



Journal of Analytical Atomic Spectrometry

## Accepted Manuscript

This article can be cited before page numbers have been issued, to do this please use: T. R. Schöberl, M. Bachmann and D. Gunther, *J. Anal. At. Spectrom.*, 2025, DOI: 10.1039/D5JA00171D.



This is an Accepted Manuscript, which has been through the Royal Society of Chemistry peer review process and has been accepted for publication.

Accepted Manuscripts are published online shortly after acceptance, before technical editing, formatting and proof reading. Using this free service, authors can make their results available to the community, in citable form, before we publish the edited article. We will replace this Accepted Manuscript with the edited and formatted Advance Article as soon as it is available.

You can find more information about Accepted Manuscripts in the Information for Authors.

Please note that technical editing may introduce minor changes to the text and/or graphics, which may alter content. The journal's standard <u>Terms & Conditions</u> and the <u>Ethical guidelines</u> still apply. In no event shall the Royal Society of Chemistry be held responsible for any errors or omissions in this Accepted Manuscript or any consequences arising from the use of any information it contains.



rsc.li/jaas

4

5 6

7 8

9

10g

115

12

13

145

44

45

46

47

48

49

50

51

52

53

54

55

56

57

58

59 60

## Quantification of Laser-Induced Aerosols by Microdroplet Calibration Vandclet Online Investigation of Matrix Effects Using LA-ICP-TOFMS

Tobias Schöberl<sup>†</sup>, Mirjam Bachmann<sup>†</sup>, Detlef Günther<sup>†,\*</sup>

<sup>†</sup>Laboratory of Inorganic Chemistry, Department of Chemistry and Applied Biosciences, ETH Zurich, 8093 Zurich, Switzerland

**ABSTRACT:** The application of monodisperse microdroplets for non-matrix-matched quantification in LA-ICP-TOFMS was investigated for inorganic and organic matrices. Suppression behavior in droplet signals caused by addition of typical major elements of geological samples (Al, Si, Ca, Ti, Fe) in the  $\mu$ g g<sup>-1</sup> range was studied using microdroplets introduced via a falling tube and compared to solution nebulization. Signal suppression patterns observed for microdroplets could be attributed to neither mass load effects nor in-plasma oxide formation, nor reproduced via solution nebulization, suggesting a fundamentally different behavior of microdroplets in the plasma. Radial diffusion profiles were acquired to assess in-plasma behavior of droplets and laser-induced aerosol from NIST SRM 610 (glass). Diffusion profiles overlapped and showed similar full width half maxima (FWHM) for microdroplets and the laser-induced aerosol, with minor spatial shifts in intensity maxima, likely due to not complete on-axis droplet introduction into the plasma. Quantification based on microdroplet calibration yielded relative deviations from reference values below ± 20 % across certified reference materials and an in-house prepared gelatine standard. Quantification of gelatine samples using NIST SRM 610 (glass) as an external standard resulted in larger deviations compared to droplet-based calibration, which yielded values in agreement with digestion data. These results demonstrate the suitability of monodisperse microdroplets for non-matrix-matched calibration in LA-ICP-MS.

#### Introduction

Since its introduction 40 years ago<sup>1</sup>, laser ablation inductively coupled plasma mass spectrometry (LA-ICP-MS) has become a widely applied analytical technique for multi-elemental analysis for major, minor and trace components, achieving detection limits at the fg to ag range<sup>2</sup>. Although LA-ICP-MS enables rapid analysis of a wide variety of solid samples with minimal material consumption, quantification can remain challenging due to matrix-dependent lasersample interactions.3 To address this, several strategies have been developed over the years. Matrix-matched quantification using certified reference materials (CRMs) or in-house prepared standards remain the most commonly employed approach<sup>4-7</sup>. Alternatively, non-matrix-matched quantification using liquid calibration standards was pioneered by Thompson et al. using ICP- atomic emission spectrometry (ICP-AES)<sup>8</sup>, followed by Cromwell et al. for ICP-MS<sup>9</sup>. Despite agreeing with literature values ( $\pm 25$  % deviation from reference values), Cromwell et al. reported that the high solvent load introduced by solution nebulization led to increased oxide formation resulting in reduced sensitivity and polyatomic interferences. To mitigate oxide formation, subsequent efforts focused on reducing solvent load by direct ablation of liquids ( $\pm$  10 % deviation from reference values)<sup>10</sup> or desolvation systems ( $\pm$  25 % deviation from reference values)<sup>11,12</sup>. However, elemental losses during desolvation impeded accurate quantification of, e.g., Cu, as reported by Halicz et al.<sup>12</sup> Such elemental losses by desolvation have also been reported for elements Ga, Ge, Zn<sup>13</sup>, Mo<sup>14</sup> and B<sup>15</sup>. Alternative strategies for the quantification of solids with liquids include ablation of micro dried droplets<sup>16</sup>, ablation from self-aliquoting micro-grooves<sup>17</sup> or laser ablation of solids in liquids (LASIL)<sup>18,19</sup>. More recently, Mervič et al.<sup>20,21</sup> demonstrated a non-matrix-matched calibration approach based on ablation volume normalization by well characterizing the sample surface morphology before and after ablation using optical profilometry, applied successfully across diverse materials. Most of these approaches employ solution nebulization to introduce the liquid calibrant, producing a polydisperse aerosol that is transported into the ICP. An alternative approach involves the use of monodisperse microdroplets, which have been used as a tool for investigation of fundamental flame<sup>22</sup> and plasma<sup>23</sup> processes and gained renewed interest in the past 15 years for applications in single-particle ICP-MS (sp-ICP-MS)<sup>24-29</sup>. Microdroplets can be introduced discretely into the ICP via a falling tube<sup>24</sup> using He/Ar as drying and transport gases<sup>30</sup> and are detected using either SFMS, QMS or TOFMS technology. Upon drying, the dissolved salts crystallize to form a nanoparticle-like salt residue<sup>28</sup>, which is expected to exhibit in-plasma behavior similar to nanoparticles and agglomerates generated by laser ablation<sup>31</sup>. Borovinskaya<sup>32</sup> demonstrated this concept on NIST SRM 610 (glass), reporting deviations of  $\pm$  20 % from certified values, highlighting the potential of this approach for non-matrix-matched quantification.

