understand the biochar aging process in soil, as biochar characteristics can be altered by a variety of processes that occur in the environment. Reliable chemical characterization can be obtained through thermoanalytical techniques, which are known to provide a trustworthy estimation of the biochar stability in soil. In the present study, the changes of labile and stable fractions of biochar after two years of application in a vineyard field were quantified before and after the amendment. Aged biochar was manually isolated from the soil-biochar mixture, and its chemical properties were measured by elemental analysis, thermogravimetric analysis (TGA) and analytical pyrolysis (Py-GC-MS). Dissolved organic matter (DOM) was analyzed by elemental and UV-vis spectroscopy analysis. A slight decrease of carbon content ($-13 \pm 5\%$) after two years was associated with a significant drop of TGA labile (from 26 to 9%) and water-soluble carbon (from 3.1 to 0.8 mg g⁻¹), concomitantly, the pyrolytic indices of aromaticity (e.g. toluene/naphthalene ratio) decreased. These results contrasted with the increase in the H/C and O/C ratios as stability indexes. The TGA stable fraction remained almost unaltered (49 vs. 52%) confirming the consistency of this index for recalcitrance. The results from Py-GC-MS and TGA techniques are consistent in highlighting significant changes in the biochar labile fraction due to the aging process. Instead, no changes were found in the stable fraction between the aged biochar and the fresh one, with important consequences on carbon cycling and soil sequestration.

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Environmental significance

Biochar is proposed as an innovative and sustainable tool to mitigate different environmental issues. Research has demonstrated that biochar could be applied for carbon sequestration, soil amelioration, greenhouse gas emissions reduction and nutrient leaching decrement, as well as waste management. The evaluation of the real environmental benefits of using biochar requires a deep knowledge of how biochar chemistry is affected by the variety of time-dependent processes that occur in the environment, which are termed “aging”. The present study provides new insights related to the effects of aging process on the biochar properties applied to agricultural soils, with regard, particularly, to the quantity and quality characteristics of labile and recalcitrant fractions. It provides an estimation of the biochar stability in soil and on changing the characteristics of biochar applied to soil systems, increasing the knowledge on the potential aging impacts on the physicochemical properties of biochar and on its connected environmental implications.

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Introduction

Biochar is a carbonaceous solid material obtained from the thermochemical conversion of biomass in an oxygen-limited environment. It has received much scientific interest due to its potential as a sustainable tool to mitigate different environmental issues. Many studies have investigated the possibility of using innovative materials such as compost, zeolites, biochar and engineered nanomaterials, in a green and circular economy loop, to improve soil fertility while providing agro-



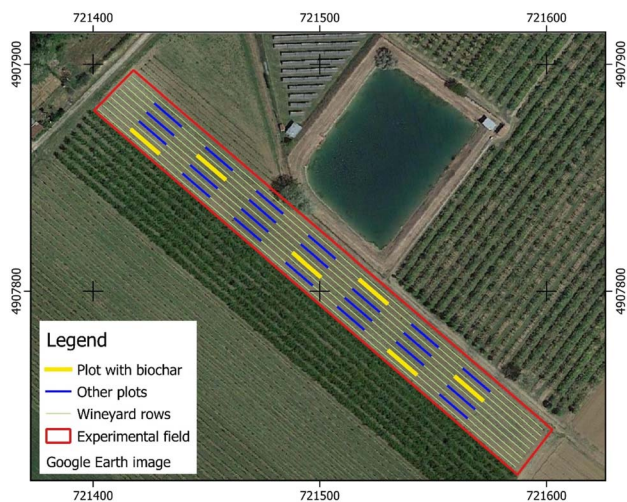


Fig. 1 Experimental field with plot location arranged on a basis of a randomized block design. Treated parcels on the same row are separated by an untreated parcel as a buffer to limit the cross-contamination during agronomical operations. Rows with treated parcels are separated by an untreated row. Each parcel consists of 15 vine plants. The yellow colour indicates the sampled parcels.

separated from the soil by the method described in Dong *et al.*,²⁰ without rinsing the samples with deionized water to remove adhering soil particles. Specifically, aged biochar particles with a diameter >0.5 mm from soil samples were separated by forceps until no visible biochar particles were present in soil samples. Then, as reported in Wang *et al.*,³⁹ each collected biochar particle was cleaned by removing the apparent soil debris from its surface. Fresh biochar particles with a diameter >0.5 mm were collected from fresh biochar samples to compare the characteristics of fresh and aged biochar. Both fresh and aged biochars were dried at 40 °C, sieved (mesh size: 2 mm), and stored at -18 °C.

