


Cite this: *RSC Adv.*, 2025, 15, 8213

Advances in isoxazole chemistry and their role in drug discovery

Glanish Jude Martis and Santosh L. Gaonkar *

Isoxazoles are a class of five-membered heterocyclic compounds that have gained significant attention in medicinal chemistry due to their diverse biological activities and therapeutic potential. Recent advances in isoxazole chemistry have led to the development of novel synthetic strategies, enabling the creation of a wide array of isoxazole derivatives with enhanced bioactivity and selectivity. This review explores the latest progress in isoxazole synthesis, highlighting key methodologies such as transition metal-catalyzed cycloadditions, green chemistry approaches, and regioselective functionalization techniques. These advances have not only improved the efficiency of isoxazole synthesis but have also facilitated the design of more complex and bioactive derivatives. In addition to their synthetic advances, isoxazoles have demonstrated a broad spectrum of biological activities, including antimicrobial, anticancer, anti-inflammatory, and neuroprotective effects, making them attractive candidates in drug discovery. This review discusses the structural modifications that enhance their pharmacological properties and their potential for developing therapies for diseases such as cancer, neurodegenerative disorders, and infections. Moreover, we examine the emerging trends in isoxazole-based drug discovery, such as the development of multi-targeted therapies and personalized medicine approaches. The evolving role of isoxazoles in drug discovery underscores their continued importance in modern pharmaceutical research and their potential to address unmet medical needs.

Received 25th November 2024
Accepted 3rd March 2025

DOI: 10.1039/d4ra08339c

rsc.li/rsc-advances

1 Introduction

Isoxazole is a five-membered heterocycle with one nitrogen and oxygen connected adjacent to each other. Analogous to this, there is another structure that is partially saturated, isoxazoline.

Department of Chemistry, Manipal Institute of Technology, Manipal Academy of Higher Education, Manipal, 576104, Karnataka, India. E-mail: sl.gaonkar@manipal.edu



Glanish Jude Martis

Glanish Jude Martis obtained his Master's degree in Organic Chemistry from Alva's College, Moodubidire, Karnataka, India (affiliated to Mangalore University) in 2023. He was the gold medallist in B.Sc (Biotechnology) in Mangalore University for the year 2021. Currently, he is pursuing his PhD as a Dr T. M. A. Pai fellow in the Department of Chemistry, Manipal Institute of Technology, Manipal Academy of Higher Education, Manipal, Karnataka, India under the supervision of Dr Santosh L. Gaonkar. His research interests include organic synthesis, green synthesis, computational design, medicinal chemistry, membrane technology and nanotechnology.



Santosh L. Gaonkar

Dr. Santosh L. Gaonkar received his PhD degree in Synthetic Organic Chemistry from the University of Mysore, India (2007). He was a JSPS Post-doctoral Fellow at AIST, Japan, where he worked on microwave-assisted synthesis of drug precursors (2008–2010). He also served as a Postdoctoral Fellow at AstraZeneca India, contributing to drug development research (2011). His research interests include Organic Synthesis, Bioorganic and Medicinal Chemistry, Drug Discovery and Development, and Material Chemistry. Currently, he is a Professor in the Department of Chemistry at Manipal Institute of Technology, MAHE, Manipal. He has over 100 research papers and six patents to his credit, with a Scopus h-index of 23.



Several biologically and pharmacologically active compounds have these moieties, which make them the best candidates for medicinal diversity.¹⁻⁴ In recent years, several strategies and

methods have been developed to synthesize isoxazole/isoxazoline derivatives. The formation of various fused heterocycles from isoxazoles is noteworthy and substantial.⁵ The

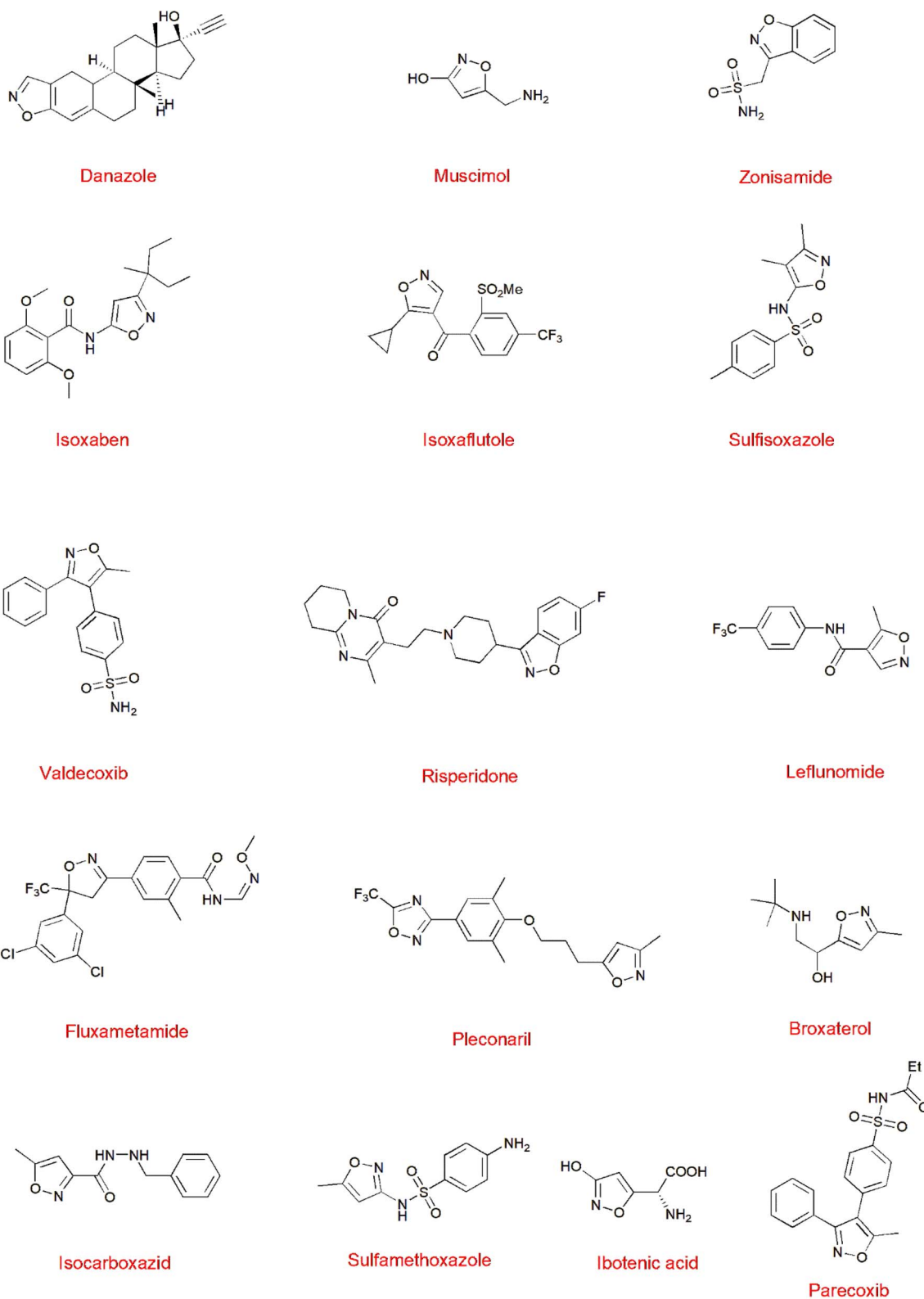
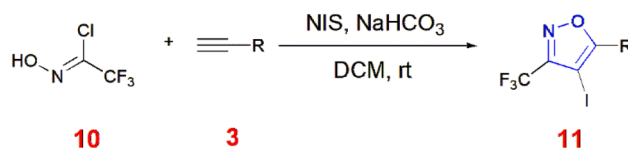


Fig. 1 Isoxazole-containing drugs.

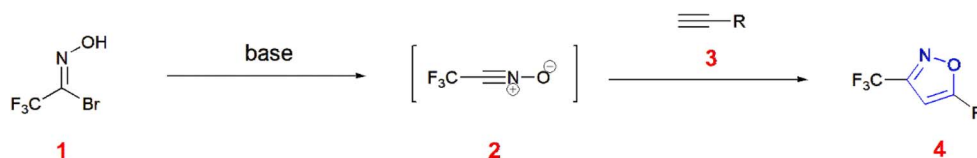
Review

reason for this approach is that these derivatives possess various biological properties such as antioxidant,^{6,7} antibacterial,^{8,9} antifungal,^{10,11} anticancer,^{12,13} insecticidal,¹⁴ anti-inflammatory,^{15,16} antidiabetic,^{17,18} and analgesic¹⁹ properties. These derivatives are also effective against Alzheimer's disease.^{20,21} In particular, isoxazole has gained much attention and importance because of its electron-rich aromatic structure. Moreover, the weak nitrogen and oxygen bond is attributed to ring cleavage reactions.^{22,23} Currently, isoxazole is present not only in pharmaceuticals, but also in natural products and agrochemicals.²⁴ Direct extraction of chemical constituents from plant sources results in low pharmacological effects. Thus, it is necessary for inducing structural modifications to improve their biological activity, pharmacological efficiency and drug



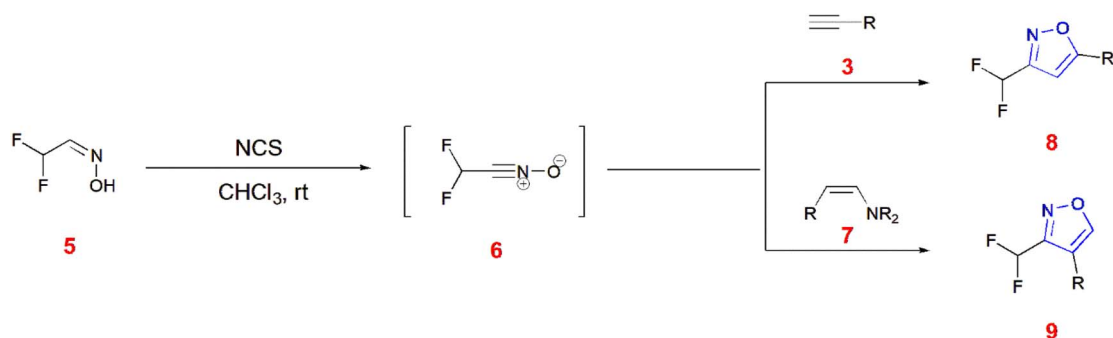
Scheme 3 Synthesis of 3-trifluoromethyl-4-iodoisoxazole 11.

selectivity. By doing so, it is also possible to have different activities from the parent source.²⁵ Isoxazoles are found in various pharmaceutical drugs such as danazol,²⁶ muscimol,^{27,28} zonisamide,²⁹ isoxaben,³⁰ isoxaflutole,³¹ sulfisoxazole,^{32,33} val-decoxib,^{34,35} fluxametamide,^{36,37} risperidone,^{38,39} leflunomide,⁴⁰

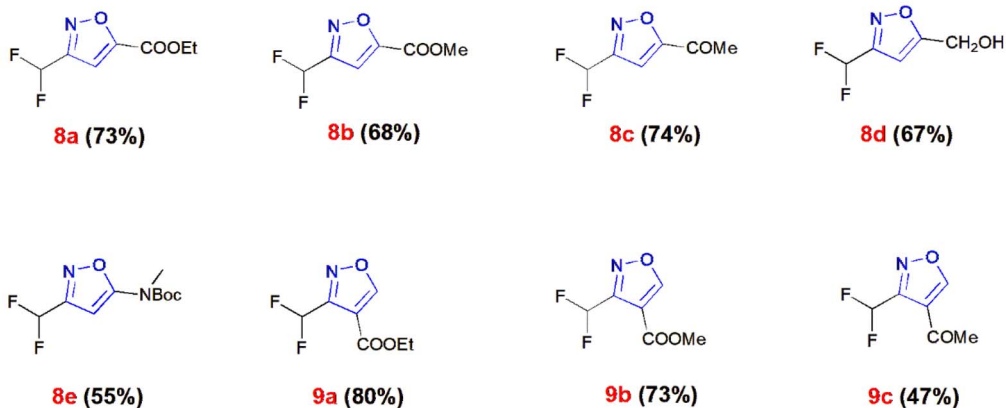


R = Aromatic: NEt₃/Toluene; 60 - 99%
R = Aliphatic: Na₂CO₃/H₂O; 54 - 77%

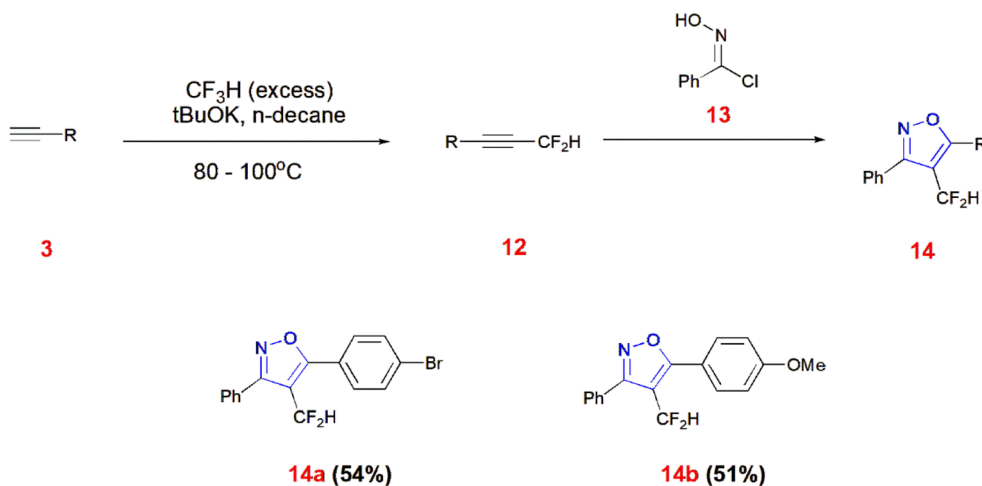
Scheme 1 Synthesis of 3-trifluoromethyl-5-substituted isoxazole derivatives 4.



Selected Examples



Scheme 2 Synthesis of 3-difluoromethyl isoxazole derivatives 8 and 9.



Scheme 4 Synthesis of 4-difluoromethyl isoxazoles 14.

pleconaril,^{41,42} broxaterol,⁴³ isocarboxazid,⁴⁴ sulfamethoxazole,⁴⁵ ibotenic acid,⁴⁶ and parecoxib.⁴⁷ The substituents present on the isoxazole moiety play a vital role in complex formation, especially, when functional groups are present.^{48,49} Thus, the structure of isoxazoles has undergone several modifications and alterations to increase their biological activity.⁵⁰ However, by including isoxazole in medicinal targets, there can be improvement in pharmacokinetic profiles, increased efficacy and decrease in toxicity.^{51,52} However, the scope of functionalized isoxazoles is wide and has delivered promising results.⁵³ (Fig. 1).

2 Synthetic strategies for isoxazoles and their derivatives

2.1 Cycloaddition reactions

2.1.1 1,3 Dipolar cycloaddition. Many heterocycles today are the result of cycloaddition reactions. Among such molecules, isoxazoles result from 1,3 dipolar cycloaddition reactions. Similarly, Ley and co-workers synthesized 3-trifluoromethylisoxazoles **4** from hydroxymoyl bromide **1**. The dipole nitrile oxide **2** is formed from the precursor hydroxymoyl bromide **1** and this dipole undergoes cycloaddition with the substituted terminal alkynes **3** to give 3-trifluoromethyl-5-substituted isoxazole derivatives **4**. The combination of the solvent systems in association with suitable bases significantly increased the yield and accelerated the cycloadditions. Triethylamine/toluene acts as a good system when phenylacetylene, 4-bromophenyl acetylene, 4-methoxy acetylene are used and sodium carbonate/water is used for cyclopropyl acetylene and cyclopentylacetylene.⁵⁴ (Scheme 1).

Mykhailiuk and co-workers generated difluoromethyl isoxazoles **8** and **9** from difluoro oxime **5** reacting with *N*-chlorosuccinimide (NCS) in the presence of chloroform as solvent to give the dipole difluoro nitrile oxide **6**. This dipole **6** reacts with terminal alkynes **3** and enamines **7** to give 3,5-disubstituted **8** and 3,4-disubstituted isoxazoles **9**, respectively.⁵⁵ (Scheme 2).

Wu *et al.*, synthesized 3-trifluoromethyl-4-iodoisoxazoles **11** from trifluoroacetoxyhydroxymoyl chloride **10**, terminal alkynes **3** and *N*-iodosuccinimide in a one-pot reaction containing sodium bicarbonate as the base and dichloromethane as the solvent at room temperature. This reaction afforded up to 81% yield.⁵⁶ (Scheme 3).

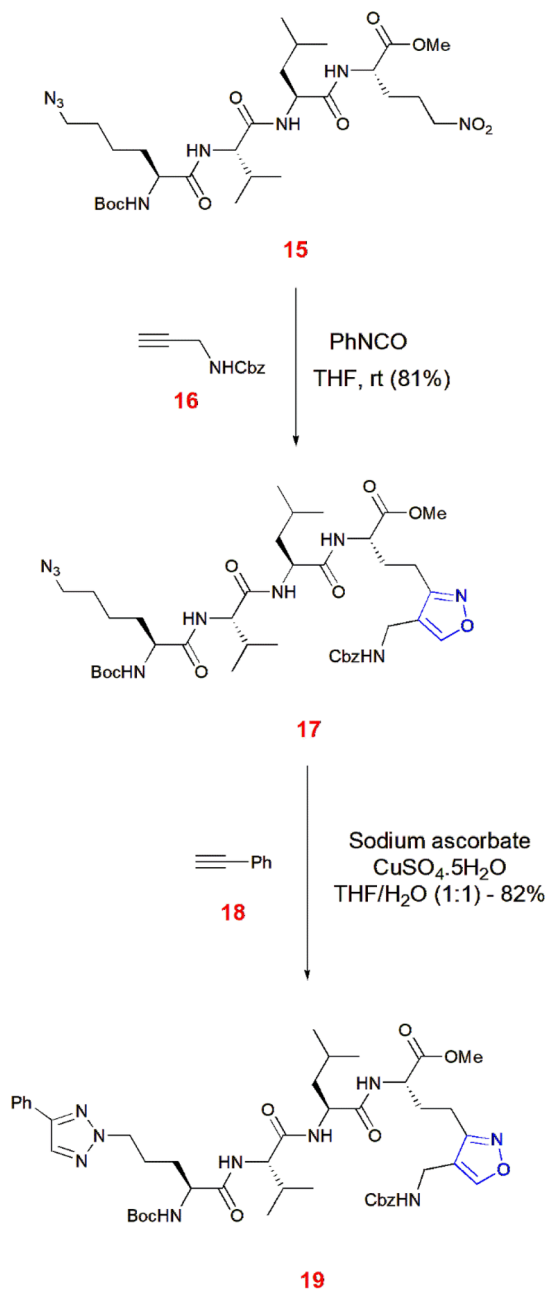
Shibata *et al.*, reported the synthesis of 4-difluoromethyl isoxazoles **14** from difluoromethyl alkynes **12** and imidoyl chloride **13** using triethylamine as the base with dichloromethane. Difluoromethyl alkynes **12** were obtained from terminal alkynes **3** via reaction with fluoroform (source of difluorocarbene) in the presence of potassium *tert*-butoxide and *n*-decane heated at 80–100 °C for 3 h.⁵⁷ (Scheme 4).

Gopi and co-workers illustrated an orthogonal cycloaddition reaction leading to the formation of doubly conjugated peptide **17** bearing isoxazole and triazole moieties. First, nitro-alkane tethered peptide **15** was treated with phenyl isocyanate and *N*-Cbz-propargylamine **16** in THF at room temperature to yield 81% of the peptide containing isoxazole **17**. This isoxazole scaffold **17** was made to undergo alkyne-azide cycloaddition with phenylacetylene **18** in the presence of sodium ascorbate, CuSO₄·5H₂O in equimolar THF/water to yield 82% of triazole-isoxazole peptide **19**.⁵⁸ (Scheme 5).

