



Cite this: *Org. Biomol. Chem.*, 2017, **15**, 7122

Received 30th June 2017,

Accepted 9th August 2017

DOI: 10.1039/c7ob01593c

rsc.li/obc

## Ratiometric electrochemical detection of $\beta$ -galactosidase†

Sam A. Spring,  Sean Goggins \* and Christopher G. Frost 

**A novel ferrocene-based substrate for the ratiometric electrochemical detection of  $\beta$ -galactosidase was designed and synthesised. It was demonstrated to be an excellent electrochemical substrate for  $\beta$ -Gal detection with sensitivity as low as  $0.1 \text{ U mL}^{-1}$ .**

$\beta$ -Galactosidase ( $\beta$ -Gal, EC 3.2.1.23) is a prominent enzyme used biologically as a reporter gene as it has been well characterised and demonstrates excellent stability.<sup>1</sup> The low level of background substrate hydrolysis and ready availability makes  $\beta$ -Gal an attractive enzyme label within enzyme-linked immunosorbent assays (ELISAs).<sup>2</sup> It can also be used in heavy-metal ion detection,<sup>3</sup> and rapid enzyme assays have been used in the detection of coliform and *E. coli* in waste water treatment.<sup>4</sup>  $\beta$ -Gal is typically detected either chromogenically, using *ortho*-nitrophenyl- $\beta$ -galactoside (ONPG),<sup>3</sup> or 5-bromo-4-chloro-3-indoyl- $\beta$ -D-galactopyranoside (X-Gal),<sup>5</sup> or is detected fluorometrically using fluorescein-di- $\beta$ -D-galactopyranoside (FDG).<sup>6</sup> Lanthanide based coumarins have also been utilised as a luminescent probe in the detection of  $\beta$ -Gal.<sup>7</sup> Optical substrates however, are limited by the use of expensive equipment, non-linear fluorescence, and potentially high levels of background signal.<sup>1</sup> The development of electrochemical enzyme substrates allows for the direct conversion of a biochemical recognition event into an electrical signal enabling facile biosensor integration within a handheld device.<sup>8</sup> 4-Aminophenyl- $\beta$ -D-galactosidase (PAPG) is standardly used as an electrochemical substrate for  $\beta$ -Gal, but high background signals prevents accurate analysis at low enzyme concentrations.<sup>9</sup> A modified PAPG-style substrate, 4-methoxyphenyl- $\beta$ -D-galactosidase (4-MPGal) has a negligible background signal, but the unfavourable use of modified graphene oxide electrodes is required.<sup>10</sup>

Due to its facile oxidation potential and excellent synthetic utility, ferrocene is implemented as the redox-active moiety in

electrochemical probes,<sup>11,12</sup> with ferrocene derivatives widely used in biological systems due to their stability in aerobic and aqueous environments.<sup>13</sup> The development of ratiometric probes has overcome the issues of reproducibility, by the ability to obtain direct conversions, minimising both sampling errors and systematic errors from instrument variation.<sup>14</sup> Ferrocene-based ratiometric chemodosimeters for enzyme detection have previously been reported for alkaline phosphatase,<sup>15</sup> and glucose oxidase,<sup>16</sup> but none currently for  $\beta$ -Gal.

Ferrocene-based electrochemical sensing is a continuing interest within our research group as it enables an inexpensive and convenient way to monitor enzyme activity.<sup>17</sup> Utilising trigger-linker-effector methodologies,<sup>18</sup> we designed ferrocenylcarbamoylphenyl- $\beta$ -D-galactosidase **1** as a ratiometric electrochemical substrate for  $\beta$ -Gal. Previously, ferrocenylamine **3** has been shown to be oxidised at a lower potential than carbamate derivatives.<sup>12</sup> Substrate **1** would have a higher oxidation potential than **3** making them electrochemically distinguishable, allowing for the ratiometric electrochemical analysis of  $\beta$ -Gal activity. We propose that in the presence of  $\beta$ -Gal, hydrolysis at the anomeric position would afford an unstable phenolate intermediate **2**. 1,6-Elimination would follow releasing ferrocenylamine **3**, quinone methide and  $\text{CO}_2$  (Scheme 2).