This work investigates the suitability of monodisperse microdroplets for non-matrix-matched quantification in LA-ICP-TOFMS across different sample matrices and wide range of elements. Matrix-induced suppression of analyte signals in droplets was studied and compared to conventional solution nebulization (SN). In addition, in-plasma behavior of microdroplets and laser-induced aerosols was compared by radial diffusion profiles of various analytes. Finally, achieved quantification results will be discussed.

#### Experimental

#### Instrumentation

#### View Article Online DOI: 10.1039/D5JA00171D

All measurements were carried out using an ArF\* 193 nm excimer laser system (GeoLas C, Lambda Physik, Germany) with an implemented modified parallel flow ablation cell (MPFAC), as described elsewhere<sup>33</sup>. The MPFAC was operated with a carrier gas mixture of He (0.65 to 0.8 L min<sup>-1</sup>) and Ar (0.55 to 0.75 L min<sup>-1</sup>) and connected to an adapted dual sample introduction setup<sup>25</sup>, allowing for the combined introduction of laser-induced aerosols and microdroplets. Aerosols were introduced into an ICP time of flight (TOF) mass spectrometer (icpTOF2R, TOFWERK AG, Switzerland), enabling quasi-simultaneous detection over a mass range of m/z = 14 to 254<sup>34</sup>. The instrument was tuned daily to ensure high sensitivity, a <sup>232</sup>Th<sup>16</sup>O<sup>+/232</sup>Th<sup>+</sup> formation rate below 1 %, a <sup>238</sup>U<sup>+/232</sup>Th<sup>+</sup> ratio of 0.9 to 1.1 and expected isotope abundance patterns for the REEs. Background correction was performed using gas blanks for LA measurements and an acidified blank solution with Cs as tracer for microdroplet measurements.

Monodisperse microdroplets were generated using an Autodrop Pipette (AD-KH-501-L6) with a nominal nozzle size of 50  $\mu$ m and an external control unit (microdrop technologies, Germany). The resulting droplets ranged in diameter from 60-75  $\mu$ m. Droplets were focused into the falling tube, dried and transported using a He flow (0.4 to 0.75 L min<sup>-1</sup>) before being merged with the LA carrier gas flow via a T-piece. No active heating was applied during droplet transport.

For quantification of gelatine digests, samples were measured employing ICP-optical emission spectrometry (ICP-OES) using a radial spectrometer (Arcos, Spectro Analytical Instruments, Germany) in Paschen-Runge geometry. The instrument was connected to a Scott-type spray chamber equipped with a crossflow nebulizer (Ar flow: 1 L min<sup>1</sup>). The instrument was tuned for maximum sensitivity, with an Ar(I) 404.442 nm intensity of 800 kcps to 1500 kcps and a Mg(II) 280.271 nm/ Mg(I) 285.213 nm ratio of >  $9.5^{35}$ .

All specific operating conditions used in this study are summarized in the SI in Tab. S 2.

#### Liquid sample preparation

All vials used were conditioned in 10 % sub-boiled HNO<sub>3</sub> (v/v) (p.a. > 65 %, Sigma Aldrich, Germany) and 10 % HCl (v/v) (p.a. > 37 %, Fisher Scientific, Germany) and stored under 1 % sub-boiled HNO<sub>3</sub>. Liquid calibration standards were prepared from single- and multi-element stock solutions (Inorganic Ventures, USA; Merck, Germany) and stabilized in diluted sub-boiled HNO<sub>3</sub> and HCl prepared in ultrapure water (18.2 MΩcm, Millipore, USA). All solutions were prepared gravimetrically with an accuracy of 0.1 mg (Mettler Toledo, Germany). For parameter studies, analytes were limited to Sr, Rh, In, the rare earth elements (REEs) (Sc, Y, La, Ce, Pr, Nd, Sm, Eu, Tb, Dy, Ho, Er, Tm, Yb, Lu), Th and U.

Gelatine standards were prepared from porcine gelatine (Sigma Aldrich, Germany). 1 g of gelatine was mixed with 9 g of ultrapure water and shaken vigorously and heated to approx. 60 °C to 70 °C. After complete dissolution, element standards (Mg, Cr, Mn, Ni, Co, Cu, Zn, Ga, Sr, Ag, Ce, Pr, Tm, Tl, Bi, Pb) were added to yield a final nominal concentration of 250  $\mu$ g g<sup>-1</sup> for each analyte. The mixture was homogenized and 10  $\mu$ L droplets were placed on a microscope slide manually using an air displacement pipette (Eppendorf, Germany) and dried overnight at 100 °C before use.

Gelatine standards were digested by weighing 30 mg of each sample (6 replicates) and adding 0.5 mL sub-boiled HNO<sub>3</sub>, 0.15 mL H<sub>2</sub>O<sub>2</sub> (35 %, Acros Organics, Netherlands) and 0.15 mL ultrapure water. All samples were digested using a turboWAVE microwave assisted digestion system (MLS GmBH, Germany). Following digestion, vessels were rinsed three times with 3 mL of ultrapure water each and the digests were diluted to a final volume of 45 mL. Elemental analysis was carried out by SN-ICP-OES. Temperature ramp and maximum power settings for the digestion are provided in Fig. S 38 and Tab. S 1.