Nitrile oxides obtained from imidoyl chloride **13** undergo 1,3-dipolar cycloaddition of mono- or di-fluorinated propargyl thioethers **21** catalyzed by a copper sulfate/sodium ascorbate line to give isoxazoles **22**. First, propargyl thioethers **20** are electrochemically reacted with HF salt to give partially fluorinated terminal alkynes **21**.⁵⁹ (Scheme 6).

Yang *et al.*, demonstrated the generation of nitrile oxide from 2-ethylazaarene **23**, KNO₃ and K₂SO₈ by selective oxidation. Then it underwent copper-catalyzed 1,3-dipolar cycloaddition reaction with terminal alkynes **3** to give quinoline-isoxazole derivatives **24**. The plausible mechanism shows the role of KNO₃ and K₂SO₈ in giving products along with Cu catalyst.⁶⁰ (Scheme 7).





Scheme 5 Orthogonal cycloaddition giving isoxazole-triazole peptide 19.

Kore and co-workers illustrated the 1,3-dipolar cycloaddition of 3'-O-propargyl guanosine with chloroximes 27 to give isoxazoles 28 in the presence of triethylamine at room temperature. Prior to this, different aldioximes 25 were chlorinated with *N*-chlorosuccinimide 26 in DMF at room temperature to generate chloroximes 27. The different derivatives and their yields are given in Table 1.⁶¹ (Scheme 8).

2.1.2 [3 + 2] cycloaddition. Chen and co-workers reported a method of Cu(I)-free [3 + 2] cycloaddition between nitrile oxide

and electron-rich terminal ynammides 29 to give 3,5-disubstituted isoxazoles 30 with proposed mechanism. Nitrile oxides are generated from chloroximes 27 upon treatment with sodium carbonate.⁶² (Scheme 9).

Isoxazoles were generated from potassium poly(heptazine imide) (K-PHI) after artificial photosynthesis to O₂ by maintaining a pressure of 1 bar at 461 nm. Next, the aldioximes 25 were quenched with ¹O₂ to form nitrile oxides, which react with alkyl nitriles 31 in a [3 + 2] cycloaddition fashion to give isoxazoles 32.⁶³ (Scheme 10).

Zhao *et al.*, proposed another method involving copper catalyzed [3 + 2] cycloaddition of phenylacetylene 18 with nitrile oxides derived from nitroso radical and copper carbene. A three component reaction of 18 with *tert*-butyl nitrite 33 and ethyl diazoacetate 34 in the presence of Cu(OAc)₂ · H₂O as catalyst and 1,4-diazabicyclo[2.2.2]octane (DABCO) as base in toluene at 130 °C gave 3,5-disubstituted isoxazole 35.⁶⁴ (Scheme 11).

2.1.3 [2 + 1 + 1 + 1] cycloaddition. Chen *et al.*, reported a multicomponent reaction involving [2 + 1 + 1 + 1] cycloaddition where fluorobutyl iodide 36 reacts with the catalyst Co(II) to generate corresponding radical A, which acts upon styrene 37 to give another radical B. Coupling this radical B with *tert*-butyl peroxy radical 39 gives β-difluoro peroxide C. The Kornblum-DeLaMare rearrangement of this peroxide C catalyzed by DABCO led to the formation of an intermediate D having a carbonyl group that undergoes DABCO-promoted HF elimination to give unsaturated compound E. This unsaturated compound when treated with sodium azide 38, gives per-fluoroalkyl isoxazole ring 40.⁶⁵ (Scheme 12).

Tang *et al.*, annulated sulfoxonium ylides 41 with *tert*-butyl nitrite 42 catalyzed by Cu(TFA)₂ with sodium acetate as the base and dioxane as the solvent heated at 80 °C for 12 h to give isoxazoles 43. This was one of the novel preparations of isoxazole cores involving [2 + 1 + 1 + 1] cycloaddition.⁶⁶ (Scheme 13).

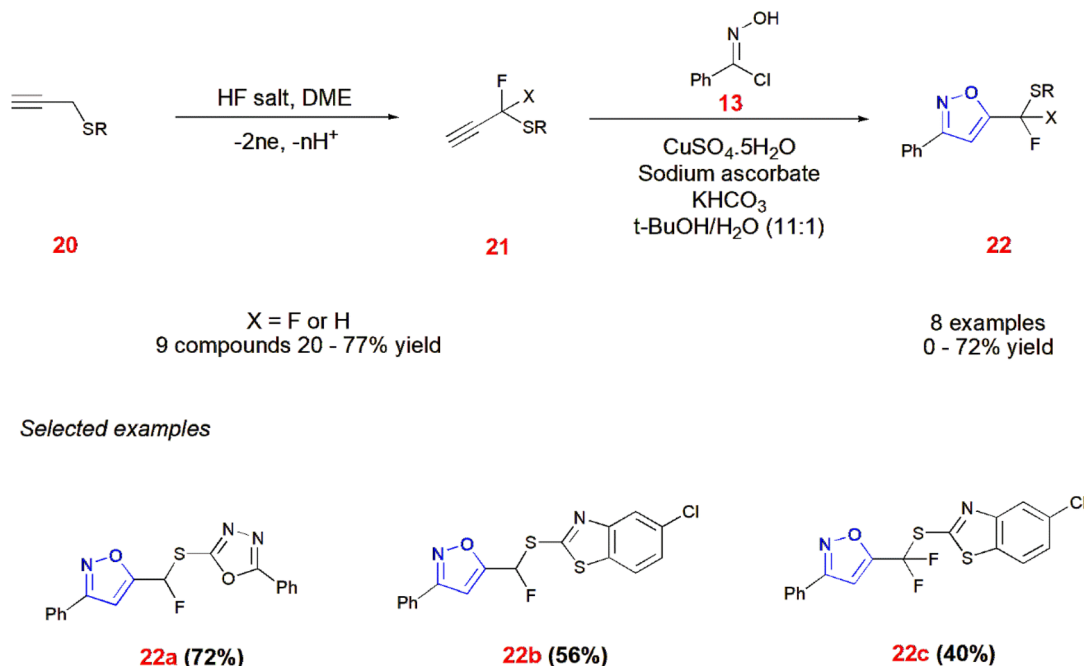
2.2 Condensation reactions

Langer and co-workers synthesized 5-trifluoromethyl isoxazoles 48 from hydrazone dianions 45, which were prepared from the reaction of *n*-BuLi with oximes 44. These dianions were then treated with trifluoroacetate 46 to give 5-trifluoromethyl isoxazoles 48 *via* 47 through reflux.⁶⁷ (Scheme 14).

Several 3-methylthio-5-aryl-isoxazoles 50 were synthesized from β-oxodithioesters 49 reacting with hydroxylamine hydrochloride *via* sodium acetate and acetic acid under acidic conditions at 90 °C for 2–10 h. One key point here is that the use of acetic acid is necessary for the formation of the isoxazole ring with possible mechanistic approach.⁶⁸ (Scheme 15).

Reddy *et al.*, proposed a synthetic strategy for 3,5-disubstituted isoxazoles 32. In this method, ynones 51 react with trimethylsilylazide 52 through *syn*-Michael addition *via* trichloroethylene in open air at room temperature to yield 3,5-disubstituted isoxazoles 32.⁶⁹ (Scheme 16).

Similarly, ynones react with azide ion to form TMS-ynammides 53 and then react to give 5-aminoisoxazoles 54 using potassium



Scheme 6 Electrochemical fluorination and Cu catalyzed cycloaddition giving isoxazoles 22.

carbonate in aqueous media at room temperature followed by the addition of sodium azide **38** in the presence of ammonium chloride. In this method, the TMS group eliminated first under basic conditions, followed by the formation of corresponding 5-aminoisoxazoles **54** from *syn*-Michael adducts.⁷⁰ (Scheme 17).

Bondarenko and co-workers reported a method of producing 5-bromoisoxazoles **58**. Here, 2 aryl-1,1-dibromocyclopropanes **55** undergo nitrosation with nitrosyl chloride **56** in the presence of nitromethane **57** to give 5-bromoisoxazoles **58** at room temperature.⁷¹ (Scheme 18) Similarly, 1,8-diazabicyclo[5.4.0]undec-7-ene (DBU)-facilitated ring opening of the aryl-cyclopropane **59** generated isoxazole-5-carboxylate **60** in the presence of nitromethane **57** as a driving force for ring cleavage maintained at 70–110 °C for 8–16 h. DBU resulted 90% of the yield with DMF as the solvent whereas acetonitrile and THF produced no yield.⁷² (Scheme 19).

2.3 Microwave-induced synthesis

Kulkarni developed a solvent free method for synthesizing 3,4-disubstituted isoxazole-5(4*H*)-ones **63** *via* microwave-induced organic synthesis. In this method, substituted aldehydes **61** were treated with hydroxylamine hydrochloride and ethyl acetoacetate **62** in the presence of catalysts such as KBr, KCl, NaOAc, MgCl₂ *etc* and irradiated at 200 W to obtain 3,4-disubstituted isoxazole-5(4*H*)-ones **63**.⁷³ (Scheme 20).

Sebbar *et al.*, illustrated microwave synthesis for 3,5-disubstituted isoxazole **66** from alkyne of benzimidazole-2-one **64** and hydroxy-4-methoxy-benzene-carboximidoyl chloride **65** in the presence of triethylamine, DMF and catalyst Cu.⁷⁴ (Scheme 21).

Qiang Gu and others gave an efficient method for synthesizing 3-substituted bis isoxazole ether **69** from chloro derived

pyridyl oxime **68** and 3-substituted phenyl-5-((prop-2-yn-1-yloxy) methyl) isoxazoles **67** in the presence of sodium bicarbonate as a base and acid-binding agent in THF and aqueous media followed by microwave irradiation of the reaction mixture.⁷⁵ (Scheme 22).

Microwave synthesis was also found to be helpful in isomerization reactions. Furfuryl ketone **70** was reacted with hydroxylamine hydrochloride to form furfuryl oxime **71** which was then treated with *m*-chloroperbenzoic acid (*m*-CPBA) at 0 °C for 1 h. The addition of trifluoroacetic acid at 0 °C and then bringing to room temperature gave 4-(3-phenylisoxazol-5-yl)but-3-en-2-one **72** as a mixture of *E* and *Z* isomers at a ratio of 1 : 30. The mixture **72** was made to undergo iodine-mediated isomerization at 140 °C in the presence of toluene in microwave synthesizer to obtain the *E* form of isomer **73**.⁷⁶ (Scheme 23).

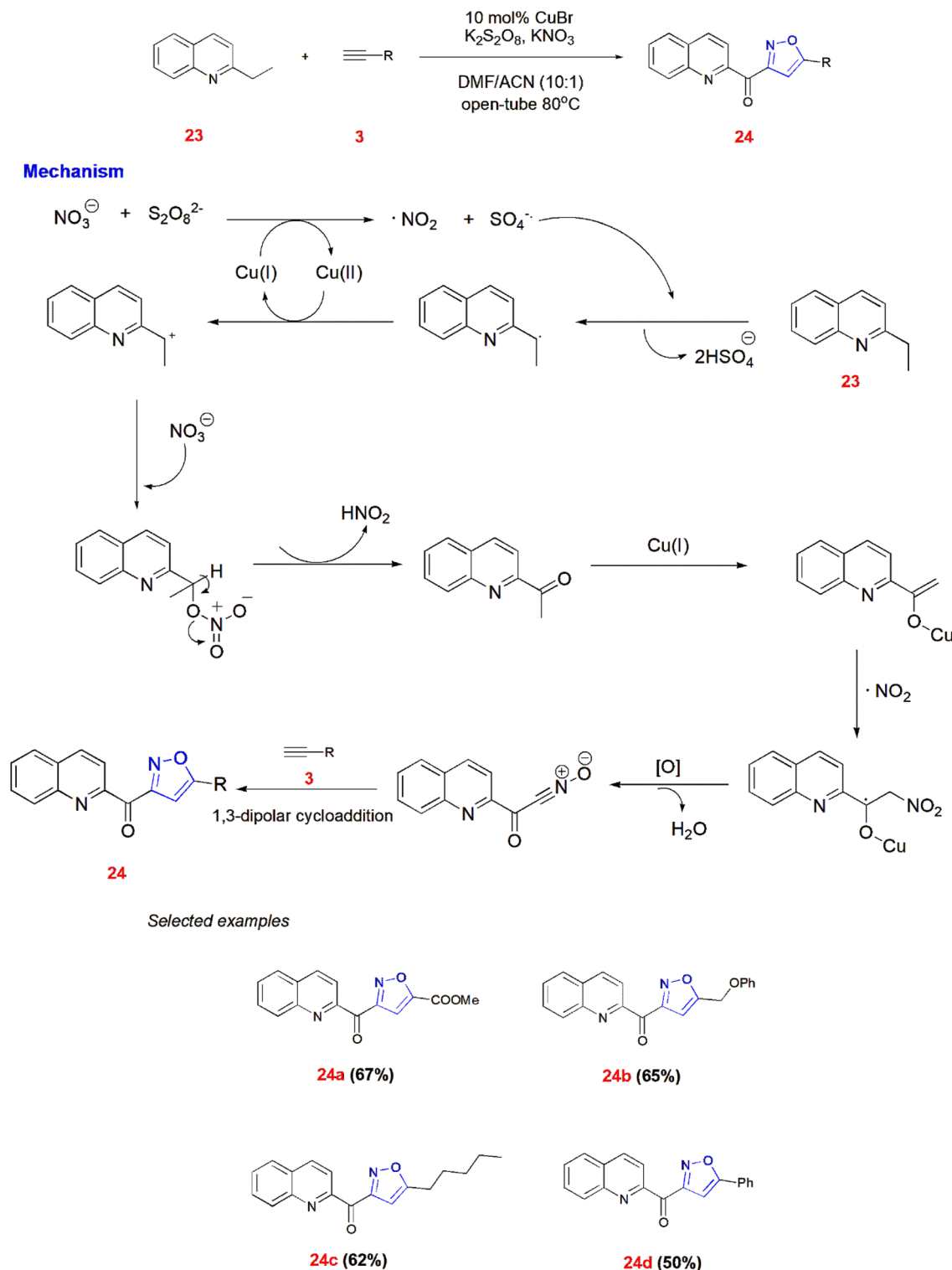
O-Hydroxyacetophenone **74** undergoes Claisen–Schmidt condensation with substituted aldehydes **61** in the presence of NaOH to give chalcones **75**. When irradiated in Microwave with hydroxylamine hydrochloride in ethanol for 10–15 min, these chalcones **75** afforded isoxazole **76**.⁷⁷ (Scheme 24).

Trifluoromethylated flavonol **77** on treatment with aromatic oxime **78** in the presence of triethylamine and CuI in DMF generated flavonoids with isoxazole ring **79**. These reactions were carried out *via* microwave irradiation (250 W) for 5 min.⁷⁸ (Scheme 25).

2.4 Cycloisomerization

Nakamura and co-workers illustrated the preparation of 4-methylated isoxazoline analogues **81** by the rearrangement of *O*-propargylic formaldoxime **80** *via* intermolecular methylene group transfer reaction using gold catalyst. Later, ene or



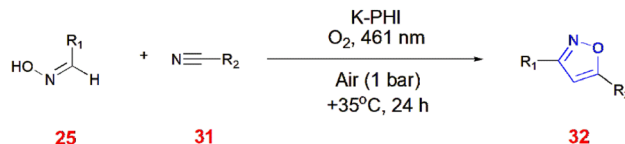
Scheme 7 Cu-catalyzed synthesis of quinoline-isoxazoles **24**.

isomerization reaction of isoxazolines **81** takes place with maleimide, glyoxalate, potassium *tert*-butoxide and azodicarboxylate to yield corresponding isoxazole derivatives **82–85**.⁷⁹ (Scheme 26) Similarly, chiral isoxazoles **88** can be made from *O*-

propargylic oxime **86** facilitated through chirality transfer. Furthermore, the chirality of *O*-propargylic oxime was retained without any changes leading to the formation of isoxazoline derivatives **87** which undergoes treatment with glyoxalate in the

Table 1 Different derivatives of isoxazoles 28

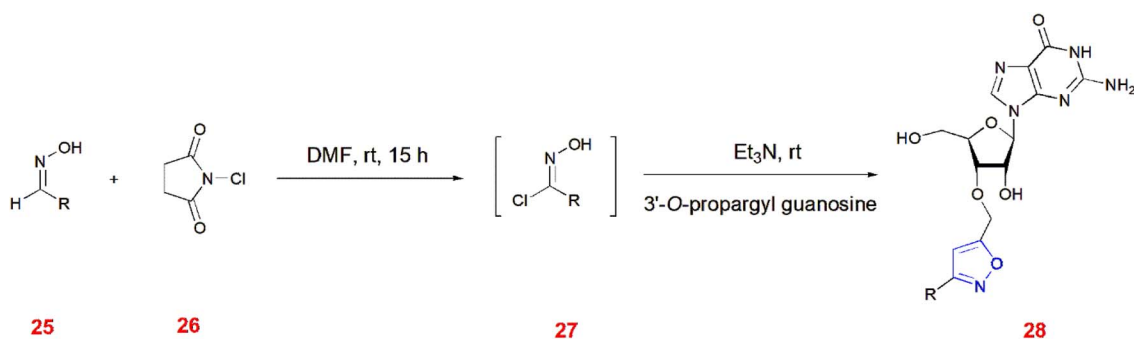
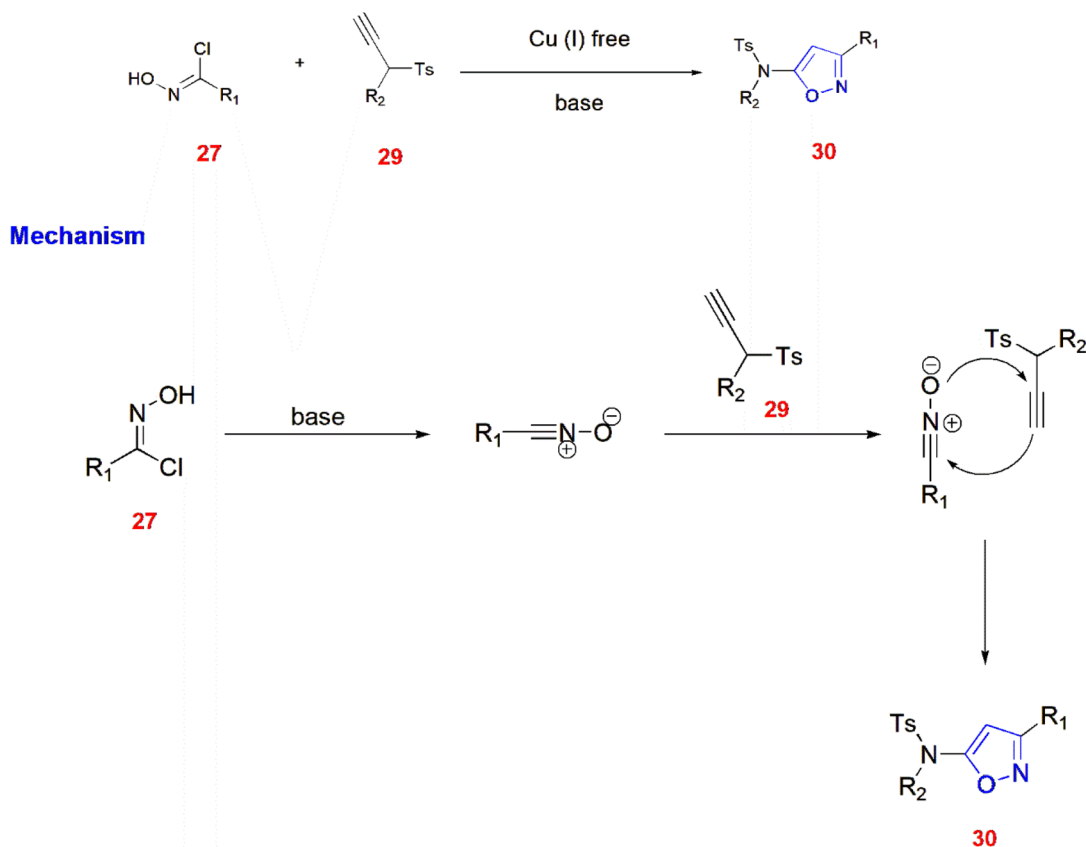
R	Yield (%)
Phenyl	84
4-Methoxy-phenyl	74
9-Anthranyl	72
3-Pyridyl	77
3-Indolyl	79
2-Chlorophenyl	70
4-Nitrophenyl	77
4-Bromo-2-thiophenyl	70



Scheme 10 Synthesis of isoxazoles 32 by artificial photosynthesis.

