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Non-classical crystallization of CeO₂ by means of *in situ* electron microscopy†

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During *in situ* liquid-phase electron microscopy (LP-EM) observations, the application of different irradiation dose rates may considerably alter the chemistry of the studied solution and influence processes, in particular growth pathways. While many processes have been studied using LP-EM in the last decade, the extent of the influence of the electron beam is not always understood and comparisons with corresponding bulk experiments are lacking. Here, we employ the radiolytic oxidation of Ce³⁺ in aqueous solution as a model reaction for the *in situ* LP-EM study of the formation of CeO₂ particles. We compare our findings to the results from our previous study where a larger volume of Ce³⁺ precursor solution was subjected to γ -irradiation. We systematically analyze the effects of the applied irradiation dose rates and the induced diffusion of Ce ions on the growth mechanisms and the morphology of ceria particles. Our results show that an eight orders of magnitude higher dose rate applied during homogeneous electron-radiation in LP-EM compared to the dose rate using gamma-radiation does not affect the CeO₂ particle growth pathway despite the significant higher Ce³⁺ to Ce⁴⁺ oxidation rate. Moreover, in both cases highly ordered structures (mesocrystals) are formed. This finding is explained by the stepwise formation of ceria particles via an intermediate phase, a signature of non-classical crystallization. Furthermore, when irradiation is applied locally using LP scanning transmission electron microscopy (LP-STEM), the higher conversion rate induces Ce-ion concentration gradients affecting the CeO₂ growth. The appearance of branched morphologies is associated with the change to diffusion limited growth.

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Radiation chemistry in aqueous solutions has been demonstrated to be a versatile method for tailored synthesis of nanomaterials.^{1–6} Upon exposure to ionizing radiation, such as high-energy electrons or gamma photons, water undergoes radiolysis producing oxidants (HO[•], HO₂[•], H₂O₂ and O₂) as well as reductants (e_{aq}[–], H[•] and H₂).^{7,8} The radical species formed upon radiolysis in general display high reactivity towards most solutes. The rate of radiolytic production of a specific species is given by the dose rate (depending on the activity or intensity of the radiation source) and the corresponding radiation chemical yield (*G*-value).^{9,10} The *G*-values depend on the type and the energy of the radiation. For gamma photons and high-energy electrons, the main products are HO[•] and e_{aq}[–], with *G*-values of 0.28 and 0.27 $\mu\text{mol J}^{-1}$, respectively. Depending on the type of ionic solution and the

targeted nanomaterial, the redox conditions can be optimized using radical scavengers. For the synthesis of a nanomaterial utilizing an oxidizing hydroxyl radical (HO[•]), the most convenient scavenger of the hydrated electron (e_{aq}[–]) is N₂O, which upon reaction with e_{aq}[–] produces HO[•] in aqueous solution. Hence, by saturating or continuously purging an aqueous solution with N₂O, the rate of hydroxyl radical production is almost doubled, while the possible interference from the strongly reducing hydrated electron on the nanomaterial synthesis is efficiently minimized. When reducing radicals are required to drive the synthesis (e.g., when synthesizing metal nanoparticles from metal ions in solution), hydroxyl radical scavengers are used. The most common scavengers for this purpose are 2-propanol and formate.¹¹

A fairly recent example of radiation induced synthesis of a nanomaterial utilizing the oxidative route is the production of CeO₂ from CeCl₃ aqueous solution.¹² In this process, hydroxyl radicals formed upon radiolysis of water, oxidize highly soluble Ce³⁺ to sparsely soluble Ce⁴⁺. The latter forms hydrated Ce(IV) hydroxides, which serve as intermediates in the liquid-to-solid phase transformation. The primary CeO₂ particles nucleate and grow (up to 3 nm in diameter) inside the intermediate gel-like phase. Thereafter, they mutually align, guided

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by the confinement of this phase, forming hierarchical structures, mesocrystals.^{13,14} It is worth mentioning that through the current manuscript, we will refer to the definition of the primary particles as nanoparticulate building blocks from which larger particles, aggregates, are formed. In the above mentioned study the aqueous solution of CeCl_3 was saturated with N_2O prior to irradiation. No other additives were used to control the growth of the particles. As the dose rate of the gamma source was relatively low (*ca.* 0.1 Gy s^{-1}), the time required to reach a significant conversion of the Ce^{3+} to Ce^{4+} was substantial (of about 20 hours). It was therefore possible to isolate samples from the solution at different stages of conversion and analyze them by *ex situ* electron microscopy (EM). Based on these measurements, a mechanism for CeO_2 mesocrystal formation was proposed.

