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Rhenium(1) complexation—dissociation strategy for synthesising fluorine-18 labelled pyridine bidentate radiotracers†

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A novel fluorine-18 method employing rhenium(i) mediation is described herein. The method was found to afford moderate to high radiochemical yields of labelled rhenium(i) complexes. Subsequent thermal dissociation of the complexes enabled the radiosynthesis of fluorine-18 labelled pyridine bidentate structures which could not be radiofluorinated hitherto. This rhenium(i) complexation—dissociation strategy was further applied to the radiosynthesis of [18F]CABS13, an Alzheimer's disease imaging agent, alongside other 2,2'-bipyridine, 1,10-phenanthroline and 8-hydroxyquinoline labelled radiotracers. Computational modelling of the reaction mechanism suggests that the efficiency of rhenium(i) activation may be attributed to both an electron withdrawal effect by the metal center and the formation of an acyl fluoride intermediate which anchors the fluoride subsequent to nucleophilic addition.

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Introduction

Radiotracers are commonly used to investigate biochemical systems in living organisms and are routinely employed as nuclear medicine imaging agents to diagnose cancers and other disease pathologies.¹ The discovery of new radiolabelling methods is critical to widen the portfolio of radiotracers available to diagnose currently untreated disease states. Positron emission tomography (PET) is the most sensitive of all nuclear imaging techniques and employs molecules with a specific biological role, radioactively tagged with a positron emitting isotope. Among the suite of available positron emitters, fluorine-18 is the most widely utilized radioisotope in existing PET medicines.²,³ However, fluorine-18 nucleophilic radiolabelling reactions (*i.e.* the main route of utilization) can be particularly challenging and low yielding due to: the limiting half-life of the radioisotope (109.7 min),⁴ the relatively few

glucosyl)-purine, was also synthesised in only 5% RCY.13 Given these difficulties associated with fluorine-18 labeling pyridinecontaining structures, it was hypothesized that a rhenium(1) activated approach could be a powerful tool towards expanding the scope of such radiotracers.14 To test this hypothesis, we selected three common bidentate structures throughout our investigation; 1,10-phenanthroline, 2,2'-bipyridine and 8hydroxyquinoline. In each of these studies we systematically tested the nucleophilic substitution of chloro, bromo and/or nitro leaving groups for [18F]fluoride on different positions of the molecule to establish the advantages and limitations of a thermally mediated rhenium(1) complexation-dissociation strategy. Additionally, this reaction fits within a limited scope of fluorine-18 labelling methods which work under partially aqueous conditions, inclusive of AIF, 15-18 boron-dipyrromethene (BODIPY)19-21 and ethenesulfonyl fluoride (ESF) agents.22-24 This

methods available for selective carbon–fluorine bond formation (*e.g.* compared to other halogens),⁵ the low tracer concentrations typically employed and the general requirement for anhydrous conditions.^{6,7} Among these reactions, pyridine structures can be yet more difficult to radiolabel with fluorine-18 and oftentimes form in very low radiochemical yields (RCYs). For example, fluorine-18 labelled acetylcholine receptor imaging agents such as 2-[¹⁸F]fluoro-3-(2(*S*)-azetidinylmethoxy) pyridine and 6-chloro-3-((2(*S*)-azetidinyl)methoxy)-5-(2-[¹⁸F]fluoropyridine-4-yl)pyridine have typically been radiosynthesis within the range of 4–10% RCY.^{8–12} A fluorine-18 labelled pyridine tracer designed for the PET imaging of O⁶-methylguanine-DNA methyltransferase (MGMT) status of tumour tissues, 2-amino-6-(2-[¹⁸F]fluoropyridine-4-ylmethoxy)-9-(octyl-β-D-

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discovery led us to compare reaction conditions employing azeotropically dried and non-dried fluorine-18 sources, the latter of which typically suppresses RCY due to the decreased nucleophilicity of the [18F]fluoride in aqueous media.³

Results and discussion

Each ligand precursor or non-radioactive standard was based on three core structures: 2,2'-bipyridine (BiPy), 1,10-phenanthroline (phen) or 8-hydroxyquinoline (8HQ), as shown in Fig. 1. A list of the substituted analogues studied is shown in Table 1. Rhenium(1) complexes of each of the ligand precursors and standards prepared facial isomers [Re(BiPy)(CO)₃Cl], as [Re(Phen)(CO)₃Cl] or [Re(8HO)(CO)₃(NCCH₃)]. Each precursor (0.08 µmol in DMSO), bearing either a chloro, bromo or nitro leaving group was reacted with azeotropically dried tetraethylammonium (TEA) [18 F]fluoride (29 \pm 10 MBq) under microfluidic conditions using an automated Advion synthesis module (ESI: Fig. S115, Section S.8†). The solutions were each eluted through a microreactor (15.6 μL) at a rate of 20 μL min⁻¹ (47 s reaction time). Reaction temperatures between 50 and 190 °C were trialled for each experiment in 20 °C increments. The solutions were subsequently analysed by radioHPLC and/or radioTLC to determine the non-isolated RCYs of the radioproducts.

Initial attempts to radiofluorinate all the BiPy ligand precursors (**L1a–L1e**) using standard radiofluorination conditions did not afford any of the labelled products across all trialled temperature ranges. Thus, an alternative synthetic route was hypothesised which first involved complexing the pyridine bidentate ligand to a source of [Re(CO) $_5$ Cl]. Heat supplied to the system then allowed for the rhenium($_1$)-mediated incorporation of fluorine-18, in concordance with a former study, whilst

$$\begin{array}{c} R_2 \\ R_1 \\ R_1 \\ R_2 \\ R_2 \\ R_2 \\ R_2 \\ R_2 \\ R_3 \\ R_2 \\ R_2 \\ R_1 \\ R_2 \\ R_2 \\ R_2 \\ R_3 \\ R_2 \\ R_3 \\ R_4 \\ R_2 \\ R_2 \\ R_3 \\ R_4 \\ R_5 \\ R_5 \\ R_6 \\$$

Fig. 1 (Top) Substitution patterns for each of the BiPy, Phen and 8HQ ligands and complexes utilised as precursors or fluorinated standards. (Bottom) Model of rhenium(i) mediation. Rather than employing poor yielding direct fluorine-18 $\langle^{18}F^-\rangle$ nucleophilic substitution, it was hypothesised that the pyridinyl bidentate ligands could first be complexed to a source of rhenium(i) $\langle Re \rangle$ then subsequently radiofluorinated and thermally dissociated (Δ) in a single step to afford the desired radiotracers in greater yield.

Table 1 Substitution patterns for each of the BiPy, Phen and 8HQ ligands and complexes utilised as precursors or fluorinated standards^a

Compound ^a	Ligand structure	R_1	R_2	R_3
L1a	BiPy	Н	Н	Н
L1b	BiPy	Cl	H	Н
L1c	BiPy	H	Cl	Н
L1d	BiPy	Н	H	Cl
L1e	BiPy	Br	H	Н
L1f	BiPy	F	H	Н
L1g	BiPy	Н	F	Н
L1h	BiPy	H	H	F
L2a	Phen	Н	H	_
L2b	Phen	Cl	H	_
L2c	Phen	Н	Cl	_
L2d	Phen	Br	H	_
L2e	Phen	Н	NO_2	_
L2f	Phen	F	H	_
L2g	Phen	Н	F	_
L3a	8HQ	H	H	_
L3b	8HQ	Cl	H	_
L3c	8HQ	H	Cl	_
L3d	8HQ	H	NO_2	_
$L3e^a$	8HQ	F	Н	_
L3f	8HQ	Н	F	_

^a [18 F]**L3e** is also known as [18 F]CABS13.

simultaneously facilitating expedient thermal decomplexation to liberate the radiolabelled ligand in considerable RCY. This rhenium(1) complexation-dissociation strategy is summarised in Fig. 1. Towards obtaining fluorine-18 labelled BiPy structures we first attempted the radiofluorination strategy on [Re(L1b)(CO)3-Cl]. Such an approach indeed allowed for the expected formation of $[^{18}F][Re(L1f)(CO)_3Cl]$ in a maximum 82% RCY at 130 °C. Furthermore, increasing the reaction temperature enabled such previously unreported thermal decomplexation and afforded the formerly unobtainable [18F]L1f ligand in a maximum 64% RCY at 190 °C. The two radiochromatograms showing the conversion of [18 F][Re(L1f)(CO) $_3$ Cl] to [18 F]L1f at 190 °C, alongside the formation of an unknown by-product, are superimposed in Fig. 2. The RCYs attained for both [18F]L1f and [18F][Re(L1f)(CO)₃Cl] as a function of the reaction temperature are plotted in Fig. 3, including the reaction pathway. The trend clearly demonstrated an increase in RCY of [18F][Re(L1f)(CO)3Cl] as the temperature approached 130 °C, and subsequently decreased using higher temperatures as the ortho-substituted [18F]L1f was liberated in greater yield. Under non-azeotropically dried conditions this trend was also noticed, forming both [18F][Re(CO)₃(L1f)Cl] and [18F]**L1f** in 29% RCY at 130 °C and 24% RCY at 190 °C, respectively (ESI: Fig. S71, Section S.4†). A similar trend was also observed for the bromo-substituted [Re(L1e)(CO)₃Cl] precursor, which formed [18F][Re(L1f)(CO)₃Cl] in 72% RCY peaking at 110 °C before decomplexing at higher temperatures to afford [18F]**L1f** in 51% RCY at 190 °C (ESI: Fig. S72, Section S.4†). On the other hand, testing the unsubstituted BiPy analogue [Re(CO)₃(-L1a)Cl] did not provide any radiofluorination product, save for the same radioactive by-product noticed with the previous orthosubstituted structure.

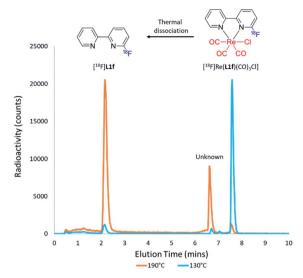


Fig. 2 Radiochromatogram profiles for the fluorine-18 labelling of $[Re(L1b)(CO)_3Cl]$ at 130 °C (blue trace) and 190 °C (orange trace). At 130 °C [^{18}F][$Re(L1f)(CO)_3Cl]$ (7.6 min) forms in the greatest yield, whereas at 190 °C the complex dissociates to form the otherwise unobtainable [^{18}F]L1f ligand (2.2 min) alongside an unknown byproduct (6.6 min).

Given the successful radiolabelling of the *ortho*-position of the pyridine ring when coordinated to the rhenium(ι) center, the [Re(L1d)(CO)₃Cl] precursor was subsequently synthesised to

assess fluorine-18 labelling in the para-position. Such S_NAr fluorinations of [Re(L1d)(CO)₃Cl] afforded the radiolabelled complex in 68% RCY at 130 °C and exhibited thermal dissociation to afford the previously unsynthesisable [18F]L1h ligand in 27% RCY at 190 °C. This trend is illustrated in Fig. 4 which also exhibits the formation of a chromatographically distinct unknown by-product at higher temperatures, which formed in a maximum of 31% RCY at 190 $^{\circ}$ C. Increasing the reaction time from 47 s to 188 s resulted in the suppression of this reaction byproduct, however, and the formation of [18F]L1h in 37% RCY. Despite the successful radiolabelling of the ortho and para positions of the pyridinyl moieties, [18F]fluoride substitution at the meta position of [Re(CO)₃(L1c)Cl] afforded negligible yields of [18F][Re(CO₃)(L1g)Cl] which were unable to be thermally dissociated to form the [18F]L1g ligand. Such results are consistent with the documented substitution efficiencies for S_NAr reactions of aryl halides.

Translation of the rhenium(ı) complexation approach to Phen ligand precursors (**L2a–L2e**) resulted first in the formation of [18 F] [Re(CO)₃(**L2f**)Cl] in 90% RCY at 90 °C from [Re(CO)₃(**L2b**)Cl]. Minor thermal dissociation liberated [18 F]**L2f** in 13% RCY at 170 °C (ESI: Fig. S62, Section S.4†), though this radioligand was found to form in 61% RCY without the aid of rhenium(ı) complexation given the same 0.08 µmol concentration of **L2b** precursor and reaction conditions. Radiofluorination of the bromo substituted [Re(CO)₃(**L2d**)Cl] precursor also mimicked this trend, forming [18 F][Re(CO)₃(**L2f**)Cl] in 69% RCY at 110 °C, though

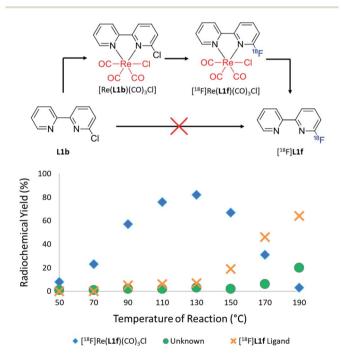


Fig. 3 [18 F]Fluoride substitution in the *ortho* position of [Re(L1b)(CO) $_3$ Cl] which afforded [18 F][Re(L1f)(CO) $_3$ Cl] (blue diamonds). Thermal decomplexation led to the subsequent formation of [18 F]L1f (orange crosses) and an unknown by-product (green circles) with the RCYs plotted as a function of reaction temperature. [18 F]L1f was unable to be synthesised from L1b without first complexing to a rhenium(I) centre.