### **Matrix Influence**

Matrix influence was first investigated using mixtures of HNO<sub>3</sub>/ HCl at varying concentrations (0.5 %/ 0.5 %, 1 %/ 1 %, 2 %/ 1 %, 1 %/ 2 %, 2 %/ 2 %). Further studies on matrix effects focused on selected matrix elements added individually into a multi-element solution (0.05  $\mu$ g g<sup>-1</sup> analytes) stabilized in 1 % sub-boiled HNO<sub>3</sub>. Equimolar fractions of Al (0-10  $\mu$ g g<sup>-1</sup>), Si (0-10  $\mu$ g g<sup>-1</sup>), Ca (0-15  $\mu$ g g<sup>-1</sup>), Ti (0-18  $\mu$ g g<sup>-1</sup>) or Fe (0-20  $\mu$ g g<sup>-1</sup>) were added individually to identical multi-element solutions. To investigate the combined effect of all selected matrix elements, solutions containing 3.7 nmol g<sup>-1</sup> to 370 nmol g<sup>-1</sup> of each element (Al, Si, Ca, Ti and Fe) were added to a multi element solution (0.05  $\mu$ g g<sup>-1</sup> analytes). Droplets were generated at 50 Hz dispensing rate and measured at a time resolution of 3 ms for a total duration of 1.5 min. Resulting signals were averaged over all droplets to obtain mean intensities from which sensitivities were calculated.

To compare droplet-based results with conventional sample introduction, all Al-containing solutions were also analyzed using solution nebulization ICP-MS (SN-ICP-MS). Measurements were carried out using a dual sample introduction setup<sup>25</sup> connected to a cyclonic double-pass spay chamber equipped with a microflow nebulizer. Data were acquired at a time resolution of 92 ms for a total of 60 s per condition.

### **Radial Diffusion Profiles**

1

Journal of Analytical Atomic Spectrometry Accepted Manuscript

4

5

6

7

8

9

10

115

12

13

14<u>5</u>

Rublished ow 7, Juliei 2025, Downloaded ea 280720, 19:26:14. 1. 1. 16 h: Reensed under a Greative Commons. Attribution-Rond Commercial 339.

Access Auticle, Rut

44

45 46

47

48

49

50

51

52

53

54

55

56

57

58 59 60

# Measurements were performed at a sampling depth of 3.5 mm, which represented the shortest achievable position/timdericle Online the given instrumental conditions. This was done to minimize sensitivity losses from radial diffusion and to replicate

typical quantification conditions. To ensure consistency, radial diffusion profiles were always recorded in the same horizontal direction (right to left). For each profile (microdroplets, laser aerosol), the scanning range was defined by first locating the position of maximum signal intensity of the  $^{238}$ U<sup>+</sup> signal and moving laterally across the plasma until signal intensity decreased to 10 % of its maximum value. From this position, data were acquired in horizontal steps of 0.3 mm until the maximum signal was reached. Beyond the maximum, step size was reduced to 0.15 mm and data acquisition continued until a 99 % signal decrease was observed or the instrument limit was reached. Diffusion profiles were recorded at 3 ms time resolution for dry laser ablation of NIST SRM 610 (glass, 44 µm spot size, 5 x 5 raster, 7 shots per spot, 10 Hz) and for microdroplets containing a nominal analyte concentration of 0.05 µg g<sup>-1</sup> and Si concentration of 20 µg g<sup>-1</sup> dispensed at a frequency of 50 Hz.

#### **Quantification of Laser-induced Aerosols**

Proof of principle and optimization studies were conducted using NIST SRM 610 (glass). Preferred literature values for all CRMs were obtained from GeoRem database<sup>36</sup>. Laser ablation sampling was performed as a 10 x 10 raster with 7 shots per spot at 10 Hz repetition rate using a 44 µm spot size at a time resolution of 3 ms. This yielded 100 individual signal peaks, which were integrated for quantification. Droplet data were acquired at a dispensing rate of 50 Hz and processed in the same data structure. Signals from approx. 6000 droplets were integrated and averaged before being used to quantify LA intensities. Quantification was performed using microdroplets containing 0.25 µg g<sup>-1</sup> of Mg, Cr, Mn, Co, Ni, Cu, Zn, Ga, Sr, Ag, Tl, Pb, and Bi, and 0.05 µg g<sup>-1</sup> of Sc, Rh, Y, La, Ce, Pr, Nd, Sm, Eu, Gd, Tb, Dy, Ho, Er, Tm, Yb, Lu, Re, Pt, Au, Th and U. For all measurements, Sr was used as the internal standard. To evaluate applicability of the method, CRMs and in-house samples with varying matrix compositions were analyzed, including NIST SRM 612 (glass), USGS BCR-2G (basalt glass), USGS MACS-3 (carbonate) and in-house prepared gelatine standards. Quantification was performed of the microdroplet-based quantification with conventional approaches, all samples were quantified using NIST SRM 610 (glass) and the droplet-based calibrant as external standards. Quantification was carried out according to Longerich<sup>37</sup> (Equ. 2), using relative sensitivity factors (RSFs). The basic assumption is that the RSF determined using microdroplets equals the RSF of the laser-induced aerosol,

$$RSF_{Droplet} = RSF_{Laser Aerosol} \tag{1}$$

which can be expressed as:

$$\begin{pmatrix} \frac{I_A}{I_{IS}} \\ \frac{C}{C_{IS}} \end{pmatrix}_{Droplet} = \begin{pmatrix} \frac{I_A}{I_{IS}} \\ \frac{C}{C_{IS}} \end{pmatrix}_{Laser Aerosol}$$
(2)

#### **Data Processing**

Data from both laser ablation and droplet measurements were processed using custom RStudio<sup>38,39</sup> scripts (RStudio version 2024.12.1) and Microsoft Excel (Microsoft 365). Peak integration, averaging, and background correction were performed prior to quantification. Relative sensitivity factors (RSFs) were calculated based on the approach described by Longerich et al.<sup>6</sup>. Data visualization was carried out using OriginPro 2024 (OriginLab, USA).

