

Cite this: *Analyst*, 2024, **149**, 5101

Local electrochemical sample acidification for the detection of Pb²⁺ traces†

 Amira Mahmoud,  Július Gajdár, Mariela Brites Helú,  Mathieu Etienne and Grégoire Herzog *

Electrochemical detection of pollutants (e.g. heavy metals) in real samples often requires the adjustment of pH to allow optimal sensitivity. Such sample pretreatment can be challenging for on-site applications as it implies the use of valves, pumps and storage of base or acid solutions. We report here the use of an electrochemical approach for the control of water sample pH. It offers the possibility for local pH adjustment while simultaneously detecting Pb²⁺, whose detection sensitivity is pH dependent. An effective electrochemical method through local electrochemical acidification is performed to detect Pb²⁺ within a desired pH range without the need to add chemical reagents. Local acidification is based on water electrolysis. An anodic potential is applied to an acidifier to rapidly electrogenerate protons. This allows the sample pH to be tailored to the optimal detection condition. Reduction of the Pt oxide layer formed on the acidifier is key to obtain repeatable results in Pb²⁺ detection. On-site sample acidification is combined with anodic stripping voltammetry to reach a detection limit of 6 ppb (30 nM), which is lower than the World Health Organization guideline value for Pb²⁺ level in drinking water.

Received 6th May 2024,
Accepted 21st August 2024

DOI: 10.1039/d4an00647j

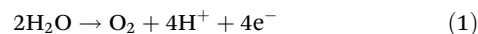
rsc.li/analyst

1. Introduction

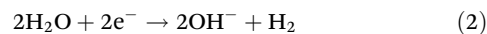
Anodic Stripping Voltammetry (ASV) is a powerful technique for the electrochemical determination of metallic cations in aqueous solutions.^{1–3} In electroanalysis, the pH of reaction medium is a fundamental parameter that needs to be adjusted for the optimal detection of the target analyte. Variation in pH can trigger dramatic changes in chemical reaction rates and thus limit metal deposition.⁴ The classic method for adjusting the sample pH is the rudimentary addition of buffers or strong acids and bases. Recently, several approaches were developed to control pH, including electrochemical redox reaction without the need of adding chemical reagents. Strakosas *et al.* developed a non-enzymatic glucose sensor based on cobalt oxide able to monitor glucose levels in neutral bodily fluids such as sweat and tears.⁵ As metal oxides require alkaline medium for glucose detection, an electrochemical strategy to control locally the pH was developed to sense glucose in fluids at neutral pH. The developed strategy was based on the integration of palladium contact that absorbs H⁺ from the neutral medium and generates a constant OH[−] concentration to increase the pH up to 10 and perform an effective glucose detection. Wiorek *et al.* recently showed that using polyaniline

PANI film acted, through an electrochemical pulse, as a proton pump for rapid sample acidification and alkalinity detection (5 min).^{6–8} Despite excellent results, it is necessary to improve the chemical composition of the proton pump to avoid the need for regeneration after acidification process. Fomina and coworker employed a combination of solution-borne quinones and galvanostatic excitation to demonstrate highly reproducible acidification or alkalisation of pH environments localized to an electrode surface.⁹ In this case, pH monitoring is realized through quinones as electroactive agents, known by their chemically reversible proton production/consumption upon electrochemical stimulation. Most frequently, controlling pH by *in situ* electrochemical approach can also involve water electrolysis.¹⁰ During this process, water is split to generate flows of protons or hydroxyl ions to radically alter medium pH. Protons generated by the oxidative water splitting reaction lead to local acidification (eqn (1)). However, hydroxyl ions produced by the reduction of water reaction serve to increase the solution pH to alkaline condition (eqn (2)).

Oxidation of water reaction:



Reduction of water reaction:



Coupling the process of water electrolysis with sensing application allows the rapid and efficient recognition of ana-

Université de Lorraine, CNRS, LCPME, F-54000 Nancy, France.

E-mail: gregoire.herzog@cnrs.fr

† Electronic supplementary information (ESI) available. See DOI: <https://doi.org/10.1039/d4an00647j>

lytes by generating protons or hydroxyl ions optimal for the detection mechanism targeted. Read *et al.* control electrochemically the pH, using a boron-doped ring disc electrode system, to detect mercury in neutral solutions.¹¹ This approach is based on water electrolysis by applying an appropriate oxidative potential to the ring electrode to generate protons. Acidification of the electrolyte solution occurs in the vicinity of the disc electrode at which mercury is detected. Similarly, Seymour and coworkers recently used gold interdigitated microband electrodes.¹² Water was electrolysed at acidifier electrodes, changing the local pH near the sensing electrodes. An effective chlorine detection was demonstrated in water samples with electrochemical pH control.¹² Similarly, Ag⁺ ions were detected in sodium acetate and in tap water with simultaneous electro-production of protons at an acidifier interdigitated electrode array to adjust the sample pH to the optimal condition.¹³ Huseinov *et al.* recently reported the use of membrane electrolysis for a reagent-free acidification.¹⁴ This approach is based on an *in situ* generation of nitric acid to produce an acidic environment in water sample enabling an effective detection of lead. On the other hand, Zhao *et al.*¹⁵ achieved optimal Pb²⁺ detection at pH 4.5 with a carbon nanotube-based electrode using a 0.1 M acetate buffer as supporting electrolyte. Similarly, Kang *et al.*¹⁶ reported pH optimization in acetate buffers with pHs in the 4.6–6.5 range. The LODs achieved in these conditions were in the nanomolar or subnanomolar range. However, this was possible in slightly acidic conditions thanks to the removal of dissolved oxygen through nitrogen bubbling.