Admittedly, this way of monitoring the reaction has certain drawbacks since all the samples are analyzed after evaporation of the solvent, which makes it difficult to confirm the exact speciation in solution, and avoid drying artifacts. A way to circumvent the problem of solvent evaporation is to use *in situ* liquid-phase electron microscopy (LP-EM)¹⁵ where the ionizing radiation from the electron beam in the electron microscope drive the radiolysis of the water contained in the liquid-cell (LC) sealed against vacuum.¹⁶ This opens up the possibility to directly investigate the primary particle formation and the subsequent phase formation in real time.^{17–23} In addition, the possibility to easily adjust dose rates (by varying the irradiation conditions) is another advantage of LP-EM, which is assumed to influence the conditions for the nucleation and growth of particles.^{24,25}

Note, that the irradiation conditions for conventional Gamma-Cell and for LP-EM are quite different. In fact, the dose rates differ by at least six orders of magnitude. As it was demonstrated previously, metal nanoparticle formation, driven by radiolytic reduction, is strongly dependent on irradiation dose rates.²⁶ Therefore, the results of the current study (using LP-EM) may differ significantly from the results of previous experiments on CeO_2 formation conducted under gamma radiolysis. Furthermore, the extremely high dose rates during LP-EM exposures will lead to rapid conversion of the dissolved species in the irradiated volume to less soluble ones, followed by sequential formation of the solid phase. Consequently, this may provoke diffusion of non-reacted ions into the irradiated volume from the surrounding non-irradiated volume.¹⁶ The diffusion will lead to inhomogeneous reaction conditions that affect the process of crystal growth.

In this work we explore a non-classical crystallization route using *in situ* LP-EM. We choose the formation of CeO_2 mesocrystals as a model process because of several reasons: (1) it allows us to use a water solution of a Ce^{3+} salt without any organic surfactants/additives; (2) radiolytic oxidation has been relatively little studied both in a LP-EM and upon gamma-radiation, and the formation mechanism in LP-EM conditions is not clear; (3) we compare a radiolytic process in a LP-EM confined cell and in a large volume of a Gamma-Cell¹² thus enabling a unique study of the influence of the irradiation

dose rate on nucleation and crystal growth by combining bulk and nanoscale approaches. Furthermore, we take advantage of the versatile irradiation conditions provided in LP-EM to study the influence of induced concentration gradients on the crystal growth during the radiolytic oxidation in solution.

In situ formation of CeO_2 mesocrystals by LP-EM applying homogeneous electron beam irradiation

A 5 mM aqueous solution of $\text{CeCl}_3 \cdot 7\text{H}_2\text{O}$ was purged with N_2O prior to liquid phase transmission electron microscopy (LP-TEM) investigations. Immediately after applying a homogeneous irradiation by the electron beam (dose rate of $16.7 \times 10^6 \text{ Gy s}^{-1}$, see ESI† for dose rate estimation) CeO_2 primary particles are formed and aggregated (Fig. 1). The aggregates grow upon continuous irradiation of solution. Fig. 1 presents images from a TEM LC with a thick layer of liquid in which many aggregates form. However, the achieved spatial resolution is insufficient to follow in detail the formation and growth of primary particles. Therefore, another experiment with a LC that contains a thinner layer of liquid is performed. The latter enables us to reach higher spatial resolution and thus, to track the development of primary particles (Fig. 2). The average diameter of primary particles develops from $3.9 \pm 0.5 \text{ nm}$ to $5.3 \pm 1.2 \text{ nm}$ (Fig. S1†), while the size of aggregates converges to approximately 25 nm (Fig. S1†).

In addition to *in situ* characterizations, LCs have been opened at the end of each experimental session and have been dried at environmental conditions in order to investigate them *ex situ* without superimposing signals from liquid and one of the two LC-membranes. Atomically resolved annular dark-field scanning transmission electron microscopy (ADF-STEM) imaging determines that the aggregates are composed by nanocrystals (Fig. 3a and S2†). The microstructure of CeO_2 formed during LP-TEM investigations strongly resembles the CeO_2 mesocrystals grown by gamma-radiation (Fig. 3b).¹² Moreover, the primary particles are found to mutually align in highly ordered structures (see Fourier transformation in Fig. S2†). Finally, energy dispersive X-ray spectroscopy (EDS) confirms the components of CeO_2 (Fig. S3†).

Radiolytic formation of CeO_2 primary particles *versus* metal nanoparticles

The size of the primary CeO_2 nanoparticles synthesized from 5 mM cerium precursor solution using gamma-radiation induced synthesis was found to be $\sim 3 \text{ nm}$, which is very close to what is found under the LP-TEM irradiation (previous section, Fig. S1† and Fig. 3b in ref. 12). This is quite surprising given the fact that for radiation synthesis of metal nanoparticles, the particle size displays strong dose rate dependence.^{11,27} The rationale for the latter is that the concen-





Fig. 1 Sequential images from an *in situ* LP-TEM movie showing the formation of aggregates (see video "V01_LP-TEM_CeO₂-nanoparticles.avi", time given as mm:ss). A dose rate of $16.7 \times 10^6 \text{ Gy s}^{-1}$ is applied.

tration of nucleation sites increases with increasing dose rate, which in turn leads to formation of more and smaller particles when those are immediately stabilized by surfactants against coalescence or other merging processes.