#### **Results & Discussion**

#### **Matrix Influence**

The influence of Al on analyte sensitivity is depicted as relative signal suppression in Fig. 1 and Fig. 2 corresponding to sampling depths of 3 mm and 9 mm, respectively. Additional data on signal suppression of other matrix elements (Fig. S 1 to Fig. S 9), sensitivity curves (Fig. S 10 to Fig. S 19), <sup>232</sup>Th<sup>16</sup>O<sup>+</sup> formation rates and <sup>238</sup>U<sup>+/232</sup>Th<sup>+</sup> ratios (Fig. S 20 to Fig. S 23) are provided in the supplementary information (SI).

At a sampling depth of 3 mm, Al exerts a pronounced effect on the sensitivity of the rare earth elements (REEs) and Th, while elements Sr, Rh, In and U exhibit minimal to no signal suppression upon Al addition. Elements Ce, Eu and Yb display sensitivity spikes, whereas Y and Lu show the strongest observed signal suppression. Although Ce, Eu and Yb are part of redox pairs, similar trends are not observed for Sm and Tb.

No apparent correlation was observed between signal suppression or spike behavior and physicochemical properties such as ionization potential (IP), melting point ( $T_m$ ), boiling point ( $T_b$ ), heat capacity ( $c_P$ ) or magnetic susceptibility ( $\chi$ ). A minor mass bias in signal suppression is visible, with decreased suppression for the lighter REEs, with the exception of Y. Based on these findings, we propose three hypotheses to explain the observed selective signal suppression:

Journal of Analytical Atomic Spectrometry Accepted Manuscript

- i) Chemical Reactions during droplet transport or within the ICP, such as matrix-analyte cluster formationticle Online resulting in selective signal suppression
- ii) Oxide formation in the plasma caused by thermal pinching due to mass load of either matrix elements or water(l)
- iii) Delayed atomization due to mass load effects as already described by Murtazin et al.<sup>40</sup>

Cluster formation can be excluded as the main mechanism. Despite the observed strong signal suppression for elements Y and Lu, no increase in signal intensity was detected at m/z = 116 and m/z = 202, which would correspond to  ${}^{89}Y^{27}Al^+$  and  ${}^{175}Lu^{27}Al^+$ , respectively. Additionally, if cluster formation would play a significant role, a sensitivity spike should have been detected for  ${}^{166}Er$  due to the potential formation of  ${}^{139}La^{27}Al^+$ , which was not the case. However, other chemical reactions in the droplet or formation of a refractory crystalline phase cannot be excluded.

Fig. 3 and Fig. 4 illustrate relative signal suppression at single-element equimolar concentrations (ca. 0.2  $\mu$ mol g<sup>-1</sup>) for Al, Si, Ca, Ti and Fe at 3 mm and 9 mm sampling depths. Despite similar molar fractions, the extent of signal suppression varies between elements. As shown in Fig. 3, Al and Ti induce the strongest signal suppression, followed by Fe and Ca. The collective addition of all selected matrix elements led to significantly stronger signal suppression compared to the individual addition of single elements at equimolar concentrations (see Fig. S 9), particularly at low matrix element concentrations. At 0.5 µg g<sup>-1</sup> or 18.5 nmol g<sup>-1</sup> Al (blue line Figure 1), analyte signals were reduced by 1 % in average. In contrast, when 3.7 nmol g<sup>-1</sup> of each matrix element was added collectively (blue line Fig. S 9), the analyte signals were reduced by 12 % in average. Addition of 3.7 nmol g<sup>-1</sup> of each matrix element corresponds to the lowest molar concentration used in a single-element concentration series (for example, 0.5 µg g<sup>-1</sup> or 18.5 nmol g<sup>-1</sup> for Al). At higher matrix concentrations (i.e., 5 µg g<sup>-1</sup> and 10 µg g<sup>-1</sup> Al or 37 nmol g<sup>-1</sup> and 74 nmol g<sup>-1</sup> per element), the magnitude of signal suppression was similar for both single-element and collective matrix addition. Mass load effects can be ruled out, as Si addition does not affect analyte sensitivity. In contrast, oxide formation trends for Al, Ca, Ti and Fe show increasing oxide rates with increasing mass fraction for both sampling depths, shown in Fig. S 20 and Fig. S 21. This increase in oxides indicates higher energy consumption within the ICP, possibly resulting from matrix-induced processes or reactions. Regarding Si, oxide rates are unaffected by increasing mass fractions. The <sup>238</sup>U<sup>+/232</sup>Th<sup>+</sup> ratios (Fig. S 22 and Fig. S 23) show increases up to a factor of 13, while <sup>232</sup>Th<sup>16</sup>O<sup>+/232</sup>Th<sup>+</sup> (Fig. S 20 and Fig. S 21) formation rates increase only by a factor of up to ca. 4. This suggests that mere in-plasma oxide formation cannot account for the extent of signal suppression observed.