Lead (Pb) is one of the heavy metals that poses damaging impacts on the environment and human health due to its extreme poisoning and bioavailability.¹⁷ Lead has been intensively used in industrial and mining activities, coal combustion, leaded paints, fuels,¹⁸ and as a material for household plumbing such as fittings, pipes, and solder.¹⁹ Lead contamination could be present on tap water that has been in contact with a lead pipe for a long time. Subsequently, lead can readily access the human body, yielding to serious health complications, such as kidney cancer, teeth disease and trouble in brain functions.²⁰ The World Health Organization (WHO) recommended a concentration below 10 µg L⁻¹ (48 nM) for Pb²⁺ in drinking water.²¹ Electroanalytical techniques are perfectly suitable to reach these concentration levels in aqueous samples to monitor drinking water.^{1,2} However, the pH level of drinking water is normally varying between 6.5 and 8.5²² with an average of 7.5.²³ For this reason, the challenge is to develop an effective lead sensor that allows the detection of lead in drinking water within this pH range without addition of reagents.

We report here on the development of a pH control approach based on an *in situ* electrochemical sample acidification. An electrochemical cell involving two working electrodes was used. The first working electrode is used for Pb sensing while the second one serves to acidify the electrolyte solution. Applying a positive potential to the acidifier, allows water oxidation and the electro-generation of a flux of protons, which diffuse to the vicinity of the sensor surface. This flux of

protons is sufficient to lower the sample pH to the optimal acidic condition. Lead is used as a model analyte to demonstrate the applicability of the electrochemical pH control method. Thus, an effective electrochemical detection with *in situ* acidification was demonstrated to detect concentration of lead in agreement with WHO recommendations.

2. Experimental section

2.1. Chemicals and reagents

Nitric acid (HNO₃, ≥99.999% trace metals basis), Sodium nitrate (NaNO₃, 99.0%) and lead standard for ICP (1000 ± 2 mg L⁻¹) were purchased from Sigma-Aldrich. All solutions were prepared using an ultrapure Millipore Milli-Q (18.2 MΩ) water. A stock solution of Pb²⁺ (10 ppm) was prepared in 10 mL of ultrapure water. Supporting electrolyte solutions of different pH values with constant ionic strength were tested for Pb²⁺ detection: 0.1 M HNO₃ (pH 1), 0.01M HNO₃ + 0.1 M NaNO₃ (pH 2), 0.001 M HNO₃ + 0.1 M NaNO₃ (pH 3), 0.1 M NaNO₃ (pH 5.6).

2.2. Electrochemical experiments

All the electrochemical measurements were carried out in a cell involving a four-electrode system: two working electrodes (i) a sensing electrode (gold screen printed electrode, Au-SPE, for lead detection), (ii) an acidifier electrode for the pH control, (iii) a reference electrode and (iv) a counter electrode. Au-SPE 220BT, used as the sensing electrode (Ø = 4 mm), and a silver/silver chloride pseudo-reference electrode, were purchased from Dropsens/Metrohm. An external platinum electrode positioned above the acidifier was used as auxiliary electrode. A platinum mesh (aperture: 0.012 cm, wire diameter: 0.004 cm) was obtained from Goodfellow France and used as acidifier. A 3D printed cell was specifically designed to introduce the SPE with the inserted Pt grid as acidifier. The Pt grid was kept at a distance of approx. 250 µm by a rubber O-ring purchased from Coruba, UK. The fabrication of the electrochemical cell by 3D printing was explained in detail in our previous publications.^{24,25} A simplified schematic representation of the designed cell and the principle of lead detection is shown in Fig. 1.

The electrochemical cell was connected to a Metrohm/Eco Chemie Autolab PGSTAT100 Bipotentiostat/Galvanostat operated by Nova 2.1.4 software. All the experiments were performed using 500 µL as sample volume. Cyclic voltammetry (CV) at scan rate of 50 mV s⁻¹ was initially performed in the presence of 100 ppm Pb²⁺ to determine the potential window and the position of the Pb deposition and stripping peak. CV analyses were also carried out to optimize the pH control conditions. Anodic stripping voltammetry using square wave voltammetry (SVW) was performed to analyse samples containing different concentrations of Pb²⁺ in NaNO₃ (0.1 M, pH 5.6) with *in situ* acidification. SWV analyses were recorded from -0.7 V to 0 V using the following parameters: -0.7 V as deposition potential, 120 s as time of deposition, 5 mV as potential step, 25 mV as amplitude and 20 Hz as frequency. During analysis, the sample was stirred to avoid the accumulation of oxygen



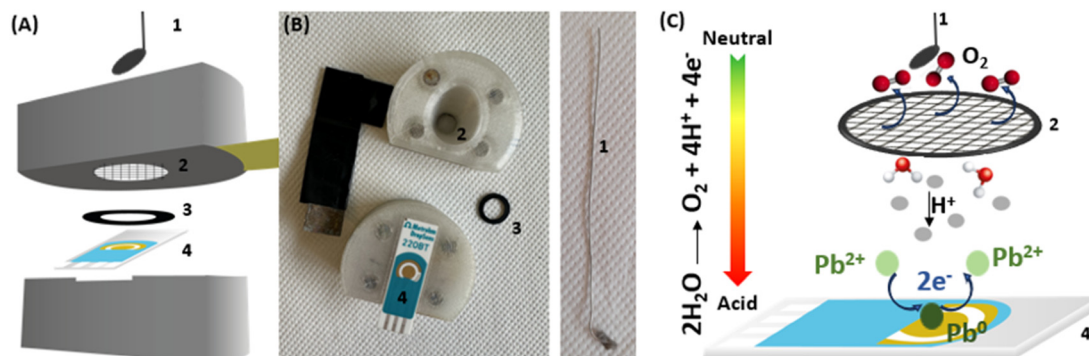


Fig. 1 (A) Schematic representation, (B) photograph, and (C) operating principle of the 3D printed electrochemical cell with integrated pieces: (1) external auxiliary Pt electrode, (2) acidifier (Pt grid), (3) spacer, (4) screen printed electrode 220BT (gold sensing electrode and silver reference electrode).

bubbles. Calibration curves were obtained by detecting different Pb^{2+} concentration going from 10 to 100 ppb. The limit of detection (LOD) was calculated using the $3.3\sigma/S$ equation, with S is the slope of the calibration curve equation and σ is the standard error of the intercept.