For CeO₂ synthesis, a dose rate difference by eight orders of magnitude does not have any considerable impact on the size of the primary particles. Thus, the rate-limiting step of the nucleation process must be different for CeO₂ compared to

metal nanoparticles and appears independent from the dose rate of the ionizing radiation in the studied range, and therefore of radical concentration. This is very well in line with the mechanism proposed for gamma-radiation induced synthesis of CeO₂ from Ce³⁺ in aqueous solution.¹² In this mechanism, nucleation takes place in a gel-like intermediate phase consisting of hydrated Ce(IV)-hydroxides formed after the radiolytic oxidation of Ce³⁺. This subsequent nucleation, followed by





Fig. 2 Sequential images from an *in situ* LP-TEM movie showing (a)–(b) formation and (c)–(f) arrangement of primary particles when a dose rate of $16.7 \times 10^6 \text{ Gy s}^{-1}$ is applied. Primary CeO_2 particles can be resolved and evaluated quantitatively, because of lower thickness of the liquid in comparison to Fig. 1. (See also according video “V02_LP-TEM- CeO_2 -nanoparticles”; time given as mm:ss).

growth of CeO_2 inside the intermediates, is not driven by radicals and, therefore, depend to a lesser extent on the irradiation dose rate.

Primary CeO_2 particle reorganization towards mesocrystals

Mutual alignment of the resolved primary CeO_2 particles can be recognized in the abovementioned observations (Fig. 1 and

2) and from *ex situ* structural and compositional analysis after opening the TEM LC (Fig. 3 and S2†). This mutual alignment of the primary particles is generally in agreement with previously presented *ex situ* interpretations.¹² In the current study, the relative orientations of primary particle in the aggregates (i), (ii) and (iii) in Fig. 2 (or in the video “V02_LP-TEM- CeO_2 -nanoparticles”) change, even though those particles are already connected to each other. Particle-particle interactions supposed to be present and may be described either by van der Waals forces, polarity forces, *etc.*,





Fig. 3 ADF-STEM images of CeO₂ grown (a) in LP-TEM (dose rate $16.7 \times 10^6 \text{ Gy s}^{-1}$) and (b) under steady-state gamma radiolysis.

or by the formation of atomic solid–solid interfaces.²⁸ If primary CeO₂ particles connect through a solid–solid interface formation, a reorganization may take place by deformation within the primary particle instead of a respective reorientation of the entire particle.^{29,30} If long-range interactions (*e.g.* van der Waals forces) are established, mutual alignment, during which entire primary particle moves, may take place. The aggregates may be kept together by a constrained environment (gel-like Ce^{IV}-hydroxide phase) in this case, as previously proposed.¹² Atomic resolution is not available within the used LC configuration *in situ*. Thus, no atomic solid–solid interface formation could be clearly stated between primary CeO₂ particles. Also, no gel-like constraining phase could be identified, because of limited contrast in LP-TEM imaging. However, single primary particles appear to be organized as a whole (Fig. 1 and 2) indicating that the latter description applies for the CeO₂ mesocrystal formation.

Comparison of CeO₂ aggregates growth in LP-TEM and low dose rate steady-state gamma radiolysis

The CeO₂ reaction pathway in LP-TEM appears to be not directly influenced by the dose rate. However, the higher concentration of radicals enables a faster consumption of Ce³⁺. This becomes obvious comparing the time difference of the mesocrystal formation under low dose rate steady-state gamma radiolysis (hours) and mesocrystal-like aggregates in LP-TEM (minutes). Nevertheless, the size saturation of aggregates obtained under steady-state gamma radiolysis and in LP-TEM (approximately 20–30 nm) is in the same order of magnitude

at the stage when all available Ce³⁺ is expected to be consumed. Generally, the size of transformed intermediate gel-like phase in solution determines the size of the formed aggregates.^{31,32} Our result suggests that these sizes of aggregates are not influenced by the different applied dose rates and therefore by radical concentrations.