If additional matrix induced processes or reactions contribute to signal suppression, increased residence time in the plasma should mitigate these suppression effects. In order to put this hypothesis to test, measurements were repeated at a sampling depth of 9 mm (Fig. 2 and Fig. 4). For all affected elements, an increase in residence time significantly reduces or eliminates signal suppression effects at low matrix mass fractions. However, in order to compensate for matrix-induced processes by an increase in sampling depth, sensitivity decreases by a factor of approx. 4.

Hypotheses ii) and iii) are expected to affect analyte behavior similarly and should also be mitigated by an increase in plasma residence time. In order to further investigate analyte suppression, Al containing solutions were measured as well via SN-ICP-MS, employing either Ar as the nebulizer gas or an Ar/He mixture (Fig. S 24 and Fig. S 25) to better simulate drying conditions in the falling tube. SN-generated droplets via pneumatic nebulization are polydisperse and if hypotheses ii) and iii) are applicable, analyte behavior should be equally influenced irrespective of sample introduction. When Al containing solutions were introduced via SN, the resulting sensitivity curves overlapped for all Al mass fractions. This indicates that drying behavior is not influenced by Al addition, which would result in an increased mass load of water(l). Based on these reported findings, all of the 3 suggested hypotheses can be ruled out. In addition, signal suppression behavior seems to be connected to the introduction of the solutions as discrete monodisperse microdroplets instead of a polydisperse aerosol.

1





Figure 1: Relative signal suppression at 3 mm sampling depth of analytes for no added Al (black),  $0.5 \ \mu g \ g^{-1}$  (blue),  $1 \ \mu g \ g^{-1}$  (red),  $3 \ \mu g \ g^{-1}$  (green),  $5 \ \mu g \ g^{-1}$  (yellow) and  $10 \ \mu g \ g^{-1}$  (purple) of Al



9 mm Sampling Depth

Figure 2: Relative signal suppression at 9 mm sampling depth of analytes for no added Al (black),  $0.5 \ \mu g \ g^{-1}$  (blue),  $1 \ \mu g \ g^{-1}$  (red),  $3 \ \mu g \ g^{-1}$  (green),  $5 \ \mu g \ g^{-1}$  (yellow) and  $10 \ \mu g \ g^{-1}$  (purple) of Al



Figure 3: Relative signal suppression at 3 mm sampling depth of analytes for no added matrix element (black), 5  $\mu$ g g<sup>-1</sup> of Al (blue), 5  $\mu$ g g<sup>-1</sup> of Si (red), 7.5  $\mu$ g g<sup>-1</sup> of Ca (green), 9  $\mu$ g g<sup>-1</sup> of Ti (yellow) and 10  $\mu$ g g<sup>-1</sup> (purple) of Fe



Figure 4: Relative signal suppression at 9 mm sampling depth of analytes for no added matrix element (black), 5  $\mu$ g g<sup>-1</sup> of Al (blue), 5  $\mu$ g g<sup>-1</sup> of Si (red), 7.5  $\mu$ g g<sup>-1</sup> of Ca (green), 9  $\mu$ g g<sup>-1</sup> of Ti (yellow) and 10  $\mu$ g g<sup>-1</sup> (purple) of Fe

View Article Online

DOI: 10.1039/D5JA00171D

Journal of Analytical Atomic Spectrometry Accepted Manuscript

To compare the in-plasma behavior of laser-induced aerosol (NIST SRM 610 (glass)) and microdroplets, radial diffusionicle Online profiles were measured at sampling conditions optimized for quantification. If the evaporation, atomization and ionization position within the plasma differs between solid standard and liquid calibrant, differences in radial diffusion widths are expected: narrower profiles in comparison would indicate delayed evaporation, while broader profiles would suggest earlier evaporation. If the evaporation position in the plasma are comparable/similar, profiles should overlap and have similar widths based on the full width half maximum (FWHM).

Radial diffusion profiles are depicted for <sup>140</sup>Ce and <sup>28</sup>Si in Fig. 5 and Fig. 6, representing analytes at vastly different concentrations in NIST SRM 610 (glass), approximately by a factor of 700. To account for evaporation and diffusion differences related to the mass load introduced into the ICP, these two elements were selected due to their differing mass load and m/z values. Plotted diffusion profiles for additional mentioned elements are available in the SI (Fig. S 29 to Fig. S 37).

For both analytes, the diffusion profiles measured for droplets and laser-induced aerosol largely overlap and exhibit comparable FWHM values which ratios lay between 0.8 and 1.3, listed in Tab. S 9, indicating similar evaporation, atomization and ionization positions in the plasma. Slight horizontal offsets in the droplet profiles were observed, likely resulting from a difference in on-axis introduction of the microdroplets, as expected for an injector diameter of 2.5 mm.

While diffusion profiles generally overlap for the measured analytes, those of Y, Gd, Tb, Tm, Dy, Ho, Er, Lu and Th (Fig. S 29 to Fig. S 37) show overlap only across half of the curve, indicating earlier evaporation or an asymmetric ion cloud. The FWHM values of these elements are 30 % higher in droplets compared to other analytes, without affecting quantifiability. Despite these shifts in maximum intensity position and slight asymmetry, the overall similarity in profile shape and width suggests that droplets and laser-induced aerosol exhibit comparable behavior in the plasma under typical measurement conditions.