To assess the influence of pH changes on the reference potential of the pseudo reference electrode, open circuit potential (OCP) measurements across a range of pH values were performed (Fig. S1†). The results indicate that the pseudo reference electrode presents relative stability with minimal potential variations across the tested pH (1; 2; 3 and 5.6), ensuring reliable sensor response with varying pH.

3. Results and discussion

3.1. Optimization of the electrochemical pH control

We have investigated the impact of the potential applied at the acidifier on the pH in the vicinity of the sensor. We used the

peak potential of the gold oxide reduction to monitor and optimize the local pH changes.²⁶ We are aiming to reach a local pH of 1 as it is reported as an optimal pH for the Pb^{2+} detection with minimised interferences from Cu^{2+} ions.¹⁹ Gold oxide was produced and then reduced by cyclic voltammetry recorded from -0.1 V to $+1$ V on Au-SPE at a scan rate of 50 mV s^{-1} . Fig. S2A† shows the cyclic voltammograms obtained at different solutions over the pH range: $1 < \text{pH} < 5.6$ as described on the Experimental section. The voltammetric response of a gold electrode immersed at pH 1 shows typical characteristic peaks in both anodic and cathodic scans (red curve of Fig. 2A).²⁷ The peaks in the anodic scan are attributed to the formation of different forms of gold oxides on the electrode surface. When reversing the scan direction, a single sharp reduction peak in the cathodic scan is shown and correspond to the reduction of the oxide layer. The position of the reduction peak is strongly pH-dependent and is shifted to lower potentials with the increase of pH. The corresponding plot illustrating this dependence is shown in Fig. S2B.† The

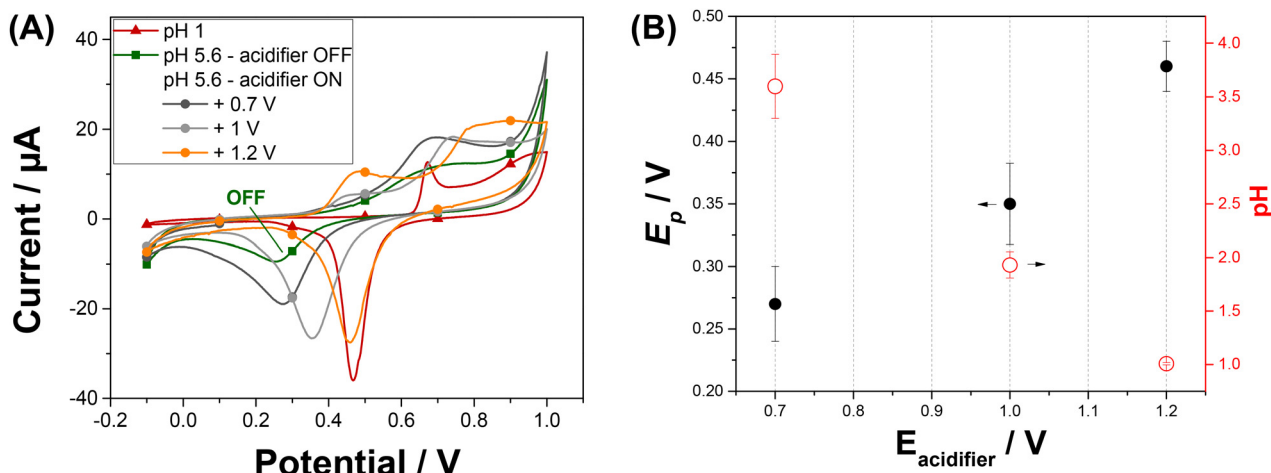


Fig. 2 (A) Cyclic voltammogram of the gold electrode obtained in 0.1 M HNO_3 (pH 1) solution and in 0.1 M NaNO_3 (pH 5.6) solution, before and after applying different potentials (+0.7; +1.0, and +1.2 V) to the acidifier for 5 s before the scan. Scan rate 50 mV s^{-1} . (B) Variation of the Au cathodic peak potential, E_p , (filled discs) and of the calculated pH (circles) as a function of the potential applied to the acidifier, $E_{\text{acidifier}}$.



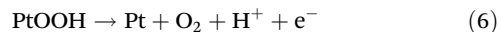
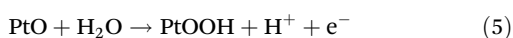
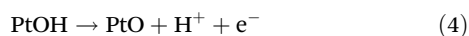
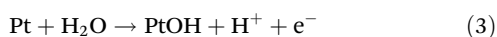
This journal is © The Royal Society of Chemistry 2024

5.6 (Fig. S4†). The resulted voltammograms at different pH values have shown that lower pH levels are more favorable for lead ion deposition and stripping using our sensor system, resulting in more defined and higher peaks. In the case of pH 5.6 with the acidifier switched off, only one cathodic peak is observed at potential of -0.33 V, which can be attributed to an overlapping of the two cathodic peaks (C_1 and C_2) of lead UPD. The disappearance of the cathodic C_3 peak confirms that the bulk deposition of lead appears only when peaks of lead UPD reach their maximal charge.³⁶ The significant change in the electrochemical behaviour of lead as a function of pH, proves that in less acidic medium, the proton deficiency makes difficult the redox reaction of lead. However, lower pH values lead to higher currents that boost the sensor response's sensitivity. Consequently, these results confirm the need to acidify the pH of the electrolyte solution to detect lead in neutral medium.