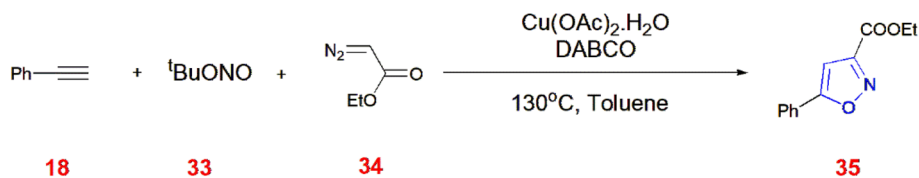
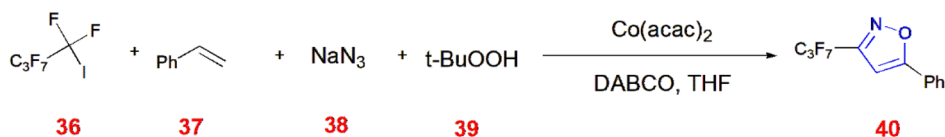
presence of boron trifluoride etherate to give expected chiral isoxazole 88.⁸⁰ (Scheme 27).

4-[Alkoxy(aryl)methyl]-substituted isoxazoles 91 were synthesized from aryl acetals 90 and alkynyl-*O*-methyl oximes 89. This is promoted by the oxocarbenium cations, which

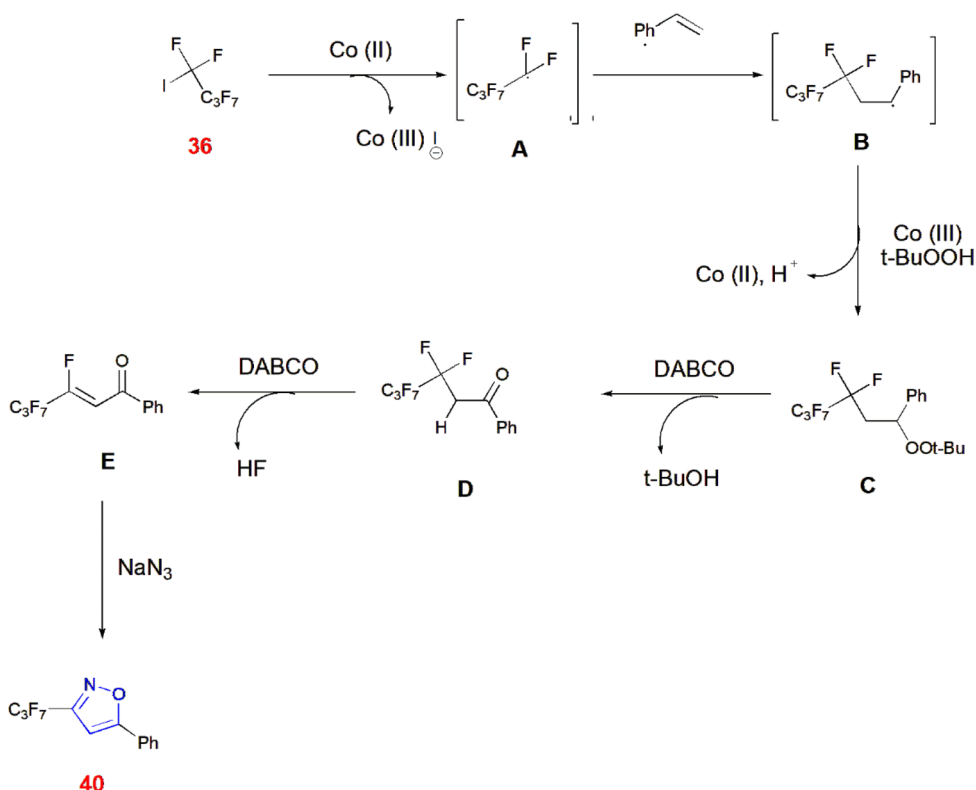
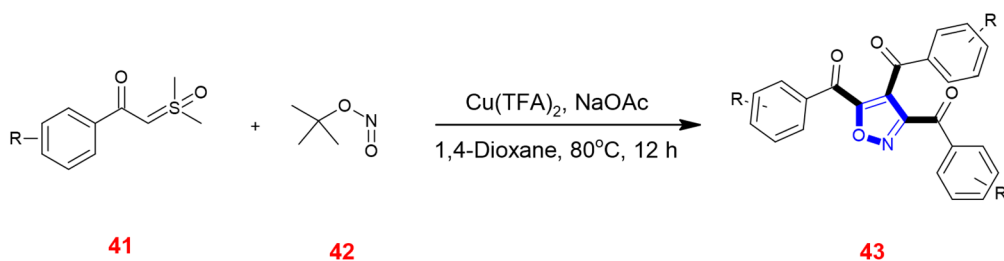
Scheme 8 Synthesis of 3-*O*-propargyl guanosyl derived isoxazoles 28.

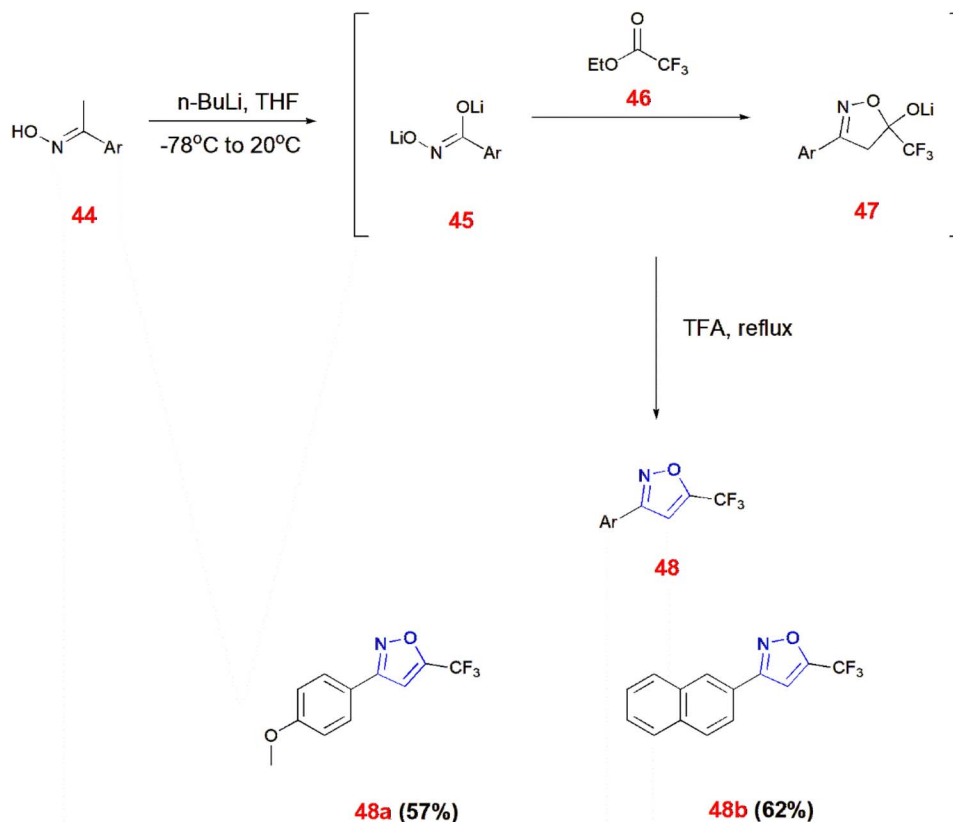
Scheme 9 Synthesis of isoxazoles 30 from nitrile oxides and ynammides 29.



Scheme 11 Synthesis of 3,5-disubstituted isoxazoles **35** via [3 + 2] cycloaddition using DABCO.

Mechanism

Scheme 12 Synthesis of perfluoroalkyl isoxazole **40** via [2 + 1 + 1 + 1] annulation.Scheme 13 Synthesis of isoxazoles **43** from sulfoxonium ylides **41**.



Scheme 14 Synthesis of 5-trifluoromethyl isoxazoles **48**.

cyclize alkynyl-*O*-methyl oximes **89** intramolecularly. For this occurrence, triple bond of alkynyl-*O*-methyl oxime **89** must be activated by oxocarbenium cations generated from aryl acetals **90** in the presence of boron trifluoride etherate, acetonitrile at room temperature for 10–15 min which led to the formation of isoxazoles **91** *via* intramolecular 5-endo cyclization.⁸¹ (Scheme 28).

Chlorinative cyclization of NCS and trimethylsilyl chloride led to the formation of 4-chloroisoxazoles **93**. (*E/Z*)-alkynyl-*O*-methyl oximes **92** were isomerized to its *Z* form to cyclize isoxazole **93** in the presence of nitromethane **57** at room temperature for 1 h.⁸² (Scheme 29).

2.5 Direct functionalization

The direct functionalization of the isoxazole ring at C-3, C-4 and C-5 is important because it is labile under basic conditions. This is accomplished either by C–H activation or transition metal cross coupling reactions. Propargyl phenyl ethers **94** were made to react with hydroxymoyl chloride **95** in the presence of CuI and potassium carbonate in diethyl ether to give isoxazole **96**. This intermediate was then cyclized using Pd(II) complex generating tricyclic fused isoxazoles **97**.⁸³ (Scheme 30).

Negishi coupling of isoxazole zinc pivalates **98** with bromopyridine derivative **99** which is highly functionalized in the

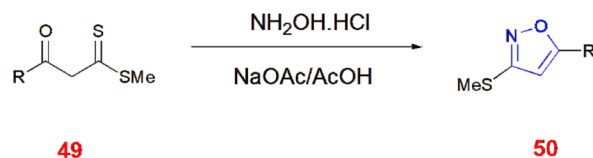
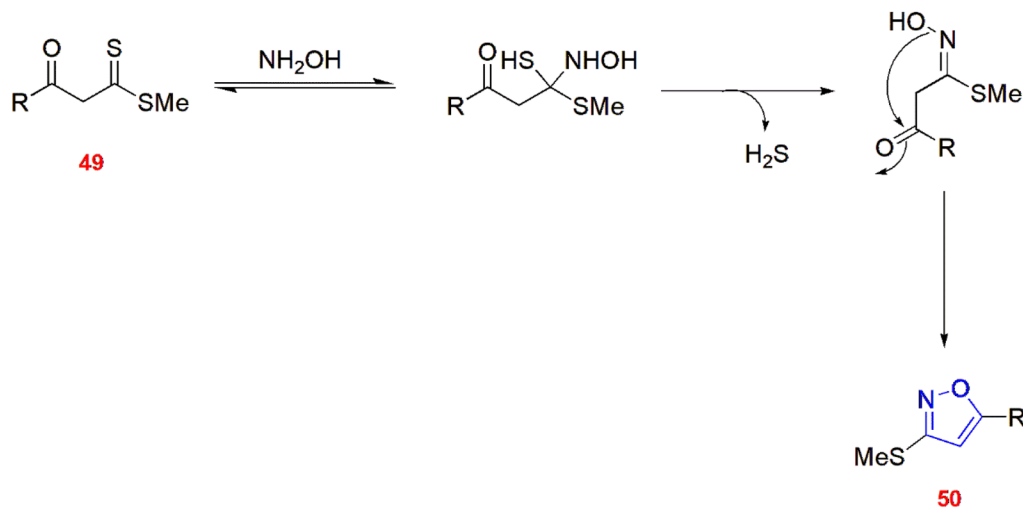
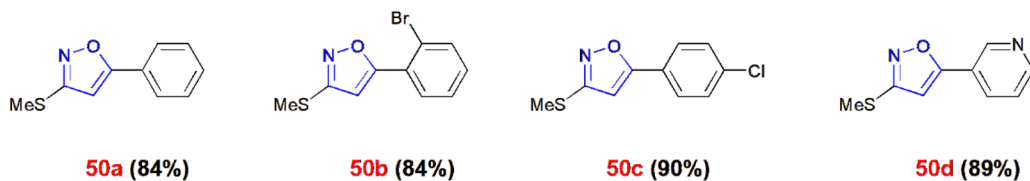
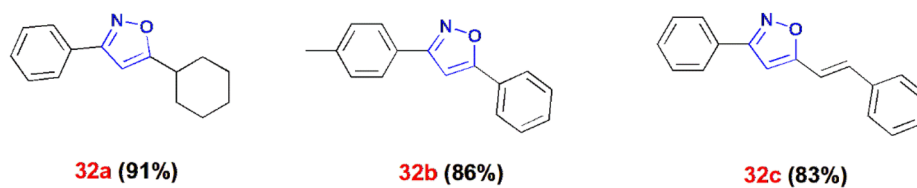
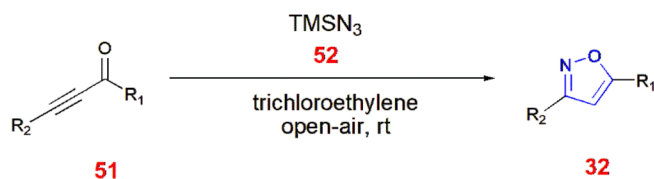
presence of the XPhos Pd G3 catalyst, produced the drug-like scaffold **100**. The reactivities of organozinc pivalates can be determined from several experimental methods.⁸⁴ (Scheme 31).

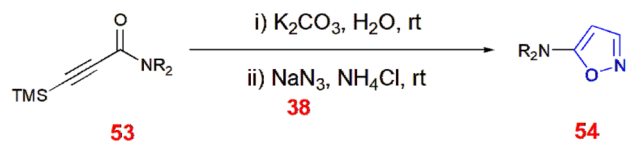
3,4-Disubstituted isoxazoles **101** underwent C–H arylation *via* 1,2-bis(diphenylphosphino)benzene (dppBz), which facilitated the coupling at C-5 of the isoxazole ring with aryl iodides **102** to yield coupled product **103**.⁸⁵ (Scheme 32).

Tang *et al.*, demonstrated various methods of fluorinating isoxazole acids **104** directly by decarboxylation in the presence of KF in 1,2-dichloroethane/water at a ratio of 2 : 1 at 70 °C for 15 h using SelectfluorTM **105** to yield fluorinated isoxazole **106**⁸⁶ (Scheme 33). Moreover, SelectfluorTM **105** was also used to fluorinate 3,5-disubstituted isoxazoles **32** at the C-4 position to give **108** in modest yields. Excess use of SelectfluorTM gave isoxazolines **109** with difluoro and monofluoro substituents at C-4 and C-5, respectively.⁸⁷ (Scheme 34).

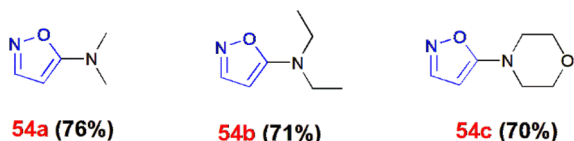
5-Aminoisoxazole **112** underwent difluoromethylthiolation using *N*-difluoromethylthiophthalimide **111** to give **113**. *N*-difluoromethylthiophthalimide **111** was prepared from reacting benzyl difluoromethylthioether **110** with chlorine in chloroform to generate dichlorocarbene at –30 °C to room temperature for 2 h. Furthermore, potassium phthalimide was added to give the desired reagent **111**.^{88,89} (Scheme 35).



**Mechanism****Selected examples**Scheme 15 Synthesis of 3-methylthio-5-aryl-isoxazoles **50**.Scheme 16 Synthesis of 3,5-disubstituted isoxazoles **32** from ynones **51**.



Selected examples



Scheme 17 Synthesis of 5-aminoisoxazoles **54** from ynamides **53**.

3 Biology of isoxazoles

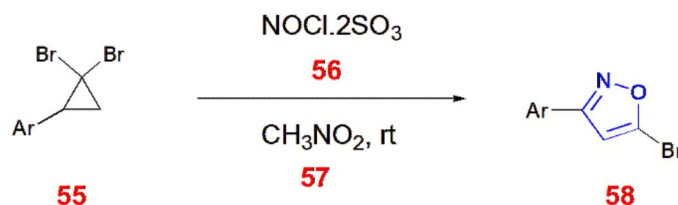
3.1 Antibacterial and antifungal activity

Antibacterial agents fall under the class of antimicrobials that can be bacteriostatic⁹⁰ or bactericidal⁹¹ in nature. Thus, several commercially available drug molecules containing isoxazoles have been approved for the treatment of acute bronchitis⁹² and infections in the urinary tract.⁹³ However, isoxazole-containing drugs such as oxacillin,⁹⁴ flucloxacillin,⁹⁵ dicloxacillin,⁹⁶

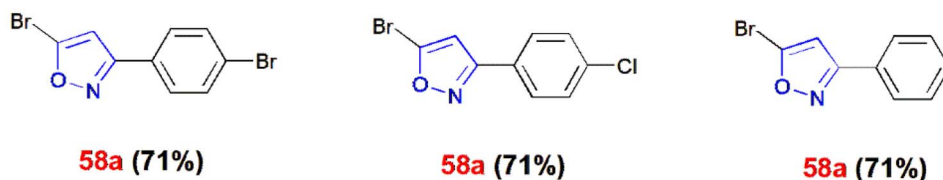
cloxacillin,⁹⁷ sulfamethoxazole⁴⁵ and sulfisoxazole,^{98,99} which are commercially available, are listed in Table 2.