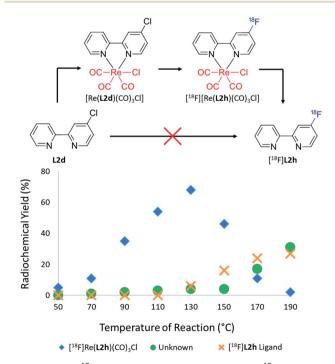


Fig. 4 RCYs of [18 F][Re(L2h)(CO) $_3$ Cl] (blue diamonds), [18 F]L2h (orange crosses) and an unknown by-product (green circles) plotted as a function of the reaction temperature for the [18 F]fluoride labelling of the para substituted [Re(CO) $_3$ (L2d)Cl] complex. [18 F]L2h was unable to be synthesised from L2d unless mediated by rhenium(i) complexation as per the above scheme.

decomplexing only 8% RCY of [18F]L2f at 170 °C (ESI: Fig. S63, Section S.4†). This was not true for the fluorine-18 labelling of [Re(CO)3(L2c)Cl], however, which consisted of a chloro leaving group in the 5-position rather than the 2-position of the phenanthroline ring. In this case, [18F][Re(CO)₃(L2g)Cl] still formed in 33% RCY (Section S.4, Fig. S64 of the ESI†) but did not dissociate to form the [18F]L2g ligand. Direct fluorine-18 labelling of both the L2c and L2e ligand precursors did not afford the [18F]L2g radio product either. Fluorine-18 labelling of the [Re(CO)₃(L2e)Cl] precursor, however, afforded not only [18F][Re(CO)3(L2g)Cl] in 39% RCY at 110 °C but also 2% RCY of the otherwise unobtainable [18F]L2g ligand. This may likely be due to the enhanced electron withdrawal effect of the nitro group which improved yields of the fluorine-18 labelled rhenium(1) complex sufficient for the dissociation of the radioligand in detectable quantities. Nonazeotropically distilled media still allowed for the syntheses of $[^{18}F]Re(CO)_3(\mathbf{L2f})Cl]$ and $[^{18}F][Re(CO)_3(\mathbf{L2g})Cl]$ from $[Re(CO)_3(\mathbf{L2b})$ Cl] and [Re(CO)₃(L2c)Cl] precursors in 59% and 2% RCY, respectively. An unknown by-product was also found to form from each of the [Re(CO)₃(phen)Cl] complexes, including the nonfunctionalized [Re(CO)3(L2a)Cl] precursor, as was observed for the suite of [Re(CO)₃(BiPy)Cl] precursor analogues. A possible explanation for the discrepancy between 2,2-bipyridine and 1,10phenanthroline rhenium-decomplexation efficiencies could be the contribution of an approximate 20-40° twist between the two pyrindyl units of [Re(BiPy)(CO)₃Cl] complexes, which result in lesser molecular rigidity and greater RCYs via dissociation.25-28 The optimum RCYs attained for each of the radiofluorinated BiPy, Phen, [Re(CO)₃(BiPy)Cl] and [Re(CO)₃(Phen)Cl] radiotracers, as dissociated from their respective rhenium(1) precursors at the listed temperatures, are itemised in Table 2.

After establishing the efficiency of the rhenium(i) complexation-dissociation method for the fluorine-18 labelling of diimine ligands, there was much interest in translating the method to 8HQ structures. This interest was due to the proposed use of 8HQ derivatives as markers of Amyloid beta

Table 2 Optimum RCYs of radiotracers obtained from each [Re(CO)₃(BiPy)Cl] and [Re(CO)₃(Phen)₃Cl] precursor exhibiting decomplexation at varying temperatures under the same reaction conditions (solvent: DMSO, reaction time: 47 s, precursor amount: 0.08 μmol, radioactivity: 29 ± 10 MBq)

Precursor	Radiotracer	T (°C)	RCY (%)
[¹⁸ F][Re(CO) ₃ (L1b)Cl]	[18F][Re(CO) ₃ (L1f)Cl]	130	82
	[18F] L1f	190	64
$[^{18}F][Re(CO)_3(\mathbf{L1d})Cl]$	[18F][Re(CO) ₃ (L1h)Cl]	130	68
	$[^{18}$ F]L1 \mathbf{h}^a	190	37
$[^{18}F][Re(CO)_3(\mathbf{L1e})Cl]$	[18F][Re(CO) ₃ (L1f)Cl]	110	69
	[¹⁸ F] L1f	190	51
$[^{18}F][Re(CO)_3(\mathbf{L2b})Cl]$	[18F][Re(CO) ₃ (L2f)Cl]	90	91
	[¹⁸ F] L2f	170	13
$[^{18}F][Re(CO)_3(\mathbf{L2d})Cl]$	[18F][Re(CO) ₃ (L2f)Cl]	110	69
	[18F] L2f	170	8
$[^{18}F][Re(CO)_3(\mathbf{L2e})Cl]$	[18F][Re(CO) ₃ (L2g)Cl]	110	39
/* / -	[¹⁸ F] L2g	130	2

^a Reaction time of 188 s used instead of 47 s.

(Aβ) plaques characteristic of Alzheimer's disease.^{29–33} In particular, it was hypothesized that the rhenium(1) complexation–dissociation approach could be applied to the $\lceil^{18}\text{F}\rceil\text{CABS13}$ ([18F]L3e) Alzheimer's disease PET diagnostic imaging agent^{34,35} Current radiosyntheses of [18F]CABS13 require a multi-step process of radiolabelling and deprotection of the alcohol, as shown in Fig. 5. By protecting the alcohol via a dative Re-O bond in a [Re(CO)₃(L3b)(NCCH₃)] precursor, as also shown in Fig. 5, we envisioned that the radiosynthesis of [18F]CABS13 could be simplified to a single step protocol. Initial fluorine-18 radiolabelling of [Re(CO)₃(L3b)(NCCH₃)] under the standard conditions afforded only 2% RCY of both the [18F][Re(CO)3(-L3e)(NCCH₃)] complex and the [18F]CABS13 radiotracer at 190 °C. Thus the precursor mass was increased ten-fold to 0.8 umol which resulted in a remarkable increase to 18% RCY of the radiolabelled [18F][Re(CO)₃(L3e)(NCCH₃)] complex and 5% of the dissociated [18F]CABS13 tracer, as shown by the radiochromatogram in Fig. 5. While the RCY of [18F]CABS13 achieved via the rhenium(1) complexation-dissociation approach is still lower than the 19 \pm 5% achieved by Liang, et al., 35 the synthesis is certainly still simplified by a single step and the 18% RCY of [18F][Re(CO)₃(L3e)(NCCH₃)] implies that more of the desired tracer could still be obtained under optimised dissociation conditions. Indeed, such results also suggest that greater RCYs from all trialled precursors herein could be attained by modifying the precursor mass. More intriguingly, the rhenium(1)

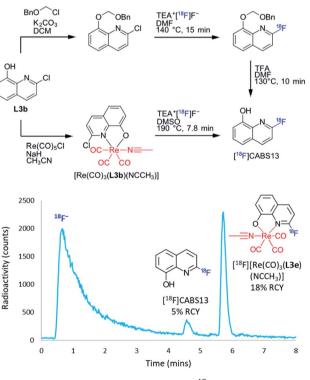


Fig. 5 (Top) Alternative radiosynthesis of [¹⁸F]CABS13 *via* the rhenium(i) complexation–dissociation method circumvented multiple-step synthesis, as reported by Liang, *et al.*³⁴ (Bottom) Radiochromatogram depicting the formation of [¹⁸F]CABS13 in 5% RCY from the thermal dissociation of [¹⁸F][Re(CO)₃(L3e)(NCCH₃)] which formed in 18% RCY.

complexation approach was also translated to the radiosynthesis of [18F]L3f. Experiments have demonstrated that L3f is more efficient at disaggregating AB plagues than CABS13 (L3e),36 however this structure was unable to be radiolabelled with fluorine-18 via traditional means. By applying the rhenium(1) complexation-dissociation strategy, it was discovered that the [18F]L3f tracer was able to be synthesised from both [Re(CO)₃(L3c)(NCCH₃)] and [Re(CO)₃(L3d)(NCCH₃)] precursors in 1% and 2% RCY, respectively. The [Re(CO)₃(L3f)(NCCH₃)] intermediate was also found to form in 3% and 6% RCY from the [Re(CO)₃(L3c)(NCCH₃)] and [Re(CO)₃(L3d)(NCCH₃)] precursors, respectively. Worthwhile mentioning was that the fluorine-18 labelling of such [Re(CO)₃(8HQ)(NCCH₃)] structures formed no unknown by-products, as had been observed in the analogous complexes of BiPy and Phen ligands. This observation was further evidenced by the radiolabelling of unsubstituted [Re(CO)₃(L3a)(NCCH₃)] which afforded no radioproducts at all. Such observations have led us to speculate that the unknown byproduct may correlate with a Re-18F bonded species, albeit nonradioactive analogues of this nature are difficult to both synthesise and characterise for the sake of confirmation (particulary given that Re-Cl & Re-F bonds readily dissociate under ESI-MS conditions).37,38 The RCYs attained for each of the labelled 8HQ structures as a function of reaction temperature can be viewed in Section S.4 of the ESI† for both azeotropically dried and non-dried conditions.

Towards understanding the mechanism by which rhenium(1) complexation-dissociation improves the fluorination of a pyridine bidentate structure, we opted to model the reaction via density functional theory (DFT) calculations at the B97-D3/def2-TZVP level of theory. The input coordinates were acquired from the XRD data of crystals of the rhenium(1) complexes grown from slow evaporation in DMSO solution. First, the coordination on the rhenium(1) centre led to a depletion of electronic density on the ligand, a phenomenon also demonstrated by the downfield chemical shifts observed in ¹H-NMR spectra of the rhenium(1) complexes, as shown by the example spectra of L2b and [Re(L2b)(CO)₃Cl] in Fig. 6. This depletion can be quantified by the Natural Orbitals for Chemical Valence-Charge Displacement (NOCV-CD),³⁹ according to which the L \rightarrow M σ -donation was 0.266e and the L \leftarrow M π back-donation was found to be 0.116e in the case of [Re(CO)₃(L1f)Cl]. Such electronic depletion favours nucleophilic addition of the fluoride on the carbon bound to the chlorine atom. The DFT computed activation energies were 6.3 and 34 kcal mol⁻¹ in the presence and absence of the metal fragment, respectively. Secondly, the fluoride was discovered to bond to the carbonyl in the cis position relative to the diimine ligand, forming an acyl fluoride intermediate as shown in Fig. 7. This intermediate was notably unstable, however, and quickly detached from the carbonyl anchor undergoing S_NAr for the chlorine atom. The proposed fluorination mechanism, occurring between the complexation and dissociation phases, is illustrated in Fig. 8. From the experimental data it was observed that [Re(CO)₃(L1f)Cl] formed in a greater fluorination yield than [Re(CO)₃(**L1h**)Cl], whereas the [Re(CO)₃(**L1g**)Cl] complex formed in almost no appreciable yield at all. This trend can be explained by calculating the atomic charges by Natural Population Analysis,

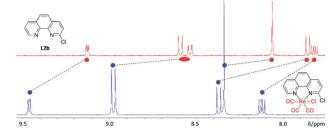


Fig. 6 Example of the downfield chemical shifts in $^1\text{H-NMR}$ aromatic signals observed when complexing L2b (top, red) with [Re(CO) $_5$ Cl] to form [Re(L2b)(CO) $_3$ Cl] (bottom, blue). The shifts suggest electron withdrawal from the π -aromatic system which likely facilitates nucleophilic substitution for [18 F]fluoride.

whereupon it was shown that the carbon atoms bound to the chlorine consisted of partial charges equal to 0.29, -0.01 and 0.04e for the [Re(CO)₃(L1b)Cl], [Re(CO)₃(L1c)Cl] and [Re(CO)₃(-L1d)Cl] precursors, respectively. These values are in line with the observed reactivity, whereby a positive partial charge facilitates fluorination and a negative partial charge inhibits nucleophilic addition of the fluoride. For example, [Re(L1b)(CO)3Cl] and [Re(L2b)(CO)₃Cl] were calculated to have the highest partial charges of 0.29e and 0.31e which correlated with high yields of 82% and 91% RCY for their radiolabelled analogues, respectively. Whereas [Re(CO)₃(L1c)Cl], for which a negative partial charge of -0.01e was calculated, did not form the radiolabelled complex in ample quantity for detection. The partial charges are plotted in Table 3 for some select Phen and 8HQ complexes as well, alongside their associated ligands. In each case the partial charge for the carbon bound to the chlorine or nitro leaving group was found to be greater in the complex than in the ligand, save for [Re(CO)₃(L2e)Cl] and L2e which differed only by 0.01e.