Soil properties

Soil at the site is a Eutric Cambisol (Loamic)⁴⁰ containing 31% sand, 47% silt, and 22% clay with a pH of 7.6 (1 : 2.5 soil : water), 1.07% total organic carbon, 0.15% total nitrogen, 25.6 $\text{cmol}_{(+)} \text{kg}^{-1}$ cation exchange capacity, slope $<2\%$. Since 2021 soil analysis are yearly performed following the legislative procedure.⁴¹ In 2021 the sample collection for the soil analysis coincided with the sample for biochar extraction.

Biochar production and characterization

The applied biochar was one of the few available biochar in the market as of 2019, with an annual industrial production rate exceeding 100 tonnes. It was compliant with Italian legislation for soil improvement, and it was listed as an approved soil amendment since 2015. The methodology employed for biochar production is of an artisanal nature and has been documented in previous studies.^{5,42,43} In brief, the vineyard pruning residues underwent a conversion process into charcoal through a slow pyrolysis procedure conducted at the “Romagna Carbone s.n.c.”

production facility in Italy. The batch pyrolysis process occurred within a transportable cast iron ring kiln with a diameter of 2.2 meters, capable of holding approximately 2 tons of dry feedstock. Following manual ignition of the combustion, the furnace was sealed at the top with a lid, and progressive combustion ensued, consuming all available oxygen, and reaching its maximum temperature in less than 20 minutes. A measured peak temperature of 700 °C was maintained for 3–4 hours, with the average heating rate of 25–30 °C min^{-1} . The pyrolysis process concluded autonomously, and the subsequent day, the biochar underwent sieving and separation based on size.

Biochar samples were oven-dried at 40 °C for 72 h and homogenized using an agate mortar and pestle and stored at -20 °C prior to analysis. Proximate analysis of biomass and biochar was carried out according to ASTM D7582 method with slight modification.³⁴ The method determines relative moisture (M), volatile matter (VM), fixed carbon (FC) content during different analysis steps. For each analysis about 0.5 g of biochar sample was introduced in aluminum oxide crucible (volume of 20 mL) and covered with the specific lid. Then the crucible was inserted in the thermogravimetric analyzer (Eltra Thermostep, Eltra GmbH, Haan, Germany).

Elemental analysis was performed using a Thermo Scientific FLASH 2000 Series CHNS/O Elemental Analyzer (Thermo Fisher Scientific, Waltham, U.S.A.) to determine C, H, and N contents of the biochar samples. The content of carbonate and ash was determined in each biochar as previously reported by Rombolà *et al.*⁴⁴ Measurements were performed in triplicate, and oxygen (O) content was determined by difference: $\text{O} (\%) = 100 - \text{C} (\%) - \text{H} (\%) - \text{N} (\%) - \text{Ash} (\%)$. Moisture contents were determined (ASTM D-3173) at 105 °C. A mixture of biochar with deionized water at 1 : 10 wt/wt ratio was prepared, thoroughly mixed and pH measured at room temperature with a digital pH meter (VWR pH 100, VWR International).

Characterization of biochar labile and stable fraction

The parameters of the thermally labile fraction were determined by thermogravimetric analysis (recalcitrant OM, T 390–600 °C) and Py-GC-MS (molecular ratios).

Analytical pyrolysis. Analytical pyrolysis was performed using a multi-shot pyrolyzer (EGA/PY-3030D, Frontier Laboratories Ltd, Japan) interfaced to a gas chromatograph (GC) coupled with mass spectrometer (MS) (7890B GC and 5977B MS, Agilent Technologies, USA). A small crucible capsule containing weighed biochar sample (*ca.* 10 mg) was introduced into the furnace and then pyrolyzed at 700 °C for 100 s using helium as carrier gas (1 mL min^{-1}) and an interface temperature of 280 °C. Pyrolysis products were introduced into the GC fused silica capillary column (HP-5MS, stationary phase 5%-diphenyl, 95%-dimethylpolysiloxane 30 m \times 0.25 mm i.d. \times 0.25 μm film thickness, Agilent Technologies, USA) with a 20 : 1 ratio. The injector temperature was 280 °C and the thermal program used was: 50 °C to 310 °C at 10 °C min^{-1} , then a hold for 5 min at 310 °C. The MS operated in electron ionisation at 70 eV with scans in the 29–600 m/z



range. The temperatures of the ion source and the transfer line were 230 and 250 °C, respectively. The quantity of evolved pyrolysis products (Py-yield) was calculated by dividing the GC peak area by sample amount (area per μg) as reported in Conti *et al.*³⁴

Thermogravimetric analyses. Thermogravimetric analysis was conducted by Eltra Thermostep thermogravimetric analyzer (Eltra GmbH, Haan, Germany) under an oxidant atmosphere (air, 90 mL min^{-1}) at 10 °C min^{-1} to 600 °C for organic material (OM) determination, then they were ramped at 25 °C min^{-1} to 850 °C for carbonate determination.⁴⁵ A sample mass of 0.5 ± 0.1 g in a 20 mL alumina crucible was used for each experiment and three replications were performed to test the reproducibility of the results. Specifically, the TGA method allows to determine mass loss relative to labile OM (220–390 °C), recalcitrant OM (390–600 °C) and carbonates (600–800 °C) fraction.⁴⁴ The mass loss (TG) and derivative curves (DTG) of the samples were represented as a function of temperature.

