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Dually-dynamic covalent tetraPEG hydrogels end-linked with boronate ester and acylhydrazone groups

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Well-defined dually dynamic hydrogels were prepared by end-linking four-armed poly(ethylene glycol) stars (tetraPEG stars) through two different types of dynamic covalent cross-links, boronates and acylhydrazones, leading to robust, self-healable materials. This required the prior end-functionalization of tetraPEG stars, originally bearing four hydroxyl terminal groups, with glucuronate, acylhydrazide and benzaldehyde groups, resulting in three differently end-functional star polymers. A first type of dually dynamic hydrogel resulted from the combination of the first two differently end-functionalized tetraPEG stars, cross-linked by 4-formylphenyl boronic acid, a small molecule bearing both an aldehyde and a boronic acid group, respectively complementary to the acylhydrazide and glucuronate end-groups of the two above-mentioned tetraPEG stars. For comparison, a singly-dynamic hydrogel cross-linked with only acylhydrazone groups was also prepared, as well as a double-like hydrogel combining the constituents of both of the above-mentioned hydrogels. All three types of hydrogels were prepared at three different pH values, 8.5, 10.5 and 12.5, leading to a total number of nine samples. All nine samples were investigated for their self-healing, mechanical, viscoelastic and aqueous swelling/degradation properties. This study sets the basis for the development of well-defined polymeric dynamic covalent hydrogels where their self-healing and stability can be readily tuned.

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Introduction

Polymer networks (thermosets and elastomers) and gels (organogels and hydrogels) represent an important part, ~25%, of the annual global polymer production of more than 300 million tons,¹ and have a similar environmental footprint to linear and other non-network commercial polymers, with the extra burden of their cross-linked structure which even further delays their disintegration in nature. Alleviating the problem of environmental pollution arising from polymer networks and gels can be accomplished by extending the service life of objects made of these materials *via* rendering them self-repairable, on the one hand, and, on the other hand, by accelerating their degradability once disposed. Both of these targets can be attained by the implementation of dynamic covalent bonds^{2–4} as cross-links^{5,6} in these materials. Although covalent in nature, dynamic covalent

bonds break and reform, *i.e.*, they display a dynamic equilibrium involving partner exchange, leading to self-healing in the bulk material, or to disintegration when the system is diluted, *e.g.*, under conditions of rain.

Several choices are available for such dynamic covalent bonds, including boronate esters exchanging at and above the pK_a of the corresponding boronic acid, imines (hydrazones and oximes) exchanging under acidic conditions, Diels-Alder adducts and nitroxides exchanging thermally, and disulfides exchanging under reduction-oxidation conditions.⁷ Boronate esters represent an attractive choice, characterized by their extreme reversibility, but limited by their consequent reduced stability.⁸ On the other hand, acylhydrazones possess high hydrolytic stability but lower reversibility, particularly the aryl-substituted ones.⁹ Combination of the two types of dynamic covalent groups, acylhydrazones and boronates, as cross-links in the same polymer network, is expected to endow the material with a superior spectrum of properties, and, in particular, good reversibility with sufficient stability, leading to self-repairable and strong polymer networks. Furthermore, fine-tuning the extent of each property, reversibility *vs.* stability, according to the demands of the application at hand, can be readily accomplished by adjusting the percentage of each of the two kinds of bonds.

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Conventional polymeric hydrogels are infamously fragile due to their pronounced spatial inhomogeneity arising from the uneven distribution of cross-links, a result of free radical cross-linking copolymerization usually employed for their preparation.¹⁰ Many successful efforts have been undertaken during the past 20 years to mechanically reinforce polymer hydrogels, including interpenetration (double-networks),¹¹ utilization of mechanical (rather than chemical) cross-links (slide-ring gels),¹² doping with inorganic nanoparticles (nanocomposite gels),¹³ and well-defined gels (tetraPEG gels) based on four-armed star poly(ethylene glycol)s (tetraPEG stars) interconnected at the tips of their arms (end-linked).¹⁴ However, only the last of these four approaches really addresses the problem of eliminating network heterogeneity, and is, consequently, the most general and promising.

Hydrogels of well-defined structure, cross-linked with a mixture of acylhydrazone and boronate dynamic covalent bonds will be readily self-healable and robust materials. This is the aim undertaken in this study. In particular, herein we employ tetraPEG stars of molar mass of 10 000 g mol⁻¹ ("TP10k") which are appropriately end-functionalized so that they bear different types of terminal groups. Appropriate combination of the various kinds star polymers leads to the formation of hydrogels bearing both boronate and acylhydrazone cross-links. Although tetraPEG gels separately end-linked with boronate^{15,16} or acylhydrazone¹⁷ groups have already been reported in the literature, the simultaneous utilization of both types of cross-links in the same material has not yet been documented. On the other hand, some hydrogels¹⁸ and organogels¹⁹ have appeared in the literature, with the simultaneous utilization of both acylhydrazone and boronate cross-links; however, the gels in these reports were randomly cross-linked. The dually dynamic tetraPEG hydrogels prepared in this investigation are characterized in terms of their self-healing, mechanical, viscoelastic and aqueous swelling/degradation properties, the last of which (swelling and degradation) are rarely studied for dynamic covalent hydrogels.

Thus, the synthetic novelty in this investigation is the preparation of well-defined hydrogels based on a mixture of a number of differently end-functionalized tetraPEG stars. These novel hydrogel materials would have a broad application potential, appropriate for cases when robust hydrogels are required, and covering a wide spectrum of needs in terms of the relative character in self-repairability *vs.* stability, which can be regulated in a straightforward manner, namely, through the fraction of each kind of cross-link in the system.

Experimental section

Materials

Diethyl ether (purity $\geq 99\%$), 4-hydroxybenzhydrazide ($\geq 97\%$), benzaldehyde ($\geq 99\%$), 4-formylphenylboronic acid ($\geq 95\%$), D-(+)-gluconic acid δ -lactone ($\geq 99\%$), 4-hydroxybenzaldehyde (98%), methanesulfonyl chloride ($\geq 99.7\%$), potassium carbonate ($\geq 99\%$), hydrazine hydrate (80% w/w solution in water),

dialysis tubing composed of benzoylated cellulose with molecular weight cut-off of 3000 g mol⁻¹ (average flat width 32 mm), methanol (99.8%), sodium carbonate ($\geq 99\%$), sodium hydrogen carbonate ($\geq 99.7\%$), sodium phosphate monobasic monohydrate ($\geq 98\%$), sodium phosphate dibasic ($\geq 99\%$), glacial acetic acid, potassium phosphate tribasic ($\geq 98\%$), sodium hydroxide ($\geq 98\%$) and 80% w/w aqueous ammonia solution were purchased from Sigma-Aldrich-Merck, Germany. Tetrahydroxy-terminated four-arm star poly(ethylene glycol) of $M_n = 10\,000$ g mol⁻¹ (TP10k-OH) was purchased from NOF Corporation, Tokyo, Japan. Dichloromethane (DCM, 99%) was purchased from Labscan, Ireland. *N,N*-dimethylformamide (DMF, 99.8%) and triethylamine (TEA, $\geq 99\%$) were purchased from Scharlau, Spain. Dimethylsulfoxide (DMSO, $\geq 99\%$) was purchased from Fisher, UK. Deuterated chloroform (CDCl₃, 99.8%) and deuterated dimethylsulfoxide (d₆-DMSO, 99.8%) were purchased from Merck, Germany. Aqueous buffer solutions (100 mM) at pH 8.5, 10.5 and 12.5 were prepared from sodium phosphate dibasic/sodium phosphate monobasic monohydrate, sodium carbonate/sodium hydrogen carbonate, and potassium phosphate tribasic, respectively.

Synthesis of benzaldehyde-protected hydrazide

For this synthesis, 3.1 g (29.9 mmol) benzaldehyde, 3.5 g (23 mmol) 4-hydroxybenzhydrazide and 60 mL DMF were first placed into a round bottomed flask. Subsequently, the mixture was refluxed at 80 °C for 24 h. Next, after most of the DMF was removed under reduced pressure, the reaction mixture was precipitated in chloroform. The precipitate obtained was dried in a vacuum oven at 60 °C for 5 h to give the product as a white powder.

Modification of TP10k-OH

Synthesis of mesylate-terminated tetraPEG (TP10k-mesyl).