Finally, quantum yields of select complexes taking the form of $[Re(CO)_3(BiPy)Cl]$ were performed to determine the utility of such complexes as multimodal PET-optical probes. The

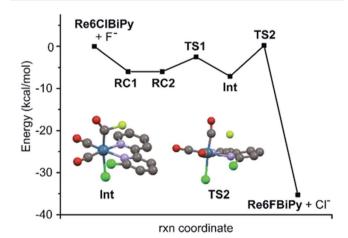


Fig. 7 DFT modelling of the reaction mechanism forming [Re(CO)₃(-L1h)Cl] (Re₆FBiPy) from [Re(CO)₃(L1b)Cl] (Re₆ClBiPy). The energies for the optimized geometries of the acyl fluoride intermediate species (Int) and the rate-determining transition states (TS) are plotted along the reaction coordinate.

Fig. 8 Example of the proposed rhenium-assisted radiolabelling mechanism: (i) precursor ligand L1b is complexed to a source of [Re(CO₅)Cl] to form [Re(CO)₃(L1b)Cl]; (ii) [¹⁸F]fluoride reacts with the cis-carbonyl of [Re(CO)₃(L1b)Cl] to from an acyl fluoride intermediate species (Int); (iii) the fluorine-18 radioisotope is in proximity of the chlorine leaving group, enabling S_NAr to form the second transition state (TS2); (iv) chloride is displaced from the aromatic ring, thus forming [¹⁸F][Re(CO)₃(L1h)Cl]; (v) thermal decomplexation of [¹⁸F] [Re(CO)₃(L1h)Cl] liberates the desired [¹⁸F]L1h radiotracer.

quantum yields were determined to be 0.5%, 1.0% and 0.6% for the [Re(CO)₃(L1b)Cl], [Re(CO)₃(L1c)Cl] and [Re(CO)₃(L1d)Cl] precursors in DMSO solution, respectively. These quantum

Table 3 Natural population analysis results for the atomic partial charge (q) of the carbon atom bonded to the Cl or NO₂ leaving group. Quantum yields (ϕ) are provided for the photophysical analyses of some complexes

Ligand	q (C)	Re(I) complex	q (C)	φ (%)
L1b	0.26	[Re(CO) ₃ (L1b)Cl]	0.29	0.5
L1e L1c	0.17 -0.03	$[Re(CO)_3(\mathbf{L1e})Cl]$ $[Re(CO)_3(\mathbf{L1c})Cl]$	$0.18 \\ -0.01$	1.0
L1d L2b	0.03 0.27	[Re(CO) ₃ (L1d)Cl] [Re(CO) ₃ (L2b)Cl]	$0.04 \\ 0.31$	0.6 0.3 (ref. 14)
L2e	0.12	$[Re(CO)_3(L2e)Cl]$	0.11	— (ICI. 14)
L3d	_	$[Re(CO)_3(L2e)(NCCH_3)]$	0.08	_

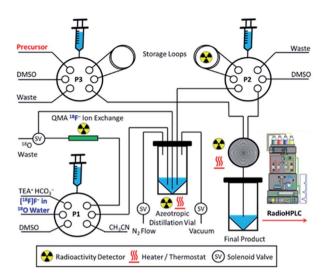


Fig. 9 Automated synthesis module for the azeotropic drying of cyclotron generated [18F]fluoride and subsequent microfluidic set-up for the rhenium(i) complexation–dissociation reactions under varying reaction environments.

yields are depicted in Table 3, alongside the $[Re(CO)_3(\mathbf{L2b})Cl]$ precursor for which a quantum yield of 0.3% was determined in one of our former studies. The fluorine substituted $[Re(CO)_3(\mathbf{L1f})Cl]$ was also found to have a quantum yield of 0.6% in DMSO solution. Emission spectra from these photophysical assessments can be found in Section S.6, Fig. S82–S85 of the ESI.† Transitions to the metal–ligand charge transfer (MLCT) excited state occurred in the UV region, with corresponding $d\pi \to \pi^*$ electronic transitions occurring at 380 nm for $[Re(CO)_3(\mathbf{L1b})Cl]$, 379 nm for $[Re(CO)_3(\mathbf{L1f})Cl]$ and at 369 nm for both $[Re(CO)_3(\mathbf{L1d})Cl]$ and $[Re(CO)_3(\mathbf{L1f})Cl]$ complexes, as depicted by the UV-Vis spectra in Section S.2 of the ESI.†

Conclusions

A novel rhenium(1) complexation-dissociation method for labelling with the fluorine-18 radioisotope is reported herein. The rhenium(1) centre improved fluorination via an electron withdrawal effect as suggested by the NMR data and, in accordance with our computational models, may facilitate radiofluorination via formation of an acyl fluoride intermediate with a carbonyl ancillary ligand. Through a series of microfluidic radiosyntheses we verified that moderate to high RCYs could be obtained by substituting [18F]fluoride in the rhenium(1) activated ortho and para positions of bidentate pyridine ligands, as well as in the 5-position of a Phen ligand. These radiolabelled rhenium(1) complexes are currently being further pursued for utility as PET-optical multimodal agents. Thermal decomplexation of such rhenium(1) complexes were shown to be particularly useful for synthesizing fluorine-18 labelled Bipy ligands, which were unable to be synthesised under analogous conditions without rhenium(1) activation. The formerly unobtainable [18F]L3f radio product was also attained from the rhenium(1) complexation-dissociation approach, which will enable its further investigation as an alternative 8HQ Alzheimer's

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disease imaging tracer to [18F]CABS13. Furthermore, the rhenium(1) complexation-dissociation strategy joins the limited suite of radiofluorinating methods which can afford radiotracers under partially aqueous conditions. Future work towards alternative decomplexation methods (e.g. photodissociation or acid dissociation), as well as attempts to fluorine-18 label monodentate pyridine ligands complexed to the rhenium(1) core, may further expand the scope of radiotracer syntheses via this rhenium(1) mediated approach.

Experimental section

Synthetic protocols for rhenium complexes

Complex derivatives of [Re(CO)₃(Phen)Cl] and [Re(CO)₃(BiPy)Cl] were prepared by refluxing rhenium(1) pentacarbonyl chloride (300 mg, 830 µmol, 1.0 eq.) and the appropriately substituted chloro, bromo or nitro 2,2-bipyridine or 1,10-phenanthroline (1.1 eq.) in a solution of anhydrous toluene (10 mL) for 4 hours in an inert nitrogen environment, resulting in a colour change from blue or colourless to yellow or orange. The solid precipitate was isolated by vacuum filtration and washed with diethyl ether. The crude material was then dissolved in a minimum of DCM and eluted over neutral alumina Brockmann grade II using a MeOH/DCM gradient mobile phase. The eluents containing the product were subsequently combined and evaporated down under reduced pressure to afford the yellow or orange powder.

Complex derivatives of [Re(CO)₃(8HQ)(NCCH₃)] were prepared by first refluxing rhenium(1) pentacarbonyl chloride (100 mg, 275 µmol, 1.0 eq.) in anhydrous acetonitrile (10 mL) for 4 hours in an inert nitrogen environment to synthesise the rhenium(1) diacetonitrilo tricarbonyl chloride intermediate, which was monitored by HPLC and IR spectroscopy. Simultaneously, the appropriate chloro or nitro substituted 8-hydroxvquinoline (1.0 eq.) was deprotonated with sodium hydride (60% wt/wt in mineral oil washed with n-hexanes) in anhydrous acetonitrile (2 mL) at room temperature under an inert nitrogen environment, resulting in a vibrant green solution. The green solution was then filtered (0.2 µm PTFE) and added dropwise to the solution containing rhenium. This solution was then left to stir for 12 hours at room temperature eventually forming an orange solution. The orange solution was added to a volume of aqueous triflic acid (0.1 M, 10 mL) and evaporated under reduced pressure to remove the acetonitrile, thus precipitating out a yellow or orange solid in aqueous suspension. The crude material was then isolated by vacuum filtration, dissolved in a minimum of acetonitrile and eluted over neutral alumina Brockmann grade II using an isocratic acetonitrile mobile phase. The eluents containing the product were subsequently combined and evaporated down under reduced pressure to afford the yellow or orange powder.

Further details on the syntheses of the rhenium(1) complexes and the fluorinated ligands can be found in Sections S.2 and S.3 of the ESI.† Characterisation data for the rhenium(1) complexes tricarbonylchloro(2,2'-bipyridine)rhenium(1) $([Re(CO)_3(L1a)]$ Cl]),40,41 tricarbonylchloro(1,10-phenanthroline)rhenium(1) ([Re(CO)₃(L2a)Cl])⁴² and acetonitriletricarbonyl(8-quinolate)

rhenium(1) ([Re(CO)₃(L3a)NCCH₃]) can be found in the existing literature.43

Characterisation data for novel complexes

Tricarbonylchloro(6-chloro-2,2'-bipyridine)rhenium(1)

([Re(CO)₃(L1b)Cl]). Yellow solid; MP: 255 °C; 86% yield. ¹H NMR (400 MHz, DMSO-d₆, δ/ppm): δ 9.06 (d, J^{d} = 4.76 Hz, 1H), δ 8.76 $(d, J^d = 8.12 \text{ Hz}, 2H), \delta 8.35 \text{ (app. td}, J^d = 1.96, J^t = 8.04 \text{ Hz}, 2H),$ δ 8.08 (dd, $J^{\rm d} = 0.68$, 8.04 Hz, 1H), δ 7.78 (app. td, $J^{\rm d} = 1.80$, $J^{\rm t} =$ 4.56 Hz, 1H). $^{13}\text{C}\{^1\text{H}\}$ NMR (100 MHz, DMSO-d₆, δ /ppm): $\delta 197.5_{C=0}, \delta 197.3_{C=0}, \delta 189.5_{C=0}, \delta 157.2, \delta 155.6, \delta 153.2,$ δ 152.8, δ 142.7, δ 140.3, δ 128.0, δ 127.7, δ 125.3, δ 123.1. FTIR (ATR corr. $\tilde{\nu}/\text{cm}^{-1}$): $\tilde{\nu}$ 3075 (w, C-H sp² str.), $\tilde{\nu}$ 2018 (s, A'(1) C \equiv O str.), $\tilde{\nu}$ 1882 (s, A'(2) & A" C \equiv O str.). UV/Vis (λ /nm, ε /L $\text{mol}^{-1} \text{ cm}^{-1}$): λ 380 (d $\pi \to \pi^*$, ϵ 3375.36), λ 301 ($\pi \to \pi^*$, ϵ 14 990.57), λ 326 ($\pi \to \pi^*$, ϵ 10 920.28), λ 233 ($\pi \to \pi^*$, ϵ 19 259.41). LRMS (ESI⁺): $[M-Cl]^+$ m/z cald: 460.97, m/z obvs 460.95. HRMS (ESI⁺): $[M-Cl]^+$ m/z cald: 460.9703, m/z obvs 460.9694 (Δ -2.0 ppm). EA (%): calc. C 31.46, H 1.42, N 5.64. Found C 31.59, H 1.17, N 5.70.

Tricarbonylchloro(5-chloro-2,2'-bipyridine)rhenium(1)

([Re(CO)₃(L1c)Cl]). Yellow solid; MP: 292 °C, 50% yield. ¹H NMR (400 MHz, DMSO-d₆, δ/ppm): δ 9.03 (d, $J^{\text{d}} = 2.28$ Hz, 2H), δ 8.80 $(t, J^{t} = 8.92 \text{ Hz}, 2H), \delta 8.55 (dd, J^{d} = 8.84, 2.28 \text{ Hz}, 1H), \delta 8.35 (td, J^{d} = 8.84, 2.28 \text{ Hz}, 2H)$ $J^{t} = 8.04 \text{ Hz}, J^{d} = 1.32 \text{ Hz}, 1\text{H}, \delta 7.77 \text{ (app. qd, } J^{q} = 5.68 \text{ Hz}, J^{d} = 5.68 \text{ Hz},$ 1.16 Hz, 1H). $^{13}\text{C}\{^1\text{H}\}$ NMR (100 MHz, DMSO-d₆, δ /ppm): δ 197.5_{C=O}, δ 197.2_{C=O}, δ 189.6_{C=O}, δ 154.2, δ 154.1, δ 153.0, δ 150.9, δ 140.4, δ 140.2, δ 133.9, δ 128.1, δ 125.3, δ 124.8. FTIR (ATR corr. \tilde{v}/cm^{-1}): \tilde{v} 3113 (w, C-H sp² str.), \tilde{v} 2021 (s, A'(1) C=O str.), $\tilde{\nu}$ 1883 (s, A'(2) & A" C \equiv O str.). UV/Vis (λ /nm, ε /L $\text{mol}^{-1} \text{ cm}^{-1}$): λ 379 (d $\pi \to \pi^*$, ϵ 7346.37), λ 299 ($\pi \to \pi^*$, ϵ 36 136.21), λ 241 ($\pi \to \pi^*$, ε 42 589.10). LRMS (ESI⁺): [M-Cl]⁺ m/ z cald: 460.97, m/z obvs 460.95. HRMS (ESI⁺): [M-Cl]⁺ m/z cald: 460.9703, m/z obvs 460.9730 (Δ 5.9 ppm). EA (%): calc. C 31.46, H 1.42, N 5.64. Found C 31.61, H 1.14, N 5.44.