Raju *et al.*, synthesized novel urea and thiourea derivatives of 6-fluoro-3-(piperidin-4-yl)benzo[d]isoxazole and evaluated their antimicrobial activity keeping tetracycline as the standard reference drug. Among the six derivatives, compounds **111–113** showed their inhibition against *Bacillus subtilis* and *Staphylococcus aureus*. Furthermore, **111** and **113** showed additional inhibition against *Escherichia coli* and *Pseudomonas aeruginosa*. Structure–activity relationship (SAR) studies revealed that the presence of electron withdrawing groups like trifluoro and chloro increased the activity to a greater extent.¹⁰⁰

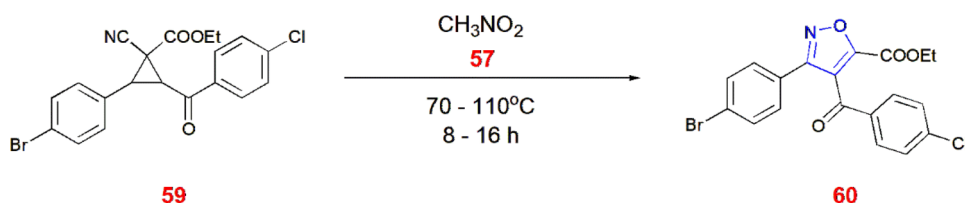
Khazi *et al.*, synthesized new sulfones and sulfides of methylene-bridged benzisoxazolyimidazo[2,1-b][1,3,4]thiadiazoles, tested them against *E. coli* ATCC 35218 and *B. subtilis* ATCC 6633 and concluded that most had significant to modest activity when compared with the standard drug ampicillin. Compounds **114–117** showed good activity against these strains. Additionally, compounds **115–117** showed activity against fungal species such as *Candida albicans* and the activity of **115** was almost equivalent to the standard drug Clotrimazole. Furthermore, compounds **114** and **118** showed good inhibitory effect against *Aspergillus fumigatus*. Coumarin and methoxy substituents made these compounds more active against fungal and bacterial strains. Especially, the delocalization of π -



Selected examples

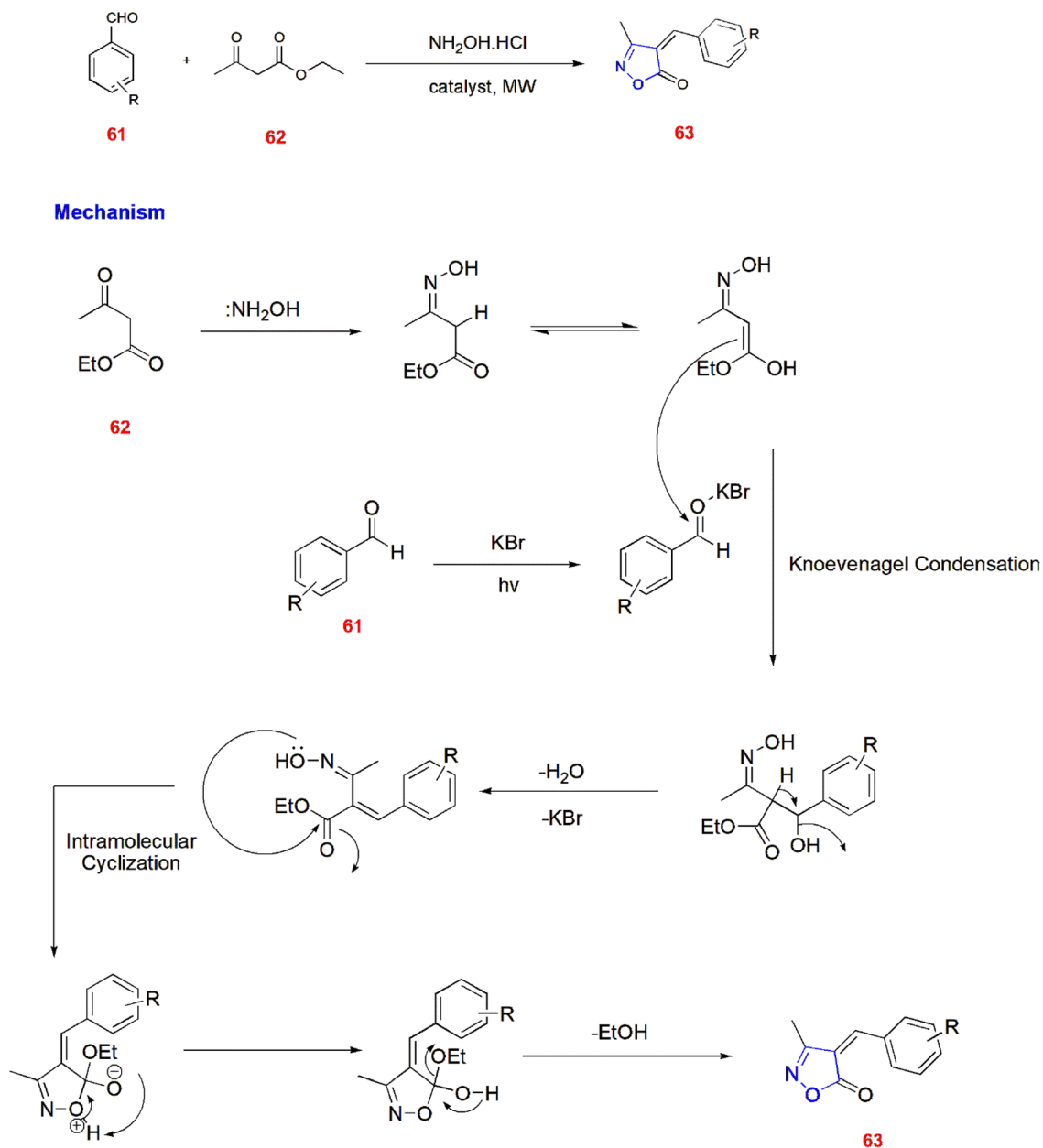
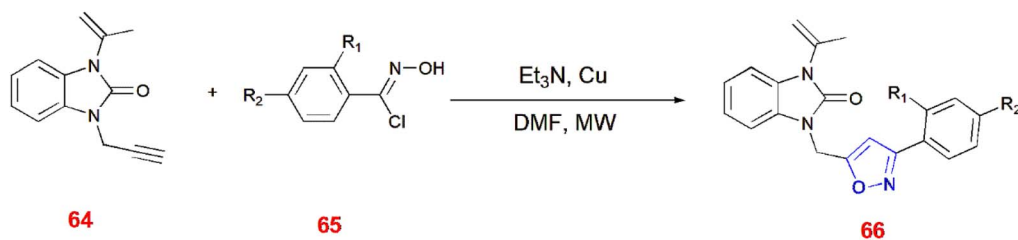


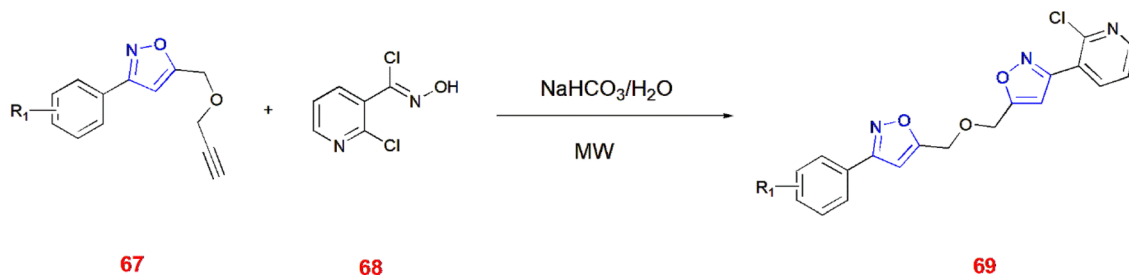
Scheme 18 Synthesis of 5-bromoisoxazoles **58**.



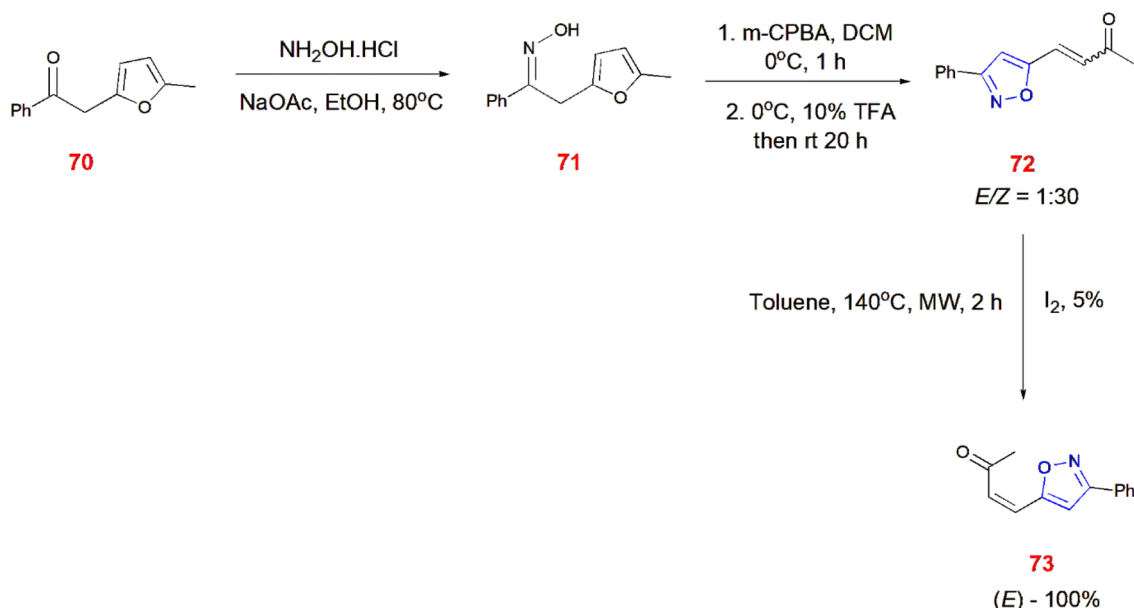
Scheme 19 Synthesis of isoxazole-5-carboxylate **60**.



Scheme 20 Microwave synthesis of 3,4-disubstituted isoxazole-5(4H)-ones **63**.Scheme 21 Microwave assisted synthesis of 3,5-disubstituted isoxazole **66**.



Scheme 22 Microwave induced synthesis of 3-substituted bis isoxazole ether 69.



Scheme 23 Microwave synthesis of (E)-4-(3-phenylisoxazol-5-yl)but-3-en-2-one 73.

electrons in **114** and **115** resulted in penetration into lipid membranes as they are lipophilic in nature.¹⁰¹

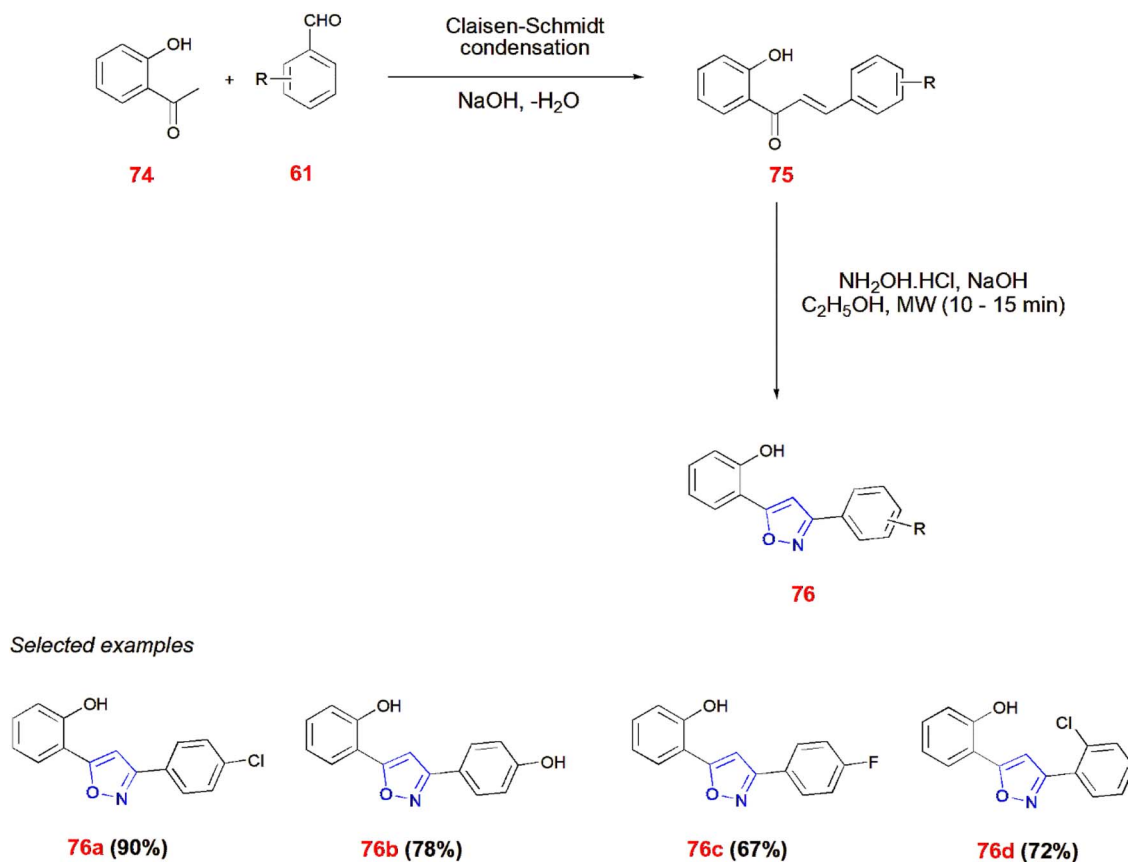
Anjani *et al.*, prepared novel derivatives of 1,3,5-triazines containing aminopyridines, isoxazoles and acetyl pyrazoline moieties, checked for their antimicrobial proficiency and determined their minimum inhibitory concentration (MIC). Compound **119** was effective against *S. aureus* bearing MIC value of 100 $\mu\text{g ml}^{-1}$, quite good when compared with ampicillin with 250 $\mu\text{g ml}^{-1}$ but not better than chloramphenicol and ciprofloxacin, both with MIC of 50 $\mu\text{g ml}^{-1}$. Compounds **121** and **123** both showed MIC values of 100 and 125 $\mu\text{g ml}^{-1}$ against *Streptococcus pyogenes*, which is equivalent to ampicillin with MIC 100 $\mu\text{g ml}^{-1}$ but moderate towards chloramphenicol and ciprofloxacin with 50 $\mu\text{g ml}^{-1}$. When Gram-negative bacteria such as *E. coli* is taken, compound **122** showed MIC value of 100 $\mu\text{g ml}^{-1}$ which was again equivalent to ampicillin but was quite higher than chloramphenicol and ciprofloxacin having MIC of 50 and 25 $\mu\text{g ml}^{-1}$, respectively. Compounds **121** and **122**

showed MIC values of 200 $\mu\text{g ml}^{-1}$ same as that of ampicillin against *S. aureus*. Notably, compound **122** showed great antifungal activity against *C. albicans* with MIC value of 250 $\mu\text{g ml}^{-1}$. Compounds **119–121** showed same activity as that of standard drug griseofulvin showing MIC value of 500 $\mu\text{g ml}^{-1}$. However, the SAR studies revealed that the presence of CF_3 in the compounds **119–123** made them active as antimicrobial scaffolds. Among them, **119** and **123** showed highest antimicrobial activity. Such electronic changes or modifications enable molecules to increase the binding energy with the microbial target site, thereby enhancing potency.¹⁰² (Fig. 2).

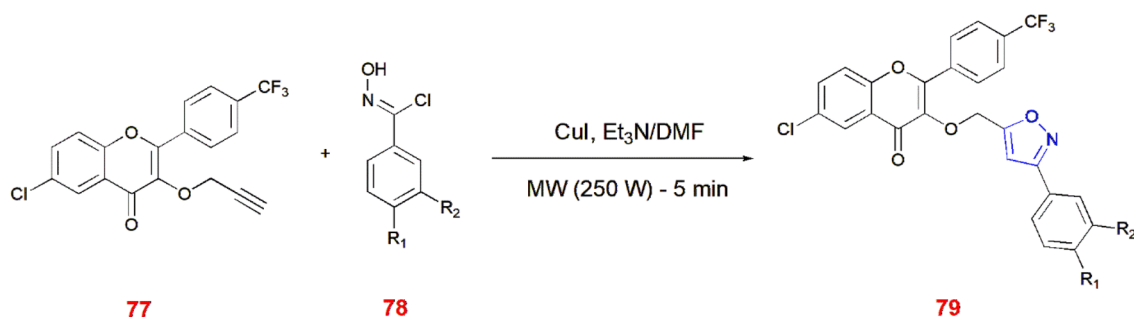
3.2 Anticancer activity

Isoxazoles have good anticancer activities and can be used to treat various carcinomic conditions. Several commercially available isoxazole-containing drug molecules such as acivicin,¹⁰³ XN05,^{104,105} PNZ5,¹⁰⁶ NVP-AUY922^{107–109} shown in Table 3.





Scheme 24 Microwave induced synthesis of isoxazole derivatives 76.

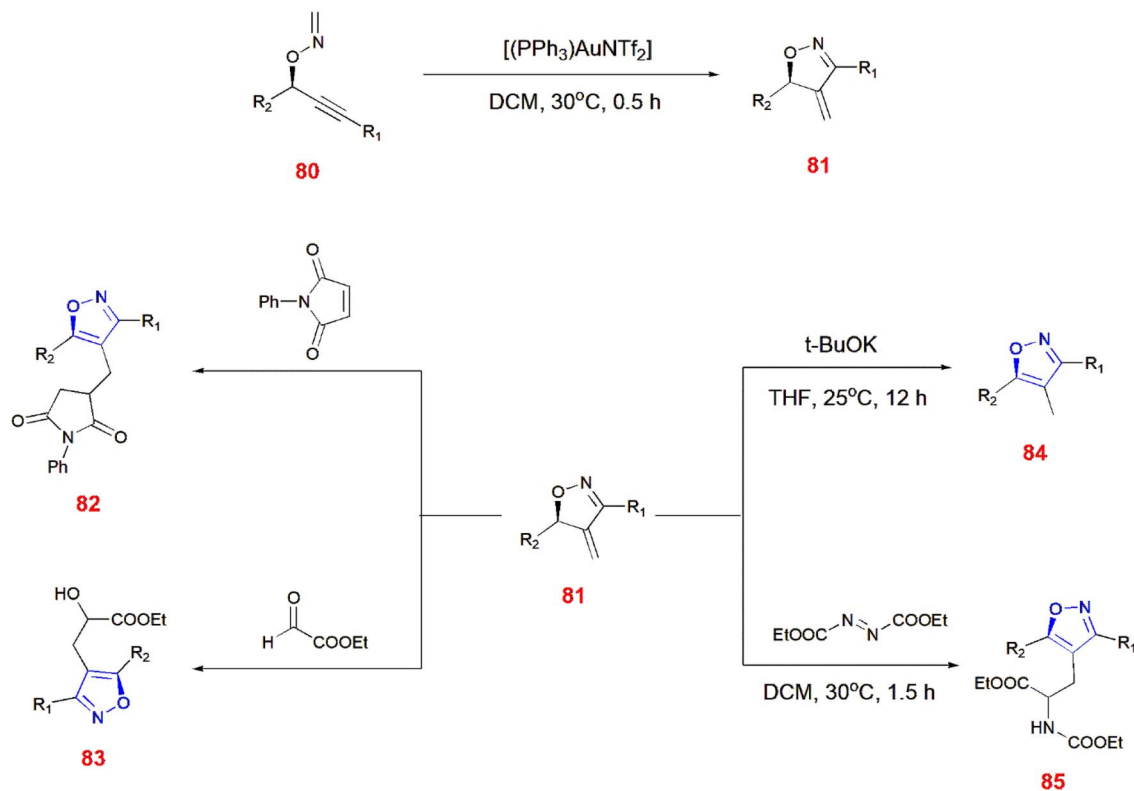


Scheme 25 Microwave synthesis of trifluoromethylated flavonoid-based isoxazoles 79.