Initially, 7 g (0.7 mmol) freeze dried TP10k-OH, 40 mL of freshly distilled DCM and 3 mL of freshly distilled TEA (21.5 mmol) were transferred into a spherical flask. Then, 2.24 g (19.6 mmol) mesyl chloride was added dropwise to the flask which was cooled down to 0 °C. The reaction mixture was allowed to react under stirring at 0 °C for 48 h. Afterwards, the solution was extracted with NaHCO₃ (5 $\times$ 50 mL) and subsequently with water (2 $\times$ 50 mL). The organic extract was condensed under reduced pressure and, subsequently, precipitated twice in cold diethyl ether. Finally, the precipitate was vacuum-dried at 60 °C for 5 h to obtain the final product as a yellow solid.

Synthesis of benzaldehyde-terminated tetraPEG (TP10k-Bz).

This synthesis started by dissolving 3.68 g (0.356 mmol) TP10k-mesyl, 0.61 g (4.98 mmol) 4-hydroxybenzaldehyde and 0.688 g (4.98 mmol) of K₂CO₃ in 40 mL of DMF placed in a round bottomed flask. The reaction mixture was allowed to react under stirring at 80 °C for 24 h. The solution was then diluted with 250 mL DCM and washed with 0.75 M NaOH aqueous solution (5 $\times$ 50 mL) and deionized water (2 $\times$ 50 mL). Finally, the organic phase was condensed and precipitated twice in cold diethyl ether to obtain TP10k-Bz as a white solid after vacuum-drying at 60 °C for 5 h.

Synthesis of benzaldehyde protected acylhydrazide-terminated tetraPEG (TP10k-ProtHz). First, 2.5 g (0.24 mmol) of TP10k-mesyl, 0.535 g (2.23 mmol) of benzaldehyde-protected acylhydrazide, 0.35 g (2.23 mmol) of K_2CO_3 and 35 mL of DMF were placed in a round-bottomed flask. The reaction mixture was allowed to react under stirring at 80 °C for 9 h. The solution was then diluted with 150 mL DCM and washed with 0.75 M NaOH aqueous solution (5×50 mL) and deionized water (2×50 mL). In the end, the polymer was isolated by precipitation twice, in cold diethyl ether, followed by vacuum drying at 60 °C for 5 h to obtain the final product as a white solid.

Synthesis of acylhydrazide-terminated tetraPEG (TP10k-Hz). First, 1 g (0.092 mmol) of TP10k-ProtHz, 1.5 mL of hydrazine hydrate and 8 mL of DMSO were placed in a round bottomed flask. The reaction mixture was allowed to react under stirring at room temperature for 24 h. The solution was then loaded into a dialysis membrane which was dialyzed against water for five days, with the external water being changed daily. The product was obtained as a white solid after lyophilization.

Synthesis of amine-terminated tetraPEG (TP10k-NH₂). Initially, 5.24 g (0.51 mmol) of TP10k-mesyl was dissolved in 79 mL of 80% w/w aqueous ammonia solution and placed in a round-bottomed flask. The flask was sealed and the reaction mixture was left under stirring at room temperature for five days. The excess ammonia solution was subsequently removed under reduced pressure, and 100 mL of 0.1 M $NaHCO_3$ solution was added to the residue. The solution was then extracted with DCM (8×50 mL). Finally, the product was isolated after the evaporation of the organic solvent, and vacuum-dried at 60 °C to give a white solid.

Synthesis of diol-terminated tetraPEG (TP10k-Gluco). First, 2.3 g (0.23 mmol) of TP10k-NH₂, 0.819 g (4.6 mmol) of D-(+)-gluconic acid δ -lactone, 115 mL of methanol, and 1 mL of freshly distilled TEA (7.17 mmol) were transferred into a round-bottomed flask. The reaction mixture was allowed to react under stirring at room temperature for three days. After that, the solution was transferred into a dialysis membrane which was dialyzed against water for four days with the external water being changed daily. After the lyophilization of the membrane contents, the product was obtained as a white solid.

Preparation of the “hydrazone” cross-linked hydrogels

These dynamic covalent hydrogels were formed by mixing aqueous solutions of TP10k-Bz and TP10k-Hz at their stoichiometric ratio. The synthesis was performed in three different buffers (100 mM) of pH values equal to 8.5, 10.5 and 12.5, and at a polymer concentration of 15% w/v each. A typical experimental procedure is given below. First, 0.034 g (3.22 μ mol) TP10k-Hz and 0.0335 g (3.22 μ mol) TP10k-Bz were dissolved separately in 225 μ L buffer each. Afterwards, the two aqueous solutions were mixed and vortexed at room temperature. The hydrogels were left for seven days to be formed completely.

Preparation of the “boronate” cross-linked hydrogels

This category of dynamic covalent hydrogels was formed by mixing solutions of TP10k-Gluco, 4-formylphenylboronic acid

(4-FPBA) and TP10k-Hz at their stoichiometric ratio. The synthesis took place in three different buffers (100 mM) of pH values equal to 8.5, 10.5 and 12.5, and at a polymer concentration of 15% w/v each. A typical experimental procedure is given below. Initially, 0.034 g (3.17 μ mol) of TP10k-Gluco and 0.0019 g (12.7 μ mol) of 4-FPBA were dissolved together in 225 μ L of aqueous buffer solution of the chosen pH. Then, 0.0334 g (3.17 μ mol) of TP10k-Hz was dissolved separately in another 225 μ L of aqueous buffer solution of the same pH as above. Subsequently, these two solutions were mixed and vortexed at room temperature. The hydrogels were left for seven days to be formed completely.

Preparation of the “double” hydrogels

These dynamic covalent hydrogels were formed by mixing solutions of TP10k-Gluco, 4-formylphenylboronic acid (4-FPBA), TP10k-Hz and TP10k-Bz at their stoichiometric ratio. The synthesis was performed in three different buffers (100 mM) of pH values equal to 8.5, 10.5 and 12.5, and at a polymer concentration of 30% w/v each. A typical experimental procedure is given below. Firstly, 0.034 g (3.17 μ mol) of TP10k-Gluco and 0.0019 g (12.7 μ mol) of 4-FPBA were dissolved together in 150 μ L of buffer. Then, 0.0674 g (6.34 μ mol) TP10k-Hz and 0.0335 g (3.17 μ mol) TP10k-Bz were dissolved separately in additional 150 μ L of buffer each, and, subsequently, the three aqueous solutions were mixed and vortexed at room temperature. The hydrogels were left for seven days to be formed completely.

Aqueous degrees of swelling of the hydrogels

The degrees of swelling of the three types of hydrogels, (“boronate” cross-linked, “hydrazone” cross-linked and “double” hydrogels) were measured in aqueous buffered solutions. Each type of hydrogel was formed in aqueous buffer solutions of three different pH values, 8.5, 10.5 and 12.5, and, later on, transferred into a vial with 10 mL of aqueous buffer solution (100 mM) of a pH value the same as that of hydrogel formation. Every hydrogel was left in the buffer to equilibrate for twenty-five days. The status of the hydrogel was observed (whether fully dissolved or not), and the swollen mass of the hydrogels was determined gravimetrically.

Characterization with ¹H NMR spectroscopy

¹H NMR spectroscopy was performed on a 500 MHz Avance Bruker spectrometer, equipped with an Ultrashield magnet, to confirm the structures (quantitative end-functionalization reactions) of the tetraPEG star building blocks of the hydrogels (TP10k-Gluco, TP10k-Bz and TP10k-Hz) and their precursors (TP10k-Ms, TP10k-NH₂ and TP10k-HzProt). The ¹H NMR spectra were recorded in the deuterated solvents $CDCl_3$ and d_6 -DMSO. The ¹H NMR spectra of the “boronate” cross-linked hydrogel in D_2O buffered (100 mM) at pD = 8.5, three and seven days after mixing its components, and that of the *in situ* formed adduct between TP10k-Gluco and FPBA at the same pD were also recorded.

Rheology

Viscoelastic characterization of the hydrogels at their as-prepared state was performed on a Discovery HR2 rheometer from Thermal Analysis Instruments (TA), operating in parallel plate geometry, with an upper plate of 20 mm in diameter and a temperature-controlled (Peltier) lower plate maintained at 20 °C. The rheometer was operated in the oscillation frequency mode, with angular frequency values from 0.1 to 100 rad s⁻¹, 1% strain, and an approximate plate separation of 3 mm. The rheological measurements provided the frequency dependence of the storage (G') and loss (G'') shear moduli of the “hydrazone” cross-linked, “boronate” cross-linked, and “double” hydrogels, prepared at a total solids concentration of 15, 15 and 30% w/v, respectively, each in aqueous buffer solutions of pH 8.5, 10.5 and 12.5. Rheological strain sweep experiments were also conducted on selected samples (“boronate” and “double” hydrogels formed at pH = 8.5) in order to more quantitatively probe their self-healing capability. In these experiments, the strain sweep first indicated the critical strain at which the sample fails. Next, a strain 20 to 50% higher than the critical was instantaneously applied to the sample, which was subsequently allowed to recover. Finally, the extent to which the sample regained its original storage shear modulus was calculated.