Tricarbonylchloro(4-chloro-2,2'-bipyridine)rhenium(1)

([Re(CO)₃(L1d)Cl]). Yellow solid; MP: 283 $^{\circ}$ C; 89% yield. 1 H NMR (400 MHz, DMSO-d₆, δ /ppm): δ 9.04 (d, J^{d} = 4.68 Hz, 1H), δ 9.01 (d, J^{d} = 2.04 Hz, 1H), δ 8.96 (d, J^{d} = 6.00 Hz, 1H), δ 8.85 (d, $J^{\rm d} = 8.16 \text{ Hz}, 1\text{H}, \delta 8.36 \text{ (app. td, } J^{\rm t} = 8.04 \text{ Hz}, J^{\rm d} = 1.32, 1\text{H}),$ δ 7.88 (dd, J^{d} = 2.12, 6.00 Hz, 1H), δ 8.36 (app. td, J^{d} = 1.12, J^{t} = 5.56 Hz, 1H). ${}^{13}\text{C}\{{}^{1}\text{H}\}$ NMR (100 MHz, DMSO-d₆, δ/ppm): δ 197.6_{C=O}, δ 197.5_{C=O}, δ 189.7_{C=O}, δ 156.9, δ 154.3, δ 153.8, δ 153.1, δ 147.0, δ 140.3, δ 128.4, δ 127.7, δ 125.0, δ 124.7. FTIR (ATR corr. \tilde{v}/cm^{-1}): \tilde{v} 3117 (w, C-H sp² str.), \tilde{v} 2019 (s, A'(1) C \equiv O str.), \tilde{v} 1884 (s, A'(2) & A" C \equiv O str.). UV/Vis (λ /nm, ϵ /L $\text{mol}^{-1} \text{ cm}^{-1}$): $\lambda 380 \text{ (d}\pi \to \pi^*, \varepsilon 7548.93), \lambda 292 \text{ (}\pi \to \pi^*, \varepsilon$ 38 022.44), λ 235 ($\pi \to \pi^*$, ε 48 347.07). LRMS (ESI⁺): [M-Cl]⁺ m/z cald: 460.97, m/z obvs 460.95. HRMS (ESI⁺): [M-Cl]⁺ m/z cald: 460.9703, m/z obvs 460.9712 (Δ 2.0 ppm). EA (%): calc. C 31.46, H 1.42, N 5.64. Found C 31.81, H 1.20, N 5.32.

Tricarbonylchloro(6-bromo-2,2'-bipyridine)rhenium(1)

([Re(CO)₃(L1e)Cl]). Yellow solid; MP: 260 °C, 86% yield. ¹H NMR (400 MHz, DMSO-d₆, δ/ppm): δ 9.06 (dd, $J^{\text{d}} = 5.48$ Hz, 0.80 Hz, 1H), δ 8.77 (q, J^{q} = 5.16 Hz, 4.08 Hz, 1H), δ 8.71 (d, J^{d} = 8.24 Hz, 1H), δ 8.33 (td, J^{t} = 8.08 Hz, J^{d} = 1.52 Hz, 1H), δ 8.20 (d, J^{d} =

1.40 Hz, 1H), δ 8.19 (s, 1H), δ 7.76 (qd, $J^{\rm q} = 5.52$ Hz, 1.24 Hz, $J^{\rm d} = 1.04$ Hz, 1H). ¹³C{¹H} NMR (100 MHz, CD₃CN, δ /ppm): δ 198.1_{C=O}, δ 197.2_{C=O}, δ 190.0_{C=O}, δ 157.6, δ 156.0, δ 152.7, δ 146.2, δ 142.0, δ 140.2, δ 131.9, δ 127.8, δ 125.3, δ 123.3. FTIR (ATR corr. $\tilde{\nu}$ /cm⁻¹): $\tilde{\nu}$ 3074 (w, C-H sp² str.), $\tilde{\nu}$ 2017 (s, A'(1) C=O str.), $\tilde{\nu}$ 1884 (s, A'(2) & A" C=O str.). LRMS (ESI⁺): [M-Cl]⁺ m/z cald: 504.92, m/z obvs 505.04. EA (%): calc. C 28.87, H 1.30, N 5.18. Found C 28.90, H 1.05, N 5.21.

Tricarbonylchloro(6-fluoro-2,2'-bipyridine)rhenium(1) ([Re(CO)₃(L1f)Cl]). Yellow solid; MP: 294 °C, 65% yield. ¹H NMR (400 MHz, DMSO-d₆, δ /ppm): δ 9.06 (app. tq, $J^{t} = 5.48$ Hz, $J^{q} =$ 0.80 Hz, 1H), δ 8.80 (d, J^{d} = 8.16 Hz, 1H), δ 8.70 (d, J^{d} = 7.92 Hz, 1H), δ 8.52 (q, $J^q = 8.04$ Hz, 1H), δ 8.36 (td, $J^t = 8.08$ Hz, $J^d =$ 1.52 Hz, 1H), δ 7.80 (m, 2H). ¹³C{¹H} NMR (100 MHz, DMSO-d₆, δ/ppm): δ 197.2_{C=O}, δ 196.6_{C=O} (d, J^{d} = 16.59 Hz), δ 188.9_{C=O}, δ 164.0, δ 161.5, δ 154.4 (d, J^{d} = 15.92 Hz), δ 153.1, δ 145.9 (d, J^{d} = 10.15 Hz), δ 140.4, δ 128.2, δ 125.1, δ 121.4 (d, J^{d} = 2.94 Hz), δ 112.7 (d, $f^{\rm d}$ = 30.19 Hz). ¹⁹F{¹H} NMR (376 MHz, DMSO-d₆, δ / ppm): δ -51.75. FTIR (ATR corr. $\tilde{\nu}/\text{cm}^{-1}$): $\tilde{\nu}$ 3040 (w, C-H sp² str.), $\tilde{\nu}$ 2016 (s, A'(1) C=O str.), $\tilde{\nu}$ 1880 (s, A'(2) & A" C=O str.). UV/Vis (λ /nm, ε /L mol⁻¹ cm⁻¹): λ 369 (d $\pi \to \pi^*$, ε 2768.36), λ 296 ($\pi \to \pi^*$, ε 12 485.88), λ 235 ($\pi \to \pi^*$, ε 15 367.33). LRMS (ESI⁺): $[M-Cl]^+ m/z$ cald: 445.00, m/z obvs 445.10. EA (%): calc. C 32.54, H 1.47, N 5.84. Found C 32.59, H 1.16, N 5.88.

Tricarbonylchloro(4-fluoro-2,2'-bipyridine)rhenium(ι) ([Re(CO)₃(L1h)Cl]). Yellow solid; MP: 302 °C; 38% yield. ¹H NMR (400 MHz, DMSO-d₆, δ /ppm): δ 9.04 (app. t, f = 6.72 Hz, 2H), δ 8.78 (m, 2H), δ 8.36 (td, f = 1.44, f = 7.92 Hz, 1H), δ 7.79 (qd, f = 1.12, f = 7.56 Hz, 1H), δ 7.69 (qd, f = 1.00, f = 7.36 Hz, 1H). ¹³C{¹H} NMR (100 MHz, DMSO-d₆, δ /ppm): δ 197.6_{C=O}, δ 197.6_{C=O}, δ 189.9_{C=O}, δ 168.3, δ 168.3, δ 158.9 (d, f = 10.26 Hz), δ 156.2 (d, f = 9.99 Hz), δ 154.6, δ 153.1, δ 140.5, δ 128.5, δ 124.9, δ 116.0 (d, f = 19.7 Hz), δ 113.0 (d, f = 22.79 Hz). ¹°F{¹H} NMR (376 MHz, DMSO-d₆, δ /ppm): δ -94.71. FTIR (ATR corr. $\tilde{\nu}$ /cm ¹): $\tilde{\nu}$ 3117 (w, C-H sp² str.), $\tilde{\nu}$ 2018 (s, f (1) C=O str.), $\tilde{\nu}$ 1884 (s, f (2) & f C=O str.). LRMS (ESI *): [M-Cl] * f f cald: 445.00, f obvs 444.92. EA (%): calc. C 32.54, H 1.47, N 5.84. Found C 32.52, H 1.33, N 5.81.

Tricarbonylchloro(2-chloro-1,10-phenanthroline)rhenium(1) ([Re(CO)₃(L2b)Cl]). Yellow solid; MP: 316 °C (dec.); 89% yield. ¹H NMR (400 MHz, DMSO-d₆, δ /ppm): δ 9.46 (dd, J^d = 5.12, 1.36 Hz, 1H), δ 8.98 (d, J^d = 8.52 Hz, 1H), δ 8.97 (d, J^d = 8.24 Hz, 1H), δ 8.37 (d, J^d = 8.56, 1H), δ 8.34 (s, 2H), δ 8.12 (dd, J^d = 8.24, 5.12 Hz, 1H). 13 C 1 H 1 NMR (100 MHz, CDCl₃, δ /ppm): δ 197.88 (C \equiv O), δ 197.65 (C \equiv O), δ 189.83 (C \equiv O), δ 154.75, δ 154.11, δ 147.19, δ 146.56, δ 142.81, δ 139.92, δ 131.69, δ 129.66, δ 128.37, δ 128.09, δ 127.30, δ 127.24. FTIR (ATR corr. $\tilde{\nu}/\text{cm}^{-1}$): $\tilde{\nu}$ 3042 (w, C-H sp² str.), $\tilde{\nu}$ 2021 (s, A'(1) C=O str.), $\tilde{\nu}$ 1889 (s, A'(2) & A" C \equiv O str.). UV/Vis (CH₃CN, λ /nm, ε /L mol⁻¹ cm⁻¹): λ 368 $(\text{Re}(d\pi) \to \pi^*, \varepsilon 374.65), \lambda 270 (\pi \to \pi^*, \varepsilon 2612.13), \lambda 218 (\pi \to \pi^*, \varepsilon 2612.13)$ π^* , ε 4703.92), λ 204 ($\pi \to \pi^*$, ε 4589.45). LRMS (ESI⁺): [M-Cl]⁺ m/z calc. 484.97, m/z obsv. 485.07. HRMS (ESI⁺): $[M + Na]^+ m/z$ calc. 540.9261 ($C_{15}H_7N_2O_3FCl_2ReNa^+$); m/z obsv. 540.9266 (Δ 0.90 ppm). EA (%): calc. C 34.62, H 1.36, N 5.38, Cl 13.63; obsv. C 34.45, H 0.95, N 5.36, Cl 13.55.

 $\label{eq:constraint} \textbf{Tricarbonylchloro(5-chloro-1,10-phenanthroline)rhenium(1)} \\ \textbf{([Re(CO)_3(L2c)Cl])}. \ Yellow \ solid; \ MP: 292 \ ^{\circ}C, 50\% \ yield. \ ^{1}H \ NMR \\ \\ \textbf{(MP: 292 \ ^{\circ}C, 50\% \ yield. \ ^{1}H \ NMR)} \\ \textbf{(MP: 292 \ ^{\circ}C, 50\% \ yiel$

(400 MHz, DMSO-d₆, δ /ppm): δ 9.03 (d, J^d = 2.28 Hz, 2H), δ 8.80 (t, J^t = 8.92 Hz, 2H), δ 8.55 (dd, J^d = 8.84, 2.28 Hz, 1H), δ 8.35 (td, J^t = 8.04 Hz, J^d = 1.32 Hz, 1H), δ 7.77 (app. qd, J^q = 5.68 Hz, J^d = 1.16 Hz, 1H). ¹³C{¹H} NMR (100 MHz, DMSO-d₆, δ /ppm): δ 197.5_{C=O}, δ 197.2_{C=O}, δ 189.6_{C=O}, δ 154.2, δ 154.1, δ 153.0, δ 150.9, δ 140.4, δ 140.2, δ 133.9, δ 128.1, δ 125.3, δ 124.8. FTIR (ATR corr. $\tilde{\nu}$ /cm⁻¹): $\tilde{\nu}$ 3113 (w, C-H sp² str.), $\tilde{\nu}$ 2021 (s, A'(1) C=O str.), $\tilde{\nu}$ 1883 (s, A'(2) & A" C=O str.). UV/Vis (λ /nm, ϵ /L mol⁻¹ cm⁻¹): λ 379 (d π \rightarrow π *, ϵ 7346.37), λ 299 (π \rightarrow π *, ϵ 36 136.21), λ 241 (π \rightarrow π *, ϵ 42 589.10). LRMS (ESI⁺): [M-Cl]⁺ m/z cald: 460.970, m/z obvs 460.9730 (Δ 5.9 ppm). EA (%): calc. C 31.46, H 1.42, N 5.64. Found C 31.61, H 1.14, N 5.44.