Diffusion effects and formations of branched structures

By expanding the view from the central parts of the irradiated volumes in LP-TEM to their rims, aggregates of larger size are identified (see Fig. 4a). Differences in their size and shape can be caused by concentration gradients (Ce³⁺ ions) and explained as follows. The extreme irradiation conditions result in almost immediate depletion of the Ce³⁺ ions in the irradiated region. This depletion limits the growth of CeO₂ mesocrystals as described above. At the same time, the depletion results in Ce³⁺ concentration gradients between the irradiated and non-irradiated volumes of solutions. This concentration gradient leads to the diffusion of Ce³⁺ into the rim of the irradiated volume. That diffusion acts as additional Ce³⁺ supply. After entering the irradiated volume, radiolytic oxidation of Ce³⁺ can occur replenishing the existing or forming additional Ce(IV)-hydroxide phase in which CeO₂ nucleation takes place. This process ultimately leads to additional crystal growth.^{26,33}

Outside the irradiated volume, we observed formation of branched structures (Fig. 4b). Following the formation mechanism described above, Ce³⁺ ions cannot be oxidized outside the irradiated volume, because of a lack of radicals that are suspected to recombine faster than to diffuse outside the irra-





Fig. 4 LP-TEM images of (a) full view on irradiated liquid volume after approximately 40 min of exposure. (b) Branched structures appear outside the previously irradiated volume.

diated volume. That raises the general question, how CeO_2 can grow into a non-irradiated volume. In general, the interface between the irradiated and non-irradiated volume has significantly different properties than either of these volumes. Assuming an atomically sharp interface between irradiated and non-irradiated volume, only a diffusion (random walk) of formed CeO_2 particles or aggregates, or Ce(IV) -hydroxide could cause formation of branched CeO_2 morphologies in the non-irradiated volume. Knowing that the investigated sample is a confined liquid of not negligible thickness, an idea of atomically sharp interface does not hold. Thus, there might be a zone outside the directly irradiated volume in which significantly less, but still enough high energy electrons are scattered into^{34,35} providing a significant concentration of radicals to initiated oxidation of Ce^{3+} . The lower radical concentration in comparison to the directly irradiated volume causes a lower oxidation rate of Ce^{3+} . The slower transformation process is thereby effected by an already established Ce^{3+} concentration gradient and diffusion between non-irradiated and irradiated volume. This may cause the observed branched morphology,³⁶ because of additional, but most likely inhomogeneous Ce^{3+} supply. As soon as CeO_2 has formed, secondary effects, such as multiple scattering of high energy electrons, can be further enhanced since CeO_2 has a higher probability for scattering of those electrons than the liquid solution. Simulations would be needed to verify these suggestions or raising the necessity to find other explanations.

A more insightful experimental proof is limited by the fact that no *in situ* information about formations of such branched aggregates at the interface is available (no direct irradiation, no measureable signal for *in situ* LP-TEM data). However, one has to re-call that imaging by liquid phase scanning trans-

mission electron microscopy (LP-STEM) provides spatially and temporally non-uniform irradiation by a small converged probe with a diameter in the order of a nanometer (or less). Thus, the growth of branched structures, as observed in LP-TEM at the rim of the irradiated volume, is expected to be inducible by LP-STEM intrinsically. The dose rate within the converged beam can be even up to five orders of magnitude higher than that in the described LP-TEM (see dose rate estimations in ESI†). The irradiation can be compared to a consecutive pulse exposure, but with fairly extreme doses per pulse (as compared to irradiation with a pulsed electron beam from an accelerator, *e.g.* ref. 37). This enables principally a systematic *in situ* study of the influence of irradiation condition on diffusion effects. The dose rate within the converged electron beam can be directly modified by a change of the current in the electron beam. Or, the scan step distance (spatial distance between pulses) can be changed, which modifies overlaps or distances between pulses and, thereby, the dose rate applied in average on an exposed volume.³⁸

When applying a frame-averaged dose rate of $5.9 \times 10^6 \text{ Gy s}^{-1}$ in LP-STEM, the aggregates form immediately during the first irradiation scan, (Fig. 5a). Their number does not change upon further irradiation (Fig. 5b). However, their initially isotropic appearance becomes anisotropic (Fig. 5c) and eventually, branched structures form (Fig. 5d). The average size of the formed branched structures is significantly larger than the sizes of the previously observed mesocrystal-like aggregates (Fig. S4†). Note, that a large uncertainty in size is given by the strong variation in size of the six grown branched structures. More importantly, the average size does not converge towards a constant value during the irradiation period. The branched structures grow continu-





Fig. 5 Sequential *in situ* ADF imaging in LP-STEM. An average dose rate of $5.9 \times 10^6 \text{ Gy s}^{-1}$ is applied. (a) Already within the first scan, the aggregates become visible. (b) Continuous growth of aggregates while no formation of the additional ones is observed. (c) The appearance of the growing aggregates changes from isotropic to branched structures. (d) Further development of branched structures to larger sizes (video "V03_LP-STEM_CeO₂-nanoparticles.avi").

ously, although they have already reached sizes larger than the mesocrystals, formed and observed in LP-TEM. The formation of larger particles can again be attributed to additional supply of Ce^{3+} by diffusion effects.^{26,33} The diffusion effects (continuous supply of Ce^{3+} ions) are known to cause branched structures.²² Similar to our very initial comment on the origin of branched structure formation, diffusion limited aggregation of CeO_2 particles could basically also describe the observed process.^{39,40} However, no individual CeO_2 primary particles or aggregates are observed that merge to the growing branched structures in LP-STEM.