Mechanical testing

An Instron 5944 mechanical testing system (Instron, Norwood, MA) was used to characterize the compressive mechanical properties of the hydrogels at their as-prepared state. The samples were cylindrical with a diameter of 9.5 mm and a height of 8.1 mm, and were compressed at a constant speed of 1.37 mm min⁻¹. “Boronate”, “hydrazone” and “double” hydrogels with a total solids concentration of 15, 15, 30% w/v, respectively, were prepared, each at three different pH values of 8.5, 10.5 and 12.5. Attempted tension experiments on the same mechanical testing system failed due to sample slippage from the instrument's grasps. However, rough tension experiments were performed on selected samples (“boronate” and “double” formed at pH = 10.5) by holding the samples using our fingers and pulling apart, which allowed an estimation of the strain at break. These semi-quantitative experiments were performed both on virgin and self-healed (cut and pushed against each other for 24 h) samples, which also allowed an estimation of the self-healing efficiency.

Self-healing

Self-repair experiments were performed on “boronate” cross-linked, “hydrazone” cross-linked, and “double” hydrogels formed at pH 8.5, 10.5 and 12.5, with a total solids concentration of 15, 15, 30% w/v, respectively. All the different types of hydrogels were prepared at the three pHs, one sample with and another without the presence of dye: rhodamine B, methylene blue, riboflavin, cresol purple, or one of two different universal pH indicators. After seven days, both the dyed and non-dyed hydrogels were cut in half, resulting in two pieces each.

Subsequently, a piece of the colored and a piece of the non-dyed hydrogel were pressed together along their cut surfaces and left for 8 h in a closed container.

Results and discussion

End-functionalized TetraPEG stars

The preparation of the designed hydrogels required the syntheses of three differently end-functionalized final tetraPEG stars, whose syntheses involved the prior preparation of three intermediates. The three final tetraPEG stars were the ones end-functionalized with glucuronate (“TP10k-Gluco”), benzaldehyde (“TP10k-Bz”) and acylhydrazide (“TP10k-Hz”) groups, whereas the intermediate products were tetraPEG stars end-functionalized with mesylate (“TP10k-Ms”), primary amine (“TP10k-NH₂”), and benzaldehyde-protected acylhydrazide (“TP10k-ProtHz”) groups. All synthetic procedures are illustrated in Fig. 1, where the employed reaction conditions, solvents, and reagents with their excesses are also provided. It must be pointed out that all end-functionalizations were complete (100%, confirmed using ¹H NMR spectroscopy), in order to secure as high cross-linking conversion in the hydrogels as possible, whereas the lower-than-perfect recovery yields, ranging between 78 and 97%, were due to material losses during the various purification procedures (extraction, precipitation, dialysis). Fig. 1 also illustrates schematics of the three differently end-functionalized final tetraPEG stars, TP10k-Gluco, TP10k-Bz and TP10k-Hz, plus the chemical structure and schematic of 4-formylphenyl boronic acid (FPBA), a small molecule subsequently used for the hydrogel synthesis.

The ¹H NMR spectra of TP10k-Bz and TP10k-Hz, as well as their intermediates, were provided in our previous work.¹⁷ Thus, here we only give the ¹H NMR spectra (Fig. 2) of TP10k-Gluco and its intermediates, synthesized following the procedure described by Yesilyurt *et al.*¹⁵

Mesylate-terminated tetraPEG (TP10k-mesyl)

The synthesis of TP10k-mesyl was performed by the mesylation of TP10k-OH, resulting in the activation of the four terminal hydroxyl groups of the tetraPEG star with mesyl groups. The final product was isolated in 78% yield. Its structure and purity were confirmed by ¹H NMR spectroscopy. Fig. 2(II) shows the ¹H NMR spectrum of the synthesized and purified TP10k-mesyl. The spectrum presented the full conversion of the hydroxyl groups (¹H NMR spectrum of starting material given in Fig. 2(I)) to mesyl groups, due to the appearance of the peak *e* at 3.1 ppm corresponding to the methyl protons of the mesyl group.

Amine-terminated tetraPEG (TP10k-NH₂)

The synthesis of TP10k-amine was achieved by the nucleophilic attack of ammonia to the mesyl terminal groups of TP10k-mesyl, resulting in their conversion to primary amine terminal groups. The final product was isolated in 96% yield, while its structure and purity were confirmed using ¹H NMR spectroscopy. Fig. 2(III)

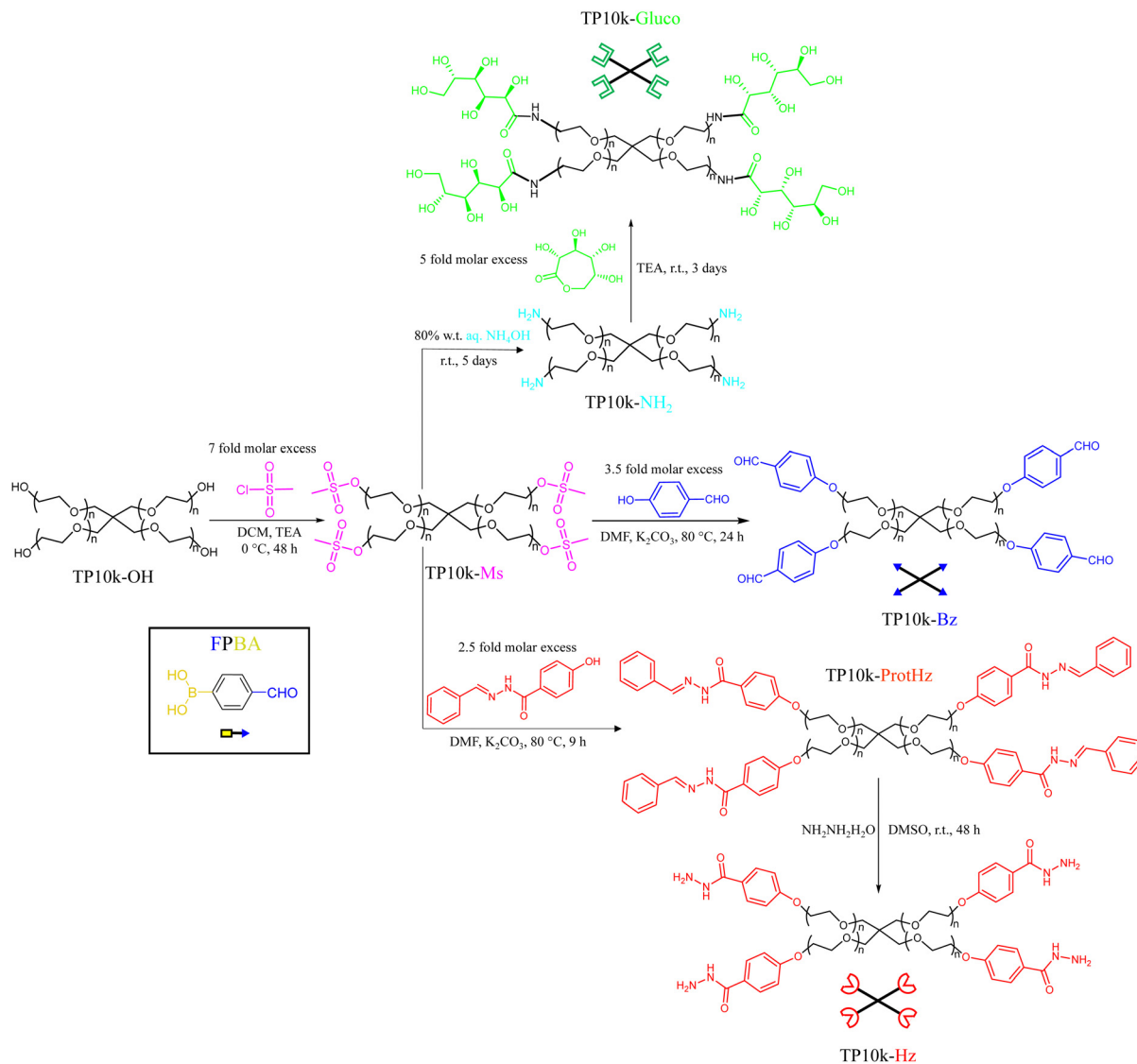


Fig. 1 Reaction schemes followed for the preparation of the three differently end-functionalized tetraPEGs, as well as their schematic representations plus that of FPBA.

shows the ¹H NMR spectrum of the purified TP10k-NH₂. The spectrum shows that the mesyl groups were completely replaced by primary amine groups, evidenced by the disappearance of the methyl protons of TP10k-mesyl at 3.1 ppm and the appearance of peak c at 2.7 ppm, corresponding to the methylene group next to the terminal amine.