The synthesis of substrate **1** (Scheme 1) started from the commercially available D-galactose pentaacetate which was converted to benzyl alcohol **5** (see ESI† for synthesis). Ferrocenyl azide **6** was synthesised according to a literature procedure,<sup>19</sup> and then coupled to the benzyl alcohol *via* a Curtius rearrangement. Zemplén deacetylation of **7** afforded the desired substrate **1** in a 16% overall yield. Once synthesised, substrate **1** was found to be a bench stable orange solid with no observable degradation over several months at room temperature. Substrate **1** was also stable to hydrolysis in Tris buffer (pH 7, 50 mM) solutions for several weeks at room temperature (see ESI, Fig. S1†). The electrochemical behaviour of substrate **1** was tested *via* differential pulse voltammetry (DPV) and compared to ferrocenylamine **3**.

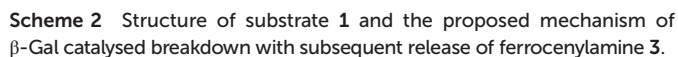
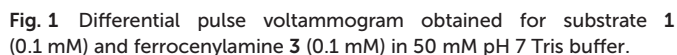
As expected, substrate **1** had a higher oxidation potential than ferrocenylamine **3**, with the difference between the two

Department of Chemistry, University of Bath, Bath, BA2 7AY, UK.

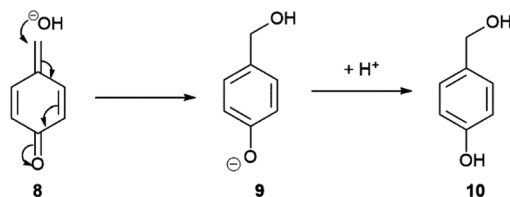
E-mail: s.goggins@bath.ac.uk; Fax: +44 (0)1225 386231; Tel: +44 (0)1225 386142

†Electronic supplementary information (ESI) available: Experimental procedures, characterisation data, and copies of NMR spectra of synthesised compounds. See DOI: 10.1039/c7ob01593c




$$\text{Conversion (\%)} = \frac{\left(\int 3\right)}{\left(\int 3 + \int 1\right)} \times 100 \quad (1)$$


Org. Biomol. Chem., 2017, 15, 7122–7126 | 7123



**Scheme 3** Proposed formation of 4-hydroxybenzyl alcohol from quinone methide.

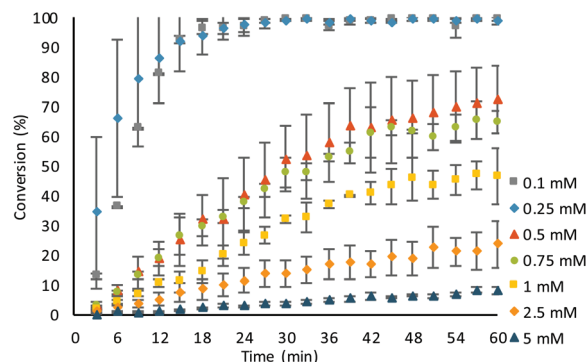
peaks showing the clear benefit of using the organometallic redox label compared with other commonly used organic ones.

Next, the effect of pH was investigated (see ESI, Fig. S4†). A  $\beta$ -Gal concentration of  $1 \text{ U mL}^{-1}$  was chosen as it allowed for both positive and negative effects due to the pH to be observed. At pH 8 there was a significant increase in the rate of conversion, with quantitative conversion observed in under 30 minutes. There was a marginal increase in rate from pH 8 to pH 7, but importantly, at the lower pH the presence of the third peak was suppressed, presumably due to protonation of the electrochemically active phenolate ion **9**. The suppression of the peak produced a cleaner voltammogram, allowing for more accurate conversion to be calculated and as a result, pH 7 was used moving forward.

According to previous literature, the optimum working temperature for  $\beta$ -Gal is  $37^\circ\text{C}$ .<sup>22</sup> A lower  $\beta$ -Gal concentration was chosen, specifically  $0.1 \text{ U mL}^{-1}$  in Tris buffer (pH 7, 50 mM) (see ESI, Fig. S5†), to allow for changes in the rate of conversion to be noticeable. Interestingly, increasing the temperature from room temperature had minimal effect on the rate of conversion, and above  $37^\circ\text{C}$  the rate was retarded. At  $57^\circ\text{C}$  negligible conversion was observed due to denaturing of the enzyme. Substrate **1**, however, remained stable to hydrolysis even at elevated temperatures, exhibiting the high stability of the substrate. With no improved rate of conversion, the assays were continued to be conducted at room temperature ( $21^\circ\text{C}$ ).