Cyclic voltammetry was then recorded in the presence of lead and with the acidifier biased at $+1.2$ V (Fig. 3). As expected, a similar electrochemical behaviour for 100 ppm Pb^{2+} is obtained when the acidifier is switched on as compared to the case of pH 1. The chemically prepared pH 1 solution gives a bulk stripping A_3 peak at $+0.59$ V with a current magnitude of 63.7 μ A, while the anodic A_3 peak is observed at $+0.58$ V with the acidifier switched on with a peak magnitude of 43.2 μ A (68% of the signal compared to the magnitude of A_3 obtained for the untreated pH 1). Hence, these results confirm the possibility to reach pH conditions to perform Pb^{2+} electroanalysis through acidification by *in situ* electrochemical method.

3.3. Pb^{2+} ASV under sample acidification

ASV of 100 ppb of Pb^{2+} was repeated three times under the same conditions (Fig. S5†). The resulting square wave voltammograms shows a lack of repeatability in the measurements. Stripping peak potential and current are dependent on the oxidation state of the electrode surface. Cyclic voltammetry of the platinum grid was performed in 0.1 M $NaNO_3$ (pH 5.6). The current shoulder around $+0.70$ V is assigned to oxidation of platinum on the forward scan and a peak at -0.07 V (on the reverse scan) is assigned to the oxide reduction (Fig. S6†). Water oxidation reaction at a platinum electrode is a multiple step reaction.³⁷ It proceeds through an adsorbate evolution mechanism. Hydroxide ions, resulting from H_2O dissociation, are adsorbed onto the Pt electroactive sites (eqn (3)). Then a deprotonation process undergoes to form PtO intermediates (eqn (4)). A nucleophilic attack by solvent water molecules leads to the formation of PtOOH at the electrode surface (eqn (5)). Finally, PtOOH goes through a deprotonation process to release H^+ and O_2 (eqn (6)).



The electro-generation of H^+ leads to the formation of Pt oxide layers, disturbing the surface atom arrangements, to which electrode reactions are sensitive to. Iizuka *et al.* have shown that oxygen evolution reaction activity of Pt electrodes dropped when an anodic potential of $+1.4$ V (vs. RHE, pH = 1) was held for 100 s. OER activity could be restored when a potential sufficiently low to reduce the Pt oxide was applied.³⁸

An electrochemical regeneration of the surface of the platinum grid is performed before each detection experiment to restore the electrode activity degraded by the prolonged oxidation of the platinum grid. The full analytical procedure involves a three-step process: (i) sensor cleaning and acidifier regeneration, (ii) deposition & acidification, (iii) detection. The cleaning step starts by applying an anodic potential of $+0.1$ V to the sensor to dissolve any metal deposits left over from a previous experiment. A potential of 0 V is applied to the acidifier to reduce the Pt oxide layer that was formed during the acidification procedure. After 60 s, the deposition step begins by the application of -0.7 V to the sensing electrode. This step involves the reduction of lead ions $Pb^{2+}(aq.)$ to its zero-valence metallic state Pb(s) on the Au-SPE surface. The acidification step starts by the application of $+1.2$ V to the acidifier for the suitable optimized time. During this step, water is oxidised and generates protons, which diffuse to the surface of the sensor. Deposition and acidification processes are then followed by the stripping step. The electrolyte solution is stirred to ensure the homogenization of the solution during the cleaning step, to improve mass transport during the deposition step and to avoid the accumulation of oxygen bubbles during the acidification step. Stirring is stopped before the start of the stripping. Fig. 4A displays the anodic stripping voltammograms recorded in the presence of 100 ppb of Pb^{2+} , at pH 1 and pH 5.6 solution, without and with the acidifier switched on. At pH 1, an anodic stripping peak is observed at -0.27 V with a peak magnitude of 14.1 ± 1.4 μ A. At pH 5.6, the application of the acidification process leads to the improvement of the anodic stripping current of Pb. Without acidification a stripping peak located at -0.28 V with a current magnitude of 9.4 ± 0.8 μ A is recorded. When the acidifier is switched on, a sharp anodic stripping peak is obtained at -0.28 V with a peak magnitude of 15.9 ± 0.7 μ A ($N = 3$). Thus, the peak current intensity increased by 59% compared to the initial obtained current when the pH of detection is dropped from 5.6 to 1. This behaviour gives direct evidence that lower pH values provide a higher current, which offers a higher sensitivity for SWV detection. These results prove the efficiency of the optimized acidification procedure. In our experimental conditions, the sample volume is 0.5 mL for a concentration of 100 ppb Pb^{2+} . If 100% of Pb^{2+} contained in the sample is deposited on a 4 mm diameter disc electrode ($A_{\text{electrode}} = 0.126$ cm^2), 0.397 μ g cm^{-2} of Pb will be deposited. Given that Pb has an atomic radius of 1.86×10^{-8} cm and an atomic mass of 207.2 g mol^{-1} , a full monolayer of Pb would correspond to 0.316 μ g cm^{-2} . It should be noted that for the simplicity of calculations,