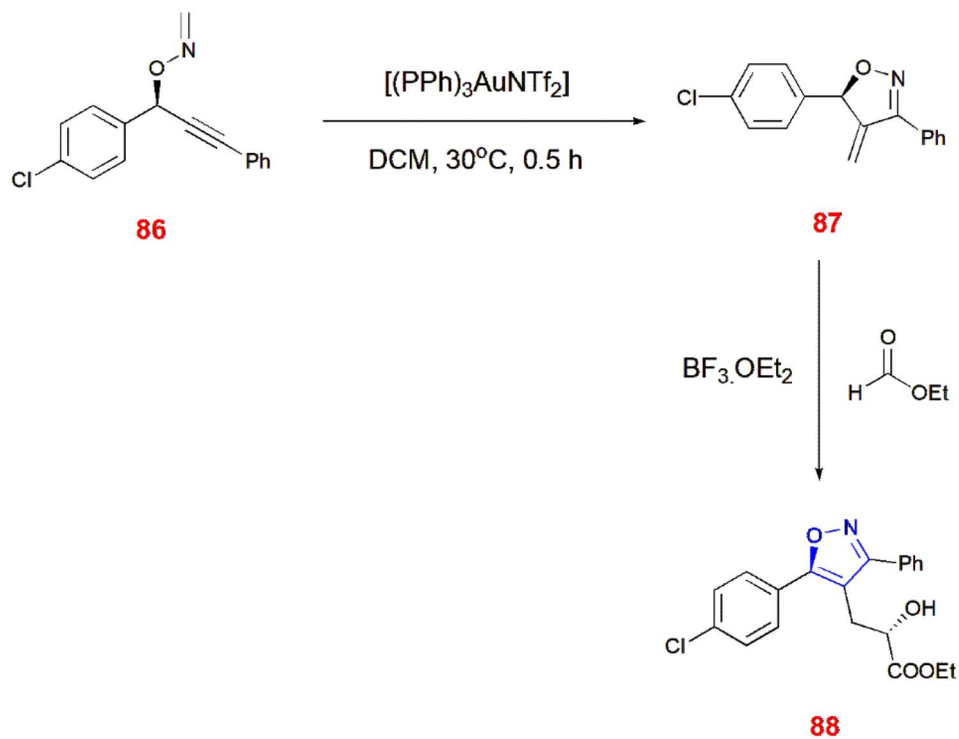
Eid *et al.*, synthesized and evaluated the biological performance of new isoxazole–amide analogues. As a result of the anticancer evaluation, these derivatives were tested against HeLa, Hep3B, and MCF-7 cell lines, their IC_{50} (half-maximal inhibitory concentration) values were compared with that of doxorubicin. It was found that, compound **124** was most active against HeLa cell line with IC_{50} value of $15:48 \pm 0:89 \mu\text{g ml}^{-1}$. However, compound **125** was considerably active against HeLa showing IC_{50} value of $18:62 \pm 0:79 \mu\text{g ml}^{-1}$. Compounds **124** and **126** showed anticancer activity against Hep3B cell line with

IC_{50} $23:98 \pm 1:83 \mu\text{g ml}^{-1}$ and $23:44 \pm 1:99 \mu\text{g ml}^{-1}$, respectively.¹¹⁰

Hawash *et al.*, synthesized and studied the biological activity of phenyl-isoxazole–carboxamide analogues. These derivatives were tested for their anticancer profiles against HeLa, MCF-7, Hep3B, HepG2, and Hek293T cell lines comparing with those of doxorubicin. Compounds **127–129** showed quite good anticancer activity against Hep3B cell lines showing the IC_{50} values (μM) of 5.96 ± 0.87 , 6.93 ± 1.88 and 8.02 ± 1.33 , respectively. Compound **130** was active against the MCF-7 cell line with IC_{50} value (μM) of 4.56 ± 2.32 . Compound **129** showed excellent

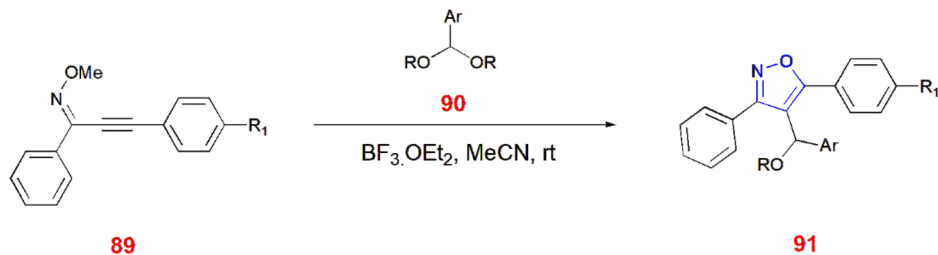
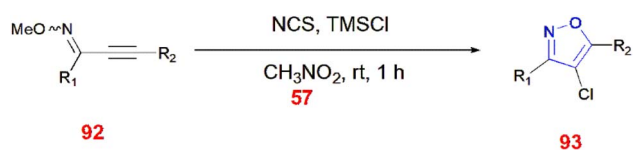


Scheme 26 Synthesis of isoxazole derivatives 82–85.



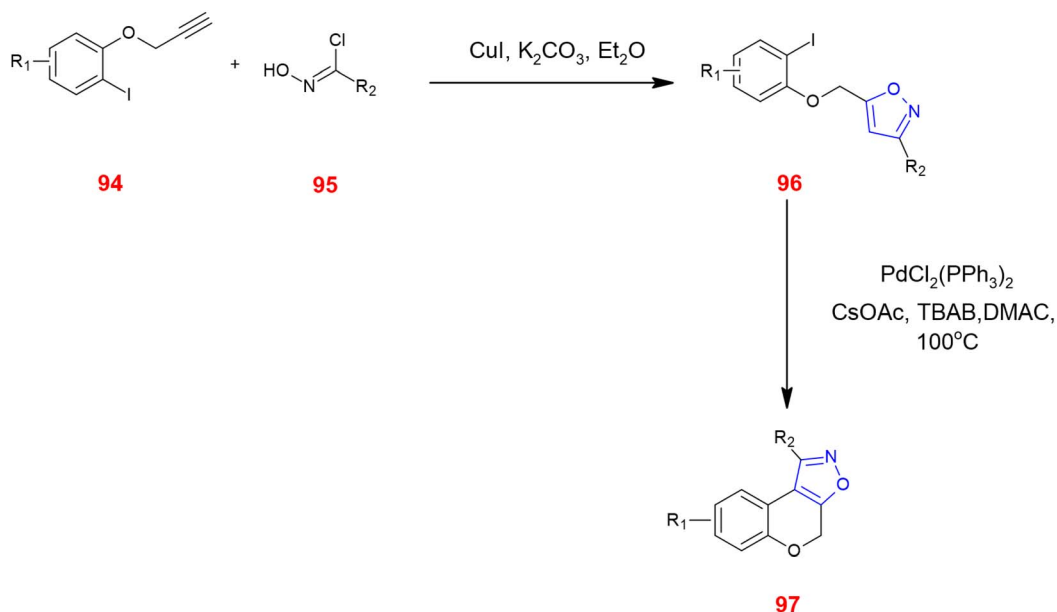
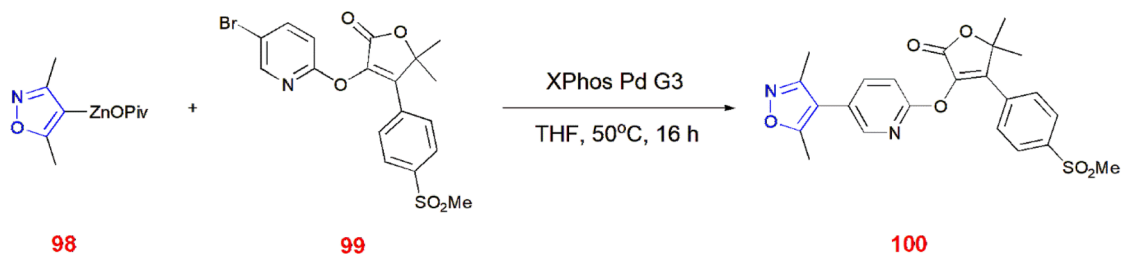
Scheme 27 Synthesis of isoxazole analogue 88.

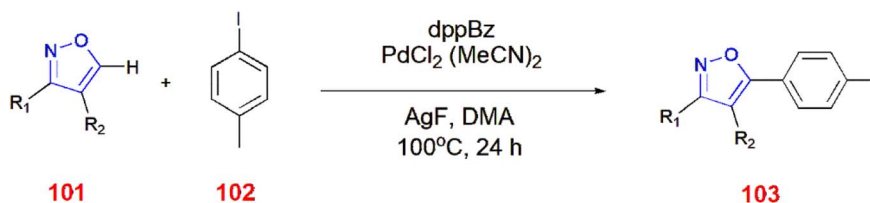
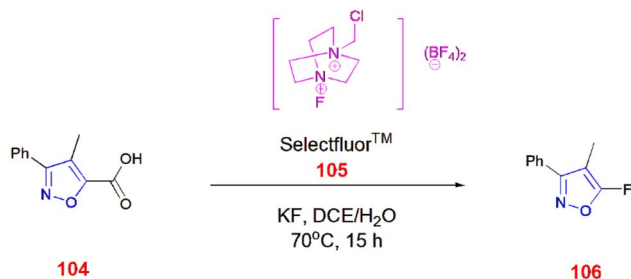


Scheme 28 Synthesis of 4-[alkoxy(aryl)methyl]-substituted isoxazoles **91**.Scheme 29 Synthesis of 4-chloroisoxazoles **93**.

anticancer activity against HeLa cell line with IC₅₀ value (μM) of 0.91 ± 1.03.¹¹¹

Ketan and co-workers produced some isoxazole derivatives with phenyl rings linked with a diazo group and evaluated their anticancer activity against PC3 and HEK normal cell lines. Compounds **131–134** showed anticancer activity against PC3 among which **134** was highly potent. However, these compounds showed anticancer activity only with high doses

Scheme 30 Synthesis of tricyclic fused isoxazoles **97** via direct functionalization.Scheme 31 Negishi cross coupling of isoxazole zinc pivalates **98**.

Scheme 32 C–H arylation of 3,4-disubstituted isoxazoles **101**.Scheme 33 Fluorination of isoxazole acids **104** via decarboxylation.

(640 μM) in normal cell lines. The SAR studies revealed the significance of the electron-withdrawing groups such as $-\text{F}$, $-\text{Cl}$, and $-\text{Br}$. The *ortho*-substituted bromo compound **134** demonstrated valuable cytotoxic effects than the rest of the halogen substituted analogues.¹¹²

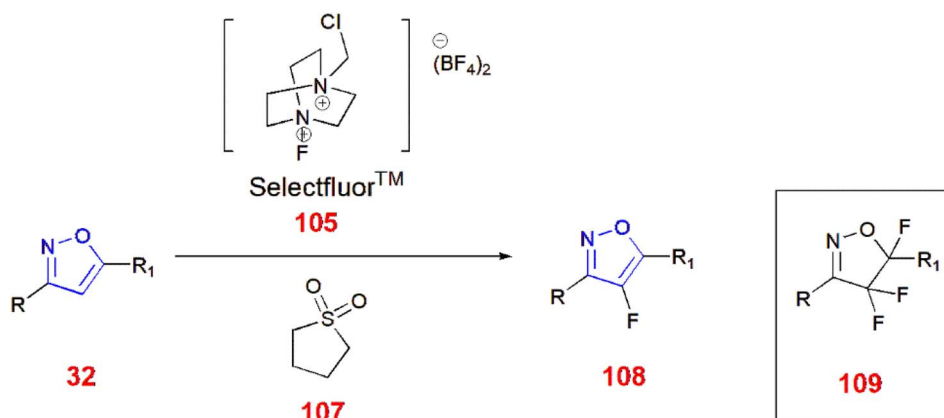
Jarina *et al.*, generated novel isoxazole derivatives, tested their anticancer activity against MCF-7 cell lines and reported their IC_{50} values ($\mu\text{g ml}^{-1}$). Among the six derivatives, compounds **135** and **136** showed good anticancer activity reflecting IC_{50} values ($\mu\text{g ml}^{-1}$) of -26.32 and -29.57 comparing with that of standard drug Adriamycin. Moreover, molecular docking studies demonstrated the binding affinity of -9 , favouring hydrophobic binding interactions with topoisomerase II.¹¹³

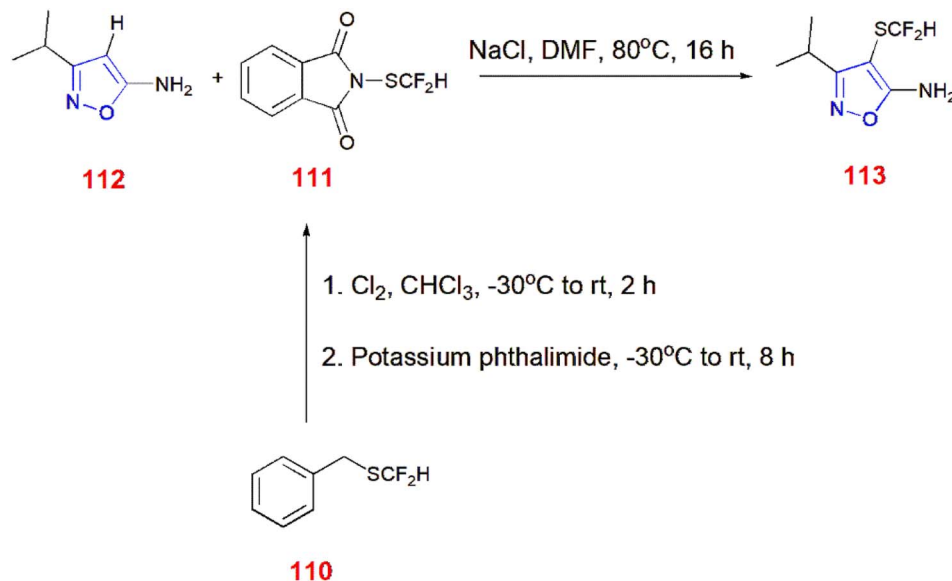
Panathur *et al.*, produced isoxazoles linked with indole and studied their antiproliferation activity against MCF-7 and HT-29 cell lines comparing with Gemcitabine. Compounds **137** and **138** were effective against MCF-7 cell lines showing IC_{50} values (μM) 7.72 and 5.51 . Compounds **139–142** were tested against HT-29 cell lines and showed interesting IC_{50} values (μM) -4.82 , -2.59 , -4.80 and -4.83 , respectively. The docking analysis proved that compound **140** containing a trifluoromethyl benzyl ether linkage was responsible for the increased cytotoxicity and is one of the lead compounds that could furnish significant anti-proliferative activity along with SIRT1 enzyme.¹¹⁴ (Fig. 3).

3.3 Anti-inflammatory activity

Isoxazoles are reported to have good anti-inflammatory activities and they control inflammation as there is a great need to reduce or treat edema. Normally, they follow two pathways namely, the cyclooxygenase (COX) and lipoxygenase (LOX) pathways.^{115–117} Some of the isoxazole-containing drugs such as Parecoxib,¹¹⁸ Valdecoxib,^{119,120} Mofezolac,^{121,122} Leflunomide¹²³ are shown in Table 4.

Abdellal worked on synthesizing novel isoxazoles derivatives and checked for their anti-inflammatory activity using rat paw edema model induced by carrageenan¹²⁴ as a suitable domain to perform experiments. Among various isoxazoles, compound **143** showed the ED_{50} (mg kg^{-1}) value 45 when compared to standard drug bearing 40 . While, other compounds **144** and **145** showed ED_{50} (mg kg^{-1}) value in the range of $48–50$. Thus, compound **143** was closer and more effective in reducing

Scheme 34 Fluorination of 3,5-disubstituted isoxazoles **32**.



Scheme 35 Difluoromethylthiolation of 5-aminoisoxazole 112.

inflammation in rat paws. The molecular docking scores with the range -14.77 to -15.63 kcal mol $^{-1}$ demonstrated the high selectivity of compounds 143–145 towards COX-2 receptor site and revealed well-established bonding interaction with active pharmacophores such as :O: of C=O and :N: of isoxazole ring.¹²⁵

Bhupinder Kumar *et al.*, synthesized indole-linked isoxazoles and tested their anti-inflammatory activity using the most accepted paw edema model. After injecting the rats with carrageenan to develop edema, these compounds were tested and the changes in the volume of paw edema at 1 h, 2 h and 4 h intervals, respectively. After thorough examination, compound 146 showed highest anti-inflammatory activity with 77.42% reduction after 4 h. Compounds 147 and 148 also showed good anti-inflammatory by reducing paw edema by 67.74 and 61.29%. Apart from these three compounds, there was an interesting result with respect to compound 149 which showed anti-inflammatory activity at 2 h interval and reduced in later hours which the biologists suggested the methoxy group at meta position had affected for such kind of deviation by inactivating the compound. However, the strong evidence was laid from the SAR studies which demonstrated the presence of 4-methoxy and 3-methoxy in compounds 147 and 148 favoured COX-2 inhibitory, anti-inflammatory and analgesic action. The selectivity index for COX-2 was highest with methoxy group at 4th position and resulted in slight decrease with the shift to 3rd position.¹²⁶

Khanum *et al.*, produced few novel 3-phenyl-5-furan isoxazole derivatives and evaluated their anti-inflammatory activity. Among the synthesized derivatives, compound 150 exhibited potency in inhibiting COX-2 with IC₅₀ value (μM) 9.16 ± 0.38 in relation with the reference drugs indomethacin and

diclofenac sodium. Through molecular docking studies, compounds 151–154 showed their affinity towards COX-1 rather than COX-2. For exhibiting COX-2 affinity, it is very essential for the isoxazole substrate to be hydrophobic and compound 150 had two chloro groups at *ortho* and *para* positions which makes them to interact with the active site. Apart from COX-2, it has also shown good 15-LOX inhibition with IC₅₀ value (μM) 8.15 ± 0.16 . These inhibitions are the great evidence for them to act as anti-inflammatory agents and lead molecules for the development of newer drug molecules.¹²⁷

Khan *et al.*, gave some isoxazole derivatives and carried out 5-LOX inhibitory assay for the designed compounds. Compound 155 showed promising 5-LOX inhibition with IC₅₀ value $3.67 \mu\text{M}$. Additionally, compounds 156 showed good 5-LOX inhibition next to compound 155. Compound 156 and 157 showed promising results in COX-2 inhibition.^{128,129} (Fig. 4).