Tricarbonylchloro(2-bromo-1,10-phenanthroline)rhenium(ι) ([Re(CO)₃(L2d)CI]). Yellow solid; MP: 328 °C (dec.), 95% yield.
¹H NMR (400 MHz, DMSO-d₆, δ/ppm): δ 9.46 (dd, J^d = 5.12 Hz, 1.36 Hz, 1H), δ 8.96 (dd, J^d = 8.24 Hz, 1.36 Hz, 1H), δ 8.82 (d, J^d = 8.56 Hz, 1H), δ 8.49 (d, J^d = 8.56 Hz, 1H), δ 8.34 (d, J^d = 1.44 Hz, 1H), δ 8.32 (d, J^d = 2.48 Hz, 1H), δ 8.11 (dd, J^d = 8.24 Hz, 5.12 Hz, 1H).
¹³C{¹H} NMR (100 MHz, CD₃CN, δ/ppm): δ 198.0_{C≡O}, δ 197.1_{C≡O}, δ 189.8_{C≡O}, δ 153.5, δ 147.8, δ 147.2, δ 146.4, δ 141.4, δ 139.4, δ 131.3, δ 130.8, δ 129.4, δ 127.8, δ 127.7, δ 126.6. FTIR (ATR corr. $\bar{\nu}$ /cm⁻¹): $\bar{\nu}$ 3043.35 (w, C–H sp² str.), $\bar{\nu}$ 2014.27 (s, A'(1) C≡O str.), $\bar{\nu}$ 1883.98 (s, A'(2) & A" C≡O str.). EA (%): calc. C 31.90, H 1.25, N 4.96. Found C 31.94, H 1.00, N 5.08.

Tricarbonylchloro(5-nitro-1,10-phenanthroline)rhenium(i) ([Re(CO)₃(L2e)Cl]). Yellow solid, MP: 190–194 °C (dec.), 26% yield. ¹H NMR (400 MHz, CD₃CN, δ /ppm): δ 9.57 (dd, f^d = 9.00, 1.28 Hz, 1H), δ 9.03 (dd, f^d = 4.84, 1.28 Hz, 1H), δ 8.63 (d, f^d = 9.24 Hz, 1H), δ 7.81 (dd, f^d = 9.00, 4.84 Hz, 1H), δ 6.88 (d, f^d = 9.24 Hz, 1H). ¹³C{¹H} NMR (100 MHz, CD₃CN, δ /ppm): δ 196.3_{C=O}, δ 194.4_{C=O}, δ 178.1_{C=O}, δ 151.6, δ 143.3, δ 137.1, δ 132.9, δ 130.8, δ 127.0, δ 126.9, δ 121.4, δ 114.2. FTIR (ATR corr. $\tilde{\nu}$ /cm⁻¹): $\tilde{\nu}$ 2939 (w, C-H sp² str.), $\tilde{\nu}$ 2918 (w, C-H sp² str.), $\tilde{\nu}$ 2328 (w, N=C str.), $\tilde{\nu}$ 2019 (s, A'(1) C=O str.), $\tilde{\nu}$ 1900 (s, A'(2) & A" C=O str.).

Tricarbonylchloro(2-fluoro-1,10-phenanthroline)rhenium(1) ([Re(CO)₃(L2f)Cl]). Yellow solid; MP: 314 °C (dec.); 65% yield. ¹H NMR (400 MHz, DMSO-d₆, δ/ppm): δ 9.47 (dd, $J^{\text{d}} = 5.04$, 1.32 Hz, 1H), δ 9.16 (dd, J^{d} = 8.84, 6.80 Hz, 1H), δ 8.99 (dd, J^{d} = 8.32, 1.32 Hz, 1H), δ 8.36 (app. q, $f^q = 14.72$, 8.84 Hz, 2H), δ 8.14 (dd, $J^{\rm d} = 8.84, 1.48 \text{ Hz}, 1\text{H}), \delta 8.12 (dd, J^{\rm d} = 8.32, 5.04 \text{ Hz}, 1\text{H}).$ ¹³C ${}^{1}H$ } NMR (100 MHz, CDCl₃, δ/ppm): δ 197.10 (C≡O), δ 196.54 (d, $J^{d} = 14$ Hz, C \equiv O), δ 189.99 (C \equiv O), δ 163.77, δ 161.18, δ 153.90, δ 145.79 (d, J^{d} = 11 Hz), δ 145.09 (d, J^{d} = 2 Hz), δ 144.07 $(d, J^d = 3 \text{ Hz}), \delta 139.48, \delta 130.99, \delta 128.29 (d, J^d = 1 \text{ Hz}), \delta 127.32$ $(d, J^d = 2 Hz), \delta 127.09 (d, J^d = 61 Hz), \delta 113.48 (d, J^d = 32 Hz).$ ¹⁹F NMR (376 MHz, DMSO-d₆, δ /ppm): δ –36.75 (dd, J^d = 6.92, 1.92 Hz), δ –106.00 (ISTD: para-DFB). FTIR (ATR corr. $\tilde{\nu}/\text{cm}^{-1}$): $\tilde{\nu}$ 3077 (w, C-H sp² str.), $\tilde{\nu}$ 2021 (s, A'(1) C=O str.), $\tilde{\nu}$ 1890 (s, A'(2) & A" C \equiv O str.). UV/Vis (CH₃CN, λ /nm, ε /L mol⁻¹ cm⁻¹): λ 375 (Re(d π) $\rightarrow \pi^*$, ε 2720.95), λ 343 ($\pi \rightarrow \pi^*$, ε 2821.73), λ 269 ($\pi \rightarrow$ π^* , ε 22 069.94), λ 262 ($\pi \to \pi^*$, ε 22 170.71), λ 215 ($\pi \to \pi^*$, ε 38 798.75), λ 200 ($\pi \to \pi^*$, ε 40 915.05). LRMS (ESI⁺): [M-Cl]⁺ m/ z calc. 469.00, m/z obsv. 469.12. HRMS (ESI⁺): $[M + Na]^+ m/z$ calc. 524.9557 ($C_{15}H_7N_2O_3FClReNa^+$); m/z obsv. 524.9579 (Δ 4.20 This article is licensed under a Creative Commons Attribution-NonCommercial 3.0 Unported Licence.

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ppm). EA (%): calc. C 3575, H 1.40, N 5.56, Cl 7.04; obsv. C 35.73, H 0.98, N 5.59, Cl 6.83.

Tricarbonylchloro(5-fluoro-1,10-phenanthroline)rhenium(1) ([Re(CO)₃(L2g)Cl]). Yellow solid, MP: 322 °C (dec.), 26% yield.

¹H NMR (400 MHz, DMSO-d₆, δ /ppm): δ 9.54 (dd, f^d = 1.28, 5.12 Hz, 1H), δ 9.42 (dd, f^d = 1.20, 5.08 Hz, 1H), δ 9.04 (dd, f^d = 1.28, 8.40 Hz, 1H), δ 8.93 (dd, f^d = 1.28, 8.40 Hz, 1H), δ 8.29 (d, f^d = 10.44 Hz, 1H), δ 8.20 (dd, f^d = 5.12, 8.40 Hz, 1H), δ 8.11 (dd, f^d = 5.08, 8.32 Hz, 1H).

¹³C{¹H} NMR (100 MHz, DMSO-d₆, δ /ppm): δ 197.4_{C=O}, δ 197.3_{C=O}, δ 189.6_{C=O}, δ 156.8, δ 154.5, δ 154.3, δ 152.9 (d, f^d = 2 Hz), δ 146.5 (d, f^d = 7 Hz), δ 143.6, δ 139.1 (d, f^d = 5 Hz), δ 133.0 (d, f^d = 4 Hz), δ 129.6 (d, f^d = 4 Hz), δ 126.9 (d, f^d = 3 Hz), δ 122.8 (d, f^d = 23 Hz), δ 109.3 (d, f^d = 22 Hz).

¹⁹F{¹H} NMR (376 MHz, CD₃CN, δ /ppm): δ −120.8.

Acetonitriletricarbonyl(2-chloro-8-quinolate)rhenium(1) ([Re(CO)₃(L3b)NCCH₃]). Amber solid; MP: 130 °C, 32% yield. ¹H NMR (400 MHz, CD₃CN, δ /ppm): δ 8.32 (d, $J^{\rm d}$ = 8.68 Hz, 1H), δ 7.58 (d, $J^{\rm d}$ = 8.68 Hz, 1H), δ 7.43 (t, $J^{\rm t}$ = 7.96 Hz, 1H), δ 7.01 (dd, $J^{\rm d}$ = 7.96, 0.96 Hz, 1H), δ 6.92 (dd, $J^{\rm d}$ = 7.96, 1.04 Hz, 1H), δ 1.96 (s, 3H). ¹³C{¹H} NMR (100 MHz, CD₃CN, δ /ppm): δ 196.6_{C=0}, δ 195.3_{C=0}, δ 195.3_{C=0}, δ 170.2, δ 152.3, δ 144.3, δ 142.6, δ 131.2, δ 130.1, δ 123.8, δ 120.8, δ 117.7, δ 112.4, δ 30.3. FTIR (ATR corr. $\tilde{\nu}$ /cm⁻¹): $\tilde{\nu}$ 2925.41 (w, C–H sp² str.), $\tilde{\nu}$ 2303.92 (w, N=C str.), $\tilde{\nu}$ 2016.96 (s, A'(1) C=O str.), $\tilde{\nu}$ 1871.84 (s, A'(2) & A" C=O str.). UV/Vis (λ /nm, ϵ /L mol⁻¹ cm⁻¹): λ 437 (d π \rightarrow π *, ϵ 235.06), λ 281 (π \rightarrow π *, ϵ 2340.84), λ 237 (π \rightarrow π *, ϵ 3104.80). EA (%): calc. C

Acetonitriletricarbonyl(5-chloro-8-quinolate)rhenium(1) ([Re(CO)₃(L3c)NCCH₃]). Yellow solid, MP: 159–170 °C (dec.), 26% yield. ¹H NMR (400 MHz, CD₃CN, δ/ppm): δ 9.98 (dd, J^d = 4.84, 1.24 Hz, 1H), δ 8.64 (dd, J^d = 8.64, 1.28 Hz, 1H), δ 7.64 (dd, J^d = 8.64, 4.84 Hz, 1H), δ 7.55 (d, J^d = 8.52 Hz, 1H), δ 6.85 (d, J^d = 8.52 Hz, 1H), δ 1.96 (s, 3H). ¹³C{¹H} NMR (100 MHz, CD₃CN, δ/ppm): δ 197.1_{C≡O}, δ 196.8_{C≡O}, δ 195.7_{C≡O}, δ 169.6, δ 151.0, δ 144.9, δ 136.5, δ 130.9, δ 128.9, δ 124.1, δ 115.4, δ 113.2. FTIR (ATR corr. $\tilde{\nu}$ /cm⁻¹): $\tilde{\nu}$ 2959 (w, C−H sp² str.), $\tilde{\nu}$ 2911 (w, C−H sp² str.), $\tilde{\nu}$ 2322 (w, N≡C str.), $\tilde{\nu}$ 2016 (s, A′(1) C≡O str.), $\tilde{\nu}$ 1874 (s, A′(2) & A″ C≡O str.).

34.32, H 1.65, N 5.72. Found C 34.89, H 1.54, N 5.60.