Diol-terminated tetraPEG (TP10k-Gluco)

The synthesis of TP10k-Gluco was performed by the nucleophilic attack of the primary amine groups of TP10k-NH₂ onto the glucuronalactone ring, leading to ring opening and incorporation of one glucuronate group at each of the terminal amine groups of TP10k-NH₂. The final product was isolated in 97% yield, and its structure and purity were confirmed using ¹H NMR spectroscopy. Fig. 2(IV) shows the ¹H NMR spectrum of the purified TP10k-Gluco. The spectrum indicates that the amine groups reacted completely with the lactone, manifested

by the appearance of the amide bond peak d at 7.7 ppm, and the peaks of the opened glucuronalactone, i, h, g, k, j, e and f, but also by the disappearance of the methylene peak at 2.7 ppm.

Hydrogel types

Fig. 3 illustrates schematically the three hydrogel types and the strategy they were prepared from the combination of the various building blocks. These building blocks include the three differently end-functionalized tetraPEG stars, TP10k-Gluco, TP10k-Bz and TP10k-Hz, as well as FPBA. Our most novel hydrogel type, henceforth to be called “boronate”, was a dually dynamic hydrogel resulting from the indirect end-linking of the glucuronate and acylhydrazone end-functionalized tetraPEG stars through FPBA whose aldehyde and boronic acid groups are reactive to acylhydrazone and glucuronate groups, respectively. In addition to that, and for comparison, a dynamic hydrogel cross-linked only *via* acylhydrazone groups was also prepared (to

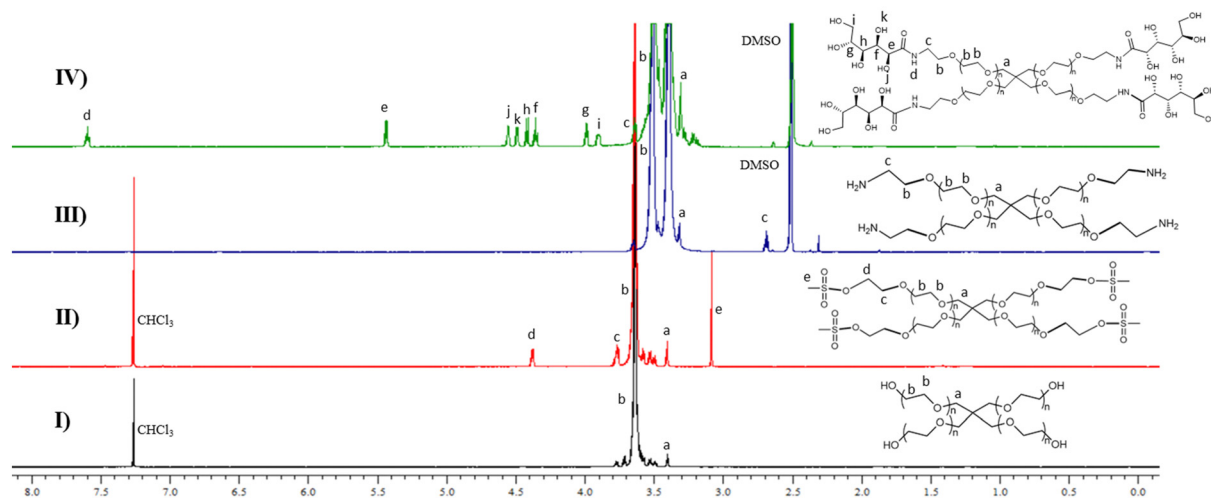


Fig. 2 ^1H NMR spectra of (I) the starting compound TP10k-OH, (II) the purified mesylate-terminated tetraPEG (TP10k-mesyl), (III) the purified amine-terminated tetraPEG (TP10k-NH₂), and (IV) the final targeted product of diol-terminated tetraPEG (TP10k-Gluco).

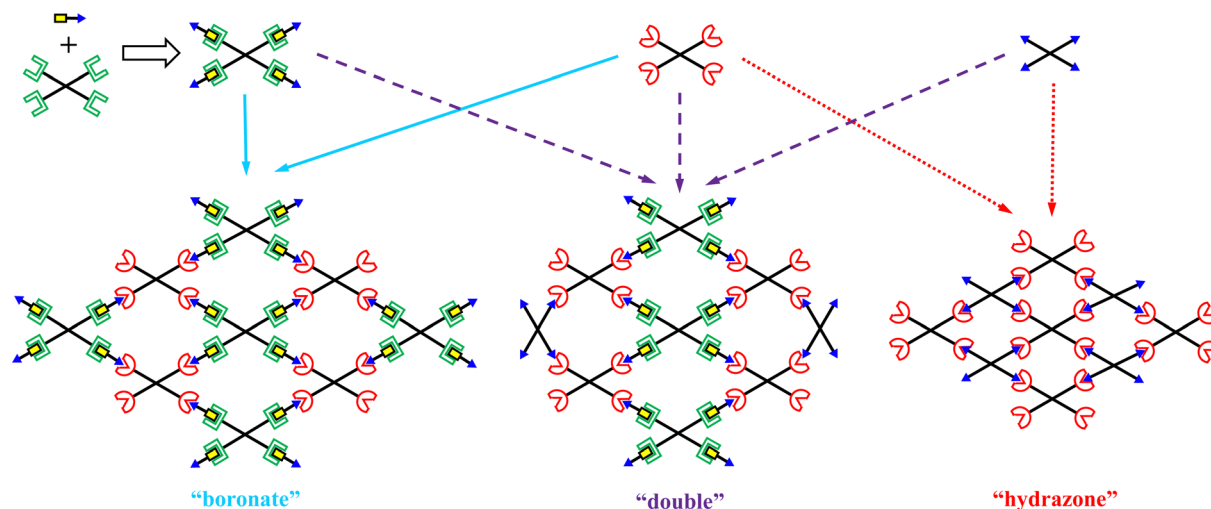


Fig. 3 Preparation of the three types of hydrogels from the four available building blocks.

be called “hydrazone” hydrogel),¹⁷ as well as a double-like hydrogel combining the constituents of both of the above-mentioned hydrogels (“double” hydrogel). Each of the three hydrogels was formed in aqueous buffers at three different pHs, 8.5, 10.5 and 12.5.

Structure of boronate hydrogel

Next, we provide ^1H NMR spectroscopy evidence for the structure of the “boronate” hydrogel prepared in two consecutive steps (see Fig. 3). We were able to record a ^1H NMR spectrum in D₂O buffered (100 mM) at pD = 8.5 not only for the product of the first step which is soluble, but also for the product of the second step which is the hydrogel. We would like to point out that NMR spectra of polymer networks, even in the swollen state, are very rare in the literature, due to peak broadening arising from low chain mobility in the semi-solid gel state.

Here, the soft nature of the system apparently allowed the recording of the high resolution proton NMR spectrum, shown in Fig. 4.