3.4 Antioxidant activity

Hawash and co-workers determined the potential of novel isoxazole derivatives with respect to their antioxidant capacity. The derivatives of fluoro-phenyl-carboxamide were synthesized previously and then checked for their antioxidant properties using 2,2-diphenyl-1-picrylhydrazyl (DPPH) assay. Trolox was used as the control for the experiments. Its IC₅₀ value ($\mu\text{g ml}^{-1}$) was found to be 3.10 ± 0.92 . After evaluation, compounds 158 and 159 showed potent antioxidant activity with IC₅₀ values ($\mu\text{g ml}^{-1}$) 0.45 ± 0.21 and 0.47 ± 0.33 , respectively. Usually, antioxidant compounds exhibit radical scavenging properties wherein the radical will abstract a proton from the antioxidant scaffold. As there are different substituents on the phenyl ring, compound 158 has *tert*-butyl group, whose absence showed very less antioxidant properties. Thus, it was made clear that the

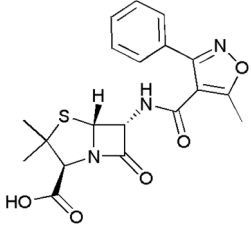
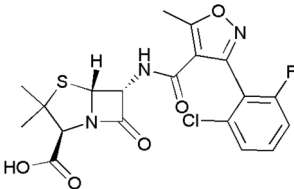
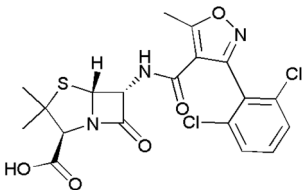
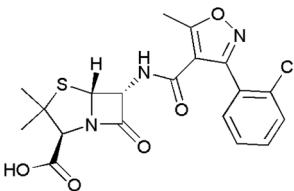
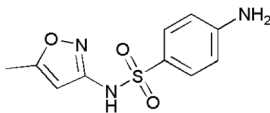
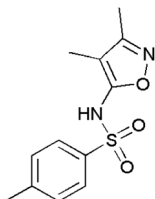
presence of electron donating groups were responsible for exhibiting antioxidant properties.¹³⁰

Chalka *et al.*, investigated the antioxidant activity of novel functionalized isoxazoles using DPPH method of screening, taking BHT as the reference standard having IC₅₀ value ($\mu\text{g ml}^{-1}$) 28.81 ± 1.84 . Among the synthesized isoxazoles, compound **160** showed great antioxidant property with IC₅₀ value ($\mu\text{g ml}^{-1}$) of 63.51 ± 1.80 . Other than this compound, some other compounds **161–163** exhibited free radical scavenging characteristics having IC₅₀ values ($\mu\text{g ml}^{-1}$) of 79.85 ± 1.90 , 83.69 ± 1.92 and 87.76 ± 1.94 , respectively. The remaining isoxazoles showed moderate DPPH activity. However, the ‘drug-

likeness’ evaluation suggested that compounds **161** and **163** were closely adhering to the Lipinski’s ‘Rule of Five’ and compound **163** persisted significant oral bioavailability threshold (>55% F). Erstwhile, all compounds **160–164** validated Egan’s rule for balanced consideration of lipophilicity and molecular weight.¹³¹

Nagaraju *et al.*, screened quinazolinone-based isoxazole derivatives for antioxidant activity using DPPH assay. Ascorbic acid was used as the control for the experiments. It was found that, compound **164** exhibited best activity with inhibitory concentrations 1.28 ± 0.33 and $1.39 \pm 0.38 \mu\text{M}$. However, compound **165** showed significant activity succeeding the prior

Table 2 Commercially available isoxazole-containing antibacterials

Name	Structure	Action
Oxacillin		Resistant to staphylococci infections
Flucloxacillin		Used in the treatment of pneumonia and endocarditis
Dicloxacillin		Acts upon penicillin resistant-staphylococci infections
Cloxacillin		Resistant to streptococcal, pneumococcal and staphylococcal infections
Sulfamethoxazole		Used to treat urinary tract and gastrointestinal tract infections
Sulfisoxazole		Used to treat meningitis, inclusion conjunctivitis



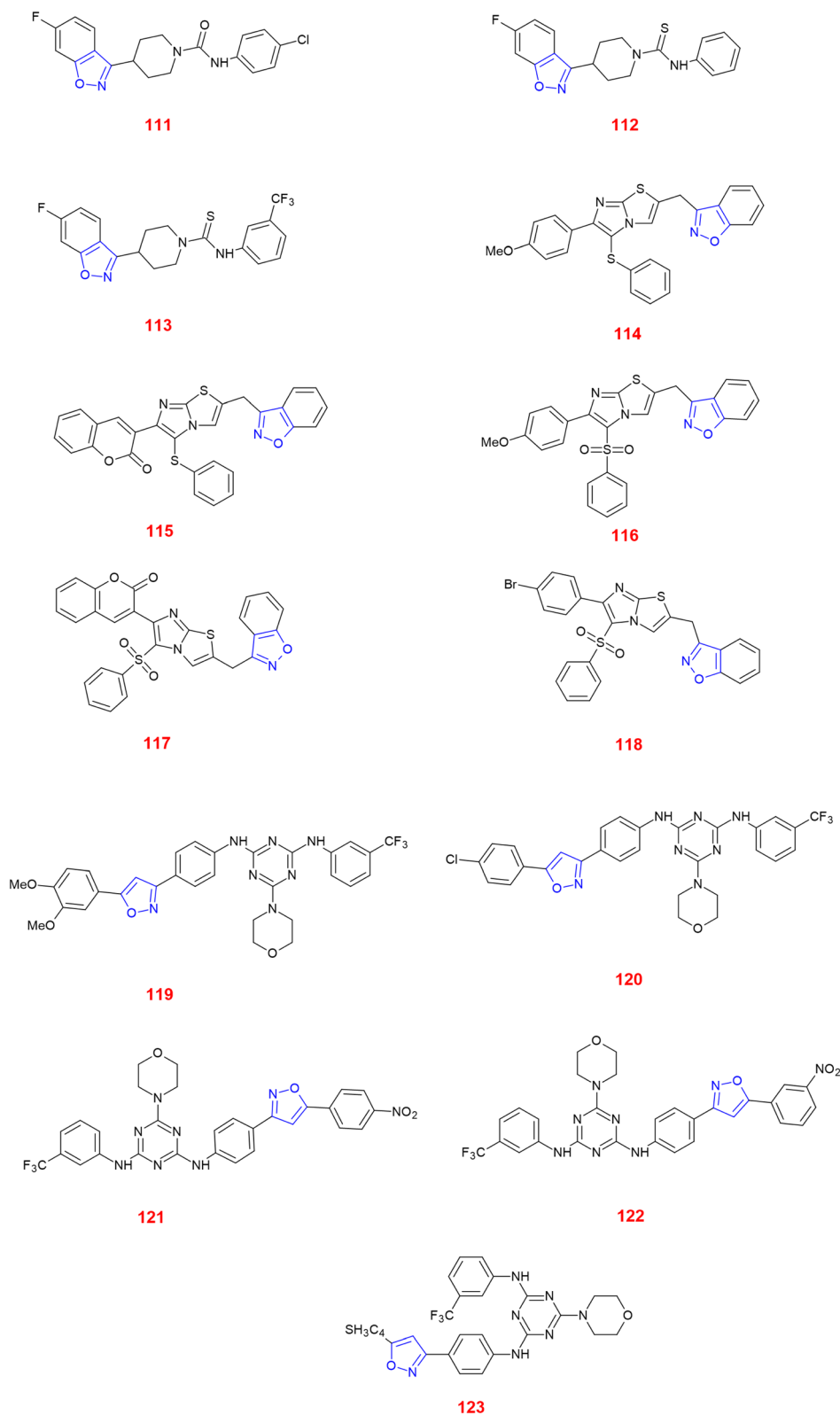
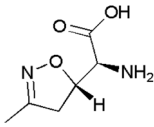
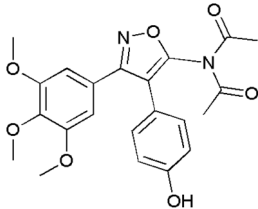
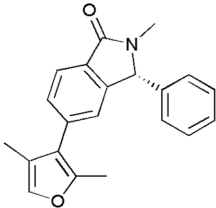
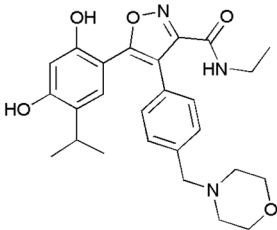


Fig. 2 Isoxazole-containing antibacterial and antifungal compounds.

Table 3 Commercially available isoxazole-containing anticancer drugs with their biological action

Name	Structure	Action
Acivicin		Cancer biomarker
XN05		Acts against hepatocellular cancer
PNZ5		Inhibits gastric cancer cell growth
NVP-AUY922		Acts against breast cancer

compound with inhibitory concentrations 2.72 ± 0.34 and $2.78 \pm 0.41 \mu\text{M}$. Furthermore, these compounds **164** and **165** were evaluated against NADP oxidase as it plays a key role in generating reactive oxygen species. As a result of this, compound **164** best fitted into the groove of NADP oxidase with quite acceptable ADME properties without violating Lipinski's rule, exploring the potential in developing into therapeutic agent.¹³²

Victor and co-workers synthesized and studied the antioxidant properties of 4-arylhydrazinylidene-isoxazoles containing polyfluoroalkyl groups. 2,2'-Azino-bis(3-ethylbenzothiazoline-6-sulfonic acid) (ABTS)^{133,134} and ferric reducing antioxidant power (FRAP)^{135,136} free radical scavenging assays were carried out for evaluating antioxidant properties taking Trolox as the reference standard with Trolox equivalent antioxidant capacity (TEAC) 1. Compounds **166–169** were found to be lead compounds in ABTS assay having TEAC 1.50 ± 0.07 , 1.80 ± 0.06 , 2.00 ± 0.10 and 1.90 ± 0.09 , respectively. Compounds **170** and **171** were found to be leads for FRAP assay as they effectively reduced the Fe^{3+} complex.¹³⁷ (Fig. 5).

3.5 Antitubercular activity

Mycobacterium tuberculosis causes Tuberculosis, an air-borne lung infection *i.e.*, contagious in nature.¹³⁸ *Mycobacterium tuberculosis* falls under the three major classes of the genus *Mycobacterium* that cause tuberculosis, leprosy and other non-tuberculous mycobacteria.^{139–141} Thus, many compounds were developed to treat tuberculosis among which isoxazole containing molecules have gained great attention and importance. Quinoline–isoxazole containing compounds have been widely studied and some of these include compounds **172–176**.^{142–146}

Sharabu and Pappula synthesized and studied the antitubercular activity of isoxazole clubbed pyrimidine derivatives. *Mycobacterium tuberculosis* H37Rv was used for this study and Pyrazinamide was used as the reference antitubercular drug and was carried out using dilution method at various concentrations ranging from 0.78 to $100 \mu\text{g ml}^{-1}$. Compound **177** and **178** showed excellent activity at MIC $0.78 \mu\text{g ml}^{-1}$. Compounds **179–183** showed better activity with MIC $1.56 \mu\text{g ml}^{-1}$ than the reference drug (MIC $3.125 \mu\text{g ml}^{-1}$).¹⁴⁷

Marco Pieroni and others evaluated the antitubercular activity of 5-(2-aminothiazol-4-yl)isoxazole-3-carboxamides



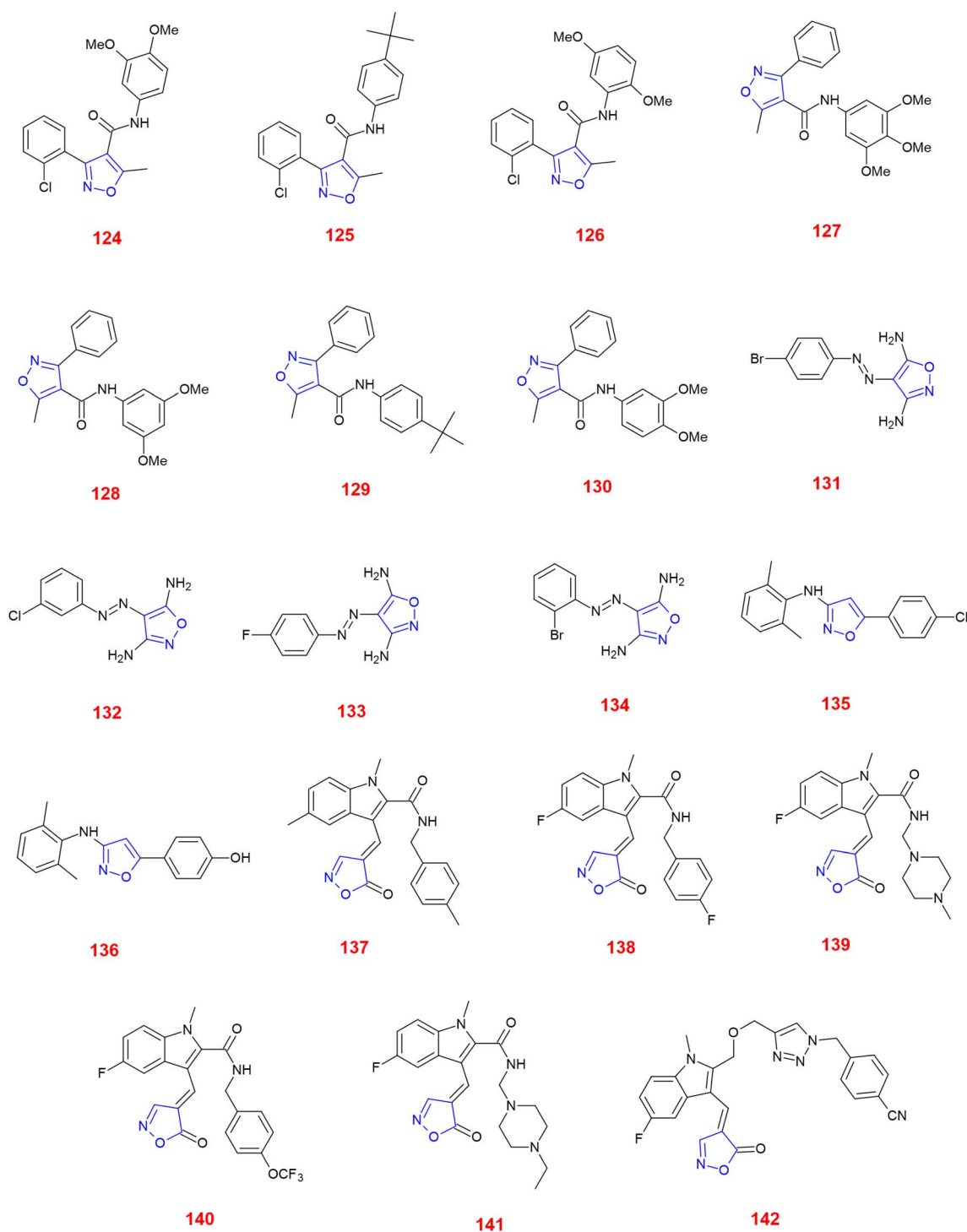
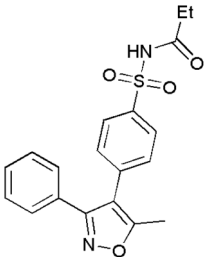
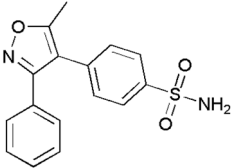
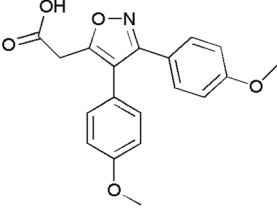
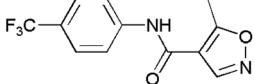


Fig. 3 Isoxazole analogues showing anticancer activity.

Table 4 Commercially available isoxazole containing anti-inflammatory drugs

Name	Structure	Action
Parecoxib		COX-2 inhibitor
Valdecoxib		Relieves arthritic inflammation
Mofezolac		Controls inflammation in rheumatoid arthritis
Leflunomide		Blocks formation of DNA thereby reducing inflammation in arthritic patients

taking Streptomycin and Isoniazid as reference standards and REMA method for determining MIC toward the tubercular strain. *M. tuberculosis* H37Rv strain was used for this study and it revealed that compounds **184** and **185** showed maximum antitubercular activity with MIC ($\mu\text{g ml}^{-1}$) 1.0 and 0.5, respectively.¹⁴⁸

Sahoo *et al.*, synthesized several novel chalcone linked 5-phenyl-3-isoxazolecarboxylic acid methyl esters and tested for their antitubercular activity against *M. tuberculosis* H37Rv strain taking isoniazid, streptomycin, rifampicin and ethambutol as reference drugs. After careful examination, they concluded that compound **186** and **187** were very potent antitubercular agents with MIC ($\mu\text{g ml}^{-1}$) 0.25 and 0.12, respectively.¹⁴⁹ (Fig. 6).

4 Conclusion

Isoxazoles are synthesized *via* unique reactions such as cyclo-additions and the inclusion of click reactions makes them more attractive. The green synthesis of isoxazoles has also attained increased attention, as the environment is a major concern for chemists working on molecules worldwide. The direct functionalization of isoxazoles illustrates the basics of organic

reactions and how unique they are upon arylation. In addition to synthetic methods, isoxazoles are also important medicinally. The biological implications of isoxazoles have made them more attributable to clinical dimensions. Most of the isoxazole analogues discussed in this review were successful in inhibiting various bacterial and fungal strains that cause various infections and other health-related diseases. However, such derivatives were also effective with anti-proliferative action when tested against various cancer cell lines such as HeLa, MCF-7, Hep3B, PC3, HEK, HT-29 *etc.* Such findings accomplish the potential of the isoxazole cores as anticancer agents with the existing standard anticancer drugs. The ability of isoxazoles to inhibit cyclooxygenase (COX) and lipoxygenase (5-LOX) pathways led for the strong anti-inflammatory action. The scope of isoxazoles is further broadened by their remarkable antioxidant and antitubercular actions supported by the 'drug-likeness' evaluation. Some isoxazole-containing compounds have shown even better results than the standard reference drugs which are commercially available. Thus, isoxazoles can be fascinating molecules in the world of organic chemistry for chemists to work in academic research and pharmaceutical research and development.



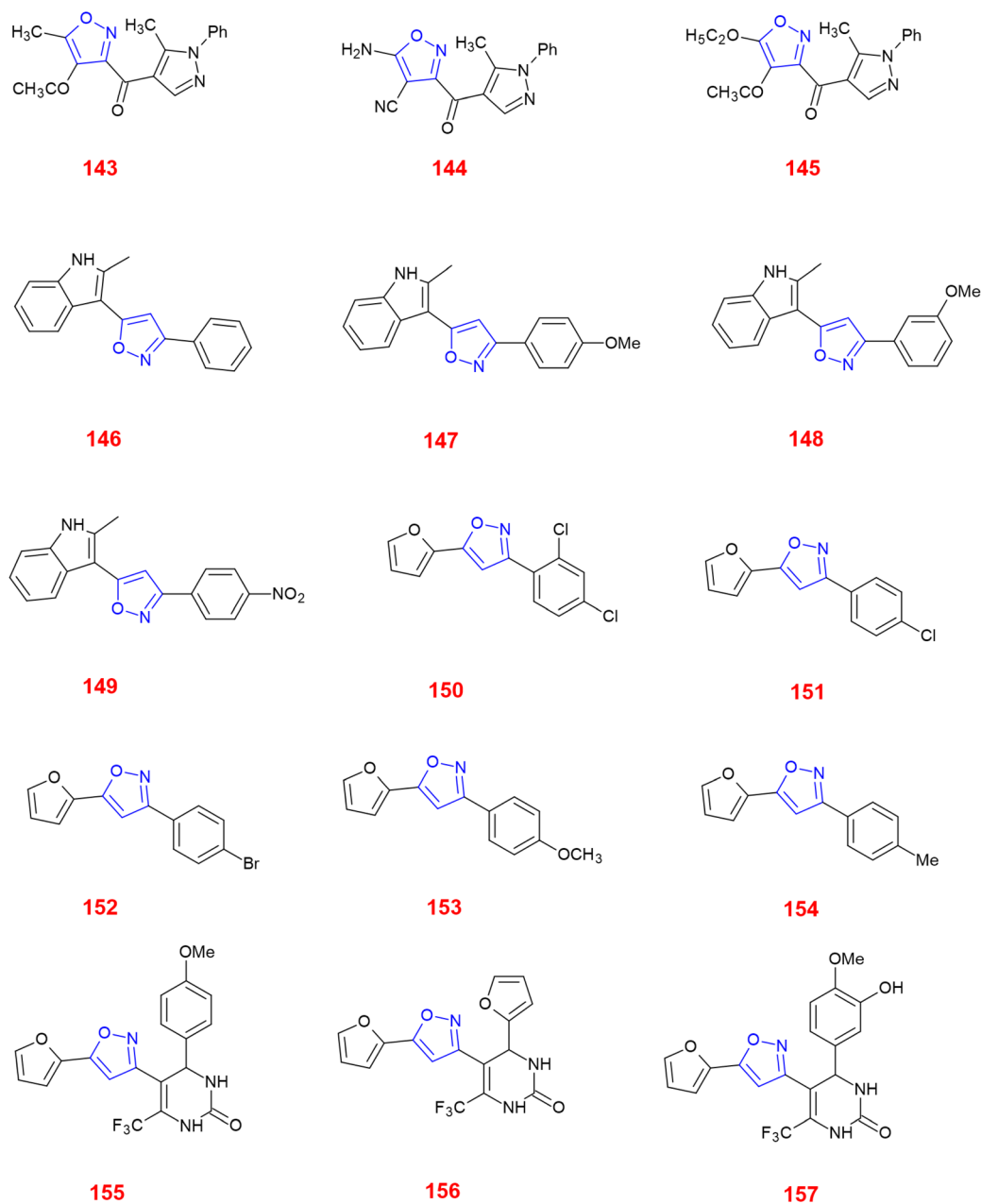


Fig. 4 Isoxazole derivatives showing anti-inflammatory activity.