The concentration of substrate **1** in the assay was then screened, utilising a  $\beta$ -Gal concentration of  $1 \text{ U mL}^{-1}$  (Fig. 2). Increasing the probe concentration to  $0.25 \text{ mM}$  from  $0.1 \text{ mM}$  had minimal effects on the rate of conversion with quantitative conversion still observed within 24 minutes. However, increasing the concentration further showed no discernible increase in the rate of reaction. When the substrate concentration was decreased to  $0.05 \text{ mM}$ , the reduced current observed was susceptible to artefacts on the voltammogram affecting accurate conversion calculations, which were unavoidable when using disposable screen-printed carbon graphite electrode cells. At concentrations above  $0.1 \text{ mM}$ , additional sample manipulation, *via* serial dilutions, was required before analysis, due to overloading of the electrodes, and therefore, an optimal substrate concentration of  $0.1 \text{ mM}$  was chosen.

The final condition to be optimised was the buffer system used. Other common  $\beta$ -Gal buffer systems such as potassium phosphate buffer (50 mM, pH 7),<sup>3</sup> and Z-buffer (50 mM, pH 7),

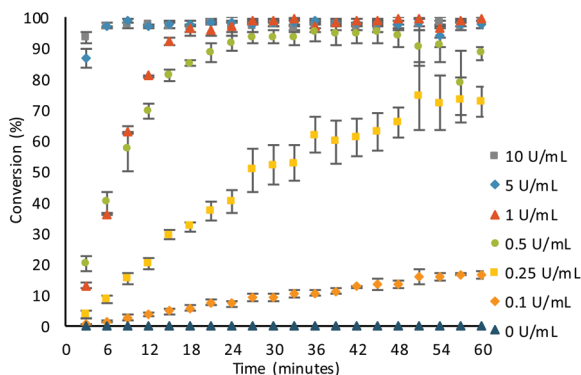


**Fig. 2** Conversion of the substrate **1** to the product after addition of  $\beta$ -Gal ( $1 \text{ U mL}^{-1}$ ) using different concentrations of the substrate at room temperature in Tris buffer (pH 7, 50 mM). Error bars represent the standard deviation where  $n = 3$ .

were tested. Z-buffer exhibited significant background signal, potentially due to the thiols present in Z-buffer, that obscured the peaks and prevented accurate electrochemical analysis. In phosphate buffer the stability of ferrocenylamine **3** was diminished with a second peak forming at a higher oxidation potential, assumed to be an electroactive by-product from ferrocenylamine decomposition.<sup>23</sup> Tris buffer (pH 7) was therefore chosen as this maintained a low background and ensured ferrocenylamine stability, and the effect of buffer concentration was investigated using a  $\beta$ -Gal concentration of  $1 \text{ U mL}^{-1}$  (see ESI, Fig. S6†). In unbuffered solution, the rate of conversion was significantly improved compared to 50 mM Tris buffer solution, however, the voltammogram peaks were shifted to a lower oxidation potential. At this lower potential, the presence of voltammogrammatic artefacts prevented accurate detection. At 25 mM Tris buffer concentration, a comparable rate of conversion to 50 mM Tris buffer concentration with quantitative conversion observed in 18 minutes. Further analysis showed that  $\beta$ -Gal was unstable at the lower buffer concentration, with increases in conversions stopping after 18 minutes. Increasing the concentration of the Tris buffer above 50 mM prevented accurate ratiometric analysis as we suspect the reduced stability of ferrocenylamine **3** resulted in unreliable peak integrations. Therefore, a Tris buffer (pH 7, 50 mM) was selected as the optimal buffer concentration.