Figure 5: Radial diffusion intensity distribution of <sup>140</sup>Ce for dry ablated NIST SRM 610 (red) and microdroplets (blue) at 50 Hz dispensing frequency



Figure 6: Radial diffusion intensity distribution of <sup>28</sup>Si for dry ablated NIST SRM 610 (red) and microdroplets (blue) at 50 Hz dispensing frequency

#### **Quantification of Laser-induced Aerosols**

Droplet-based quantification was evaluated against NIST SRM 610 (glass) to assess performance across both matrixmatched standards (NIST SRM 612 (glass), USGS BCR-2G (basalt glass)) and non-matrix matched standards (USGS MACS-3 (carbonate), gelatine). The goal was to determine whether droplets can serve as a viable external standard for non-matrix matched quantification in LA-ICP-MS. Relative deviations from the preferred values listed in the GeoRem database<sup>36</sup> are shown in Fig. 7 to Fig. 10.

Across all analyzed matrices, the determined mass fractions were in agreement with literature values, with relative deviations below  $\pm$  20 %. This demonstrates that the microdroplet-based approach is suitable for both inorganic and proteinaceous matrices. However, internal standardization remains essential and total consumption approaches may be limited by differing detection efficiencies between microdroplets and laser-induced aerosols, which was beyond the scope of this study. Accuracy of the microdroplet-based calibration was comparable to NIST SRM 610 (glass) for all inorganic matrices investigated.

When used as external standard for gelatine quantification, NIST SRM 610 (glass) showed a deviation of  $\pm 30$  %, while microdroplet-based quantification resulted in  $\pm 15$  % of analyzed reference values. This highlights the applicability of droplets for non-matrix matched quantification when an internal standard can be used.

10g

In all discussed results, <sup>88</sup>Sr was used as an internal standard despite being a trace component in the used CRMs and the online gelatine standard. Ideally, a matrix element should serve as the internal standard, but this was initially avoided due to concerns regarding matrix-induced effects and potential HF traces in element standards, resulting in precipitation of targeted analytes. Since the quantification approach relies on the signal ratio between analyte and internal standard, and Sr may also be affected by signal suppression from certain matrix constituents, such effects inherently influence the ratio and cannot be fully corrected by internal standardization. As Si showed little to no influence on analyte behavior in earlier experiments, it was evaluated as an internal standard. However, signal-to-noise ratios remained poor due to low sensitivity in the low mass range, high background levels for all Si isotopes and necessary notch-filters, even when high amounts of Si (20  $\mu$ g g<sup>-1</sup>) were added to the calibrant. Consequently, accuracy decreased but was still in good agreement with literature values. Results for Si as internal standard can be found in the SI (Fig. S 28).



Figure 7: Deviation from reference values for NIST SRM 612 for quantification using NIST SRM 610 as external standard (black) and microdroplets (yellow); Microdroplets contained only target analytes (see Experimental Section)





View Article Online DOI: 10.1039/D5JA00171D



Figure 9: Deviation from reference values for USGS MACS-3 for quantification using NIST SRM 610 as external standard (black) and microdroplets (yellow); Microdroplets contained only target analytes (see Experimental Section)



Figure 10: Deviation from digestion data for in-house prepared gelatine standards for quantification using NIST SRM 610 as external standard (black) and microdroplets (yellow); Microdroplets contained only target analytes (see Experimental Section)

#### Conclusion

10g

Influence of matrix-induced effects for microdroplets was investigated and signal suppression behavior for Al, Si, Ca, Ti and Fe were evaluated. The observed suppression patterns can neither be explained by common effects such as mass load nor oxide formation. To determine whether these patterns are also apparent using other liquid sample introduction techniques, Al containing solutions were introduced via solution nebulization. No comparable signal suppression was observed, suggesting a unique behavior of discretely introduced microdroplets in the ICP. To further investigate potential differences in the in-plasma behavior between microdroplets and the laser-induced aerosol, radial diffusion profiles were measured without addition of matrix elements known to cause signal suppression. Microdroplets and laser-induced aerosol of NIST SRM 610 (glass) exhibit comparable in-plasma behavior based on the radial diffusion profiles. Although horizontal shifts between microdroplets and laser-induced aerosol suggest a difference in on-axis introduction of the droplets, which however, does not affect quantification accuracy. Quantification results of inorganic matrices using droplets show general deviations below  $\pm 20$  % and perform comparable to NIST SRM 610 (glass) when employed as matrix-matched and non-matrix-matched standard. While quantification of the in-house gelatine standard using NIST SRM 610 (glass) showed deviations  $\pm 30$  %, microdroplets yielded values  $\pm 15$  % compared to digestion data. However, an internal standard is required, as detection efficiencies cannot be assumed to be comparable. Despite the general agreement of  $\pm 20$  %, possible limitations of the proposed microdroplet-based quantification approach should be

4

5

6

7

8

9

10

115

12

13

145

44

45

46

47 48

49

50 51

52

53

54 55

56

57

58 59

60

considered. First, if laser-induced aerosols contain a significant fraction of large particles, incomplete transport to the cle Online plasma, e.g., due to particle loss within the tubing, or incomplete vaporization in the plasma can lead to inaccurate quantification. This is particularly relevant for aerosols generated under argon atmosphere or with longer wavelength lasers, both of which are known to produce broader particle size distributions and contribute to elemental fractionation.<sup>31,41,42</sup> Second, analytes that partially enter the plasma in gaseous form may lead to biased results, as differences in transport and ionization behavior between gaseous species and the particulate-phase are not accounted for in the current approach. Lastly, quantification may be affected if matrix-induced effects affect the in-plasma behavior of the laser aerosol such that the analyte and the internal standard no longer exhibit similar ionization characteristics compared to the microdroplets. These first two limitations can be mitigated by using short-pulsed, flat-top deep-UV lasers in combination with fast-washout ablation cells and helium as the carrier gas, which have been shown to reduce particle size<sup>41,42</sup>, improve transport characteristics<sup>43-45</sup>, and minimize elemental fractionation<sup>31,46,47</sup>. This study demonstrates that monodisperse microdroplets can be employed as a non-matrix-matched external standard for quantification of laser-induced aerosols in ICP-TOFMS. Microdroplets can be utilized for the quantitative analysis of a range of matrices, including inorganic and proteinaceous samples and offer great flexibility in analytes and concentration ranges.