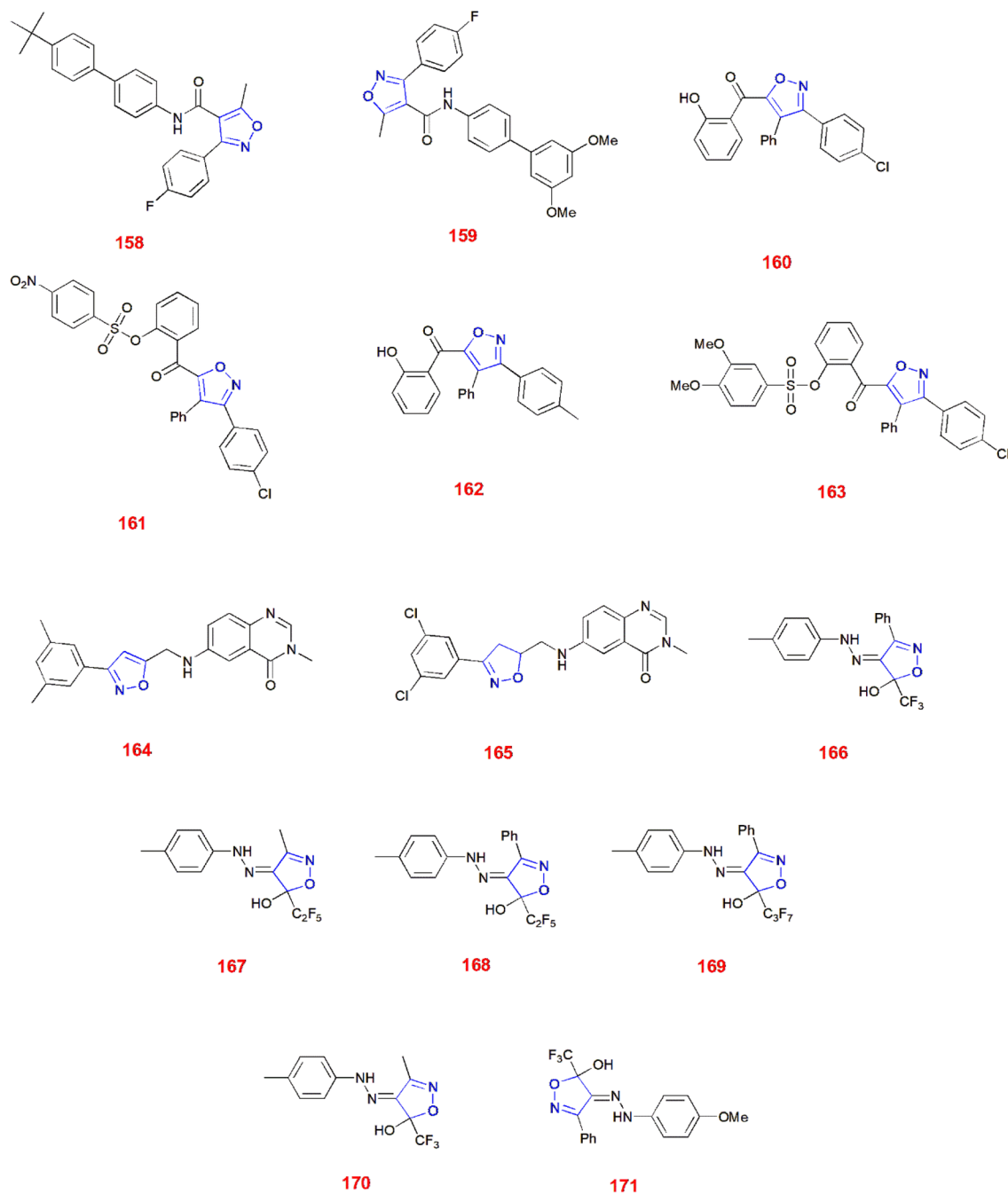


Fig. 5 Isoxazole derivatives showing antioxidant activity.

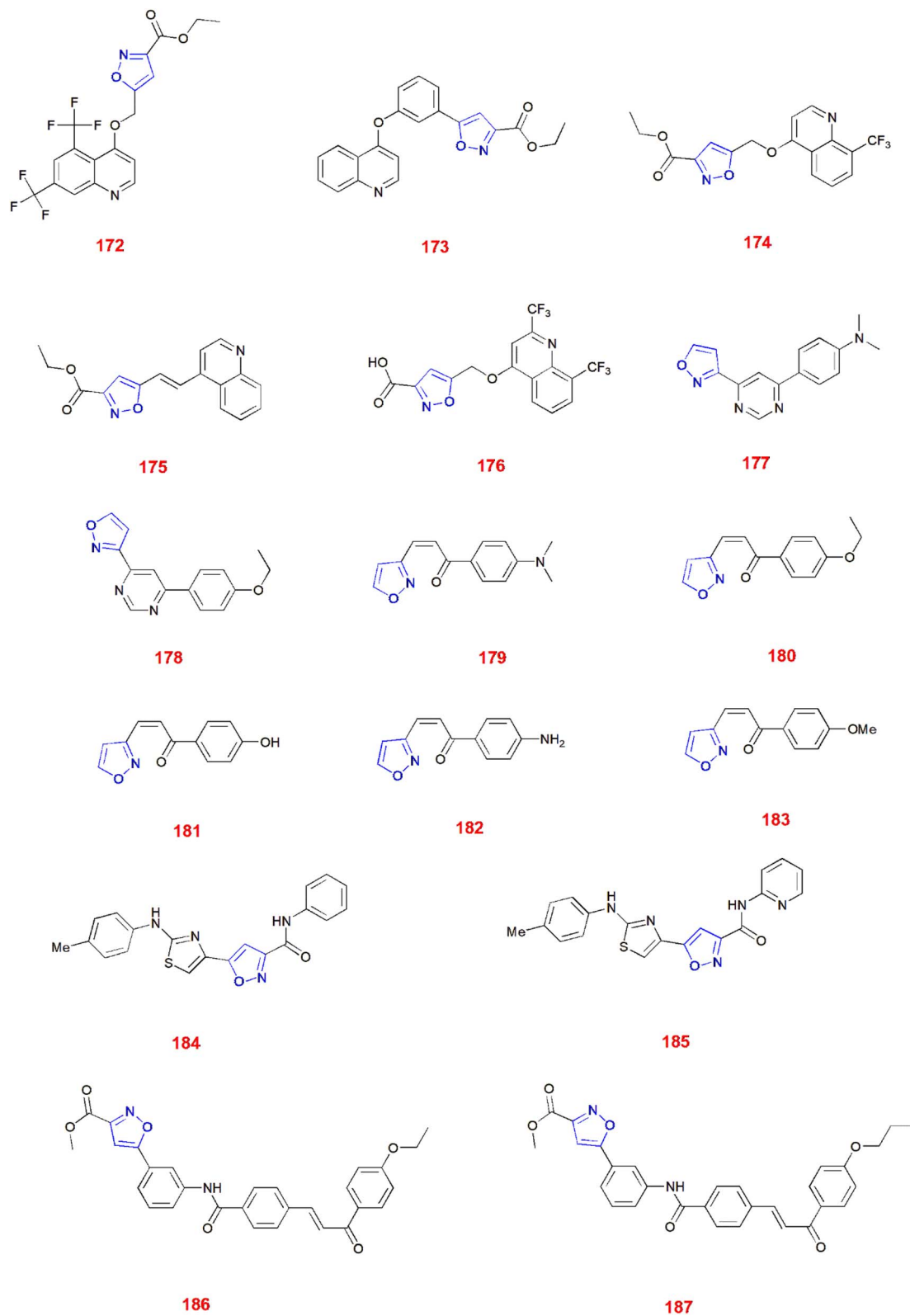


Fig. 6 Isoxazoles analogues showing antitubercular activity.

Abbreviations

NCS:	<i>N</i> -Chlorosuccinimide
NIS:	<i>N</i> -Iodosuccinimide
DCM:	Dichloromethane
DCE:	Dichloroethane
THF:	Tetrahydrofuran
DME:	Dimethyl ether
ACN:	Acetonitrile
DMF:	Dimethylformamide
TBAB:	Tetrabutylammonium bromide
DMAC:	Dimethylacetamide
K-PHI:	Potassium poly(heptazine imide)
DABCO:	1,4-Diazabicyclo[2.2.2]octane
TMSN ₃ :	Trimethylsilyl azide
TMSCl:	Trimethylsilyl chloride
DBU:	1,8-Diazabicyclo[5.4.0]undec-7-ene
MW:	Microwave
<i>m</i> -CPBA:	<i>meta</i> -Chloroperbenzoic acid
TFA:	Trifluoroacetic acid
SAR:	Structure–activity relationship
ATCC:	American type culture collection
MIC:	Minimum inhibitory concentration
HeLa:	Henrietta lacks
MCF-7:	Michigan cancer foundation-7
PC3:	Human prostate cancer cell line
IC ₅₀ :	Half-maximal inhibitory concentration
ED ₅₀ :	Median effective dose 50
HEK:	Human embryonic kidney cell line
SIRT1:	Silent information regulator sirutin1
COX:	Cyclooxygenase
LOX:	Lipoxygenase
DNA:	Deoxyribonucleic acid
DPPH:	2,2-Diphenyl-1-picrylhydrazyl
NADP:	Nicotinamide adenine dinucleotide phosphate
ADME:	Absorption, distribution, metabolism, excretion
ABTS:	2,2'-Azino-bis(3-ethylbenzothiazoline-6-sulfonic acid)
FRAP:	Ferric reducing antioxidant power
TEAC:	Trolox equivalent antioxidant capacity
REMA:	Resazurin microtiter assay plate

Ethical statement

This study doesn't involve the use of any humans or animals.

Data availability

All data has been obtained from peer-reviewed articles cited in the reference list, with no additional datasets utilized.

Author contributions

Glanish Jude Martis: writing – original draft, software.
Santosh L. Gaonkar: writing – review & editing, supervision.

Conflicts of interest

On behalf of the authors, the corresponding author declare no interests.

References

- G. Kumar and R. Shankar, *ChemMedChem*, 2021, **16**, 430–447.
- S. Tilvi and K. S. Singh, *Curr. Org. Chem.*, 2016, **20**, 898–929.
- C. P. Pandhurnekar, H. C. Pandhurnekar, A. J. Mungole, S. S. Butoliya and B. G. Yadao, *J. Heterocycl. Chem.*, 2023, **60**, 537–565.
- J. Zhu, J. Mo, H. Lin, Y. Chen and H. Sun, *Bioorg. Med. Chem.*, 2018, **26**, 3065–3075.
- A. V. Galenko, A. F. Khlebnikov, M. S. Novikov, V. V. Pakalnis and N. V. Rostovskii, *Russ. Chem. Rev.*, 2015, **84**, 335–377.
- M. Gul and S. Eryilmaz, *Lett. Org. Chem.*, 2019, **16**, 501–510.
- V. V. Pothuri, P. V. S. Machiraju and V. S. S. Rao, *Russ. J. Gen. Chem.*, 2020, **90**, 889–894.
- M. Aarjane, S. Slassi, A. Ghaleb, B. Tazi and A. Amine, *Arabian J. Chem.*, 2021, **14**, 103057.
- A. Shaik, R. R. Bhandare, K. Palleapati, S. Nissankararao, V. Kancharlapalli and S. Shaik, *Molecules*, 2020, **25**, 1047.
- O. S. Trefzger, N. V. Barbosa, R. L. Scapolatempo, A. R. das Neves, M. L. F. S. Ortale, D. B. Carvalho, A. M. Honorato, M. R. Fragoso, C. Y. K. Shuiguemoto, R. T. Perdomo, M. F. C. Matos, M. R. Chang, C. C. P. Arruda and A. C. M. Baroni, *Arch. Pharm.*, 2020, **353**, 1900241.
- T. Zhang, M. Dong, J. Zhao, X. Zhang and X. Mei, *J. Pestic. Sci.*, 2019, **44**, 181–185.
- G. C. Arya, K. Kaur and V. Jaitak, *Eur. J. Med. Chem.*, 2021, **221**, 113511.
- K. Kaur, V. Kumar, A. K. Sharma and G. K. Gupta, *Eur. J. Med. Chem.*, 2014, **77**, 121–133.
- S. Huang, B. Zhu, K. Wang, M. Yu, Z. Wang, Y. Li, Y. Liu, P. Zhang, S. Li, Y. Li, A. Liu and Q. Wang, *Pest Manage. Sci.*, 2022, **78**, 2011–2021.
- A. A. Abu-Hashem and M. El-Shazly, *Med Chem*, 2018, **14**, 356–371.
- F. V. B. Mota, M. S. de Araújo Neta, E. de Souza Franco, I. V. G. A. Bastos, L. C. C. da Araújo, S. C. da Silva, T. B. de Oliveira, E. K. Souza, V. M. de Almeida, R. M. Ximenes, M. B. de Sousa Maia, F. J. B. M. Junior, P. Marchand, A. R. de Faria and T. G. da Silva, *MedChemComm*, 2019, **10**, 1916–1925.
- Z. Li, C. Liu, W. Shi, X. Cai, Y. Dai, C. Liao, W. Huang and H. Qian, *Bioorg. Med. Chem.*, 2018, **26**, 703–711.
- S. Fettach, F. Z. Thari, Z. Hafidi, K. Karrouchi, K. Bouathmany, Y. Cherrah, M. El Achouri, L. Benbacer, M. El Mzibri, H. Sefrioui, K. Bougrin and M. E. A. Faouzi, *J. Biomol. Struct. Dyn.*, 2023, **41**, 1072–1084.
- K. V. Chikkula and R. Sundararajan, *Med. Chem. Res.*, 2017, **26**, 3026–3037.



- 20 A. Rastegari, M. Safavi, F. Vafadarnejad, Z. Najafi, R. Hariri, S. N. A. Bukhari, A. Iraj, N. Edraki, O. Firuzi, M. Saeedi, M. Mahdavi and T. Akbarzadeh, *Mol. Diversity*, 2022, **26**, 409–428.
- 21 P. Patil, A. Thakur, A. Sharma and S. J. S. Flora, *Drug Dev. Res.*, 2020, **81**, 165–183.
- 22 A. Sysak and B. Obmińska-Mrukowicz, *Eur. J. Med. Chem.*, 2017, **137**, 292–309.
- 23 M. Tugrak, H. I. Gul, K. Bandow, H. Sakagami, I. Gulcin, Y. Ozkay and C. T. Supuran, *Bioorg. Chem.*, 2019, **90**, 103095.
- 24 A. Sysak and B. Obmińska-Mrukowicz, *Eur. J. Med. Chem.*, 2017, **137**, 292–309.
- 25 H. Yao, J. Liu, S. Xu, Z. Zhu and J. Xu, *Expert Opin. Drug Discovery*, 2017, **12**, 121–140.
- 26 R. L. Barbieri, S. Evans and R. W. Kistner, *Fertil. Steril.*, 1982, **37**, 737–746.
- 27 H. A. Ramawad, P. Paridari, S. Jabermoradi, P. Gharin, A. Toloui, S. Safari and M. Yousefifard, *Korean J. Pain*, 2023, **36**, 425–440.
- 28 D. Rivera-Illanes and G. Recabarren-Gajardo, *ACS Chem. Neurosci.*, 2024, **15**, 3257–3269.
- 29 I. E. Leppik, *Seizure*, 2004, **13**, S5–S9.
- 30 C. Lamberth, *J. Heterocycl. Chem.*, 2018, **55**, 2035–2045.
- 31 K. E. Pallett, S. M. Cramp, J. P. Little, P. Veerasekaran, A. J. Crudace and A. E. Slater, *Pest Manage. Sci.*, 2001, **57**, 133–142.
- 32 S. Costanzi, T. Santhosh Kumar, R. Balasubramanian, T. Kendall Harden and K. A. Jacobson, *Bioorg. Med. Chem.*, 2012, **20**, 5254–5261.
- 33 Y. Kanda, T. Takahashi, Y. Araki, T. Konoike, S. Mihara and M. Fujimoto, *Bioorg. Med. Chem. Lett.*, 2000, **10**, 1875–1878.
- 34 S. Dadiboyena and A. Nefzi, *Eur. J. Med. Chem.*, 2010, **45**, 4697–4707.
- 35 M. L. Chavez and C. J. DeKorte, *Clin. Ther.*, 2003, **25**, 817–851.
- 36 S. Jeanmart, A. J. F. Edmunds, C. Lamberth, M. Pouliot and J. A. Morris, *Bioorg. Med. Chem.*, 2021, **39**, 116162.
- 37 M. Asahi, M. Kobayashi, T. Kagami, K. Nakahira, Y. Furukawa and Y. Ozoe, *Pestic. Biochem. Physiol.*, 2018, **151**, 67–72.
- 38 D. Kim, M.-S. Kang, J. S. Kim and J.-H. Jeong, *Arch. Pharmacol. Res.*, 2005, **28**, 1019–1022.
- 39 M. Agrawal, S. Saraf, S. Saraf, S. G. Antimisariar, M. B. Chougule, S. A. Shoyele and A. Alexander, *J. Controlled Release*, 2018, **281**, 139–177.
- 40 R. I. Fox, M. L. Herrmann, C. G. Frangou, G. M. Wahl, R. E. Morris, V. Strand and B. J. Kirschbaum, *Clin. Immunol.*, 1999, **93**, 198–208.
- 41 J. R. Romero, *Expert Opin. Invest. Drugs*, 2001, **10**, 369–379.
- 42 J. K. Aronson, In *Meyler's Side Effects of Drugs*, Elsevier, 2016, p. 834.
- 43 D. Della Bella, *Respiration*, 1989, **55**, 10–14.
- 44 J. Davidson and C. Turnbull, *J. Affective Disord.*, 1983, **5**, 183–189.
- 45 S. Rostamizadeh, Z. Daneshfar and H. Moghimi, *Eur. J. Med. Chem.*, 2019, **171**, 364–371.
- 46 S. Obermaier and M. Müller, *Angew. Chem.*, 2020, **132**, 12532–12535.
- 47 R. J. Noveck and R. C. Hubbard, *J. Clin. Pharmacol.*, 2004, **44**, 474–480.
- 48 L. Johnson, J. Powers, F. Ma, K. Jendza, B. Wang, E. Meredith and N. Mainolfi, *Synthesis*, 2012, **45**, 171–173.
- 49 C.-J. Hsieh, K. Xu, I. Lee, T. J. A. Graham, Z. Tu, D. Dhavale, P. Kotzbauer and R. H. Mach, *ACS Omega*, 2018, **3**, 4486–4493.
- 50 N. Agrawal and P. Mishra, *Med. Chem. Res.*, 2018, **27**, 1309–1344.
- 51 P. Anand and B. Singh, *Mini-Rev. Med. Chem.*, 2014, **14**, 623–627.
- 52 M. A. Barmade, P. R. Murumkar, M. Kumar Sharma and M. Ram Yadav, *Curr. Top. Med. Chem.*, 2016, **16**, 2863–2883.
- 53 T. Morita, S. Yugandar, S. Fuse and H. Nakamura, *Tetrahedron Lett.*, 2018, **59**, 1159–1171.
- 54 J.-S. Poh, C. García-Ruiz, A. Zúñiga, F. Meroni, D. C. Blakemore, D. L. Browne and S. V. Ley, *Org. Biomol. Chem.*, 2016, **14**, 5983–5991.
- 55 A. Khutorianskyi, B. Chalyk, P. Borysko, A. Kondratiuk and P. K. Mykhailiuk, *Eur. J. Org. Chem.*, 2017, **2017**, 3935–3940.
- 56 Y. Guo, X. Wang, Z. Zhu, J. Zhang and Y. Wu, *Synlett*, 2016, **27**, 2259–2263.
- 57 S. Okusu, E. Tokunaga and N. Shibata, *Org. Lett.*, 2015, **17**, 3802–3805.
- 58 R. M. Reja, S. Sunny and H. N. Gopi, *Org. Lett.*, 2017, **19**, 3572–3575.
- 59 S. Kuribayashi, N. Shida, S. Inagi and T. Fuchigami, *Tetrahedron*, 2016, **72**, 5343–5349.
- 60 G.-W. Wang, M.-X. Cheng, R.-S. Ma and S.-D. Yang, *Chem. Commun.*, 2015, **51**, 6308–6311.
- 61 M. Shanmugasundaram, A. Senthilvelan and A. R. Kore, *Tetrahedron Lett.*, 2020, **61**, 152464.
- 62 C. Chen and S. Cui, *J. Org. Chem.*, 2019, **84**, 12157–12164.
- 63 A. Savateev, N. V. Tarakina, V. Strauss, T. Hussain, K. ten Brummelhuis, J. M. Sánchez Vadillo, Y. Markushyna, S. Mazzanti, A. P. Tyutyunnik, R. Walczak, M. Oschatz, D. M. Guldi, A. Karton and M. Antonietti, *Angew. Chem., Int. Ed.*, 2020, **59**, 15061–15068.
- 64 X.-D. Wang, L.-H. Zhu, P. Liu, X.-Y. Wang, H.-Y. Yuan and Y.-L. Zhao, *J. Org. Chem.*, 2019, **84**, 16214–16221.
- 65 Y. Chen, L. Li, X. He and Z. Li, *ACS Catal.*, 2019, **9**, 9098–9102.
- 66 Z. Tang, Y. Zhou and Q. Song, *Org. Lett.*, 2019, **21**, 5273–5276.
- 67 T. N. Ngo, S. A. Ejaz, T. Q. Hung, T. T. Dang, J. Iqbal, J. Lecka, J. Sévigny and P. Langer, *Org. Biomol. Chem.*, 2015, **13**, 8277–8290.
- 68 J. Li, W. Ma, W. Ming, C. Xu, N. Wei and M. Wang, *J. Org. Chem.*, 2015, **80**, 11138–11142.
- 69 G. R. Kumar, Y. K. Kumar and M. S. Reddy, *Chem. Commun.*, 2016, **52**, 6589–6592.
- 70 M. V. Andreev, A. S. Medvedeva, L. I. Larina and M. M. Demina, *Mendeleev Commun.*, 2017, **27**, 175–177.