Biochar-derived DOM extraction and analysis

Dissolved Organic Matter (DOM) was extracted from the fresh and aged biochar according to method reported in Ghidotti *et al.*,⁴⁶ and in Rombolà *et al.*⁴⁴ Dissolved organic carbon (DOC) and dissolved total nitrogen (DTN) in each DOM extracted from fresh and aged biochar were determined with a total organic carbon analyzer (TOC-L, Shimadzu, Japan) coupled to a TN module (TNM-L, Shimadzu, Japan). DOC and DTN in fresh and aged biochar were analyzed in triplicate, and their standard deviations are presented in Table 5.

The concentration of water-soluble organic carbon (WSOC) ($\text{mg g}^{-1} \text{TOC}$) in fresh and aged biochar was determined as the proportion of biochar OC pool extracted by the water extraction method. In particular, as reported in Rombolà *et al.*,⁴⁴ the values were calculated using the following eqn (1):

$$\text{WSOC} \left(\frac{\text{mg DOC}}{\text{g SOC}} \right) = \frac{V \text{ (L)} \times C \left(\frac{\text{mg}}{\text{L}} \right)}{\text{TOC} \left(\frac{\text{g}}{\text{kg}} \right) \times M \text{ (kg)}} \quad (1)$$

Spectroscopic characterization. The biochar-derived DOM was characterized using UV-visible spectroscopy. Absorbance was measured using a Cary 300 UV-visible spectrophotometer (Agilent Technologies, USA) within a spectrum of 250–800 nm, at 1 nm increments, using a 1 cm quartz cuvette. Ultrapure water (18.2 M Ω cm) was used as a reference.

The specific UV absorbance at 254 nm (SUVA₂₅₄, L $\text{mg}^{-1} \text{m}^{-1}$), a proxy for DOM aromaticity, was calculated as follows: $\text{SUVA}_{254} = a_{254}/\text{DOC}$. The a_{254} indexes is the absorption coefficient at wavelength 254 nm and was calculated using the eqn (2).

$$a_{254} \text{ (m}^{-1}\text{)} = \frac{\text{UV}_{254} \times 2.303}{l \text{ (m)}} \quad (2)$$

where UV_{254} is the absorbance at 254 nm, 2.303 is the transform coefficient, and l the path length of the cuvette in meters.⁴⁷

Statistical analysis

Quantitative data of fresh and aged biochar used to compare results are presented as the mean and standard deviation of three replicates. Data were analyzed using an analysis of variance (ANOVA) procedure for independent samples followed by Tukey's post hoc tests using R Statistical Software (Ver. 4.2.2, 2022, The R Foundation for Statistical Computing). The different properties between fresh and aged biochar were analyzed by t -test ($p < 0.05$).

Results and discussion

Soil modification due to biochar amendments

After two years of experimentation, the soil properties were slightly modified by the biochar amendment. In Table 1 the main properties of control and biochar-amended soil are resumed.

The soil pH has been slightly increased by the biochar amendment (from 7.77 to 7.93), even if the high standard deviation in the control parcels highlights the heterogeneity of the soil. The addition of biochar to soil results in a weak increase in EC and a larger increase in TOC and in available P and K, while it does not modify TN content. These modifications are perfectly aligned with outcomes from Schmidt *et al.*,⁴⁸ as a result of global meta-analysis on biochar effects on agricultural soils.

Effect of aging on biochar properties

Elemental composition and basic features of biochar with aging at field. The biochar used in the experiment for soil amendment had a total content of C, N, H and O 80%, 1.7%, 0.5%, 2.5% (dry weight), respectively; 15.2% ash and pH 9.96. The molar H/C and O/C ratios of 0.25 and 0.02, respectively, indicate a high degree of carbonization and aromaticity.^{24,49,50}

The C content decreased from $80 \pm 1.9\%$ in the fresh to an average of $67 \pm 3.3\%$ with aging (Table 2). Therefore, through the aging process, biochar C content significantly decreased by $13 \pm 5\%$ ($p < 0.05$). Such effect can be due to decomposition of labile C through abiotic and biotic processes⁵¹ that take place in the environment, including abiotic and biotic redox reactions, solubilization and interactions with microbes, OM, minerals and solutes in the soil environment.⁵²

Microorganisms likely played an important role in the biochar aging process. As an example, the C lost from abiotic laboratory incubations of biochar was 50–90% less than that of microbially inoculated incubations.¹⁶ Moreover, a fraction of biochar-derived C was likely lost through leaching of DOC. Mukherjee and Zimmerman⁵³ measured a significantly high amount of DOC in the soil leachate of biochar amended soils. When in soil environments biochars are exposed to leaching often coupled with mild acid or base hydrolysis.