Acetonitriletricarbonyl(5-nitro-8-quinolate)rhenium(1) ([Re(CO)₃(L3d)NCCH₃]). Yellow solid, MP: 190–194 °C (dec.), 26% yield. ¹H NMR (400 MHz, CD₃CN, δ /ppm): δ 9.57 (dd, f^d = 9.00, 1.28 Hz, 1H), δ 9.03 (dd, f^d = 4.84, 1.28 Hz, 1H), δ 8.63 (d, f^d = 9.24 Hz, 1H), δ 7.81 (dd, f^d = 9.00, 4.84 Hz, 1H), δ 6.88 (d, f^d = 9.24 Hz, 1H). ¹³C{¹H} NMR (100 MHz, CD₃CN, δ /ppm): δ 196.3_{C=O}, δ 194.4_{C=O}, δ 178.1_{C=O}, δ 151.6, δ 143.3, δ 137.1, δ 132.9, δ 130.8, δ 127.0, δ 126.9, δ 121.4, δ 114.2. FTIR (ATR corr. $\tilde{\nu}$ /cm⁻¹): $\tilde{\nu}$ 2939 (w, C–H sp² str.), $\tilde{\nu}$ 2918 (w, C–H sp² str.), $\tilde{\nu}$ 2328 (w, N=C str.), $\tilde{\nu}$ 2019 (s, A'(1) C=O str.), $\tilde{\nu}$ 1900 (s, A'(2) & A" C=O str.).

Acetonitriletricarbonyl(2-fluoro-8-quinolate)rhenium(ι) ([Re(CO)₃(L3e)NCCH₃]). Amber solid; MP: 134 °C (dec.), 14% yield. ¹H NMR (400 MHz, CD₃CN, δ /ppm): δ 8.51 (dd, $J^{\rm d}$ = 8.92, 7.04 Hz, 1H), δ 7.43 (t, $J^{\rm t}$ = 7.96 Hz, 1H), δ 7.33 (dd, $J^{\rm d}$ = 8.92, 1.24 Hz, 1H), δ 7.06 (d, $J^{\rm d}$ = 8.00 Hz, 1H), δ 6.94 (d, $J^{\rm d}$ = 7.92 Hz, 1H). ¹³C{¹H} NMR (100 MHz, CD₃CN, δ /ppm): δ 195.9_{C=O}, δ 194.9_{C=O}, δ 194.2_{C=O}, δ 168.0 (d, $J^{\rm d}$ = 2 Hz), δ 161.5 (d, $J^{\rm d}$ = 256 Hz), δ 145.0 (d, $J^{\rm d}$ = 2 Hz), δ 139.7, δ 129.6 (d, $J^{\rm d}$ = 2 Hz),

δ 128.6, δ 116.5, δ 111.5, δ 109.8 (d, $J^d = 33 \text{ Hz}$). $^{19}\text{F}\{^1\text{H}\}$ NMR (376 MHz, CD₃CN, δ/ppm): δ −56.94. FTIR (ATR corr. $\tilde{\nu}/\text{cm}^{-1}$): $\tilde{\nu}$ 2925 (w, C–H sp² str.), $\tilde{\nu}$ 2853 (w, C–H sp² str.), $\tilde{\nu}$ 2322 (w, N≡C str.), $\tilde{\nu}$ 2020 (s, A'(1) C≡O str.), $\tilde{\nu}$ 1875 (s, A'(2) & A" C≡O str.).

Acetonitriletricarbonyl(5-fluoro-8-quinolate)rhenium(1) ([Re(CO)₃(L3e)NCCH₃]). Yellow solid, MP: 171–184 °C (dec.), 26% yield. ¹H NMR (400 MHz, CD₃CN, δ /ppm): δ 8.97 (dd, f^d = 4.84, 1.08 Hz, 1H), δ 8.55 (dd, f^d = 8.56, 1.12 Hz, 1H), δ 7.58 (dd, f^d = 8.56, 4.84 Hz, 1H), δ 7.26 (q, f^q = 10.52, 8.72 Hz, 1H), δ 6.78 (dd, f^d = 4.40, 8.72 Hz, 1H). ¹³C{¹H} NMR (100 MHz, CD₃CN, δ /ppm): δ 197.3_C= $_{\odot}$, δ 196.9_C= $_{\odot}$, δ 196.0_C= $_{\odot}$, δ 166.4 (d, f^d = 2 Hz), δ 151.3, δ 147.1 (d, f^d = 235 Hz), δ 143.0 (d, f^d = 5 Hz), δ 133.1 (d, f^d = 3 Hz), δ 123.3 (d, f^d = 2 Hz), δ 121.1 (d, f^d = 21 Hz), δ 144.5 (d, f^d = 20 Hz), δ 113.0 (d, f^d = 7 Hz). ¹°F{¹H} NMR (376 MHz, CD₃CN, δ /ppm): δ −144.8. FTIR (ATR corr. $\tilde{\nu}$ /cm⁻¹): $\tilde{\nu}$ 2954 (w, C-H sp² str.), $\tilde{\nu}$ 2909 (w, C-H sp² str.), $\tilde{\nu}$ 2298 (w, N=C str.), $\tilde{\nu}$ 2019 (s, A'(1) C=O str.), $\tilde{\nu}$ 1892 (s, A'(2) & A" C=O str.).

Synthetic protocols for fluorinated ligands

Fluorinated ligands (L1h, L1f, L2f, L2g and L3e-OBn) were prepared by azeotropically drying masses of potassium fluoride (12–25 eq.) and 18-crown-6 ether (4–10 eq.) via the dropwise addition of anhydrous acetonitrile in an inert nitrogen gas environment. A mass of the chlorinated or nitrated precursor (1.0 eq.) was dissolved in anhydrous DMSO and added to the complex. The solution was then heated to between 90-250 °C and stirred for 96–120 h. The reaction mixture was then cooled and filtered through a PTFE membrane (0.2 μ m). The filtrate was eluted over C₁₈ (40 g) using an acetonitrile/TFA (0.1% in water) gradient mobile phase. The eluents containing the product were subsequently combined and lyophilised to afford a pure white, pink or orange powder.

6-Fluoro-2,2'-bipyrdine (L2c). White solid; 13% yield. 1 H NMR (400 MHz, CDCl₃, δ /ppm): δ 8.74 (d, J^{d} = 4.20 Hz, 1H), δ 8.50 (d, J^{d} = 6.48 Hz, 1H), δ 8.45 (d, J^{d} = 8.04 Hz, 1H), δ 7.96 (q, J^{q} = 15.84, 8.00 Hz, 2H), δ 7.45 (d, J^{t} = 6.52 Hz, 1H), δ 7.00 (dd, J^{d} = 8.08, 2.80 Hz, 1H). 13 C{ 1 H} NMR (100 MHz, CDCl₃, δ /ppm): δ 163.3 (d, J^{d} = 237 Hz), δ 155.1 (d, J^{d} = 13 Hz), δ 154.7, δ 149.4, δ 142.0 (d, J^{d} = 7 Hz), δ 137.2, δ 124.3, δ 121.4, δ 118.3 (d, J^{d} = 4 Hz), δ 109.6 (d, J^{d} = 37 Hz). 19 F{ 1 H} NMR (376 MHz, CDCl₃, δ /ppm): δ -66.77. LRMS (ESI $^{+}$): [M + H] $^{+}$ m/z cald: 175.07, m/z obvs 175.14.

4-Fluoro-2,2'-bipyrdine (L1h). White solid; 42% yield. 1 H NMR (400 MHz, CDCl₃, δ /ppm): δ 8.87 (s, 1H), δ 8.79 (s, 1H), δ 8.62 (d, J^d = 5.58 Hz, 1H), δ 8.34 (t, J^t = 7.28 Hz, 1H), δ 7.70 (s 1H), 7.28 (s 1H). 13 C{ 1 H} NMR (100 MHz, CDCl₃, δ /ppm): δ 127.0, δ 169.4, δ 159.5 (d, J^d = 39 Hz), δ 158.7 (d, J^d = 39 Hz), δ 151.3, δ 146.7, δ 143.2, δ 125.7 (d, J^d = 239 Hz), δ 116.7, δ 113.8. 19 F{ 1 H} NMR (376 MHz, CDCl₃, δ /ppm): δ -75.99. LRMS (ESI⁺): [M + H]⁺ m/z cald: 175.07, m/z obvs 175.12.

2-Fluoro-1,10-phenanthroline (L2f). White solid; MP: 123 °C; 71% yield. ¹H NMR (400 MHz, CDCl $_3$, δ /ppm): δ 9.21 (dd, f^d = 4.32, 0.43 Hz, 1H), δ 8.37 (app. t, f^t = 8.44, 1H), δ 8.28 (dd, f^d = 8.12, 0.43 Hz, 1H), δ 7.83 (d, f^d = 21.16, 1H), δ 7.83 (d, f^d = 3.52 Hz, 1H), δ 7.66 (dd, f^d = 8.12, 1.08 Hz, 1H), δ 7.32 (dd, f^d = 8.56, 0.79 Hz, 1H). ¹³C{¹H} NMR (100 MHz, CDCl $_3$, δ /ppm):

δ 163.2, δ 160.8, δ 150.6, δ 145.2, δ 141.8 (d, $J^d=10.06$ Hz), δ 136.0, δ 129.2, δ 127.0, δ 126.0 (d, $J^d=2.01$ Hz), δ 125.6, δ 123.4, δ 111.3 (d, $J^d=41.25$ Hz). ¹⁹F NMR (376 MHz, CDCl₃, δ/ppm): δ -46.79 (dd, $J^d=3.27$, 10.05 Hz). FTIR (ATR corr. $\tilde{\nu}/\text{cm}^{-1}$): $\tilde{\nu}$ 3053 (w, C–H sp² str.), $\tilde{\nu}$ 3026 (w, C–H sp² str.), $\tilde{\nu}$ 1588 (s, C=C π_{Ar} str.), $\tilde{\nu}$ 1497 (s, C=C π_{Ar} str.), $\tilde{\nu}$ 1416 (s, C=C π_{Ar} str.). UV/Vis (CH₃CN, λ /nm, ϵ /L mol⁻¹ cm⁻¹): λ 262 ($\pi \to \pi^*$, ϵ 28 595.96), λ 227 ($\pi \to \pi^*$, ϵ 47 402.32), λ 200 ($\pi \to \pi^*$, ϵ 16 444.83). LRMS (ESI⁺): [M + H]⁺ m/z calc. 199.06660 (C₁₂H₈N₂F⁺); m/z obsv. 199.06673 (Δ 0.64 ppm); [M–F]⁺ m/z calc. 179.06092 (C₁₂H₇N₂⁺); m/z obsv. 179.06050 (Δ 2.36 ppm).

5-Fluoro-1,10-phenanthroline (L2g). White solid; MP, 7% yield. 1 H NMR (400 MHz, CDCl $_3$, δ /ppm): δ 9.48 (d, f^d = 4.68 Hz, 1H), δ 9.37 (dd, f^d = 4.56, 1.52 Hz, 1H), δ 8.82 (d, f^d = 8.08 Hz, 1H), δ 8.72 (dd, f^d = 8.36, 1.52 Hz, 1H), δ 8.12 (dd, f^d = 8.20, 5.04 Hz, 1H), δ 7.98 (dd, f^d = 8.36, 4.56 Hz, 1H), δ 7.73 (d, f^d = 9.44 Hz, 1H). 13 C{ 1 H} NMR (100 MHz, CDCl $_3$, δ /ppm): δ 155.8, δ 151.6, δ 145.7, δ 142.4 (f^d = 6 Hz), δ 140.8, δ 136.5, δ 131.8 (f^d = 4 Hz), δ 129.5 (f^d = 11 Hz), δ 125.8, δ 125.2, δ 122.8, δ 107.7 (f^d = 22 Hz). 19 F{ 1 H} NMR (376 MHz, CDCl $_3$, δ /ppm): δ -75.90. LRMS (ESI $^+$): [M-Cl] $^+$ m/z calc. 199.07, m/z obsv. 199.23.

2-Fluoro-8-benzoxyquinoline (L3e-OBn). Pink solid; MP: 107 °C, 26% yield. ¹H NMR (400 MHz, CDCl₃, δ /ppm): δ 8.22 (d, $f^{\rm d}=8.40$ Hz, 1H), δ 7.51 (d, $f^{\rm d}=7.40$ Hz, 1H), δ 8.35 (m, 5H), δ 7.10 (m, 2H), δ 5.42 (s, 1H). ¹³C{¹H} NMR (100 MHz, CDCl₃, δ /ppm): δ 162.0, δ 159.5, δ 153.6 ($f^{\rm d}=2$ Hz), δ 142.1 ($f^{\rm d}=9$ Hz), δ 137.7 ($f^{\rm d}=15$ Hz), δ 137.0, δ 128.8, δ 128.2 ($f^{\rm d}=2$ Hz), δ 128.0, δ 127.2, δ 126.3 ($f^{\rm d}=9$ Hz), δ 111.8, δ 110.9, δ 110.4. ¹⁹F {¹H} NMR (376 MHz, DMSO-d₆, δ /ppm): δ -61.07.