#### **AUTHOR INFORMATION & CONTRIBUTION**

#### **Corresponding Author**

Detlef Günther – Laboratory of Inorganic Chemistry, Department of Chemistry and Applied Biosciences, ETH Zurich, Zurich 8093, Switzerland

Conceptualization, Formal analysis, Investigation, Project administration, Supervision, Funding acquisition, Writing – review & editing.

#### Authors

Tobias Schöberl – Laboratory of Inorganic Chemistry, Department of Chemistry and Applied Biosciences, ETH Zurich, Zurich 8093, Switzerland

 $Conceptualization, Formal \ analysis, Investigation, Writing-original \ draft, Writing-review \ \& \ editing.$ 

Mirjam Bachmann – Laboratory of Inorganic Chemistry, Department of Chemistry and Applied Biosciences, ETH Zurich, Zurich 8093, Switzerland Formal analysis, Investigation, Writing – review & editing.

#### **Declaration of competing interest**

There are no conflicts of interest to declare.

#### Data availability

The data supporting this article have been included as part of the SI.

#### Acknowledgements

The authors acknowledge the funding of this project by the Swiss National Science Foundation by project number 200021-231340.

The authors would like to thank Barbara Umfahrer, Bodo Hattendorf, Pascal Becker and Thomas Nauser for valuable discussions.

The authors gratefully acknowledge the anonymous reviewers for their constructive comments and suggestions, which helped to improve the clarity and quality of the manuscript.

#### References

- 1 A. L. Gray, *Analyst*, 1985, **110**, 551–556.
- 2 T. Van Acker, S. Theiner, E. Bolea-Fernandez, F. Vanhaecke and G. Koellensperger, *Nature Reviews Methods Primers*, 2023, **3**, 52.
- B. J. Fryer, S. E. Jackson and H. P. Longerich, *The Canadian Mineralogist*, 1995, **33**, 303–312.
- 4 J. S. Becker, M. V. Zoriy, C. Pickhardt, N. Palomero-Gallagher and K. Zilles, *Anal Chem*, 2005, 77, 3208–3216.
- 5 S. E. Jackson, in Laser Ablation ICP-MS in the Earth Sciences: Current Practices and Outstanding Issues, 2008, vol. 40, pp. 169–188.
- 6 A. Schweikert, S. Theiner, D. Wernitznig, A. Schoeberl, M. Schaier, S. Neumayer, B. K. Keppler and G. Koellensperger, *Anal Bioanal Chem*, 2022, **414**, 485–495.

Spectrometry Accepted Manuscript

ournal of Analytical Atomic

4	
5	
6	
7	
, 0	
0	
9	_ ·
1	0 <u>ខ</u>
1	19
1	25
1	3Ĕ
1	4 <sup>Ĕ</sup>
1	с. Д
1	ی. ۳
1	LCIED
4	/a
9	88
7	ଖୁ
3	0Ž
Ź	រុទ្ធ
3	2 gau
1 0	2 Little
रू द्र	JS SA
<del>g</del>	45 -
2	5를
2	യ്
2	7Å
2	<b>8</b> <u></u>
Ż	9~
Ę	0 <sup>j</sup>
3	1
റ്റ	sed
Sec	Zeo
3 J	3≝
3	4 <u>-</u>
3	5Ĕ
Ž	6≊
ૈજી	F
ŝ	
₹ a	
പ്പ	12
<del>ا</del> ف	Ϋ́
4	()
4	ڰ
Δ	_
	3
4	3 4
4	3 4 5
4 4 4	3 4 5 6
4 4 4 4	3 4 5 6 7
4 4 4 4 4	3 4 5 6 7 8
4 4 4 4 4	3 4 5 6 7 8 0
4 4 4 4 4	3 4 5 6 7 8 9
4 4 4 4 4 4 5	3 4 5 6 7 8 9 0
4 4 4 4 4 5 5	3 4 5 6 7 8 9 0
4 4 4 4 4 5 5 5	3 4 5 6 7 8 9 0 1 2
444455555	3 4 5 6 7 8 9 0 1 2 3
444445555555	3 4 5 6 7 8 9 0 1 2 3 4
444445555555555555555555555555555555555	3 4 5 6 7 8 9 0 1 2 3 4 5
444445555555555555555555555555555555555	3 4 5 6 7 8 9 0 1 2 3 4 5 6
444445555555555555555555555555555555555	3 4 5 6 7 8 9 0 1 2 3 4 5 6 7
444445555555555555555555555555555555555	3 4 5 6 7 8 9 0 1 2 3 4 5 6 7 8
444445555555555555555555555555555555555	3 4 5 6 7 8 9 0 1 2 3 4 5 6 7 8 9
444445555555555555555555555555555555555	34567890123456789