However, the $13 \pm 5.2\%$ reduction of total C concentration in aged biochar after two years soil incubation appears comparable to mean averages reported in literature.^{19,54,55} In particular, Sorrenti *et al.*,¹⁹ reported losses of total C by 14.5% for aged biochar in a 4 year field experiment, which was partly attributed



Table 1 Main soil properties for biochar amended and control soils, mean values and standard deviation in parentheses ($n = 3$). Data refer to 2021 sample campaign

Sample	H ₂ O reaction	EC 1 : 2.5	TOC	TN	Available P	Available K
	pH	dS m ⁻¹	%	%	mg kg ⁻¹	mg kg ⁻¹
Biochar amended soil	7.93 (0.13)	0.33 (0.09)	1.51 (0.06)	0.15 (0.015)	17.6 (2.7)	140.4 (48.0)
Control soil	7.77 (0.32)	0.29 (0.10)	1.07 (0.02)	0.15 (0.002)	15.6 (3.2)	106.3 (43.9)

Table 2 Proximate and ultimate analysis (oxygen by difference), molar ratios and pH of fresh and aged biochar applied in the field experimental. Mean values \pm s.d. ($n = 3$)

Parameters	Units	Fresh biochar	Aged biochar
TC	%	80 \pm 1.9	67 \pm 3.3
TOC	%	79 \pm 1.9	66 \pm 3.4
TIC	%	0.70 \pm 0.04	0.29 \pm 0.08
N	%	1.66 \pm 0.11	1.77 \pm 0.08
H	%	0.50 \pm 0.01	0.94 \pm 0.08
O	%	2.5 \pm 1.5	10.6 \pm 2.1
Ash	%	15.2 \pm 0.4	20.1 \pm 1.5
H/C	Atomic	0.25	0.32
O/C	Atomic	0.02	0.11
VM	%	26 \pm 0.5	20.5 \pm 1.4
FC	%	57 \pm 0.9	61 \pm 1.6
pH	—	9.96 \pm 0.07	7.50 \pm 0.07

to mineralization. The impacts of field aging on biochar were evidenced by Spokas *et al.*,⁵⁴ who observed that in the pre and post weathered wood-derived biochars (~3 years after application) there was a ~16% loss of C, dropping from 78.3 to 62.5% carbon. However, this difference in C content could be related not solely to weathering losses but also to the trapped soil in the pores of post-weathering biochar samples and the formation of an organic coating on aged biochar. The formation of an organic coating on biochar was identified in soil-aged biochars that strengthens biochar–water interactions and thus enhances nutrient retention.^{56,57} This could imply a reduction of total C in the aged biochar due to increased uptake of mineral and organic compounds into the pore system or *via* organomineral coating in the aged biochar. Also noteworthy is the observation that all weathered biochar samples possessed higher analytical variability (increase in standard deviations; Table 2). Previous studies reported that one of the causes of this variability can be the presence of entrapped soil into the biochar pores.^{54,55}

The decrease in C content is accompanied by a slight increase of ash, H and N contents, whereas O content of aged biochar (13.7%) was significantly higher than fresh biochar (2.5%). Similar increase of O was found by Campos *et al.*,⁵⁸ which detected an increase of aryl C and O-containing functional groups after 2 year of soil application. Increase of O content can be due to abiotic or biotic oxidation, with a number of studies suggesting that abiotic oxidation plays the dominant role.^{16,59,60} Abiotic biochar oxidation has been observed to occur through various processes. Atmospheric oxygen-induced oxidation can introduce additional oxygen-containing functional groups, such as hydroxyl, carbonyl, and carboxyl, to the

biochar surface. This mild oxidation process is slow at ambient temperature.⁶¹ Rainfall events can also result in biochar oxidation owing to the dissolved oxygen and nitrogen oxides in rainwater.^{62,63} Rainfall events can also lead to physical disintegration and acidification, thus causing labile carbon to be released as DOM, minerals to leach out and additional oxygen-containing functional groups, such as hydroxyl, carbonyl, and carboxyl, to be introduced to the biochar surface.