The ^1H NMR spectrum in part (II) of the figure clearly shows the complete conversion of the starting TP10k-Gluco (its ^1H NMR spectrum is shown in part (I) of the figure) to its adduct with FPBA after 1 h of reaction (with a stoichiometric amount of FPBA), as evidenced by the downfield shift of the signals from the methine protons a and b, and the appearance of aromatic protons c, d and e at the correct positions and with the correct integrals in the spectrum. Parts (III) and (IV) of the figure display the ^1H NMR spectra for the “boronate” hydrogel recorded three and seven days, respectively, after the addition of a stoichiometric amount of TP10k-Hz to the previously mentioned adduct. While near-complete conversion can already be detected within three days of the reaction, as

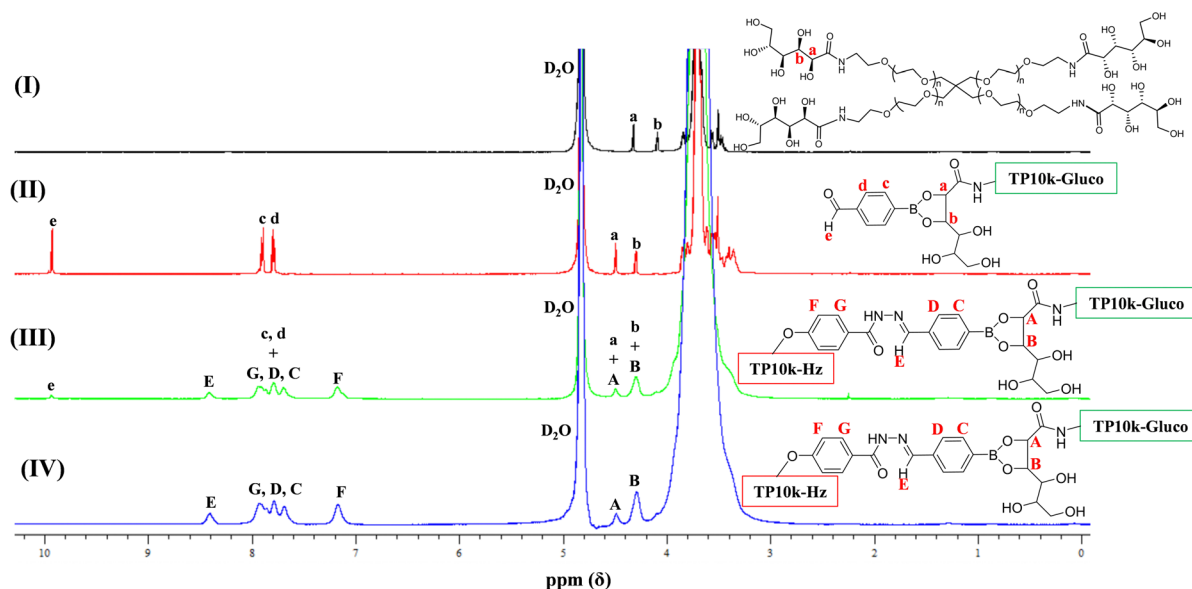


Fig. 4 ^1H NMR spectra in D_2O buffered at $\text{pD} = 8.5$ of (I) the starting TP10k-Gluco, (II) the fully-formed (within 1 h) adduct of TP10k-Gluco with a stoichiometric amount of FPBA, and (III and IV) the “boronate” hydrogel formed from the reaction of the TP10k-Gluco-FPBA adduct with a stoichiometric amount of TP10k-Hz after three days (III) and after seven (IV).

evidenced from the large reduction in the signal from the benzaldehyde proton e at 9.8 ppm, complete conversion is achieved in seven days, when proton e totally disappears from the spectrum in part (IV).

Apparent degrees of swelling and degradation of the hydrogels

The degrees of swelling of hydrogels represent a very important thermodynamic property and easy to measure, usually gravimetrically.^{20–22} However, many studies do not report it, as they focus on the (bio)applications of the as-prepared hydrogel samples. Furthermore, regarding the “boronate”-cross-linked hydrogels, these would totally degrade upon their equilibration with excess water,¹⁵ due to the very fast dynamics of the boronate bond. Still, it would be very useful to obtain the time of complete gel dissolution. Thus, in this first part of hydrogel characterization, we study the swelling behavior of all gel samples, to obtain the relevant swelling properties and dissolution times. Fig. 5 displays the temporal evolution of the apparent aqueous degrees of swelling for the three hydrogel

types, each at the three different pH values. The nine swelling profiles can be grouped into two types, according to their shape: six of them, which form a maximum, and three of them that tend to level off. The latter type corresponds to the most stable samples, which are the “hydrazone” hydrogels at the two lower pH values, 8.5 and 10.5, plus the “double” hydrogel at pH 8.5, whereas the former type corresponds to the rest of the samples. The maximum swelling value in the former type is created by the tendency of the hydrogel to absorb more solvent from the as-prepared state in order to attain equilibrium swelling leading to an increase in the degree of swelling at the beginning of the process, on the one hand, and, on the other hand, the eventual degradation of the sample arising from the dynamic nature of the cross-links in combination with the presence of a large excess of solvent, ultimately leading to the complete dissolution of the hydrogel.

The time of complete gel dissolution for the six curves (also presenting a maximum) was taken from Fig. 5 and is listed in Table 1. For the three remaining samples, no gel dissolution

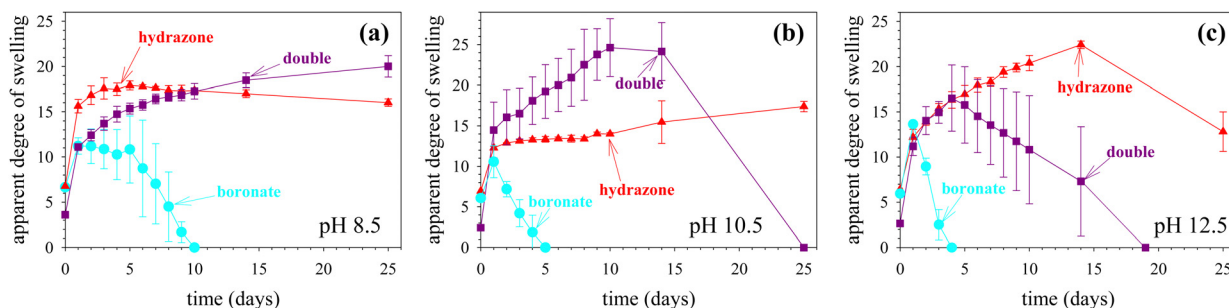


Fig. 5 Temporal evolution of the apparent aqueous degrees of swelling of the three types of hydrogels, each prepared at three different pH values, (a) 8.5, (b) 10.5 and (c) 12.5, and equilibrated in an aqueous buffer of pH the same as that at its preparation.

was observed within the timespan of the observation of ~ 50 days. In the same table, we also list the value of the maximum apparent degree of swelling observed. For the two samples not presenting a clear swelling maximum, the apparent swelling degree on the last day of the measurement is taken instead. For each hydrogel type, the hydrogel dissolution times in Table 1 decrease as the pH increases. This is due to the increased instability of both the acylhydrazone²³ and boronate²⁴ bonds as the pH increases above 8.5. At a given pH value, the dissolution times for the “hydrazone” cross-linked hydrogels are much longer than the corresponding values for the “boronate”-cross-linked hydrogels, consistent with the higher stability of the hydrazone bond, whereas the dissolution times for the “double” hydrogels are expectedly intermediate between those of the two single counterparts.

The values of the maximum apparent degrees of swelling are also listed in Table 1 and their relative values can be explained using similar arguments to those used for the dissolution times. The “boronate”-cross-linked gels present the lowest values of maximum apparent degrees of swelling, ~ 11 – 14 , because these samples degrade fast, faster than the rate of solvent absorption, thereby not being capable of attaining equilibrium swelling; an estimation of the equilibrium degree of swelling may be ~ 17 – 18 , *i.e.*, the maximum apparent degrees of swelling of the non-degrading “hydrazone”-cross-linked hydrogels, discussed next. At the other extreme, the “hydrazone”-cross-linked gels present high maximum apparent degrees of swelling because they do not degrade at all or they degrade very slowly. Finally, the “double” hydrogels, containing both types of dynamic covalent cross-links, display the highest values of maximum degrees of swelling at two out of the three pH values of the study, 8.5 and 10.5, probably a result of the slow degradation, where the partially degraded gels can absorb more solvent due to their decreasing cross-linking density.

Self-healing capabilities of the hydrogels

As mentioned in the introduction, one of the most important properties of hydrogels cross-linked with dynamic covalent bonds is their self-healing capability, thereby extending their service life and reducing environmental impact from early disposal. To this end, all nine hydrogels, arising from the three hydrogel types, each prepared at three different pH values, were tested for their ability to self-mend. For each hydrogel, two samples were employed, one colored with the addition of a small amount of a dye to the reaction mixture of the polymer

solution before gel formation. Each sample was subsequently cut in half, and two halves, one from each of the two different samples, were brought together and slightly compressed against each other by placing them in a glass vial with the same dimensions as that where gel preparation was performed. The results of these experiments are shown in Fig. 6 below, showing pictures of the two cut samples for each hydrogel before and after the attempted self-healing procedure.