Isotropic growth in LP-STEM at lower average dose rates

When the frame-averaged dose rate is reduced by one order of magnitude to $2.4 \times 10^5 \text{ Gy s}^{-1}$ by expanding the scanning steps in LP-STEM, CeO_2 aggregates form as well (Fig. 6). However, their sizes are significantly smaller than the sizes of branched structures (Fig. S5†). Furthermore, their appearance remains isotropic and does not evolve to branched structures with longer exposure (Fig. 6d). The spatial separation of pulses





Fig. 6 Sequential *in situ* ADF imaging in LP-STEM. An averaged dose rate of $2.4 \times 10^5 \text{ Gy s}^{-1}$ is applied. (a) Initial scan. (b) Nano-sized aggregates become visible in the second scan and (c) their contrast increases during exposure. (d) The projected size of the aggregates increases constantly while almost no new ones are formed. (see video "V04_LP-STEM_CeO₂-nanoparticles.avi").

might reduce significantly the concentration gradients, and thereby diffusion effects.¹⁶ Potentially, conditions similar to those for uniform irradiation in LP-TEM are established. This statement is supported by the fact that the formed aggregates possess uniform size of about 30–40 nm, that is comparable to the size of mesocrystals (of about 25 nm) obtained in LP-TEM. However, the average size of aggregates appears to have not reached a constant value with increasing cumulative radiation dose (Fig. S5†). Thus, further Ce³⁺ diffusion from the outside of the irradiated liquid volume (also not previously irradiated) may still take place and effect the growth. The transition from the growth of branched structures to growth of spherical struc-

tures for lower dose rates in LP-STEM has been observed previously and described in the literature.²²

Interestingly, while only one kind of aggregates (spherical or branched structures) are observed in the LP-STEM depending on the applied dose rate (Fig. S6†), aggregates of various morphology are formed simultaneously in LP-TEM (Fig. 4). The latter is argued by the presence of both, negligible diffusion effects on the growth in the central part of homogeneously irradiated liquid volume and significant diffusion effects at the rim of the irradiated liquid volume. In contrast, diffusion effects may always play a key role in LP-STEM and growth of spherical-like or branched structures



can be adjusted in dependence of the in average applied dose rate.

Conclusions

CeO₂ nucleation and growth from aqueous CeCl₃·7H₂O solutions using ionizing electron irradiation have been studied using LP-EM. A direct comparison of experiments performed in LP-TEM and in a Gamma-Cell on identical material systems has been carried out. It is found that the orders of magnitudes higher dose rates in LP-TEM do not alter the CeO₂ multistep crystallization pathway (typical for non-classical crystallization) established earlier using low dose rate steady-state gamma-radiation.

The conversion rate of Ce³⁺ to Ce⁴⁺ is affected by the higher dose rates. Consequently, the aggregates grow much faster during LP-TEM experiments than during gamma-radiation induced synthesis. However, the sizes of the formed primary CeO₂ particles are close to each other, indicating a dose-rate-independent nucleation process of primary CeO₂ particles for the investigated dose rate range.

Furthermore, a gradient in size of the formed CeO₂ aggregates is determined from the rim of irradiated liquid volume to its center in LP-TEM. This is caused by diffusion of Ce³⁺ from the outside of irradiated liquid volume towards its center. Thus, an additional supply of Ce³⁺ can lead to continuous growth of the metal oxide aggregates and is therefore a parameter to tune their size.

Finally, we propose that depending on induced Ce³⁺ concentration gradients a change of the growth mode can be triggered from approximately spherical to complex branched nanostructure morphologies. The growth of those morphologies can be controlled and uniquely produced employing periodic pulsed irradiation in LP-STEM.

Methods

In the LP-EM experiments 5 mM aqueous solution of CeCl₃·7H₂O (98.5% purity, Merck), saturated with N₂O, were used. The solutions were load into a LC following Protochips' standard routine.⁴¹ The used LC consists of Protochips' "No spacer" bottom E-chip and Protochips' "Microwell" 8 × 16 top E-chip. Protecting layers on the E-chips were removed by washing in acetone and methanol. Then, E-chips were gently dried by compressed air. Then, E-chips have been plasma cleaned in order to make their surfaces hydrophilic and facilitate spreading of the dropped liquid. A liquid volume of 5 µl was deposited on a bottom chip. A fraction of solution was confined within the "microwell" chambers. LP-EM observations have been performed after successful leak check of the assembled LC EM-holder (Protochips' Poseidon).

LP-EM investigations were performed on a double C_s-corrected JEOL JEM-ARM200F operated at 200 kV and equipped with a cold-field emission gun (Emission current set to 5 µA).