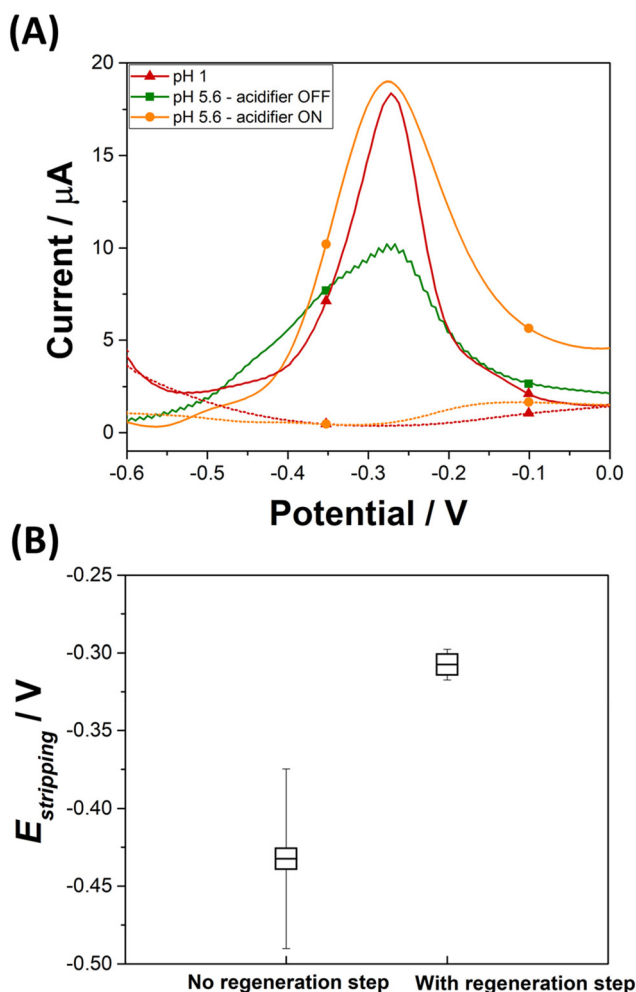


Fig. 4 (A) SWV of 100 ppb Pb^{2+} on a gold electrode in HNO_3 (pH 1) (red triangle curve) and in 0.1 M NaNO_3 (pH 5.6) with the acidifier switch off (green square curve) and on (orange circle curve). (B) Average of stripping peak potentials values obtained with ($N = 20$) and without ($N = 80$) regeneration step of the acidifier.

geometric surface area was considered. These results suggest that, in our experimental conditions, a single monolayer of Pb is formed during the preconcentration process. Such calculation agrees with our decision to use underpotential deposition instead of the bulk deposition for the lead detection. Considering the shapes of the Pb ASV peaks obtained in pH 1 and in pH 5.6 electrochemically acidified samples, the differences observed can be attributed to several factors, including variations in the local pH environment, the presence of different species already present in the electrolytes or formed during electrochemical acidification, and different kinetics of the stripping process under these conditions.

The average and the standard deviation of the stripping peak potential $E_{\text{stripping}}$ from a hundred measurements are estimated before and after applying the optimized SWV procedure (Fig. 4B). In the absence of the regeneration of the acidifier surface, the average potential value for the stripping of Pb(s) to $\text{Pb}^{2+}(\text{aq.})$ is -0.43 ± 0.06 V. This wide distribution of

$E_{\text{stripping}}$ values can be corrected by the application of a reductive potential that regenerates the acidifier surface. Once a regeneration step is included in the analytical procedure, the average $E_{\text{stripping}}$ value is -0.31 ± 0.01 V. This shows that applying the optimized SWV procedure, which includes the cleaning step, results in achieving stability in the stripping peak potential.

3.4. Analytical sensing of Pb^{2+} with *in situ* acidification

SWV analysis were performed in the presence of various concentrations of Pb^{2+} in 0.1 M NaNO_3 (pH 5.6) with *in situ* acidification, using the optimal procedure to build a calibration curve (Fig. 5A). In the presence of Pb^{2+} , the peak current indicative of the anodic stripping of lead is observed at -0.28 V. A proportional increase of the stripping current response is observed when the concentration of Pb^{2+} increased from 10 to 100 ppb. The Pb stripping peaks shift to higher potentials when increasing Pb^{2+} concentration and consequently increasing the Pb^{2+}/Pb ratio, which is in agreement with the Nernst equation. Furthermore, it is obvious that when the lead concentration reaches 100 ppb, the SWV stripping response shows the appearance of a small shoulder around -0.48 V. This can be explained by the fact that from this concentration, the second UPD stripping peak of Pb monolayer starts to be visible. The variation of the shape of the stripping peaks for various Pb^{2+} concentrations may be due to the several factors that may include the surface coverage of the electrode by Pb^{2+} , the co-adsorption of nitrate anions and the variation of dissolved oxygen concentration linked to the acidification process. The dependency of the stripping peak current with Pb^{2+} concentration is linear as shown in Fig. 5B. All analyses were performed in triplicate to obtain standard deviation (shown as error bars). The correlation equation is given by $i_{\text{stripping}} = (0.086 \pm 0.003) [\text{Pb}^{2+}] + (4.24 \pm 0.164)$. This variation shows a wide range of linearity which extends from 10 ppb to 60 ppb with a correlation coefficient of $R^2 = 0.996$ and a LOD of 6 ppb, which is below the WHO recommendation for Pb^{2+} concentration in drinking waters. As a comparison, the calibration curve when the acidifier is OFF gives a correlation equation of $i_{\text{stripping}} = (0.034 \pm 0.002) [\text{Pb}^{2+}] - (0.138 \pm 0.055)$. The equation for the calibration curve at pH 1 is $i_{\text{stripping}} = (0.188 \pm 0.017) [\text{Pb}^{2+}] - (0.662 \pm 0.436)$. The sensitivity of the Pb^{2+} stripping when the acidifier is switched ON is higher than when it is OFF. However, it does not achieve the same sensitivity as when the pH is close to one. The high value of the intercept found when the acidifier is ON can be explained by the high dissolved oxygen concentrations linked to the acidification process discussed beforehand.