With optimal conditions obtained, the sensitivity of  $\beta$ -Gal was tested (Fig. 3). In the optimised conditions, there was no background hydrolysis observed, allowing for detection of low  $\beta$ -Gal concentration of  $0.1 \text{ U mL}^{-1}$  within 60 minutes. Utilising pseudo-first order kinetics (see ESI, Fig. S7†), a rate constant of  $2.91 \times 10^{-3} \text{ s}^{-1}$  was calculated for a  $\beta$ -Gal concentration of  $1 \text{ U mL}^{-1}$ . At higher concentrations of 10 and  $5 \text{ U mL}^{-1}$ , quantitative conversions were exhibited within just 6 minutes, and at a low concentration of  $0.25 \text{ U mL}^{-1}$ , a  $73 \pm 5\%$  conversion was achieved within 60 minutes, with an observed rate constant of  $0.044 \times 10^{-3} \text{ s}^{-1}$ . The small error bars afforded, indicate the good reliability showing the benefit of using ratiometric electrochemical analysis.

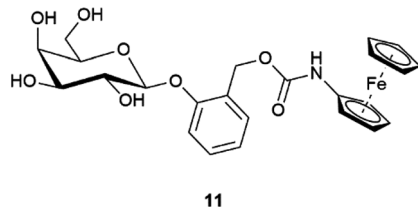




**Fig. 3** Conversion of the substrate **1** (0.1 mM) to the product after addition of varying concentration of  $\beta$ -Gal in Tris buffer (pH 7, 50 mM) at room temperature. Error bars represent the standard deviation where  $n = 3$ .

It has been previously shown that  $\beta$ -Gal has a large tolerance to the aglycon, as long as the D-galactose moiety remains untouched.<sup>24</sup> To further explore how the sterics of the substrate could impact  $\beta$ -Gal activity, substrate **11** was synthesised (Fig. 4), and tested utilising the optimal conditions (see ESI, Fig. S8†). Comparatively, substrate **11**, was significantly slower than substrate **1**. At low  $\beta$ -Gal concentration of  $0.1 \text{ U mL}^{-1}$  a conversion of  $12 \pm 6\%$  within 60 minutes, compared to a conversion of  $16 \pm 1\%$  for substrate **1**. The calculated rate constants of  $0.04 \times 10^{-3} \text{ s}^{-1}$  and  $0.06 \times 10^{-3} \text{ s}^{-1}$  for substrate **11** and **1** respectively (see ESI, Fig. S9†), show only a small difference in rate between the two regioisomers. This difference in rate of hydrolysis, is more significant at  $1 \text{ U mL}^{-1}$  concentrations, where the rate constant for substrate **1**,  $2.91 \times 10^{-3} \text{ s}^{-1}$ , is an order of magnitude higher than for substrate **11**,  $0.14 \times 10^{-3} \text{ s}^{-1}$ . The increased steric bulk around the anomeric position in substrate **11** inhibits the rate of hydrolysis, indicating the suitability of substrate **1**.

In conclusion, we have developed a new ferrocene-based electrochemical substrate for the detection of  $\beta$ -galactosidase activity. The substrate with a D-galactopyranoside trigger was synthesised and was distinguishable from the product electrochemically *via* differential pulse voltammetry. The substrate was shown to be stable to background hydrolysis, was demonstrated to be sensitive to low concentrations of  $\beta$ -galactosidase and shown to be both reproducible and reliable which makes  $\beta$ -galactosidase sensing applicable to electrochemical point-of-care biosensors.



**Fig. 4** Substrate **11**.

## Conflicts of interest

There are no conflicts to declare.