It can be seen from the figure that all “boronate”-cross-linked hydrogels self-healed. This can be attributed to the high reversibility of the boronic ester bond at the three pH values employed, as all three were above the pK_a value of the phenylboronic acid of 7.6.²⁵ In contrast, none of the “hydrazone”-cross-linked hydrogels could self-heal due to the low reversibility of the hydrazone bond in this mildly alkaline to alkaline pH range used.¹⁷ On the other hand, all the “double” hydrogels exhibited self-healing capability because of the presence of the boronate reversible cross-links as well. More quantitative self-healing tests were also performed on selected samples, as these are described in the end of the two following sections of the manuscript, “Viscoelastic Properties” and “Compressive Mechanical Properties”.

Viscoelastic properties of the hydrogels

The study of the viscoelastic properties of dynamic hydrogels is very important for understanding their reversibility and self-healing behavior. We studied these properties for all nine hydrogels using dynamic rheology and measuring the shear storage modulus, G' , and the shear loss modulus, G'' , as a function of the angular frequency, ω . Fig. 7 plots the values of the two shear moduli, G' and G'' , against angular frequency, ω . With the exception of the “boronate” and “double” hydrogels at pH 8.5, the remaining seven hydrogels exhibit G' values which are frequency independent, consistent with a totally elastic behavior within the employed angular frequency range. In contrast, the two boronate-containing hydrogels (“boronate” and the “double” hydrogels) at pH 8.5 display G' values which increase with ω in the lower ω region, levelling off in the higher ω range. This is consistent with a hybrid behavior, with more viscous characteristics at low frequencies, and more elastic nature at higher frequencies.

In contrast to the two types of frequency-dependence of the G' values, three types of frequency-dependence are observed with the G'' values. The first type concerns the three “hydrazone”-cross-linked hydrogels which present almost constant G'' values (*i.e.*, frequency-independent), indicating little or no viscous behavior, which is also consistent with the trends in their corresponding G' values discussed in the previous paragraph above. The second type concerns the other two hydrogels, “boronate” and “double”, at the two higher pHs, 10.5 and 12.5, in which G'' appears to decrease linearly as the frequency increases in the double-logarithmic plots. This suggests an increasingly viscous behavior as frequency is reduced, revealing that the samples display reversible characteristics at longer timescales, matching the time required for the dynamic cross-link to open up. However, as it will be explained in the

Table 1 Main swelling characteristics of the three dynamic hydrogel types, each at the three different pH values, including gel dissolution time and degree of swelling maximum

pH	Dissolution time (days)			Maximum degree of swelling		
	Boronate	Double	Hydrazone	Boronate	Double	Hydrazone
8.5	10	>50	>50	11.2 ± 0.9	20.0 ± 1.2	17.9 ± 0.5
10.5	5	25	>50	10.6 ± 2.0	24.6 ± 3.6	17.4 ± 0.6
12.5	4	19	35	13.6 ± 0.4	16.3 ± 3.7	22.4 ± 0.4

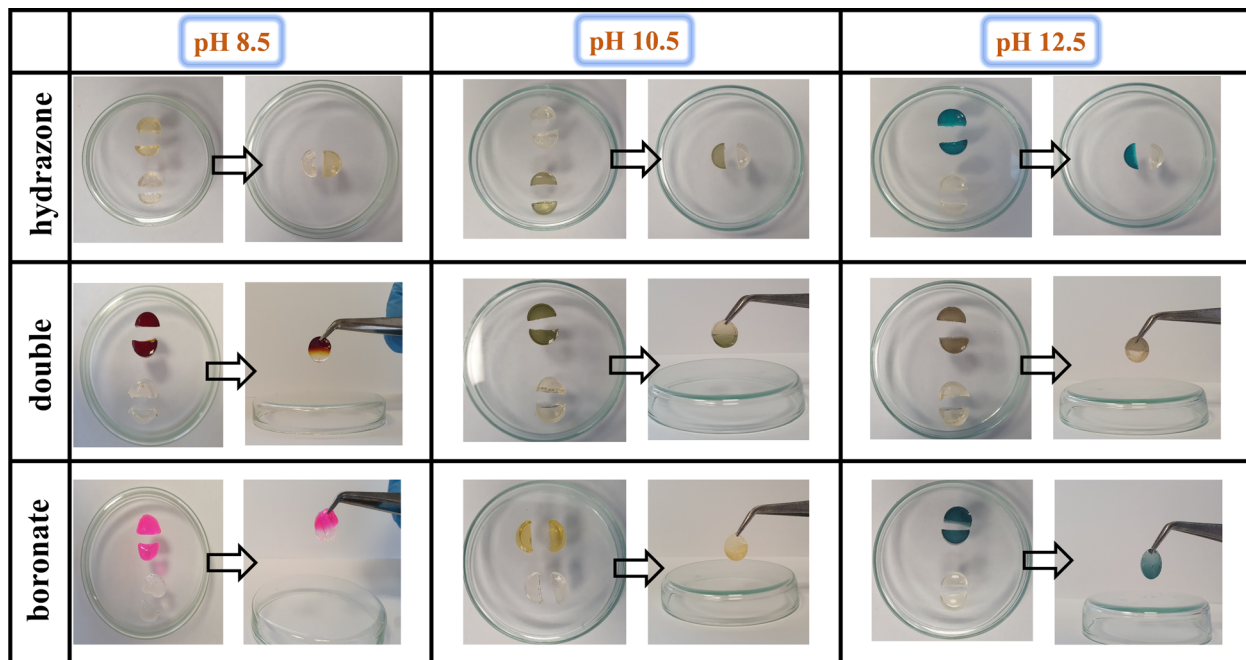


Fig. 6 Self-healing tests performed on the three hydrogel types, each prepared at three different pH values, 8.5, 10.5 and 12.5, without the addition of catalyst.

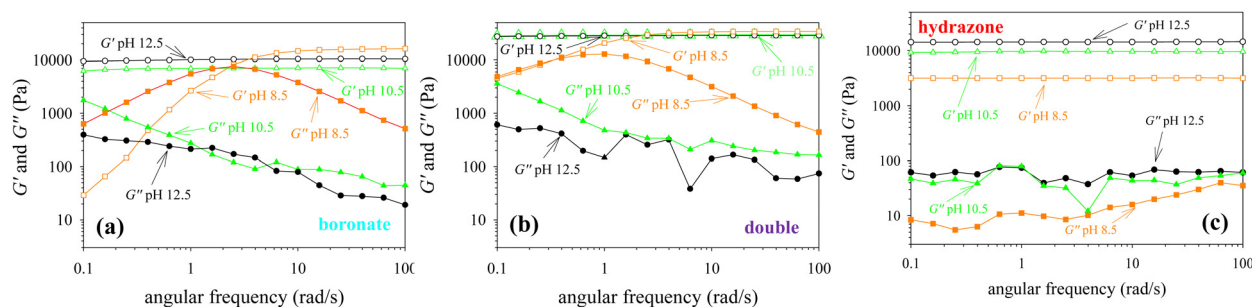


Fig. 7 Viscoelastic properties of the three hydrogel types, (a) “boronate”, (b) “double” and (c) “hydrazone”, each prepared at three different pH values, 8.5, 10.5 and 12.5, studied by dynamic rheology. It is noted that G'' values lower more than 100 compared to the corresponding G' values (mainly the case in part (c) of the figure) are of limited accuracy and should, therefore, be considered as rough estimations of G'' .

following, the behavior in this region is still dominated by elastic rather than viscous character. Finally, the third type concerns the “boronate” and “double” hydrogels at pH 8.5, whose G'' values exhibit a maximum with frequency. The frequency value at which G'' is maximized, $\omega_{\max}$, delineates the regions of mainly viscous (for $\omega < \omega_{\max}$) and mainly elastic (for $\omega > \omega_{\max}$) behavior. This hybrid viscous/elastic behavior shown by the G'' vs. ω profiles of these two hydrogels is also consistent with their G' vs. ω profiles discussed in the previous paragraph.

Importantly, the G' vs. ω curves intersect (or “cross with”; that is the reason why $\omega_{\max}$ is frequently called ω_{crossing}) the G'' vs. ω curves at (or near) $\omega_{\max}$, which is the characteristic relaxation frequency, the inverse of which can give the relaxation time. It is noteworthy that the second type of G'' vs. ω profile is expected to also intersect the corresponding G' vs. ω

profile at a frequency lower than those studied herein. In fact, such an extrapolation (called superposition) can be employed to determine the projected intersection point and the relaxation frequency $\omega_{\max}$. For the two boronate-containing hydrogels at pH 8.5, the intersection of the curves of the two moduli takes place within the studied frequency range. The determined $\omega_{\max}$ values are 2.31 and 0.339 rad s^{−1}, corresponding to relaxation times, τ ($=2\pi/\omega_{\max}$), equal to 2.72 and 18.52 s, for the “boronate”-cross-linked and “double” hydrogels, respectively. The longer relaxation time for the “double” hydrogel compared to the “boronate” can be attributed to the higher polymer concentration and the co-presence of the more stable acylhydrazone cross-links in the “double” hydrogel.