Concerning the N content, numerous studies have reported increases in organic and inorganic N content of aged biochar.^{14,19,54} However, in a 2 year field experiment, de la Rosa *et al.*,⁵⁵ reported a decrease in N content for not wood-derived aged biochar. The impact of aging has been hypothesized as a critical factor for the interaction of biochar with the plant and soil systems, particularly for the sorption of nitrogen containing compounds.^{55,64} A relative enrichment could be due to contributions of dead vegetation, which were incorporated into the pores or adsorbed to the surface of the biochar.⁵⁵ Similarly, previous studies reported that H content of aged biochar was significantly higher than fresh biochar.^{20,54}

The molar H/C and O/C ratios of 0.25 and 0.02, respectively, indicate a lower degree of carbonization and aromaticity of aged biochar.^{24,50} As a result of weathering, the aged biochar had a significant alteration in the H/C and O/C ratios, which increased from 0.25 to 0.32 and from 0.02 to 0.11, respectively. After incubation in soil for 240 days, Huang *et al.*,⁶⁵ reported that biochar of rice husk increased H/C and O/C ratios from 0.46 to 0.50 and from 0.26 to 0.33, respectively. Joseph *et al.*,¹⁴ observed a decrease in the total C of biochar samples (poultry waste and green waste biochar) and an increase in the oxygen content leading to an alteration in the O/C ratio from <0.2 to ~0.75 after 2 years of field weathering. In the present study, the increase in the O/C ratio for aged biochar was not as large as the alteration documented in Joseph *et al.*,¹⁴ but it is the same as the increase (from 0.02 to 0.11) reported by Spokas.⁵⁴ Both ratios reflect physical–chemical properties of biochar related to stability, as the proportion of elemental compounds (H and O) relative to C present in biochar, and the molar ratio of H/C and of both ratios have been adopted by IBI and EBC, respectively. The present study showed that after the aging period in soil results of biochar molar H/C and O/C ratios are inconsistent with those obtained by thermoanalytical techniques as discussed in the next section.

The pH value of fresh biochar through the aging process decreased from 9.96 to 7.50 and it is in accordance with the results of several studies.^{14,19,54} These studies have reported that the pH values of the biochars decreased with an aging period in



the field as a consequence of the field weathering. Weathering typically results in the increased abundance of carbonyl, carboxylic, and phenolic functional groups on the biochar surface,^{58,66,67} which leads to a pH decrease in aged biochar.⁶⁷ This suggests that biochar liming potential is limited to a few years after its application. Hence, biochar-induced benefits in nutrient availability in acid soils may be more pronounced in the first seasons following application.

Characterization of fresh and aged biochar by analytical pyrolysis. Examples of TIC pyrograms related to the fresh and aged biochars obtained at 600 °C are depicted in Fig. 2. The results of Py-GC-MS analysis were used to estimate of thermal stability of fresh and aged biochars.⁶⁸ In particular, the degree of thermal stability was estimated using specific molecular ratios (toluene/benzene (T/B), toluene/naphthalene (T/NAP), methylnaphthalene/naphthalene (MeNAP/NAP) and methylphenanthrene/phenanthrene (MePHE/PHE)). The degree of alkylation detected by Py-GC-MS is widely regarded as a molecular index of biochar thermal stability.^{30,34,36,38,69,70} Kaal *et al.*,³⁶ proposed that the decrease in T/B and MeNAP/NAP ratios can be representative of dealkylation associated with polycondensation and aromatization that increase with charring. The validity of T/NAP and MePHE/PHE molecular ratio as proxies of charring extent was confirmed in previous research.³⁴ These molecular indices were applied to characterize fresh and aged biochars. Results reported in Table 3 showed that the ratios were lower for the aged biochar in accordance with a decrease in the relative abundance of O-alkyl C and alkyl C.^{55,68,71} As reported in Table 3, Py-yields (area per μg) decreased from $390\,000 \pm 20\,000$ to $170\,000 \pm 48\,000$ showing effects of aging in soil on the thermally labile fraction of aged biochar. The changes in biochar composition with age can be summarized as a reduced diversity of products, a decrease in markers

of non/weakly-charred biomass and a relative increase in the products of relatively condensed black carbon. These results suggest that, unsurprisingly, the more condensed aromatic compounds of biochar are selectively preserved during long-term storage in soil.

Table 3 shows that the variability of molecular ratios between fresh and aged biochar was much greater (45% MePHE/PHE, 38% MeNAP/NAP, 39% T/NAP, 48% T/B) than that of TOC (13%), VM (6%) and labile OM (17%). Therefore, the molecular ratios may explain not only the mass loss or C loss alone during field aging but also the aging process that induces primary changes in the aromatic structure of biochar.