To study the impact of interference such as Cu^{2+} on the detection of Pb^{2+} , SWV analysis was then undertaken with *in situ* acidification in the presence of a fixed concentration of 100 ppb ($0.48 \mu\text{M}$) of Pb^{2+} and an increasing concentrations of Cu^{2+} : 200 ppb ($3.2 \mu\text{M}$), 500 ppb ($7.9 \mu\text{M}$), and 1000 ppb ($15.7 \mu\text{M}$). The average percentage of the stripping Pb^{2+} signal ($N = 3$) in the presence of Cu^{2+} is presented in Fig. S7.† Despite the presence of high concentrations of the interferent, Cu^{2+}



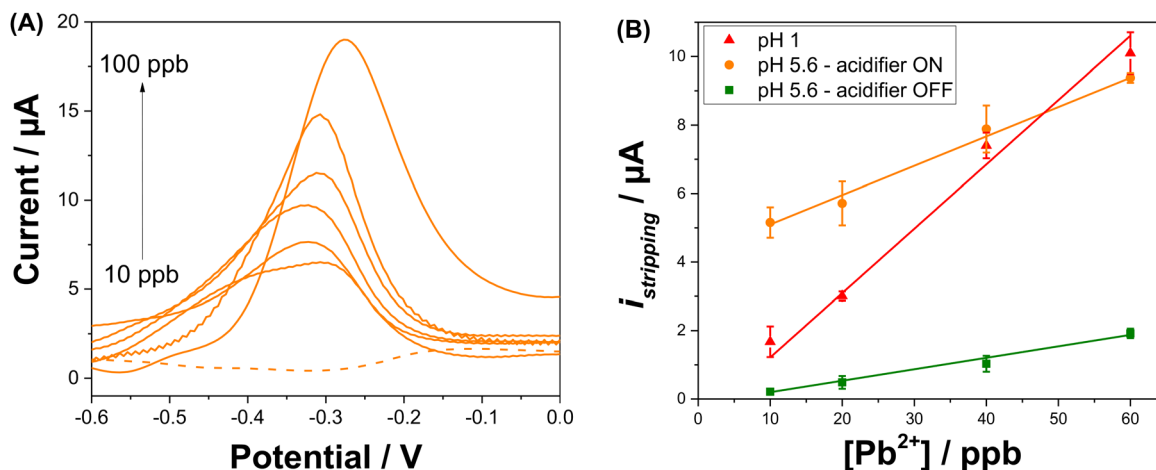


Fig. 5 (A) SWV of different Pb^{2+} concentrations (10, 20, 40, 60, 80, 100 ppb) recorded in 0.1 M NaNO_3 (pH 5.6) with *in situ* acidification, according to the optimized procedure. (B) Calibration curves for anodic stripping peak current vs. Pb^{2+} concentration at pH 1 (\blacktriangle), and at pH 5.6 when the acidifier is ON (\bullet) and OFF (\blacksquare).

slightly affects the stripping signal of Pb^{2+} , resulting in a low decrease in the Pb^{2+} signal by approximately 22%, 15%, and 19% in the presence of 200, 500, and 1000 ppb of Cu^{2+} , respectively.

4. Conclusions

We have proceeded to local electrochemical acidification involving water electrolysis for the detection of Pb^{2+} traces. An electrochemical cell was designed to integrate an acidifier close to a gold sensor. By applying an optimal positive potential to the acidifier, water oxidation leads to the electro-generation of protons, which diffuse to the vicinity of the sensing electrode and locally acidify the sample. Under the optimized conditions, the pH in the vicinity of the sensor surface was dropped from the initial value (pH 5.6) to the optimal detection value (pH 1). Anodic stripping voltammetry was then carried out, to successfully detect concentrations down to 10 ppb of Pb^{2+} , in agreement with the WHO guidelines. The developed approach proves that the control of pH *in situ* paves the way for on-field analysis, whether for lead detection or other pH-dependent analytes. In this sense, experiments are underway to develop an innovative electrochemical device for real-time lead detection in water utility systems using the *in situ* pH control method developed in this work.

Data availability

Data for this article, including raw voltammograms used to draw Fig. 2–5 and Fig. S1–S7† will be available, when the manuscript is accepted for publication at Recherche Data Gouv (<https://entrepot.recherche.data.gouv.fr/>) repository at <https://doi.org/10.57745/F82FDN>.

Conflicts of interest

There are no conflicts to declare.

Acknowledgements

The authors extend their appreciation to SATT Sayens, ERDF and the National Research Agency (ANR) for providing funding to this study as a part of aQuaLead project (ANR-22-CE42-0028-01).

References

- 1 A. J. Borrill, N. E. Reily and J. V. Macpherson, Addressing the Practicalities of Anodic Stripping Voltammetry for Heavy Metal Detection: A Tutorial Review, *Analyst*, 2019, **144**(23), 6834–6849, DOI: [10.1039/C9AN01437C](https://doi.org/10.1039/C9AN01437C).
- 2 C. Ariño, C. E. Banks, A. Bobrowski, R. D. Crapnell, A. Economou, A. Królicka, C. Pérez-Ràfols, D. Soulis and J. Wang, Electrochemical Stripping Analysis, *Nat. Rev. Methods Primers*, 2022, **2**(1), 1–18, DOI: [10.1038/s43586-022-00143-5](https://doi.org/10.1038/s43586-022-00143-5).
- 3 G. Herzog and V. Beni, Stripping Voltammetry at Micro-Interface Arrays: A Review, *Anal. Chim. Acta*, 2013, **769**, 10–21, DOI: [10.1016/j.aca.2012.12.031](https://doi.org/10.1016/j.aca.2012.12.031).
- 4 W. Chen, M.-K. Zhang, B.-Y. Liu, J. Cai and Y.-X. Chen, Challenges and Recent Progress in Unraveling the Intrinsic pH Effect in Electrocatalysis, *Curr. Opin. Electrochem.*, 2022, **34**, 101003, DOI: [10.1016/j.coelec.2022.101003](https://doi.org/10.1016/j.coelec.2022.101003).
- 5 X. Strakosas, J. Selberg, P. Pansodtee, N. Yonas, P. Manapongpun, M. Teodorescu and M. Rolandi, A Non-Enzymatic Glucose Sensor Enabled by Bioelectronic pH Control, *Sci. Rep.*, 2019, **9**(1), 10844, DOI: [10.1038/s41598-019-46302-9](https://doi.org/10.1038/s41598-019-46302-9).