As the G' values in Fig. 7 are either frequency-independent or level off at a sufficiently high frequency, one may extract from the figure these high-frequency (“plateau”) shear storage

modulus values and compare them with the theoretical ones. This is done in Table 2 which lists both the experimental, at the three hydrogel formation pH values, for all three network types, *i.e.*, the two “single” (“boronate” and “hydrazone”) and the “double” networks, and the corresponding theoretical ones calculated from the phantom network model.²⁶ Expectedly, the experimental G' plateau values are lower (by a factor of 2.5–3.5, for most cases) than the corresponding experimental values. This difference can be attributed to incomplete or/and reversible end-linking reactions, and secondary or higher loop formation. The fair proximity of the experimental “plateau” values to the theoretical ones may suggest that the G' plateau values from rheology are fairly consistent with the network (expected) structure.

Rheology may also be utilized to further explore the self-healing capability of the hydrogels. To this end, we performed large amplitude oscillatory shear tests (“strain sweeps”) on the two self-mendable network types, the “boronate” and the “double”, focusing our attention on those prepared at pH = 8.5, where these materials have the most liquid-like behavior, so as to better show their self-repair, and also identify any differences between the two network types regarding this self-healing behavior. The results collected from these strain sweeps and subsequent hydrogel destruction followed by self-repair are plotted in Fig. 8.

In these experiments, systematic strain sweeps were first performed to determine the critical strains at which the hydrogels would start failing (“flowing”; parts (a) and (c) of the figure, for the “boronate” and “double” hydrogels, respectively; “flow” occurs when G'' becomes greater than G'). These experiments indicated that the less stiff “boronate” hydrogel required the higher critical strain of $\sim 400\%$ to start failing, as compared to 33% for the stiffer “double” hydrogel. Knowing these critical strain values, we then equilibrated both hydrogels at the low oscillatory strain, γ , of 1%, and, subsequently, subjected each hydrogel to a step increase in strain, reaching a value of at least 20% higher than the corresponding critical strain, causing the destruction and “flow” of each hydrogel (parts (b) and (d) of the figure, for the “boronate” and “double” hydrogels, respectively). After that, we returned the strain to the low value of 1%, and waited for the recovery of each hydrogel. Both hydrogels fully recovered their initial elastic shear modulus values, with the recovery time for the stiffer “double” hydrogel being expectedly shorter, at ~ 3 min, than that of the less stiff “boronate” hydrogel whose full recovery time was ~ 9 min.

Table 2 Plateau shear storage moduli for the nine hydrogels extracted from rheology, and comparison with the theoretical values predicted using the phantom network model

	Experimental shear storage modulus, G' (kPa)			Theoretical shear storage modulus, G' (kPa)
Network type	pH = 8.5	pH = 10.5	pH = 12.5	
“boronate”	15	7.1	11	37
“hydrazone”	3.2	9.6	14	37
“double”	34	30	29	73

Compressive mechanical properties of the hydrogels

The mechanical properties of hydrogels are very important for their efficient use in various biomedical or technological applications. Although most hydrogels are infamously fragile, usually due to spatial inhomogeneities arising from uneven distribution of cross-links within the hydrogel volume,¹⁰ relatively recent research¹⁴ has shown that preparing hydrogels from large, well-defined building blocks, such as star polymers, and then end-linking them, can lead to mechanically robust materials, even when highly swollen in water.²⁷ This was the reason why we chose tetraPEG stars as building blocks for our hydrogels in the present work. Fig. 9 and 10 show representative stress–strain curves in compression of the nine hydrogel samples, and the mechanical properties calculated from these curves, respectively. These mechanical properties are the elastic modulus, E , the stress at break, $\sigma_{\max}$, and the strain at break, $\epsilon_{\max}$.

At all three pHs, the values of the elastic moduli, E , decrease in the order: “double” > “hydrazone” > “boronate”. The highest values exhibited by the “double” hydrogels are due to the fact that we chose that each “double” hydrogel contains the same concentration of boronate and hydrazone cross-links as in the respective “boronate” and “hydrazone” hydrogels. This resulted in twice as high overall cross-link concentration in the “double” hydrogel relative to the two above-mentioned hydrogels (and also twice as high polymer concentration). The lower values of elastic modulus displayed by the “boronate”-cross-linked hydrogels as compared to the “hydrazone”-cross-linked hydrogels can be attributed to the greater reversibility of the boronate bond than the hydrazone bond.

The elastic modulus of the “boronate” hydrogels gradually increases with the pH. This is because the boronate bond becomes less reversible as the pH value rises above the pK_a value of the boronic acid of 7.6.²⁵ On the other hand, the elastic modulus of the “double” hydrogel presents a maximum with respect to pH (although the error bars of the measurements are relatively large especially at the two highest pH values). The low value of the elastic modulus of the “double” hydrogel at pH 8.5 can be attributed to the boronate bonds which become more reversible at this pH, whereas the relatively low value of the elastic modulus at pH 12.5 is due to the reversibility increase of the hydrazone bond, which is also the reason why the “hydrazone” hydrogel also exhibits a slight decrease in its elastic modulus upon the increase in pH from 10.5 to 12.5. Interestingly, one may observe in the figure an approximately additive relationship between the elastic moduli of the “double” hydrogel with the “boronate” and “hydrazone” constituent hydrogels because the “double” hydrogel is essentially the product of “adding” the two of them together. Finally, upon comparing the values of the elastic moduli, E , plotted in Fig. 10(a) with the values of the plateau shear storage moduli, G' , listed in Table 2, one may conclude that the well-known relationship $E \sim 3 G'$ approximately holds, with the exception of all three samples prepared at pH 8.5 which are “too liquid”. The agreement is particularly good for the “boronate” cross-linked samples

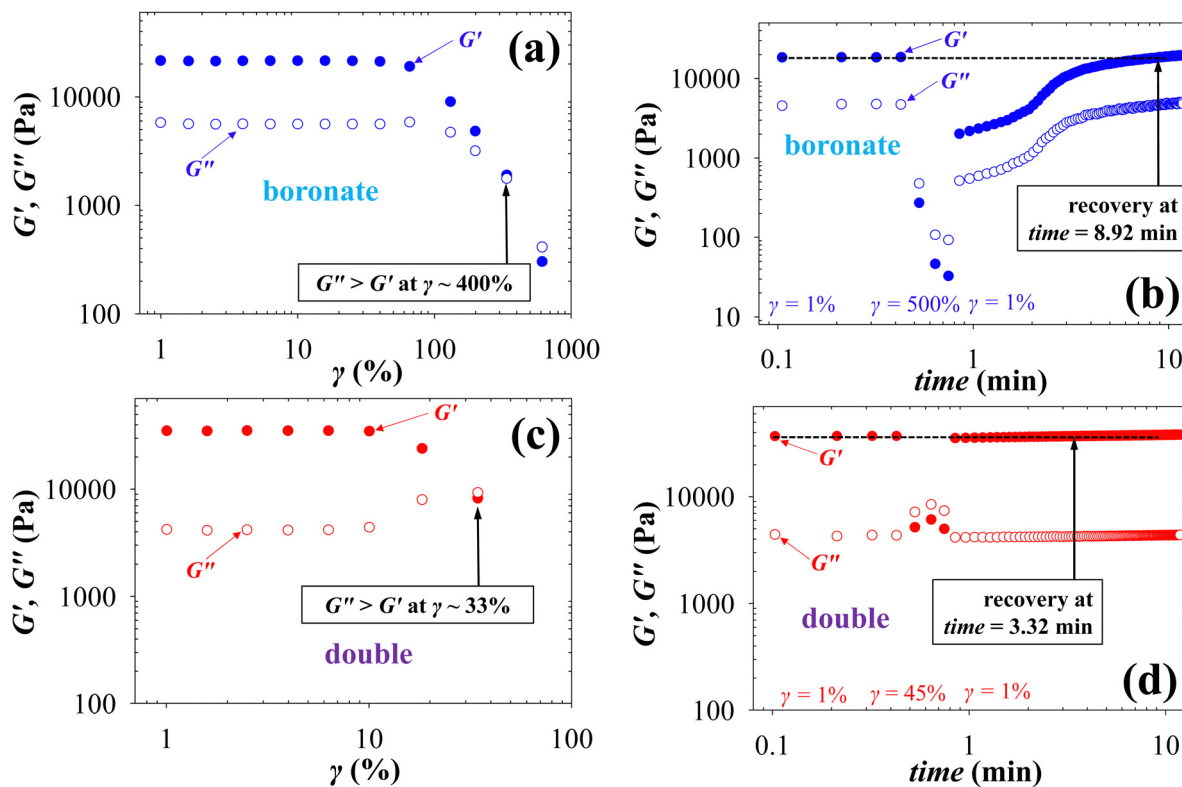


Fig. 8 Strain sweep (parts (a) and (c)) and destruction followed by self-repair (parts (b) and (d)) rheological experiments for the “boronate” (parts (a) and (b)) and “double” (parts (c) and (d)) hydrogels, both prepared at pH 8.5.