Good correlations between Py-GC-MS results and ratio of H/C were found in several research.^{24,34} In the present study, however, the depicted Py-GC-MS derived stability molecular ratios are not aligned with molar H/C and O/C ratios for fresh and aged biochars. The molar H/C and O/C ratios of 0.25 and 0.02 for fresh biochar and of 0.32 and 0.11 for aged biochar, respectively, indicate a greater degree of aromaticity and stability of fresh biochar than aged. Nevertheless, the degree of thermal stability estimated by Py-GC-MS using specific molecular ratios suggested that in aged biochar, through the aging process, the labile fraction significantly decreases. As discussed above, the data reported in this study suggest that molar H/C and O/C ratios do not seem to efficiently distinguish between the biochar with a higher level of labile C fraction such as that fresh from the aged biochar with a lower level of labile C fraction.

The data obtained by Py-GC-MS are coupled with the quantitative information on the weight loss with temperature obtained by TGA.

Thermal characterisation by TGA. TG and DTG curves obtained from fresh and aged biochar are shown in Fig. SM2 and

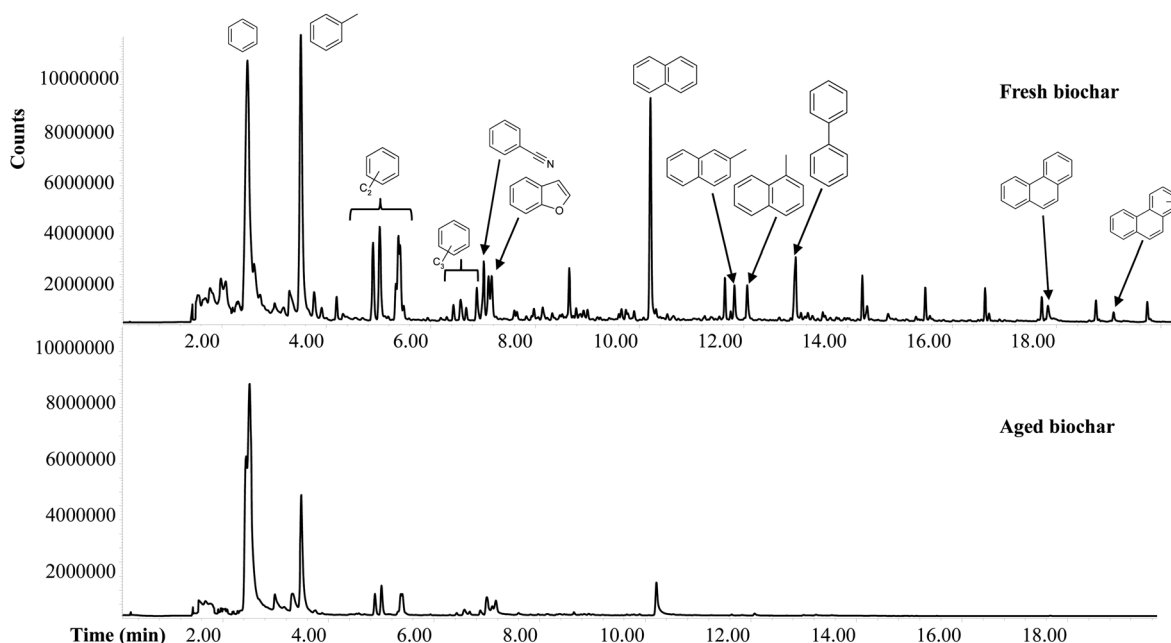


Fig. 2 Total ion chromatograms from Py-GC-MS at 600 °C of fresh and aged biochar.



Table 3 Yields of pyrolysis products (peak area per μg) and molecular ratios MePHE/PHE (methylphenanthrene/phenanthrene), methylnaphthalene/naphthalene (MeNAP/NAP) toluene/naphthalene (T/NAP) and toluene/benzene (T/B) from Py-GC-MS at 600 °C of fresh and aged biochar. Mean values \pm s.d. ($n = 3$)

Samples	MePHE/PHE	MeNAP/NAP	T/NAP	T/B	Py-yield
Fresh biochar	0.256 \pm 0.002	0.26 \pm 0.04	1.8 \pm 0.2	0.5 \pm 0.1	390 000 \pm 20 000
Aged biochar	0.14 \pm 0.01	0.16 \pm 0.03	1.1 \pm 0.3	0.26 \pm 0.03	170 000 \pm 48 000