2-Chloro-8-benzoxyquinoline (L3b-OBn). A mass of L3b (2.4 g, 13 mmol, 1.0 eq.) was dissolved in a volume of anhydrous DMF (2.5 mL). A mass of potassium carbonate (3.7 g, 27 mmol, 2.0 eq.) was added. A volume of benzyl chloride (3.1 mL, 27 mmol, 2.0 eq.) was added dropwise to the solution. The reaction was stirred at 60 °C in an inert nitrogen environment (3 h). The solution was then cooled and extracted with DCM (20 imes10 mL) and the combined organic extracts were washed with brine (3 \times 20 mL). The organic layer was then dried over sodium sulphate, filtered and evaporated under reduced pressure. Recrystallisation from hot ethanolic solution afforded a pure pink product (3.0 g). 85% yield; MP: 101 °C; ¹H NMR (400 MHz, CDCl₃, δ /ppm): δ 8.06 (d, J^{d} = 8.60 Hz, 1H), δ 7.50 (d, J^{d} = 7.36 Hz, 1H), δ 7.40 (d, J^{d} = 8.56 Hz, 1H), δ 7.36 (m, 1H), δ 7.30 $(d, J^d = 7.06 \text{ Hz}, 1\text{H}), \delta 7.06 (dd, J^d = 6.08, 3.00 \text{ Hz}, 1\text{H}).$ ¹³C{¹H} NMR (100 MHz, CDCl₃, δ /ppm): δ 153.7, δ 150.0, δ 140.0, δ 138.7, δ 137.0, δ 128.7, δ 128.3, δ 127.9, δ 127.2, δ 127.1, δ 127.1, δ 123.1, δ 119.7, δ 111.8, δ 71.0.

2-Fluoro-8-hydroxyquinoline (L3e). A mass of L3e-OBn (218 mg, 0.859 mmol, 1.0 eq.) was dissolved in a volume of acetonitrile (6 mL). Masses of 10% wt palladium on activated carbon (123 mg) and 20% wt palladium hydroxide on activated carbon (125 mg) were added to the reaction in an inert nitrogen environment. The reaction flask was purged with hydrogen gas and left to stir (1 h). The solution was filtered over diatomaceous earth to afford a yellow filtrate. The filtrate was evaporated under reduced pressure and recrystallised from hot ethanolic

solution to afford a white powder (106 mg). 75% yield; MP: 62 °C; ¹H NMR (400 MHz, DMSO-d₆, δ /ppm): δ 9.89 (br. s, 1H), δ 8.51 (t, J^{t} = 8.60 Hz), δ 7.45 (m, 2H), δ 7.32 (dd, J^{d} = 8.80, 2.76 Hz, 1H), δ 7.14 (dd, J^{d} = 7.20, 1.64 Hz, 1H). ¹³C{¹H} NMR (100 MHz, DMSO-d₆, δ /ppm): δ 159.6 (d, J^{d} = 238 Hz), δ 152.5, δ 143.0 (d, J^{d} = 10 Hz), δ 135.4 (d, J^{d} = 15 Hz), δ 127.9, δ 126.9 (d, J^{d} = 2 Hz), δ 117.9, δ 113.5, δ 110.2 (d, J^{d} = 42 Hz). ¹°F{¹H} NMR (376 MHz, DMSO-d₆, δ /ppm): δ -63.53. FTIR (ATR corr. $\tilde{\nu}$ /cm⁻¹): $\tilde{\nu}$ 2473 (br. m, O-H str.).

All other ligands were acquired through commercially available sources (Sigma Aldrich or BePharm Ltd). Further details regarding ligand syntheses, including the experimental spectra, can be found in Section S.7 of the ESI.†

Radiolabelling protocols

Aqueous [18F]fluoride was produced on an IBA Cyclone 18 Twin cyclotron via the $^{18}O(p,n)^{18}F$ nuclear reaction. The $[^{18}F]$ fluoride was trapped by a QMA cartridge and eluted off with tetraethylammonium bicarbonate in 10% v/v water/acetonitrile for wet conditions, and azeotropically dried and reconstituted in anhydrous DMSO for dry conditions. Fig. 9 depicts the set-up of the synthesis module used to automate the radiolabelling reactions. Microfluidic radiosyntheses were performed in Discovery Mode using a NanoTek LF Microfluidic Synthesis System (Advion) connected to a standard laptop using NanoTek software v1.4.0 GMP Lite. Microreactors were made of fused silica tubing (100 μ m \times 2 m, 15.6 μ L) coiled tightly into a brass ring and held by a thermoresistant polymer. Radio HPLC analyses were carried out using a Shimadzu system comprised of a CBM-20 controller, LC-20AD pump, SIL-20AHT autoinjector, SPD-M20A PDA detector and a Lablogic Posi-RAM gamma detector. Analyses were performed using gradient conditions over 15 minutes, consisting of 95:5 water-: acetonitrile for the first 3 minutes followed by a linear transposition to 5:95 water: acetonitrile over the remaining 12 minutes using a 2 mL min⁻¹ flow rate. A Chromolith RP column (monolith system, Merck 50 × 4.6 mm), demonstrated to have <8% [18F]fluoride retention in low pH ranges,44 was employed as the stationary phase. Radio HPLC derived non-isolated RCYs were calculated via the integrated peak area ratio between the radio product and other radioactive species present, inclusive of any unreacted [18F]fluoride, using Laura V4.1.70 SP2 HPLC data analysis software. Further information on the radiochemical syntheses can be found in Section S.9 of the ESI.†

Computational protocols

Geometry optimisations and bond analyses were computed using ORCA software⁴⁵ at the DFT level using def2-TZVP basis sets and with the B97 density functional⁴⁶ corrected using the Grimme D3 correction with Becke–Johnson damping⁴⁷ to the DFT energy. The resolution of identity (RI) approximation⁴⁸ and the Effective Core potential LANL2DZ^{49,50} were also applied. The structures were confirmed to be local energy minima except for the transition states (one imaginary frequency rather than none). Thermodynamic properties were computed at the def2-TZVP/B97-D3 level of theory and calculated at 298 K. Zero-

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point energy corrections were included and the DMSO solvent was accounted for through the Continuum Polarizable Medium (PCM) model. ⁵¹ Natural Bond Order (NBO) analyses were performed using the NBO6 suite of programs. ⁵² The bond analyses between the rhenium(I) and diimine ligands were performed with the combined NOCV ⁵³ and CD ⁵⁴ frameworks. Within the NOCV framework, ^{39,55} the interaction between the [Re(CO)₃Cl] fragment (A) and the N,N diimine ligand fragment (B) were analysed by comparing the wavefunction of the adduct AB (ψ^{AB})

$$\Delta
ho = \sum_i \left|\psi_i^{
m (AB)}
ight|^2 - \sum_i \left|\psi_i^0
ight|^2$$

with the antisymmetrized and renormalized product of the

wavefunctions of the isolated fragments (ψ^0). The charge rear-

rangement $(\Delta \rho)$ upon formation of the interaction between A

and B fragments is defined through the following equation:

The charge displacement was separated in diagonal contributions through the valence operator (\hat{V}) , ⁵⁶⁻⁵⁸ producing pairs of complementary orbitals $(\varphi_k, \varphi_{-k})$ corresponding to eigenvalues with the same absolute value but opposite sign $(\pm v_k)$. Accordingly, $\Delta \rho$ can be expressed in terms of NOCV pairs via the following equation:

$$\Delta \rho = \sum_{k} v_{k} \left(\left| \varphi_{k} \right|^{2} - \left| \varphi_{-k} \right|^{2} \right) = \sum_{k} \Delta \rho_{k}$$

The different $\Delta \rho_k$ were then also integrated along the axis (z) aligned with the Re(1) and diimine N atom in order to quantify the fraction of electrons (Δq) involved in the $\varphi_{-k} \to \varphi_k$ transition and, therefore, in the bond component, throughout the whole molecule.

$$\Delta q_k = \int_{-\infty}^{z'} \mathrm{d}z \int_{-\infty}^{\infty} \Delta \rho_k \mathrm{d}x \, \mathrm{d}y$$

Atomic charges were calculated through the NBO. Further computational details can be found in Section S.5 of the ESI.†

Photophysical protocols

Photophysical analyses were performed using an Edinburg FLS980 photospectrometer. Light from a 450 W xenon arc lamp source was passed through excitation and emission monochromators to acquire the corresponding spectra. The quantum yields (ϕ) were calculated against a [Ru(BiPy)₃]Cl₂ reference solution (ϕ_r) *via* the following equation:

$$\phi = \phi_{ ext{r}} \left[rac{(I)(A_{ ext{r}})(n^2)}{(I_{ ext{r}})(A)(n_{ ext{r}}^2)}
ight]$$

where correction factors are applied for the integrated area of the emission spectra (*I*), absorbances of the solutions at the MLCT excitation wavelengths (*A*) and the refractive indices of the solvents (*n*) utilised.⁵⁹ Further information on the photophysical analyses can be found in Section S.6 of the ESI.†

Single crystal X-ray diffraction

Data were with collected using either an Oxford Rigaku Synergy-S employing confocal mirror monochromated Mo-K_{\alpha} radiation generated from a microfocus source (0.71073 Å) with ω and ψ scans at 100(2) K or at the MX1 beamline of the Australian Synchrotron with silicon double crystal monochromated radiation at 100(2) K ($\lambda = 0.7108 \text{ Å}$). Oata integration and reduction were undertaken with CrysAlisPro or XDS.61 Subsequent computations were carried out using Olex2.62 Structures were solved with ShelXT⁶³ and refined and extended with ShelXL⁶⁴ Carbon-bound hydrogen atoms were included in idealised positions and refined using a riding model. The instrument design of the Synchrotron beamline restricts data collection to rotation around a single axis which limits available completeness for low symmetry crystal systems. Ellipsoid plots for the five rhenium(1) complexes can be viewed in the ESI: Fig. S116-S120, Section S.10.† Crystallographic data is summarised below and the CIFs have been deposited at the Cambridge Crystallographic Data Centre with CCDC 1970989-1970993.

[Re(L3d)(CO)₃(NCCH₃)]; C₁₄H₈N₃O₆Re (M =500.44 g mol⁻¹): monoclinic, space group $P2_1/c$ (no. 14), a = 13.0671(4) Å, b = 15.9871(5) Å, c = 7.5731(2) Å, β = 96.415(3)°, V = 1572.14(8) Å³, Z = 4, T = 100.00(10) K, μ (MoKα) = 7.764 mm⁻¹, $D_{\rm calc}$ = 2.123 g cm⁻³, 19 869 reflections measured (5.096° ≤ 2 θ ≤ 56.562°), 3893 unique ($R_{\rm int}$ = 0.0967, $R_{\rm sigma}$ = 0.0600) which were used in all calculations. The final R_1 was 0.0390 (I > 2 σ (I)) and wR_2 was 0.0969 (all data).

[Re(L2b)(CO)₃Cl]; $C_{15}H_7Cl_2N_2O_3Re$ (M=520.33 g mol⁻¹): triclinic, space group $P\bar{1}$ (no. 2), a=8.1930(16) Å, b=8.6870(17) Å, c=10.559(2) Å, $\alpha=95.04(3)^\circ$, $\beta=93.13(3)^\circ$, $\gamma=97.76(3)^\circ$, V=740.1(3) Å³, Z=2, T=100(2) K, μ (synchrotron) = 8.586 mm⁻¹, $D_{\rm calc}=2.335$ g cm⁻³, 13 499 reflections measured ($3.882^\circ \le 2\theta \le 63.35^\circ$), 3590 unique ($R_{\rm int}=0.0627$, $R_{\rm sigma}=0.0533$) which were used in all calculations. The final R_1 was 0.0331 ($I>2\sigma(I)$) and wR_2 was 0.0858 (all data).

[Re(L2c)(CO)₃Cl]; $C_{13}H_7Cl_2N_2O_3$ Re ($M=496.3159 \text{ g mol}^{-1}$): triclinic, space group $P\bar{1}$ (no. 2), a=11.318(2) Å, b=12.872(3) Å, c=15.669(3) Å, $\alpha=102.17(3)^\circ$, $\beta=99.94(3)^\circ$, $\gamma=102.50(3)^\circ$, V=2120.8(8) Å³, Z=6, T=100(2) K, μ (synchrotron) = 8.981 mm⁻¹, $D_{\text{calc}}=2.331 \text{ g cm}^{-3}$, 31 430 reflections measured (3.354° $\leq 2\theta \leq 54.97^\circ$), 8464 unique ($R_{\text{int}}=0.0480$, $R_{\text{sigma}}=0.0438$) which were used in all calculations. The final R_1 was 0.0382 ($I>2\sigma(I)$) and w R_2 was 0.0975 (all data)

[Re(L1b)(CO)₃Cl]; $C_{13}H_7Cl_2N_2O_3Re$ (M=496.31 g mol⁻¹): monoclinic, space group $P2_1/c$ (no. 14), a=14.655(3) Å, b=10.996(2) Å, c=9.0920(18) Å, $\beta=107.76(3)^\circ$, V=1395.3(5) Å³, Z=4, T=100(2) K, μ (synchrotron) = 9.032 mm⁻¹, $D_{\rm calc}=2.363$ g cm⁻³, 22 464 reflections measured (2.918° $\leq 2\theta \leq 56.54^\circ$), 3218 unique ($R_{\rm int}=0.0448$, $R_{\rm sigma}=0.0259$) which were used in all calculations. The final R_1 was 0.0554 ($I>2\sigma(I)$) and w R_2 was 0.1411 (all data).