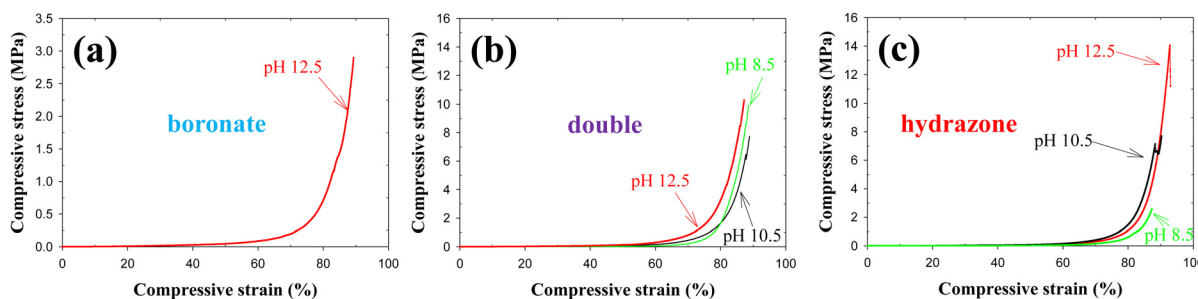


Fig. 9 Representative stress–strain curves in compression for the three hydrogel types, “boronate”, “double” and “hydrazone”, prepared at the three different pH values, 8.5, 10.5 and 12.5.

prepared at pHs 10.5 and 12.5, and satisfactory for the four other samples prepared at these two pHs, particularly if one takes into account the sizable error bars in Fig. 10(a).

Next, we discuss together the stress and strain at break of all samples. We must first point out that it was not possible to determine the values of the stress and strain at break for the “boronate”-cross-linked hydrogels at pH 8.5 and 10.5 because those samples did not fracture but rather deformed plastically. This was not the case for the “boronate”-cross-linked hydrogel at pH 12.5, which did present fracture. The former hydrogels do not fracture because of the high reversibility of the boronate bonds at pH 8.5 and 10.5, and in which the boronate cross-links exchange very fast, with exchange/relaxation times of 2.72 and

314 s, well within the timeframe of the compressive experiment, thereby immediately mending any incurring hydrogel damage in the course of the compression. The stress at break for the “boronate”-cross-linked hydrogel at pH 12.5 was lower than the corresponding values for the two other hydrogels at the same pH because of their decreased reversibility leading to hydrogels with more permanent cross-links. The stress at break values for the “double” hydrogel are pH-independent, within experimental error, and these are lower than those for the “hydrazone”-cross-linked hydrogels (at the two higher pHs), which can be attributed to the contribution of the boronate bonds to the dynamics of the double hydrogel. Nonetheless, all the hydrogels are very robust, most of them displaying stress at

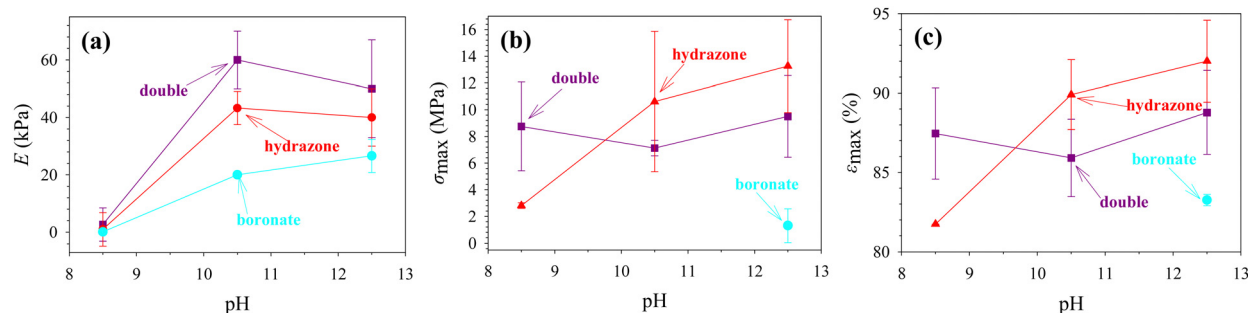


Fig. 10 Compressive mechanical properties, (a) elastic modulus, E , (b) stress at break, σ_{max} , and (c) strain at break, ϵ_{max} , of the three hydrogel types, boronate, hydrazone and double, each prepared at three different pH values, 8.5, 10.5 and 12.5.

break values within the range between 7 and 13 MPa. Finally, the trends in the strain at break behavior of the three hydrogel types *versus* pH are identical to those observed in the stress at break, and, therefore, similar explanations may apply.

It is to be noted that we unsuccessfully attempted several times to perform tension experiments on the hydrogels using the same mechanical testing instrument, which were always hampered by sample slippage from the grasps of the instrument. Thus, we tried to perform tension experiments in a “more qualitative” (“semi-quantitative”) way, in the sense that we held the samples using our fingers and pulled them to verify their extensibility and approximately determine their tensile strain at break, missing to determine their tensile stress at break and tensile elastic modulus. These “semi-quantitative” experiments were performed both on virgin and self-healed samples, focusing on the two self-healable samples prepared at pH = 10.5, choosing them over the ones at pH = 8.5 which are “too liquid”, and also preferring them to those at pH 12.5 where self-healing capability may be reduced. Our results are summarized in Table 3.

The results in the table above show that virgin samples from both network types displayed good extension at break, being able to be elongated about 4 (the “boronate”) and 3 (the “double”) times their initial length before failure. The less stiff “boronate” network expectedly presented the higher ϵ_{max} value. Furthermore, cut samples from both network types could self-heal upon pressing the two cut surfaces against each other for 24 h. The self-healed sample of the less stiff “boronate” gel again attained a higher strain at break (220%) in comparison with the stiffer “double” counterpart (120%), with the former network type also exhibiting a higher self-healing efficiency (55%) than the latter (40%). We did not attempt to determine

the self-healing efficiency *via* mechanical testing in compression, because we anticipated a reduced accuracy in compressive mode where the sample is self-supported, thereby making any defects or cracks more difficult to appear.

Conclusions

The “boronate”-linked hydrogels developed in this study represent the first example of well-defined dually-dynamic tetraPEG gels. These hydrogels exhibit moderate aqueous stability, disintegrating within 4–10 days, depending on the pH, display excellent self-healing capability due to their short bond lifetimes, and show mechanical properties which improve only at the highest investigated pH of 12.5. On the other hand, the “hydrazone”-linked hydrogels, also investigated herein, exhibit excellent aqueous stability, poor self-healing capability, and very good mechanical properties. The “double” hydrogels expectedly display properties intermediate between the two above-mentioned hydrogels, as they are their hybrids. In particular, these hybrid hydrogels present good aqueous stability (compared to the moderate stability of the “boronate”), good self-healing capability (compared to non-self-healable “hydrazone”), and good mechanical properties. This study opens the way for the macromolecular engineering of more hybrid materials, where the ratio of the “boronate” and “hydrazone” components is changed, so as to optimize the property of interest, whether aqueous stability, self-healing capability, and mechanical properties, while keeping the remaining properties at acceptable levels.

Conflicts of interest

There are no conflicts to declare.

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Table 3 Results from the semi-quantitative elongation experiments on original and self-healed “boronate” and “double” hydrogels prepared at pH = 10.5

Network type	Tensile strain at break, ϵ_{max} (%)		Self-healing efficiency based on tensile strain at break (%)
	Virgin	Healed	
“boronate”	390	220	55
“double”	300	120	40

the rheometer and the NMR spectrometer (500 MHz), respectively, at the University of Cyprus, used in this investigation. Finally, we thank our colleague Prof. E. Leontidis of the Chemistry Department at the University of Cyprus for kindly providing access to his rheometer used in our experiments.

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