- 6 A. Wiorek, M. Cuartero, R. De Marco and G. A. Crespo, Polyaniline Films as Electrochemical-Proton Pump for Acidification of Thin Layer Samples, *Anal. Chem.*, 2019, **91**(23), 14951–14959, DOI: [10.1021/acs.analchem.9b03402](https://doi.org/10.1021/acs.analchem.9b03402).
- 7 A. Wiorek, G. Hussain, A. F. Molina-Osorio, M. Cuartero and G. A. Crespo, Reagentless Acid–Base Titration for Alkalinity Detection in Seawater, *Anal. Chem.*, 2021, **93**(42), 14130–14137, DOI: [10.1021/acs.analchem.1c02545](https://doi.org/10.1021/acs.analchem.1c02545).
- 8 F. Steininger, A. Wiorek, G. A. Crespo, K. Koren and M. Cuartero, Imaging Sample Acidification Triggered by Electrochemically Activated Polyaniline, *Anal. Chem.*, 2022, **94**(40), 13647–13651, DOI: [10.1021/acs.analchem.2c03409](https://doi.org/10.1021/acs.analchem.2c03409).
- 9 N. Fomina, C. A. Johnson, A. Maruniak, S. Bahrampour, C. Lang, R. W. Davis, S. Kavusi and H. Ahmad, An Electrochemical Platform for Localized pH Control on Demand, *Lab Chip*, 2016, **16**(12), 2236–2244, DOI: [10.1039/C6LC00421K](https://doi.org/10.1039/C6LC00421K).
- 10 H. Zhou, G. Li and S. Yao, A Droplet-Based pH Regulator in Microfluidics, *Lab Chip*, 2014, **14**(11), 1917–1922, DOI: [10.1039/C3LC51442K](https://doi.org/10.1039/C3LC51442K).
- 11 T. L. Read, E. Bitziou, M. B. Joseph and J. V. Macpherson, In Situ Control of Local pH Using a Boron Doped Diamond Ring Disk Electrode: Optimizing Heavy Metal (Mercury) Detection, *Anal. Chem.*, 2014, **86**(1), 367–371, DOI: [10.1021/ac403519p](https://doi.org/10.1021/ac403519p).
- 12 I. Seymour, B. O'Sullivan, P. Lovera, J. F. Rohan and A. O'Riordan, Electrochemical Detection of Free-Chlorine in Water Samples Facilitated by *in situ* pH Control Using Interdigitated Microelectrodes, *Sens. Actuators, B*, 2020, **325**, 128774, DOI: [10.1016/j.snb.2020.128774](https://doi.org/10.1016/j.snb.2020.128774).
- 13 L. Wasiewska, I. Seymour, B. Patella, R. Inguanta, C. Burgess, G. Duffy and A. O'Riordan, Reagent Free Electrochemical-Based Detection of Silver Ions at Interdigitated Microelectrodes Using *in situ* pH Control, *Sens. Actuators, B*, 2021, **333**, 129531, DOI: [10.1016/j.snb.2021.129531](https://doi.org/10.1016/j.snb.2021.129531).
- 14 A. Huseinov, A. Hoque, K. A. Ruble, B. J. Dee and N. T. Alvarez, Reagentless Dissolution and Quantification of Particulate Lead in Tap Water via Membrane Electrolysis, *Anal. Chem.*, 2023, **95**(24), 9297–9303, DOI: [10.1021/acs.analchem.3c01201](https://doi.org/10.1021/acs.analchem.3c01201).
- 15 D. Zhao, D. Siebold, N. T. Alvarez, V. N. Shanov and W. R. Heineman, Carbon Nanotube Thread Electrochemical Cell: Detection of Heavy Metals, *Anal. Chem.*, 2017, **89**(18), 9654–9663, DOI: [10.1021/acs.analchem.6b04724](https://doi.org/10.1021/acs.analchem.6b04724).
- 16 W. Kang, X. Pei, C. A. Rusinek, A. Bange, E. N. Haynes, W. R. Heineman and I. Papautsky, Determination of Lead with a Copper-Based Electrochemical Sensor, *Anal. Chem.*, 2017, **89**(6), 3345–3352, DOI: [10.1021/acs.analchem.6b03894](https://doi.org/10.1021/acs.analchem.6b03894).
- 17 Y. Zhang, H. Jiang and X. Wang, Cytidine-Stabilized Gold Nanocluster as a Fluorescence Turn-on and Turn-off Probe for Dual Functional Detection of Ag⁺ and Hg²⁺, *Anal. Chim. Acta*, 2015, **870**, 1–7, DOI: [10.1016/j.aca.2015.01.016](https://doi.org/10.1016/j.aca.2015.01.016).
- 18 C. Wang, X. Cui, Y. Li, H. Li, L. Huang, J. Bi, J. Luo, L. Q. Ma, W. Zhou, Y. Cao, B. Wang and F. Miao, A Label-Free and Portable Graphene FET Aptasensor for Children Blood Lead Detection, *Sci. Rep.*, 2016, **6**(1), 21711, DOI: [10.1038/srep21711](https://doi.org/10.1038/srep21711).
- 19 D. Dragoie, N. Spătaru, R. Kawasaki, A. Manivannan, T. Spătaru, D. A. Tryk and A. Fujishima, Detection of Trace Levels of Pb²⁺ in Tap Water at Boron-Doped Diamond Electrodes with Anodic Stripping Voltammetry, *Electrochim. Acta*, 2006, **51**(12), 2437–2441, DOI: [10.1016/j.electacta.2005.07.022](https://doi.org/10.1016/j.electacta.2005.07.022).
- 20 H. Needleman, Lead Poisoning, *Annu. Rev. Med.*, 2004, **55**(1), 209–222, DOI: [10.1146/annurev.med.55.091902.103653](https://doi.org/10.1146/annurev.med.55.091902.103653).
- 21 J. Strandberg, *Lead in Drinking-Water, Health Risks, Monitoring and Corrective Actions – Technical Brief*, 2022.
- 22 Background documents for development of WHO Guidelines for drinking-water quality and Guidelines for safe recreational water environments. <https://www.who.int/publications/m/item/background-documents-for-development-of-who-guidelines-for-drinking-water-quality-and-guidelines-for-safe-recreational-water-environments> (accessed 2023-12-01).
- 23 K. Kulthanan, P. Nuchkull and S. Varothai, The pH of Water from Various Sources: An Overview for Recommendation for Patients with Atopic Dermatitis, *Asian Pac. Allergy*, 2013, **3**(3), 155–160, DOI: [10.5415/apallergy.2013.3.3.155](https://doi.org/10.5415/apallergy.2013.3.3.155).
- 24 J. Gajdár, G. Herzog and M. Etienne, Amperometric Sensor for Selective On-Site Analysis of Free Sulfite in Wines, *ACS Sens.*, 2022, **7**(8), 2209–2217, DOI: [10.1021/acssensors.2c00611](https://doi.org/10.1021/acssensors.2c00611).
- 25 M. Etienne, T. X. H. Le, T. Nasir and G. Herzog, Electrochemical Filter To Remove Oxygen Interference Locally, Rapidly, and Temporarily for Sensing Applications, *Anal. Chem.*, 2020, **92**(11), 7425–7429, DOI: [10.1021/acs.analchem.0c00395](https://doi.org/10.1021/acs.analchem.0c00395).
- 26 S. Yang and D. G. H. Hetterscheid, Redefinition of the Active Species and the Mechanism of the Oxygen Evolution Reaction on Gold Oxide, *ACS Catal.*, 2020, **10**(21), 12582–12589, DOI: [10.1021/acscatal.0c03548](https://doi.org/10.1021/acscatal.0c03548).
- 27 J. P. Hoare, A Cyclic Voltammetric Study of the Gold–Oxygen System, *J. Electrochem. Soc.*, 1984, **131**(8), 1808, DOI: [10.1149/1.2115966](https://doi.org/10.1149/1.2115966).
- 28 M. Pourbaix and J. Burbank, Atlas D-Equilibres Electrochimiques, *J. Electrochem. Soc.*, 1964, **111**(1), 14C, DOI: [10.1149/1.2426051](https://doi.org/10.1149/1.2426051).
- 29 D. W. M. Arrigan, L. L. Bihan and M. J. Pickup, Apparent Enhanced Underpotential Voltammetry of Lead(II) at a Spontaneously Adsorbed Monolayer-Coated Gold Electrode, *Analyst*, 1999, **124**(12), 1797–1802, DOI: [10.1039/A906125H](https://doi.org/10.1039/A906125H).
- 30 I. Mazerie and F. Geneste, Coupling of Anodic Stripping Voltammetry with Sampled-Current Voltammetry on an Electrode Array: Application to Lead Detection, *Sensors*, 2020, **20**(5), 1327, DOI: [10.3390/s20051327](https://doi.org/10.3390/s20051327).