## Notes and references

- (a) J. Alam and J. L. Cook, *Anal. Biochem.*, 1990, **188**, 245; (b) L. H. Naylor, *Biochem. Pharmacol.*, 1999, **58**, 749.
- Z. Liu, T. Gurlo and H. von Grafenstein, *J. Immunol. Methods*, 2000, **234**, 153.
- S. M. Z. Hossain and J. D. Brenna, *Anal. Chem.*, 2011, **83**, 8772.
- (a) L. Fiksdal and I. Tryland, *Curr. Opin. Biotechnol.*, 2008, **19**, 289; (b) J. C. Jokerst, J. A. Adkins, B. Bisha, M. M. Mentele, L. D. Goodridge and C. S. Henry, *Anal. Chem.*, 2012, **84**, 2900.
- J. Kiernan, *Biotech. Histochem.*, 2007, **82**, 73.
- B. Rotman, J. A. Zderic and M. Edelstein, *Proc. Natl. Acad. Sci. U. S. A.*, 1963, **50**, 1.
- E. Pershagen and K. E. Borbas, *Angew. Chem., Int. Ed.*, 2015, **54**, 1787.
- J. Wang, *Biosens. Bioelectron.*, 2006, **21**, 1887.
- D. Wang, J. Chen and S. R. Nugen, *Anal. Chem.*, 2017, **89**, 1650.
- K. Manibalan, V. Mani, C.-H. Huang, S.-T. Huang and P.-C. Chang, *Analyst*, 2015, **140**, 6040.
- (a) P. D. Beer, *Chem. Soc. Rev.*, 1989, **18**, 409; (b) P. D. Beer, P. A. Gale and G. Z. Chen, *Coord. Chem. Rev.*, 1999, **185**, 3; (c) D. Kang, F. Ricci, R. J. White and K. W. Plaxco, *Anal. Chem.*, 2016, **88**, 10452.
- A. Sagi, J. Rishpon and D. Shabat, *Anal. Chem.*, 2005, **78**, 1459.
- D. R. van Staveren and N. Metzler-Nolte, *Chem. Rev.*, 2004, **104**, 5931.
- Y. Du, B. J. Lim, B. Li, Y. S. Jiang, J. L. Sessler and A. D. Ellington, *Anal. Chem.*, 2014, **86**, 8010.
- S. Goggins, C. Naz, B. J. Marsh and C. G. Frost, *Chem. Commun.*, 2015, **51**, 561.
- S. Goggins, E. A. Apsey, M. F. Mahon and C. G. Frost, *Org. Biomol. Chem.*, 2017, **15**, 2459.
- (a) S. C. Hillier, C. G. Frost, A. T. A. Jenkins, H. T. Braven, R. W. Keay, S. E. Flower and J. M. Clarkson, *Bioelectrochemistry*, 2004, **63**, 307; (b) S. C. Hillier, S. E. Flower, C. G. Frost, A. T. A. Jenkins, R. Keay, H. Braven and J. M. Clarkson, *Electrochem. Commun.*, 2004, **6**, 1227; (c) B. J. Marsh, L. Hampton, S. Goggins and C. G. Frost, *New J. Chem.*, 2014, **38**, 5260.
- (a) R. J. Amir, N. Pessah, M. Shamis and D. Shabat, *Angew. Chem., Int. Ed.*, 2003, **42**, 4494; (b) E. Sella and D. Shabat, *J. Am. Chem. Soc.*, 2009, **131**, 9934.
- D. C. D. Butler and C. J. Richards, *Organometallics*, 2002, **21**, 5433.
- $1 \text{ U mL}^{-1}$  = Units per millilitre. Where one unit will hydrolyse 1.0 micromole of *O*-nitrophenyl- $\beta$ -D-galactoside to *O*-nitrophenol and D-galactose per minute at pH 7.3 at 37 °C.



- 21 M. G. Willcockson, M. M. Toteva and V. J. Stella, *J. Pharm. Sci.*, 2013, **102**, 3579.
- 22 K. Wallenfels and R. Weil, in *The Enzymes*, ed. P. D. Boyer, Academic Press, New York, 3rd edn, 1972, vol. 20, p. 617.
- 23 (a) H. Hagen, P. Marzenell, E. Jentsch, F. Wenz, M. R. Veldwijk and A. Mokhir, *J. Med. Chem.*, 2012, **55**, 924; (b) P. Marzenell, H. Hagen, L. Sellner, T. Zeng, R. Grinyte, V. Pavlov, S. Daum and A. Mokhir, *J. Med. Chem.*, 2013, **56**, 6935; (c) M. Schikora, A. Reznikov, L. Chaykovskaya, O. Sachinska, L. Polyakova and A. Mokhir, *Bioorg. Med. Chem. Lett.*, 2015, **25**, 3447.
- 24 P. Fernandez, F. J. Cañada, J. Jiménez-Barbero and M. Martín-Lomas, *Carbohydr. Res.*, 1995, **271**, 31.