1 2 3

 7
 M. Šala, V. S. Šelih and J. T. Van Elteren, Analyst, 2017, 142, 3356–3359.
 View Article Online

 ODI: 10.1039/D5JA00171D

- 8 M. Thompson, S. Chenery and L. Brett, *J Anal At Spectrom*, 1989, 4, 11–16.
- 9 E. F. Cromwell and P. Arrowsmith, *Anal. Chem*, 1995, **67**, 131–138.
- 10 D. Günther, R. Frischknecht, H.-J. Müschenborn and C. A. Heinrich, *Fresenius J Anal Chem*, 1997, **359**, 390–393.
- 11 D. Günther, H. Cousin, B. Magyar and I. Leopold, *J Anal At Spectrom*, 1997, **12**, 165–170.
- 12 L. Halicz and D. Günther, J Anal At Spectrom, 2004, 19, 1539–1545.
- 13 P. W. J. M. Boumans and F. J. De Boer, *Spectrochimica Acta Part B*, 1976, **31B**, 355–375.
- 14 E. T. Luong, R. S. Houka and R. E. Serfass, *J Anal At Spectrom*, 1997, **12**, 703–708.
- 15 M. Kuonen, B. Hattendorf and D. Günther, *J Anal At Spectrom*, 2024, **39**, 1388–1397.
- 16 L. Yang, R. E. Sturgeon and Z. Mester, *J Anal At Spectrom*, 2005, **20**, 431–435.
- 17 W. Nischkauer, F. Vanhaecke and A. Limbeck, *Anal Bioanal Chem*, 2016, **408**, 5671–5676.
- 18 D. N. Douglas, J. L. Crisp, H. J. Reid and B. L. Sharp, *Journal of Analytical Atomic Spectrometry*, 2011, 26, 1294– 1301.
- 19 C. Herzig, J. Frank, A. K. Opitz, J. Fleig and A. Limbeck, *Talanta*, 2020, 217, 1–7.
- 20 K. Mervič, J. T. van Elteren, M. Bele and M. Šala, *Talanta*, 2024, 269.
- 21 K. Mervič, V. S. Šelih, M. Šala and J. T. van Elteren, *Talanta*, 2024, 271.
- 22 G. M. Hieftje and H. V. Malmstadt, Anal Chem, 1968, 40, 1860–1867.
- 23 J. W. Olesik, Appl Spectrosc, 1997, 51, 158A-175A.
- 24 S. Gschwind, L. Flamigni, J. Koch, O. Borovinskaya, S. Groh, K. Niemax and D. Günther, J Anal At Spectrom, 2011, 26, 1166–1174.
- B. Ramkorun-Schmidt, S. A. Pergantis, D. Esteban-Fernández, N. Jakubowski and D. Günther, *Anal Chem*, 2015, 87, 8687–8693.
- 26 L. Hendriks, B. Ramkorun-Schmidt, A. Gundlach-Graham, J. Koch, R. N. Grass, N. Jakubowski and D. Günther, *J* Anal At Spectrom, 2019, **34**, 716–728.
- A. Gundlach-Graham and K. Mehrabi, J Anal At Spectrom, 2020, 35, 1727–1739.
- 28 C. C. Garcia, A. Murtazin, S. Groh, M. Becker and K. Niemax, Spectrochimica Acta Part B, 2010, 65, 80–85.
- 29 C. C. Garcia, A. Murtazin, S. Groh, V. Horvatic and K. Niemax, J Anal At Spectrom, 2010, 25, 645–653.
- 30 J. Koch, L. Flamigni, S. Gschwind, S. Allner, H. Longerich and D. Günther, J Anal At Spectrom, 2013, 28, 1707– 1717.
- 31 H. R. Kuhn, M. Guillong and D. Günther, *Anal Bioanal Chem*, 2004, **378**, 1069–1074.
- 32 O. Borovinskaya, PhD Thesis, ETH Zurich, 2014.
- 33 P. Becker and D. Günther, *J Anal At Spectrom*, 2023, **38**, 1704–1712.
- M. Burger, G. Schwarz, A. Gundlach-Graham, D. Käser, B. Hattendorf and D. Günther, *J Anal At Spectrom*, 2017, 32, 1946–1959.
- 35 J. M. Mermet, *Spectrochimica Acta Part B*, 1989, **44B**, 1109–1989.
- 36 K. P. Jochum, U. Nohl, K. Herwig, E. Lammel, B. Stoll and A. W. Hofmann, *Geostand Geoanal Res*, 2005, **29**, 333–338.
- 37 H. P. Longerich, S. E. Jackson and D. Günther, *J Anal At Spectrom*, 1996, **11**, 899–904.
- 38 R Foundation for Statistical Computing, *R Core Team*, R Foundation for Statistical Computing, Vienna, 2021.
- 39 RStudio, *RStudio*, PBC, Boston MA, 2025.

40 A. Murtazin, S. Groh and K. Niemax, *Spectrochim Acta Part B At Spectrosc*, 2012, **67**, 3–16.

- 41 I. Horn and D. Günther, *Appl Surf Sci*, 2003, **207**, 144–157.
- 42 M. Guillong, I. Horn and D. Günther, J Anal At Spectrom, 2003, 18, 1224–1230.
- 43 E. L. Gurevich and R. Hergenröder, *J Anal At Spectrom*, 2007, **22**, 1043–1050.
- 44 M. B. Fricker, D. Kutscher, B. Aeschlimann, J. Frommer, R. Dietiker, J. Bettmer and D. Günther, *Int J Mass Spectrom*, 2011, **307**, 39–45.
- 45 C. Neff, P. Becker and D. Günther, J Anal At Spectrom, 2022, **37**, 677–683.
- 46 M. Guillong and D. Günther, J Anal At Spectrom, 2002, 17, 831–837.
- 47 P. Becker, J. Koch and D. Günther, *J Anal At Spectrom*, 2022, **37**, 1846–1854.

The data supporting this article have been included as part of the SI.

View Article Online DOI: 10.1039/D5JA00171D

Journal of Analytical Atomic Spectrometry Accepted Manuscript