SM3.† As reported in previous works, the OM decomposition takes place into two main temperature intervals associated to labile (220–390 °C) and recalcitrant (390–600 °C) components.^{44,45,72} A third mass loss in the range 500–650 °C, associated to carbonate fraction was also proposed.⁴⁵ Results of thermal analysis by TGA showed that labile OM of fresh biochar was 26.1 \pm 0.6%, and it decreased to 9.1 \pm 0.3% with aging (Table 4). Moreover, as reported in Table 4, recalcitrant OM slightly increased not showing significant impacts of aging in soil on the recalcitrant fraction of aged biochar. Therefore, these data show that, as expected, the field aging of biochar did not affect the mass loss in the thermal region associated to recalcitrant OM (390–600 °C), while it caused a significant decrease in the thermal region of labile OM (220–390 °C). Total labile OM concentration was reduced by 17% by aging, not consistent with the decomposed amount of the labile fraction estimated at roughly 6–7% after 2 years by the double first-order exponential decay model.²⁹ Similar to other organic materials added into soils, biochar decomposition can be approximated well by biphasic patterns. However, the decomposition rates for the labile pool ($\sim 0.0093\%$ day⁻¹) and recalcitrant pools ($\sim 0.0018\%$ year⁻¹) estimated by the double first-order exponential decay model vary considerably for biochars from different feedstocks and pyrolysis conditions.²⁹

Results from several studies reported that the decomposition rate of biochar showed a clear decrease pattern with experimental duration, with intensive initial decomposition which disappeared after 2 years and was maintained at a very low level for prolonged periods.^{18,21,73} In the field, therefore, optimal incubation conditions in the short-term period often involves the uppermost decomposition rates of the biochar labile fraction with respect to those estimated by the models.^{23,27,29,74} For instance, this might be caused by the physical breakdown of biochar structure during aging in the environment that would accelerate the degradation process.¹⁵

The trend resulting from TGA was confirmed by elemental analysis of fresh and aged biochar, with a decrease during the field aging in the C content and a slight increase in the stable structures. The results of Py-GC-MS analysis, as for TGA results,

showed that the field aging process clearly influenced levels and molecular composition of thermally labile fraction. However, a variable fraction of the carbon in biochar is labile (degradable on annual to decadal timescales) and hence only a proportion of total carbon in biochar allows long-term storage of carbon in soils. Therefore, understanding the real or possible benefits and drawbacks of using biochar as tool for carbon sequestration requires knowledge of recalcitrance of carbon in biochar. According to results shown in this study, decreased molar ratio MePHE/PHE, MeNAP/NAP, T/NAP and T/B, as well as decreased TGA losses at low temperature, confirmed to be correlated with the stable (2 year aged) fraction of the biochar. Therefore, TGA and Py-GC-MS confirmed to be useful tools for the estimation of biochar stability.

DOM analysis

Elemental analysis of fresh and aged biochar DOM. The changes in DOC contents during the aging period in the field are shown in Table 5. Significant differences (ANOVA, $p < 0.05$) were observed in DOC and in WSOC due to aging in the field of biochar (Table 5), indicating that the biochar aging gave rise to a sharp decrease in DOC. Specifically, after 29 months of field aging, the biochar-derived DOC concentrations significantly decreased from 500 \pm 34 mg L⁻¹ in fresh biochar to 114 \pm 14 mg L⁻¹ in aged biochar, representing a loss of 77%.

In biochar-derivate DOM, moreover, more marked differences are observed between fresh and aged biochar in the dissolved fraction of total OC (WSOC = mg DOC g⁻¹ TOC). The

Table 5 Total nitrogen (TN) in water-soluble matter, dissolved organic carbon (DOC) and water-soluble organic carbon (WSOC) concentration of fresh and aged biochar from field experiment. Mean values \pm s.d. ($n = 3$)

Samples	TN (mg L ⁻¹)	DOC (mg L ⁻¹)	WSOC (mg g ⁻¹ TOC)
Fresh biochar	0.5 \pm 0.1	500 \pm 34	3.1 \pm 0.2
Aged biochar	34.7 \pm 0.4	114 \pm 14	0.8 \pm 0.1

Table 4 Weight loss of fresh and aged biochar samples determined by the TGA method. Labile OM from 220 to 390 °C, recalcitrant OM from 390 to 600 °C and carbonates from 600 to 800 °C. Mean values \pm s.d. ($n = 3$)

Samples	105–220 °C weight loss (%)	220–390 °C weight loss (%)	390–600 °C weight loss (%)	600–800 °C weight loss (%)	Residue (%)
Fresh biochar	6.9 \pm 0.1	26.1 \pm 0.6	48.7 \pm 0.7	2.76 \pm 0.05	15.5 \pm 0.1
Aged biochar	7.0 \pm 0.1	9.1 \pm 0.3	51.8 \pm 0.3	3.26 \pm 0.07	28.0 \pm 0.7