Image acquisition in LP-TEM was done on Gatan's Oneview camera. Consecutive series of images (videos) have been acquired applying "in situ tools" within Gatan's Microscopy Suite (GMS) version 3.43. "In situ Editor" toolbox in the same software was used for post treatments. In LP-STEM, the sample was scanned by a probe with a convergence semi-angle of approximately 23 mrad. ADF-STEM images were collected with an angular collection semi angle range from 33 mrad to 125 mrad.

After, *in situ* investigations in the microscope, LCs were opened and samples were dried at air enabling *ex situ* investigations. For *ex situ* studies, E-chips have been re-assembled in the LC EM-holder with complementary (bottom or top) E-chips without SiN membrane in order to minimize thickness of the samples which causes scattering of the electron beam. EDS signal was collected on a high-angle silicon drift Energy Dispersive X-ray detector (solid angle up to 0.98 steradians with a detection area of 100 mm²).

Conflicts of interest

There are no conflicts to declare.

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References

- 1 Y. Lattach, C. Coletta, S. Ghosh and S. Remita, Radiation-Induced Synthesis of Nanostructured Conjugated Polymers in Aqueous Solution: Fundamental Effect of Oxidizing Species, *ChemPhysChem*, 2014, **15**, 208–2018, DOI: [10.1063/5.0019490](https://doi.org/10.1063/5.0019490).
- 2 K. Cubova and V. Cuba, Synthesis of inorganic nanoparticles by ionizing radiation - a review, *Radiat. Phys. Chem.*, 2020, **169**, 108774, DOI: [10.1016/j.radphyschem.2019.02.022](https://doi.org/10.1016/j.radphyschem.2019.02.022).
- 3 C. Dispenza, N. Grimaldi, M. A. Sabatino, I. L. Soroka and M. Jonsson, Radiation-Engineered Functional Nanoparticles in Aqueous Systems, *J. Nanosci. Nanotechnol.*, 2015, **15**(5), 3445–3467, DOI: [10.1166/jnn.2015.9865](https://doi.org/10.1166/jnn.2015.9865).
- 4 H. Remita, I. Lampre, M. Mostafavi, E. Balanzat and S. Bouffard, Comparative study of metal clusters induced in aqueous solutions by gamma-rays, electron or C6+ ion beam irradiation, *Radiat. Phys. Chem.*, 2005, **72**, 575–586, DOI: [10.1016/j.radphyschem.2004.03.042](https://doi.org/10.1016/j.radphyschem.2004.03.042).
- 5 A. Bharti, R. Bhardwaj, A. K. Agrawal, N. Goyal and S. Gautam, Monochromatic X-ray induced novel synthesis