- 71 O. B. Bondarenko, A. A. Vinogradov, P. A. Danilov, S. N. Nikolaeva, A. Yu. Gavrilova and N. V. Zyk, *Tetrahedron Lett.*, 2015, **56**, 6577–6579.
- 72 S. Xue, J. Liu and C. Wang, *Eur. J. Org. Chem.*, 2016, **2016**, 2450–2456.
- 73 P. Kulkarni, *J. Indian Chem. Soc.*, 2021, **98**, 100013.
- 74 M. Zouhair, L. El Ghayati, H. El Monfalouti, H. Abchihi, T. Hökelek, M. Ahmed, J. T. Mague and N. K. Sebbar, *Acta Crystallogr., Sect. E: Crystallogr. Commun.*, 2023, **79**, 1179–1182.
- 75 R. Zheng, F. Feng, Z. Zhang, J. Fu, Q. Su, Y. Zhang and Q. Gu, *Mol. Diversity*, 2020, **24**, 423–435.
- 76 N. Sawengngan, A. A. Kolodina and O. V. Serdyuk, *Molbank*, 2019, **2019**, M1081.
- 77 K. C. Panda, B. V. V. R. Kumar and B. M. Sahoo, *Nat. Volatiles Essent. Oils*, 2021, **8**, 11503–11510.
- 78 F. K. Algethami, I. Saidi, H. N. Abdelhamid, M. R. Elamin, B. Y. Abdulkhair, A. Chrouda and H. Ben Jannet, *Molecules*, 2021, **26**, 5214.
- 79 I. Nakamura, S. Gima, Y. Kudo and M. Terada, *Angew. Chem., Int. Ed.*, 2015, **54**, 7154–7157.
- 80 S. Gima, I. Nakamura and M. Terada, *Eur. J. Org. Chem.*, 2017, **2017**, 4375–4378.
- 81 M. Jonušis, L. Šteinys, R. Bukšnaitienė and I. Čikotienė, *Synthesis*, 2016, **49**, 1122–1130.
- 82 W. Kaewsri, C. Thongsornkleeb, J. Tummatorn and S. Ruchirawat, *RSC Adv.*, 2016, **6**, 48666–48675.
- 83 D.-C. Guo, C. Zhang, F. Li, F. Zhang, F. Yu and Y.-P. He, *Synthesis*, 2016, **49**, 1356–1370.
- 84 T. J. Greshock, K. P. Moore, R. T. McClain, A. Bellomo, C. K. Chung, S. D. Dreher, P. S. Kutchukian, Z. Peng, I. W. Davies, P. Vachal, M. Ellwart, S. M. Manolikakes, P. Knochel and P. G. Nantermet, *Angew. Chem., Int. Ed.*, 2016, **55**, 13714–13718.
- 85 M. Shigenobu, K. Takenaka and H. Sasai, *Angew. Chem., Int. Ed.*, 2015, **54**, 9572–9576.
- 86 X. Zhang, G. Liu, X. Sun, L.-S. Wan and Y. Zhou, *J. Org. Chem.*, 2024, **89**, 14591–14595.
- 87 K. Sato, G. Sandford, K. Shimizu, S. Akiyama, M. J. Lancashire, D. S. Yufit, A. Tarui, M. Omote, I. Kumadaki, S. Harusawa and A. Ando, *Tetrahedron*, 2016, **72**, 1690–1698.
- 88 D. Zhu, Y. Gu, L. Lu and Q. Shen, *J. Am. Chem. Soc.*, 2015, **137**, 10547–10553.
- 89 D. Zhu, X. Hong, D. Li, L. Lu and Q. Shen, *Org. Process Res. Dev.*, 2017, **21**, 1383–1387.
- 90 D. F. Basri, L. W. Xian, N. I. Abdul Shukor and J. Latip, *BioMed Res. Int.*, 2014, **2014**, 1–8.
- 91 F. Baquero and B. R. Levin, *Nat. Rev. Microbiol.*, 2021, **19**, 123–132.
- 92 C. Llor, A. Moragas, C. Bayona, R. Morros, H. Pera, O. Plana-Ripoll, J. M. Cots and M. Miravittles, *BMJ*, 2013, **347**, f5762.
- 93 T. Jancel, *West. J. Med.*, 2002, **176**, 51–55.
- 94 M. Pasticci, A. Moretti, G. Stagni, V. Ravasio, L. Soavi, A. Raglio, F. Vailati, A. Cardaccia, A. Santucci, R. Papili, A. Sgrelli, C. Pallotto and F. Baldelli, *Ann. Clin. Microbiol. Antimicrob.*, 2011, **10**, 26.
- 95 M. N. de Menezes, B. A. de Marco, F. A. M. Fiorentino, A. Zimmermann, A. C. Kogawa and H. R. N. Salgado, *Crit. Rev. Anal. Chem.*, 2019, **49**, 67–77.
- 96 S. S. Castle, in *xPharm: The Comprehensive Pharmacology Reference*, Elsevier, 2007, pp. 1–5.
- 97 T. Akbarzadeh, A. Fallah Tafti, N. Samadi, A. Foroumadi, M. Amanlou, M. A. Faramarzi and A. Shafiee, *Daru, J. Fac. Pharm., Tehran Univ. Med. Sci.*, 2010, **18**, 118–123.
- 98 E.-J. Im, C.-H. Lee, P.-G. Moon, G. G. Rangaswamy, B. Lee, J. M. Lee, J.-C. Lee, J.-G. Jee, J.-S. Bae, T.-K. Kwon, K.-W. Kang, M.-S. Jeong, J.-E. Lee, H.-S. Jung, H.-J. Ro, S. Jun, W. Kang, S.-Y. Seo, Y.-E. Cho, B.-J. Song and M.-C. Baek, *Nat. Commun.*, 2019, **10**, 1387.
- 99 P. Fonseka, S. V. Chitti, R. Sanwani and S. Mathivanan, *Nat. Commun.*, 2021, **12**, 977.
- 100 H. Sudhamani, S. T. Basha, N. Venkateswarlu, T. Vijaya and C. N. Raju, *J. Chem. Sci.*, 2015, **127**, 1739–1746.
- 101 N. S. Belavagi, M. G. Sunagar, R. S. Lamani, N. Deshapande and I. A. M. Khazi, *Phosphorus, Sulfur Silicon Relat. Elem.*, 2015, **190**, 1580–1587.
- 102 A. Solankee and R. Tailor, *Elixir Org. Chem.*, 2015, **87**, 35620–35627.
- 103 A. Galbiati, A. Zana, C. Borsari, M. Persico, S. Bova, O. Tkachuk, A. I. Corfu, L. Tamborini, N. Basilico, C. Fattorusso, S. Bruno, S. Parapini and P. Conti, *Molecules*, 2023, **28**, 3172.
- 104 J. Zhu, J. Mo, H. Lin, Y. Chen and H. Sun, *Bioorg. Med. Chem.*, 2018, **26**, 3065–3075.
- 105 R. Wu, W. Ding, T. Liu, H. Zhu, Y. Hu, B. Yang and Q. He, *Cancer Lett.*, 2009, **285**, 13–22.
- 106 R. C. Montenegro, P. G. K. Clark, A. Howarth, X. Wan, A. Ceroni, P. Siejka, G. A. Nunez-Alonso, O. Monteiro, C. Rogers, V. Gamble, R. Burbano, P. E. Brennan, C. Tallant, D. Ebner, O. Fedorov, E. O'Neill, S. Knapp, D. Dixon and S. Müller, *Oncotarget*, 2016, **7**, 43997–44012.
- 107 G. Augello, M. R. Emma, A. Cusimano, A. Azzolina, S. Mongiovi, R. Puleio, G. Cassata, A. Gulino, B. Belmonte, R. Gramignoli, S. C. Strom, J. A. McCubrey, G. Montalto and M. Cervello, *Int. J. Cancer*, 2019, **144**, 2613–2624.
- 108 M. R. Jensen, J. Schoepfer, T. Radimerski, A. Massey, C. T. Guy, J. Brueggen, C. Quadt, A. Buckler, R. Cozens, M. J. Drysdale, C. Garcia-Echeverria and P. Chène, *Breast Cancer Res.*, 2008, **10**, R33.
- 109 K. Lee, J. Lee, S. Han, S. Im, T. Kim, D. Oh and Y. Bang, *Cancer Sci.*, 2011, **102**, 1388–1395.
- 110 A. M. Eid, M. Hawash, J. Amer, A. Jarrar, S. Qadri, I. Alnimer, A. Sharaf, R. Zalmoot, O. Hammoudie, S. Hameedi and A. Mousa, *BioMed Res. Int.*, 2021, **2021**, 1–9.
- 111 M. Hawash, N. Jaradat, N. Bawwab, K. Salem, H. Arafat, Y. Hajjousef, T. Shtayeh and S. Sobuh, *Heterocycl. Commun.*, 2021, **21**, 133–141.
- 112 K. Vashisht, P. Sethi, A. Bansal, T. Singh, R. Kumar, H. S. Tuli and S. Saini, *Eur. J. Clin. Exp. Med.*, 2024, **22**, 376–387.



- 113 A. Jarina, S. Kavimani, V. M. Mounnissamy and J. Abdul, *J. Pharm. Negat. Results*, 2022, **13**, 2634–2642.
- 114 N. Panathur, N. Gokhale, U. Dalimba, P. V. Koushik, P. Yogeewari and D. Sriram, *Bioorg. Med. Chem. Lett.*, 2015, **25**, 2768–2772.
- 115 S. Fiorucci, R. Meli, M. Bucci and G. Cirino, *Biochem. Pharmacol.*, 2001, **62**, 1433–1438.
- 116 C. D. Funk, *Science*, 2001, **294**, 1871–1875.
- 117 C. Charlier and C. Michaux, *Eur. J. Med. Chem.*, 2003, **38**, 645–659.
- 118 A. Urdaneta, A. Siso, B. Urdaneta, R. Cardenas, L. Quintero, R. Avila and H. Suarez-Roca, *Brain Res. Bull.*, 2009, **80**, 56–61.
- 119 J. J. Talley, D. L. Brown, J. S. Carter, M. J. Graneto, C. M. Koboldt, J. L. Masferrer, W. E. Perkins, R. S. Rogers, A. F. Shaffer, Y. Y. Zhang, B. S. Zweifel and K. Seibert, *J. Med. Chem.*, 2000, **43**, 775–777.
- 120 N. A. Nussmeier, A. A. Whelton, M. T. Brown, R. M. Langford, A. Hoeft, J. L. Parlow, S. W. Boyce and K. M. Verburg, *N. Engl. J. Med.*, 2005, **352**, 1081–1091.
- 121 M. L. Pati, P. Vitale, S. Ferorelli, M. Iaselli, M. Miciaccia, A. Boccarelli, G. D. Di Mauro, C. G. Fortuna, T. F. Souza Domingos, L. C. Rodrigues Pereira da Silva, M. de Pádula, L. M. Cabral, P. C. Sathler, A. Vacca, A. Scilimati and M. G. Perrone, *Eur. J. Med. Chem.*, 2019, **164**, 59–76.
- 122 G. Cingolani, A. Panella, M. G. Perrone, P. Vitale, G. Di Mauro, C. G. Fortuna, R. S. Armen, S. Ferorelli, W. L. Smith and A. Scilimati, *Eur. J. Med. Chem.*, 2017, **138**, 661–668.
- 123 M. Cutolo, S. Capellino, P. Montagna, A. Sulli, B. Serio and B. Villaggio, *Ann. Rheum. Dis.*, 2006, **65**, 728–735.
- 124 M. Zimmermann, *Pain*, 1983, **16**, 109–110.
- 125 E. K. A. Abdelall, *Bioorg. Chem.*, 2020, **94**, 103441.
- 126 R. Bhatia, A. Vyas, S. M. El-Bahy, M. M. Hessien, G. A. M. Mersal, M. M. Ibrahim, R. Dogra and B. Kumar, *ChemistrySelect*, 2022, **7**, e202200800.
- 127 H. M. Pallavi, F. H. Al-Ostoot, H. K. Vivek and S. A. Khanum, *J. Mol. Struct.*, 2022, **1250**, 131812.
- 128 W. Alam, H. Khan, M. S. Jan, H. W. Darwish, M. Daglia and A. A. Elhenawy, *PLoS One*, 2024, **19**, e0297398.
- 129 W. Alam, H. Khan, M. Saeed Jan, U. Rashid, A. Abusharha and M. Daglia, *Front. Chem.*, 2023, **11**, 1222047.
- 130 M. Hawash, N. Jaradat, M. Abualhasan, M. Thaher, R. Sawalhi, N. Younes, A. Shanaa, M. Nuseirat and A. Mousa, *Sci. Rep.*, 2022, **12**, 18223.
- 131 A. Arzine, O. Abchir, M. Chalkha, K. Chebbac, Y. Rhazi, N. Barghady, I. Yamari, A. EL Moussaoui, A. Nakkabi, M. Akhazzane, M. Bakhouch, S. Chtita and M. EL Yazidi, *Comput. Biol. Chem.*, 2024, **108**, 107993.
- 132 N. Myakala, V. Thumma, K. Kandula, N. Rayala, L. S. Boddu and K. D. B. Anagani, *Mol. Diversity*, 2024, DOI: [10.1007/s11030-024-11032-2](https://doi.org/10.1007/s11030-024-11032-2).
- 133 R. Re, N. Pellegrini, A. Proteggente, A. Pannala, M. Yang and C. Rice-Evans, *Free Radical Biol. Med.*, 1999, **26**, 1231–1237.
- 134 G. F. Makhaeva, N. A. Elkina, E. V. Shchegolkov, N. P. Boltneva, S. V. Lushchekina, O. G. Serebryakova, E. V. Rudakova, N. V. Kovaleva, E. V. Radchenko, V. A. Palyulin, Y. V. Burgart, V. I. Saloutin, S. O. Bachurin and R. J. Richardson, *Bioorg. Chem.*, 2019, **91**, 103097.
- 135 I. F. F. Benzie and J. J. Strain, *Methods Enzymol.*, 1999, **299**, 15–27.
- 136 G. F. Makhaeva, N. V. Kovaleva, E. V. Rudakova, N. P. Boltneva, S. V. Lushchekina, I. I. Faingold, D. A. Poletaeva, Y. V. Soldatova, R. A. Kotelnikova, I. V. Serkov, A. K. Ustinov, A. N. Proshin, E. V. Radchenko, V. A. Palyulin and R. J. Richardson, *Molecules*, 2020, **25**, 5891.
- 137 N. A. Elkina, E. V. Shchegolkov, Y. V. Burgart, N. A. Agafonova, A. N. Perminova, N. A. Gerasimova, G. F. Makhaeva, E. V. Rudakova, N. V. Kovaleva, N. P. Boltneva, O. G. Serebryakova, S. S. Borisevich, N. P. Evstigneeva, N. V. Zilberberg, N. V. Kungurov and V. I. Saloutin, *J. Fluorine Chem.*, 2022, **254**, 109935.
- 138 Q. Chai, Y. Zhang and C. H. Liu, *Front. Cell. Infect. Microbiol.*, 2018, **8**, 158.
- 139 M. A. De Groote and G. Huitt, *Clin. Infect. Dis.*, 2006, **42**, 1756–1763.
- 140 M. W. Bratschi, P. Steinmann, A. Wickenden and T. P. Gillis, *Lepr. Rev.*, 2015, **86**, 142–155.
- 141 V. N. Dahl, M. Mølhave, A. Fløe, J. van Ingen, T. Schön, T. Lillebaek, A. B. Andersen and C. Wejse, *Int. J. Infect. Dis.*, 2022, **125**, 120–131.
- 142 N. Scheinfeld, *Dermatol. Online J.*, 2016, **22**(6), 1–9.
- 143 J. Mao, H. Yuan, Y. Wang, B. Wan, M. Pieroni, Q. Huang, R. B. van Breemen, A. P. Kozikowski and S. G. Franzblau, *J. Med. Chem.*, 2009, **52**, 6966–6978.
- 144 A. Lilienkampf, J. Mao, B. Wan, Y. Wang, S. G. Franzblau and A. P. Kozikowski, *J. Med. Chem.*, 2009, **52**, 2109–2118.
- 145 P. P. Jain, M. S. Degani, A. Raju, A. Anantram, M. Seervi, S. Sathaye, M. Ray and M. G. R. Rajan, *Bioorg. Med. Chem. Lett.*, 2016, **26**, 645–649.
- 146 R. S. Keri and S. A. Patil, *Biomed. Pharmacother.*, 2014, **68**, 1161–1175.
- 147 N. Pappula and R. Sharabu, *J. Pharm. Res. Int.*, 2021, 69–79.
- 148 M. Girardini, F. Ferlenghi, G. Annunziato, G. Degiacomi, B. Papotti, C. Marchi, J. C. Sammartino, S. S. Rasheed, A. Contini, M. R. Pasca, F. Vacondio, J. C. Evans, T. Dick, R. Müller, G. Costantino and M. Pieroni, *Eur. J. Med. Chem.*, 2023, **245**, 114916.
- 149 S. K. Sahoo, B. Rani, N. B. Gaikwad, M. N. Ahmad, G. Kaul, M. Shukla, S. Nanduri, A. Dasgupta, S. Chopra and V. M. Yaddanapudi, *Eur. J. Med. Chem.*, 2021, **222**, 113580.