Furthermore, the physiochemical properties of biochar, as well as soil properties (*e.g.*, soil organic carbon, minerals, microorganisms, pH, temperature, and moisture) and the crops and plants growing on soil, can have a decisive influence on biochar environmental behavior and its degree of aging.^{21,88,91,92} Therefore, the behavior of weathering is not universal across all biochar types and the effects of aging on the biochar properties need to be extensively explored, possibly through long term field experiments.⁹³ Previous studies reported microbial decomposition and leaching of biochar DOC as the two predominant mechanisms that likely explain biochar C loss, with a C loss from biochar of the 0.5–1% year⁻¹ by microbial degradation^{94,95} and of the 0.5% year⁻¹ by leaching.⁹⁴

Noteworthy is that no significant differences were found in the levels of stable fraction between the aged biochar and the fresh one, indicating that the biochar used in this study was relatively recalcitrant even under field conditions. Therefore, it seems reasonable to assume that after the relatively rapid mineralization of labile OC, the remaining ~50% recalcitrant fraction of biochar (TGA data; Table 4) is not degradable on annual to decadal timescales and hence, only this proportion of total carbon in biochar provides long-term carbon storage. The persistence of OC biochar demonstrated by TGA and Py-GC-MS analysis supports the role played by biochar as a sustainable soil management and carbon sequestration technique, but potential losses due to SOC dynamics should be carefully evaluated especially in long-term field experiments where abiotic and biotic components drive the carbon stability rather than the inherent biochar recalcitrance.

In addition to the potential accumulation of the stable C in soil, the present study confirms that biochar also increases labile forms of soil C. When biochar is applied to soil, the labile C fractions evolved from biochar and can impact microbial activity, including the balance of indigenous labile pools, and therefore on the overall soil ecosystem.^{26,96} As summarized in Table 4, the amount of biochar labile fraction in fresh biochar was $26.1 \pm 0.6\%$, and it decreased to an average of $9.1 \pm 0.3\%$ with aging. Therefore soil fertility can be improved thanks to the labile carbon loss from biochar.

Conclusions

The knowledge of the recalcitrance of carbon in biochar and the behavior of the labile C fraction are crucial issues for evaluating the agro-environmental impact of biochar. This work reported the first comprehensive and quantitative study exploring the multi-annual environmental behavior of biochar by Py-GC-MS and TG analysis of aged samples. It provides insights related to the effects of aging on the biochar properties applied to agricultural soils, particularly regarding the quantity and quality characteristics of labile and recalcitrant fractions after two years of field experiment. Biochar used in the field experiment contained two pools of OM, namely, labile and recalcitrant OM. Results highlighted the effects of aging on these two pools of OM. Specifically, comparing Py-GC-MS and TGA data, similar trends were observed for the % labile and recalcitrant fraction (TGA), and molecular ratios (Py-GC-MS) values. Results

showed lower labile fraction (TGA) and increased aromaticity (Py-GC-MS) in aged biochar. This finding may explain the lower proportion of DOM released by aged biochar, but with a higher degree of aromaticity (UV-vis) in comparison to fresh biochar. In the case of aged biochar, results showed that the use of H/C and O/C ratios as stability indexes will not offer an estimation of the aged biochar real stability. Instead, the qualification of aged biochar C structures such as aromaticity or degree of aromatic condensation by Py-GC-MS analysis and the thermal quantification of the labile and recalcitrant C by TGA provide comprehensive detailed information on biochar behavior in soil. In summary, with regard to the objectives of this study, it is concluded that: (1) no changes in the stable fraction of biochar between the aged and fresh biochar; (2) significant decrease of the biochar labile fraction and of total C concentration in biochar from 26% to 9.1% and from 80% to 67%, respectively, during the aging process in soil; (3) usefulness of Py-GC-MS and TGA to shed light on the characteristics of biochar applied to soil systems, providing consistent information concerning the stability of biochar under real environmental conditions. Therefore, the present work increases the knowledge on the potential aging impacts on the physicochemical properties of biochar and on its connected environmental implications. However, more efforts are needed to better understand the causes of these biochar changes as well as their effects on biochar performance over time.

Author contributions

The manuscript was written through the contributions of all authors. All authors have approved the final version of the manuscript. Conception and design of study: A. G. R., N. G.; acquisition of data: A. G. R., N. G., A. B.; analysis and/or interpretation of data: A. G. R., N. G., D. F.; drafting the manuscript: A. G. R.; revising the manuscript critically for important intellectual content: A. G. R., N. G., D. F., A. B., C. T.; approval of the version of the manuscript to be published: A. G. R., N. G., D. F., A. F., C. T., R. P., C. C., E. B., D. M., D. Z., A. B.

Conflicts of interest

There are no conflicts to declare.

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