[Re(L1f)(CO)₃Cl]; $C_{13}H_7$ ClFN₂O₃Re ($M=479.86 \text{ g mol}^{-1}$): monoclinic, space group $P2_1/n$ (no. 14), a=7.9600(16) Å, b=17.171(3) Å, c=10.245(2) Å, $\beta=99.25(3)^\circ$, V=1382.1(5) Å³, Z=4, T=100(2) K, μ (synchrotron) = 9.007 mm⁻¹, $D_{\text{calc}}=2.306 \text{ g cm}^{-3}$, 19 995 reflections measured (4.674° $\leq 2\theta \leq 10.000$

54.968°), 2754 unique ($R_{\rm int}=0.0324$, $R_{\rm sigma}=0.0191$) which were used in all calculations. The final R_1 was 0.0226 ($I > 2\sigma(I)$) and w R_2 was 0.0683 (all data).

Conflicts of interest

The authors have no conflicts of interest to declare.

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References

- 1 S. M. Ametamey, M. Honer and P. A. Schubiger, *Chem. Rev.*, 2008, **108**, 1501–1516.
- 2 D. van der Born, A. Pees, A. J. Poot, R. V. A. Orru, A. D. Windhorst and D. J. Vugts, *Chem. Soc. Rev.*, 2017, 46, 4709–4773.
- 3 O. Jacobson, D. O. Kiesewetter and X. Chen, *Bioconjugate Chem.*, 2015, **26**, 1–18.
- 4 A. F. Brooks, J. J. Topczewski, N. Ichiishi, M. S. Stanford and P. J. H. Scott, *Chem. Sci.*, 2014, 5, 4545–4553.
- 5 T. Furuya, J. E. Klein and T. Ritter, Synthesis, 2010, 11, 1804– 1821.
- 6 A. J. Plamer, J. C. Clark and R. W. Goulding, Int. J. Appl. Radiat. Isot., 1977, 28, 53-65.
- 7 O. Jacobson and X. Chen, Med. Chem., 2010, 10, 1048–1059.
- 8 Y. Zhang and A. G. Horti, *J. Label. Compd. Radiopharm.*, 2004, 47, 947–952.
- 9 Y. Zhang, A. W. Hall and A. G. Horti, *J. Label. Compd. Radiopharm.*, 2004, 47, 385–392.
- 10 A. G. Horti, A. O. Koren, H. T. Ravert, J. L. Musachio, W. B. Mathews, E. D. London and R. F. Dannals, J. Label. Compd. Radiopharm., 1998, 41, 309–318.
- 11 Y. Zhang, O. A. Pavlova, S. I. Chefer, A. W. Hall, V. Kurian, L. L. Brown, A. S. Kimes, A. G. Mukhin and A. G. Horti, *J. Med. Chem.*, 2004, 47, 2453–2465.
- 12 Y.-S. Ding, N. Liu, T. Wang, J. Marecek, V. Garza, I. Ojima and J. S. Fowler, *Nucl. Med. Biol.*, 2000, 27, 381–389.
- 13 R. Schrirrmacher, U. Muhlhausen, B. Wangler, E. Schirrmacher, J. Reinhard, G. Nagel, B. Kaina, M. Piel, M. Wiebler and F. Rosch, *Tetrahedron Lett.*, 2002, 43, 6301–6304.
- 14 M. A. Klenner, G. Pascali, B. Zhang, T. R. Sia, L. K. Spare, A. M. Krause-Heuer, J. R. Aldrich-Wright, I. Greguric, A. J. Guastella, M. Massi and B. H. Fraser, *Chem.-Eur. J.*, 2017, 23, 6499–6503.

- 15 F. Cleeran, J. Lecina, J. Bridoux, N. Devoogdt, T. Tshibangu, C. Xavier and G. Bormans, *Nat. Protoc.*, 2018, 13, 2330–2347.
- 16 P. Laverman, W. J. McBride, R. M. Sharkey, D. M. Goldenberg and O. C. Boerman, J. Labelled Compd. Radiopharm., 2014, 57, 219–223.
- 17 S. H. Hausner, B. Bauer and J. L. Sutcliffe, *Nucl. Med. Biol.*, 2014, **41**, 43–50.
- 18 N. Malik, B. Baur, G. Winter, S. N. Reske, A. J. Beer and C. Solbach, *Mol. Imag. Biol.*, 2015, 17, 777–785.
- 19 Z. Li, T.-P. Lin, S. Liu, C.-W. Huang, T. W. Hudnall, F. P. Gabbi and P. S. Conti, *Chem. Commun.*, 2011, 47, 9324–9326.
- 20 S. Liu, D. Li, Z. Zhang, G. K. Surya-Praskash, P. S. Conti and Z. Li, *Chem. Commun.*, 2014, **50**, 7371–7373.
- 21 H. Kim, K. Kim, S.-H. Son, J. Y. Choi, K.-H. Lee, B.-T. Kim, Y. Byun and Y. S. Choe, ACS Chem. Neurosci., 2019, 10, 1445–1451.
- 22 B. Zhang, G. Pascali, N. Wyatt, L. Matesic, M. A. Klenner, T. R. Sia, A. J. Guastella, M. Massi, A. J. Robinson and B. H. Fraser, J. Labelled Compd. Radiopharm., 2018, 61, 847–856.
- 23 B. Zhang, B. H. Fraser, M. A. Klenner, Z. Chen, S. H. Liang, M. Massi, A. J. Robinson and G. Pascali, *Chem. - Eur. J.*, 2019, 25, 7613–7617.
- 24 M. A. Klenner, G. Pascali, B. Zhang, G. Ciancaleoni, M. Massi and B. H. Fraser, *Aust. J. Chem.*, 2019, 72, 288–294.
- 25 L. N. Winslow, D. P. Rillema, J. H. Welch and P. Singh, *Inorg. Chem.*, 1989, 28, 1596–1599.
- 26 J. R. Kirchhoff and K. Kirschbaum, *Polyhedron*, 1998, 17, 4033–4039.
- 27 A. Kochel, M. Holynska and K. Twarog, *Inorg. Chem. Commun.*, 2012, 24, 47–49.
- 28 M. Zhang, P. Lu, Y. Ma and J. Shen, *J. Phys. Chem. B*, 2003, **107**, 6535–6538.
- 29 M. I. Fernandez-Bachiller, C. Perez, G. C. Gonzalez-Munoz, S. Conde, M. G. Lopez, M. Villarroya, A. G. Garcia and M. I. Rodriguez-Franco, *J. Med. Chem.*, 2010, 53, 4927–4937.
- 30 C. Deraeve, M. Pitie, H. Mazarguil and B. Meuiner, *New J. Chem.*, 2007, **31**, 193–195.
- 31 C. Deraeve, C. Boldron, A. Maraval, H. Mazarguil, H. Gornitzka, L. Vendier, M. Pitie and B. Meuiner, *Chem.–Eur. J.*, 2008, **14**, 682–696.
- 32 F. Prati, C. Bergamini, R. Fato, O. Soukup, J. Korbecny, V. Andrisano, M. Bartolini and M. L. Bolognesi, *ChemMedChem*, 2016, 11, 1284–1295.
- 33 D. Antequera, M. Bolos, C. Spuch, C. Pascual, I. Ferrer, M. I. Fernandez-Bachiller, M. I. Rodriguez-Franco and E. Carro, *Neurobiol. Dis.*, 2012, 46, 682–691.
- 34 N. Vasdev, P. Cao, E. M. van Oosten, A. A. Wilson, S. Houle, G. Hao, X. Sun, N. Slavine, M. Alhasan, P. P. Antich, F. J. Bonte and P. Kulkarni, *MedChemComm*, 2012, 3, 1228– 1230.
- 35 S. H. Liang, J. P. Holland, N. A. Stephenson, A. Kassenbrock, B. H. Rotstein, C. P. Diagnault, R. Lewis, L. Collier, J. M. Hooker and N. Vasdev, ACS Chem. Neurosci., 2015, 6, 535–541.

- 36 S. H. Liang, A. G. Southon, B. H. Fraser, A. M. Krause-Heuer, B. Zhang, T. M. Shoup, R. Lewis, I. Volitakis, Y. Han, I. Greguric, A. I. Bush and N. Vasdev, ACS Med. Chem. Lett., 2015, 6, 1025–1029.
- 37 E. Horn and M. R. Snow, Aust. J. Chem., 1984, 37, 35-45.
- 38 E. Horn and M. R. Snow, Aust. J. Chem., 1981, 34, 737-743.
- 39 G. Bistoni, S. Rampino, F. Tarantelli and L. Belpassi, *J. Chem. Phys.*, 2015, **142**, 084112.
- 40 E. Portenkirchner, A. Apaydin, G. Aufischer, M. Havlicek, M. White, M. C. Scharber and N. S. Sariciftci, *ChemPhysChem*, 2014, 15, 3634–3638.
- 41 K. Kalyanasundaram, J. Chem. Soc., Faraday Trans. 2, 1986, 82, 2401–2415.
- 42 M. Wrighton and D. L. Morse, *J. Am. Chem. Soc.*, 1974, **96**, 998–1003.
- 43 H. C. Zhao, B. Mello, B.-L. Fu, H. Chowdhury, D. J. Szalda, M.-K. Tsai, D. C. Grills and J. Rochford, *Organometallics*, 2013, 32, 1832–1841.
- 44 D. Ory, J. Van den Brande, T. de Groot, K. Serdons, M. Bex, L. Declercq, F. Cleeren, M. Ooms, K. Van Laere, A. Verbruggen and G. Bormans, J. Pharm. Biomed. Anal., 2015, 111, 209–214.
- 45 F. Nesse, Wiley Interdiscip. Rev.: Comput. Mol. Sci., 2017, e1327.
- 46 A. D. Becke, J. Chem. Phys., 1997, 107, 8554-8560.
- 47 S. Grimme, J. Antony, S. Ehrlich and H. A. Krieg, J. Chem. Phys., 2010, 132, 154104.
- 48 R. A. Kendall and H. A. Fruchtl, *Theor. Chim. Acta*, 1997, **97**, 158–163
- 49 P. J. Hay and W. R. Wadt, J. Chem. Phys., 1985, 82, 299-310.
- 50 W. R. Wadt and P. J. Hay, J. Chem. Phys, 1985, 82, 284-298.

- 51 A. Klamt and G. Schuurmann, J. Chem. Soc., Perkin Trans. 2, 1993, 799–805.
- 52 E. D. Glendening, C. R. Landis and F. Weinhold, *J. Comput. Chem.*, 2019, **40**, 2234–2241.
- 53 M. Mitoraj and A. Michalak, J. Mol. Model., 2007, 13, 347–355.
- 54 L. Belpassi, I. Infante, F. Tarantelli and L. Visscher, *J. Am. Chem. Soc.*, 2008, **130**, 1048–1060.
- 55 A. Altun, F. Nesse and G. Bistoni, *J. Chem. Theory Comput.*, 2019, **15**, 215–228.
- 56 R. F. Nalewajski, A. M. Koster and K. Jug, *Theor. Chim. Acta*, 1993, 85, 463–484.
- 57 R. F. Nalewajski and J. Ozek, *Int. J. Quantum Chem.*, 1994, 51, 187–200.
- 58 R. F. Nalewajski, J. Mrozek and A. Michalak, *Int. J. Quantum Chem.*, 1997, **61**, 589–601.
- 59 P. J. Wright, S. Muzziloi, M. V. Werrett, P. Raiteri, B. W. Skelton, D. S. Silvester, S. Stagni and M. Massi, Organometallics, 2012, 31, 7566-7578.
- 60 T. M. McPhillips, S. E. McPhillips, H. J. Chiu, A. E. Cohen, A. M. Deacon, P. J. Ellis, E. Garman, A. Gonzalez, N. K. Sauter, R. P. Phizackerley, S. M. Soltis and P. J. Kuhn, J. Synchrotron Radiat., 2002, 9, 401–406.
- 61 W. J. Kabsch, J. Appl. Crystallogr., 1993, 26, 795-800.
- 62 O. V. Dolomanov, L. J. Bourhis, R. J. Gildea, J. A. K. Howard and H. J. Puschmann, J. Appl. Crystallogr., 2009, 42, 339–341.
- 63 G. M. Sheldrick, Acta Crystallogr., Sect. A: Found. Adv., 2015, 71, 3–8.
- 64 G. M. Sheldrick, Acta Crystallogr., Sect. C: Struct. Chem., 2015, 71, 3–8.