- of plasmonic nanostructure for photovoltaic application, *Sci. Rep.*, 2016, **6**, 22394, DOI: [10.1038/srep22394](https://doi.org/10.1038/srep22394).
- 6 J. Belloni, Nucleation, growth and properties of nano-clusters studied by radiation chemistry Application to catalysis, *Catal. Today*, 2006, **113**, 141–156, DOI: [10.1016/j.cattod.2005.11.082](https://doi.org/10.1016/j.cattod.2005.11.082).
 - 7 C. D. Jonah, A short history of the radiation chemistry of water, *Radiat. Res.*, 1995, **144**(2), 141–147, <https://www.jstor.org/stable/3579253>.
 - 8 C. Ferradini and J.-P. Jay-Gerin, La radiolyse de l'eau et des solutions aqueuses: historique et actualité, *Can. J. Chem.*, 1999, **77**, 1542–1575, DOI: [10.1139/v99-162](https://doi.org/10.1139/v99-162).
 - 9 J. W. Spinks and R. J. Woods, *An Introduction to Radiation Chemistry*, Wiley, New York, 1990.
 - 10 M. Mostafavi and T. Douki, *Radiation Chemistry: From basics to applications in material and life sciences*, EDP Sciences, 2008.
 - 11 J. Belloni and M. Mostafavi, Radiation chemistry of nano-colloids and clusters, in *Radiation Chemistry: Present Status and Future Trends*, Elsevier, Amsterdam, 2001, pp. 411–452.
 - 12 Z. Li, D. Piankova, Y. Yang, Y. Kumagai, H. Zschiesche, M. Jonsson, N. V. Tarakina and I. L. Soroka, Radiation Chemistry Provides Nanoscopic Insights into the Role of Intermediate Phases in CeO₂ Mesocrystal Formation, *Angew. Chem., Int. Ed.*, 2022, **61**, 1–9, DOI: [10.1002/ange.202112204](https://doi.org/10.1002/ange.202112204).
 - 13 E. V. Sturm and H. Cölfen, Mesocrystals: Past, Presence, Future, *Crystals*, 2017, **7**, 207–224, DOI: [10.3390/cryst7070207](https://doi.org/10.3390/cryst7070207).
 - 14 H. Cölfen and M. Antonietti, *Mesocrystals and Nonclassical Crystallization*, John Wiley & Sons, Chichester, 2008.
 - 15 F. M. Ross, *Liquid Cell Electron Microscopy*, Cambridge University Press, New York, 2016.
 - 16 N. M. Schneider, M. M. Norton, B. J. Mendel, J. M. Grogan, F. M. Ross and H. H. Bau, Electron - Water Interactions and Implications for Liquid Cell Electron Microscopy, *J. Phys. Chem. C*, 2014, **118**, 22373–22382, DOI: [10.1021/jp507400n](https://doi.org/10.1021/jp507400n).
 - 17 E. A. Sutter and P. W. Sutter, Determination of Redox Reaction Rates and Orders by In Situ Liquid Cell Electron Microscopy of Pd and Au Solution Growth, *J. Am. Chem. Soc.*, 2014, **136**, 16865–16870, DOI: [10.1021/ja508279v](https://doi.org/10.1021/ja508279v).
 - 18 G. Zhu, Y. Jiang, F. Lin, H. Zhang, C. Jin, J. Yuan, D. Yang and Z. Zhang, In situ study of the growth of two-dimensional palladium dendritic nanostructures using liquid-cell electron microscopy, *Chem. Commun.*, 2014, **50**, 9447–9450, DOI: [10.1039/c4cc03500c](https://doi.org/10.1039/c4cc03500c).
 - 19 T.-Y. Ahn, S.-P. Hong, S.-I. Kim and Y.-W. Kim, In situ liquid-cell transmission electron microscopy for direct observation of concentration-dependent growth and dissolution of silver nanoparticles, *R. Soc. Chem. Adv.*, 2017, **5**, 82342–82345, DOI: [10.1039/c5ra14879k](https://doi.org/10.1039/c5ra14879k).
 - 20 T. J. Woehl and P. Abellan, Defining the radiation chemistry during liquid cell electron microscopy to enable visualization of nanomaterial growth and degradation, *J. Microsc.*, 2016, **265**, 135–147, DOI: [10.1111/jmi.12508](https://doi.org/10.1111/jmi.12508).
 - 21 Y. Wang, S. Wang and X. Lu, In Situ Observation of the Growth of ZnO Nanostructures Using Liquid Cell Electron Microscopy, *J. Phys. Chem. C*, 2018, **122**, 875–879, DOI: [10.1021/acs.jpcc.7b10064](https://doi.org/10.1021/acs.jpcc.7b10064).
 - 22 B. Jin, Z. Liu, R. Tang and C. Jin, Quantitative investigation of the formation and growth of palladium fractal nanocrystals by liquid-cell transmission electron microscopy, *Chem. Commun.*, 2019, **55**, 8186–8189, DOI: [10.1039/c9cc03161h](https://doi.org/10.1039/c9cc03161h).
 - 23 J. Cookman, V. Hamilton, L. S. Price, S. R. Hall and U. Bangert, Visualising early-stage liquid phase organic crystal growth via liquid cell electron microscopy, *Nanoscale*, 2020, **12**, 4636, DOI: [10.1039/c9nr08126g](https://doi.org/10.1039/c9nr08126g).
 - 24 B. Ambrozic, A. Prasnikar, N. Hodnik, N. Kostevsek, B. Likozar, K. Z. Rozman and S. Sturm, Controlling the radical-induced redox chemistry inside a liquid-cell TEM, *Chem. Sci.*, 2019, **10**, 8735–8743, DOI: [10.1039/c9sc02227a](https://doi.org/10.1039/c9sc02227a).
 - 25 D. Alloyeau, W. Dachraoui, Y. Javed, H. Belkhal, G. Wang, H. Lecoq, S. Ammar, O. Ersen, A. Wisnet, F. Gazeau and C. Ricolleau, Unravelling Kinetics and Thermodynamic Effects on the Growth of Gold Nanoplates by Liquid Transmission Electron Microscopy, *Nano Lett.*, 2015, **15**, 2574–2581, DOI: [10.1021/acs.nanolett.5b00140](https://doi.org/10.1021/acs.nanolett.5b00140).
 - 26 J. H. Park, N. M. Schneider, J. M. Grogan, M. C. Reuter, H. H. Bau, S. Kodambaka and F. M. Ross, Control of Electron Beam-Induced Au Nanocrystal Growth Kinetics through Solution Chemistry, *Nano Lett.*, 2015, **15**, 5314–5320, DOI: [10.1021/acs.nanolett.5b01677](https://doi.org/10.1021/acs.nanolett.5b01677).
 - 27 H. Fujita, M. Izawa and H. Yamazaki, Gamma-ray-induced Formation of Gold Sol from Chloroauric Acid Solution, *Nature*, 1962, **196**, 666–667, DOI: [10.1038/196666a0](https://doi.org/10.1038/196666a0).
 - 28 H. Cölfen and S. Mann, Higher-Order Organization by Mesoscale Self-Assembly and Transformation of Hybrid Nanostructures, *Angew. Chem., Int. Ed.*, 2003, **42**, 2350–2365, DOI: [10.1002/anie.200200562](https://doi.org/10.1002/anie.200200562).
 - 29 H. Zheng, R. K. Smith, Y.-w. Jun, C. Kisielowski, U. Dahmen and A. P. Alivisatos, Observation of Single Colloidal Platinum Nanocrystal Growth Trajectories, *Science*, 2009, **324**, 1309–1312, DOI: [10.1126/science.1172104](https://doi.org/10.1126/science.1172104).
 - 30 M. Yeadon, M. Ghaly, J. C. Yang, R. S. Averback and J. Gibson, “Contact epitaxy” observed in supported nanoparticles, *Appl. Phys. Lett.*, 1998, **73**, 3208–3210, DOI: [10.1063/1.122720](https://doi.org/10.1063/1.122720).
 - 31 M. Jehannin, A. Rao and H. Cölfen, New Horizons of Nonclassical Crystallization, *J. Am. Chem. Soc.*, 2019, **141**, 10120–10136, DOI: [10.1021/jacs.9b01883](https://doi.org/10.1021/jacs.9b01883).
 - 32 I. L. Soroka, K. K. Norrfor and N. V. Tarakina, Nonconvex Morphology of Metallic Copper Obtained via Nonclassical Crystallization of Copper Hydride, *Cryst. Growth Des.*, 2019, **19**, 5275–5282, DOI: [10.1021/acs.cgd.9b00689](https://doi.org/10.1021/acs.cgd.9b00689).
 - 33 M. R. Hauwiller, X. Zhang, W.-I. Liang, C.-H. Chiu, Q. Zhang, W. Zheng, C. Ophus, E. M. Chan, C. Czarnik, M. Pan, F. M. Ross, W.-W. Wu, Y.-H. Chu, M. Asta, P. W. Voorhees, A. P. Alivisatos and H. Zheng, Dynamics of Nanoscale Dendrite Formation in Solution Growth Revealed Through in Situ Liquid Cell Electron Microscopy, *Nano Lett.*, 2018, **18**(10), 6427–6433, DOI: [10.1021/acs.nanolett.8b02819](https://doi.org/10.1021/acs.nanolett.8b02819).