- 31 G. Herzog and D. W. M. Arrigan, Underpotential Deposition and Stripping of Lead at Disorganized Monolayer-Modified Gold Electrodes, *Electroanalysis*, 2005, **17**(20), 1816–1821, DOI: [10.1002/elan.200503318](https://doi.org/10.1002/elan.200503318).
- 32 M. Brand, I. Eshkenazi and E. Kirowa-Eisner, The Silver Electrode in Square-Wave Anodic Stripping Voltammetry. Determination of Pb²⁺ without Removal of Oxygen, *Anal. Chem.*, 1997, **69**(22), 4660–4664, DOI: [10.1021/ac970420f](https://doi.org/10.1021/ac970420f).
- 33 A. J. Bard, *Standard Potentials in Aqueous Solution*, Routledge, New York, 2017. DOI: [10.1201/9780203738764](https://doi.org/10.1201/9780203738764).
- 34 P. Atkins, *Physical Chemistry for the Life Sciences*, Oxford University Press, Oxford, 2010.
- 35 G. Herzog and D. W. M. Arrigan, Determination of Trace Metals by Underpotential Deposition–Stripping Voltammetry at Solid Electrodes, *TrAC, Trends Anal. Chem.*, 2005, **24**(3), 208–217, DOI: [10.1016/j.trac.2004.11.014](https://doi.org/10.1016/j.trac.2004.11.014).
- 36 E. Kirowa-Eisner, Y. Bonfil, D. Tzur and E. Gileadi, Thermodynamics and Kinetics of Upd of Lead on Polycrystalline Silver and Gold, *J. Electroanal. Chem.*, 2003, **552**, 171–183, DOI: [10.1016/S0022-0728\(03\)00181-5](https://doi.org/10.1016/S0022-0728(03)00181-5).
- 37 B. E. Conway, Electrochemical Oxide Film Formation at Noble Metals as a Surface-Chemical Process, *Prog. Surf. Sci.*, 1995, **49**(4), 331–452, DOI: [10.1016/0079-6816\(95\)00040-6](https://doi.org/10.1016/0079-6816(95)00040-6).
- 38 K. Iizuka, T. Kumeda, K. Suzuki, H. Tajiri, O. Sakata, N. Hoshi and M. Nakamura, Tailoring the Active Site for the Oxygen Evolution Reaction on a Pt Electrode, *Commun. Chem.*, 2022, **5**(1), 126, DOI: [10.1038/s42004-022-00748-7](https://doi.org/10.1038/s42004-022-00748-7).