- 34 A. A. Sousa, M. F. Hohmann-Marriott, G. Zhang and R. D. Leapman, Monte Carlo electron-trajectory-simulations-in-bright-field-and-dark-field-STEM: Implications-for-tomography of thick biological sections, *Ultramicroscopy*, 2009, **109**, 213–221, DOI: [10.1016/j.ultramic.2008.10.005](https://doi.org/10.1016/j.ultramic.2008.10.005).
- 35 F. Wang, H.-B. Zhang, M. Cao, R. Nishi and A. Takaoka, Image blurring of thick specimens due to MeV transmission electron scattering: a Monte Carlo study, *J. Electron Microsc.*, 2011, **60**(5), 315–320, DOI: [10.1093/jmicro/dfp054](https://doi.org/10.1093/jmicro/dfp054).
- 36 L. M. Sander, Fractal growth processes, *Nature*, 1986, **322**, 789–793, DOI: [10.1038/322789a0](https://doi.org/10.1038/322789a0).
- 37 B. Dahlgren, M. A. Sabatino, C. Dispenza and M. Jonsson, Numerical Simulations of Nanogel Synthesis Using Pulsed Electron Beam, *Macromol. Theory Simul.*, 2020, **29**, 1900046, DOI: [10.1002/mats.201900046](https://doi.org/10.1002/mats.201900046).
- 38 J. Lee, D. Nicholls, N. D. Browning and B. L. Mehdi, Controlling radiolysis chemistry on the nanoscale in liquid cell scanning transmission electron microscopy, *Phys. Chem. Chem. Phys.*, 2021, **23**, 17766–17773, DOI: [10.1039/d0cp06369j](https://doi.org/10.1039/d0cp06369j).
- 39 H. Müller-Krumbhaar, M. Zimmer, T. Ihle and Y. Saito, Morphology and selection processes in diffusion-controlled growth patterns, *Phys. A*, 1996, **224**, 322–337, DOI: [10.1016/0378-4371\(95\)00390-8](https://doi.org/10.1016/0378-4371(95)00390-8).
- 40 N. Ahmad, Y. Le Bouar, C. Ricolleau and D. Alloyeau, Growth of dendritic nanostructures by liquid-cell transmission electron microscopy: a reflection of the electron irradiation history, *Adv. Struct. Chem. Imaging*, 2016, **2**, 1–10, DOI: [10.1186/s40679-016-0023-0](https://doi.org/10.1186/s40679-016-0023-0).
- 41 “Protochips”, 2023, [Online]. Available: <https://www.protochips.com>. [Accessed 02 04 2023].

